Research Paper

Head to head evaluation of second generation ALK inhibitors brigatinib and alectinib as first-line treatment for ALK+ NSCLC using an *in silico* systems biology-based approach

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ABSTRACT

Around 3–7% of patients with non-small cell lung cancer (NSCLC), which represent 85% of diagnosed lung cancers, have a rearrangement in the ALK gene that produces an abnormal activity of the ALK protein cell signaling pathway. The developed ALK tyrosine kinase inhibitors (TKIs), such as crizotinib, ceritinib, alectinib, brigatinib and lorlatinb present good performance treating ALK+ NSCLC, although all patients invariably develop resistance due to ALK secondary mutations or bypass mechanisms. In the present study, we compare the potential differences between brigatinib and alectinib's mechanisms of action as first-line treatment for ALK+ NSCLC in a systems biology-based *in silico* setting.

Therapeutic performance mapping system (TPMS) technology was used to characterize the mechanisms of action of brigatinib and alectinib and the impact of potential resistances and drug interferences with concomitant treatments.

The analyses indicate that brigatinib and alectinib affect cell growth, apoptosis and immune evasion through ALK inhibition. However, brigatinib seems to achieve a more diverse downstream effect due to a broader cancer-related kinase target spectrum. Brigatinib also shows a robust effect over invasiveness and central nervous system metastasis-related mechanisms, whereas alectinib seems to have a greater impact on the immune evasion mechanism.

Based on this *in silico* head to head study, we conclude that brigatinib shows a predicted efficacy similar to alectinib and could be a good candidate in a first-line setting against ALK+ NSCLC. Future investigation involving clinical studies will be needed to confirm these findings. These *in silico* systems biology-based models could be applied for exploring other unanswered questions.

INTRODUCTION

Lung cancer (LC) remains the leading cause of death worldwide, with an estimated 1.6 million deaths each year [1, 2]. Despite significant therapeutic advances over the last decade, over half of patients diagnosed

with LC die within one year of diagnosis and the fiveyear survival is around 18% [3]. About 85% of LCs are diagnosed as the subtype non-small cell lung cancer (NSCLC), adenocarcinoma being one of the most common histological subtypes. In adenocarcinoma, several driver mutations have been identified, including mutations/alterations of the epidermal growth factor receptor (EGFR), anaplastic lymphoma kinase (ALK), and ROS1, among others; most of them are therapeutically targetable [4]. Around 3–7% of NSCLC cases present active ALK rearrangement (ALK+ NSCLC) that produces an abnormal activity of the ALK protein cell signaling pathway and causes the cancer cells to grow and metastasize [5, 6]. Central nervous system (CNS) metastasis is a common finding in NSCLCs, occurring in 10% of patients, and even more frequent in ALK+ NSCLCs, were the frequency of CNS metastasis is around 20–30% at the time of diagnosis [7, 8]. CNS is also the most common site of relapse [9]. Thus far, three generations of ALK tyrosine kinase

Thus far, three generations of ALK tyrosine kinase inhibitors (TKIs) have been developed. Some of the drugs that target the abnormal ALK protein are crizotinib (first generation), ceritinib, alectinib, brigatinib, ensartinib (second generation) and lorlatinib (third generation) [10]. However, despite their effectiveness in ALK+ NSCLC cases, all patients invariably develop treatment resistance at some point. Consequently, it is of the upmost importance to adequately use the currently available treatments in the correct order to maximise the life span of NSCLC patients. The most common progression mechanisms for all ALKi are: 1) ALK secondary mutations, which affect not only crizotinib-treated patients (around 20-30% of patients [11]), but also second and third generation ALKi [12, 13]; and 2) bypass mechanisms (i.e., activation of other parallel pro-proliferative signaling pathways [14]). Crizotinib has been used as first-line since its approval in 2011 in the United States [15] and in 2015 in Europe [11]. The results of its phase III trial (PROFILE 1014) demonstrated that crizotinib was superior to standard chemotherapy [16]. Second and third generation ALK TKIs are effective in treating numerous crizotinib-resistant ALK mutations and are used after crizotinib, and some of them have even replaced crizotinib as first-line option among patients with ALK-rearranged NSCLC [17-19]. Second generation ALKi ceritinib, alectinib and brigatinib have been approved for the treatment of ALK+ NSCLC patients after treatment with crizotinib (ceritinib in 2014 [20] and in 2015 [21]; alectinib in 2015 [22] and in 2017 [23], while brigatinib in 2017 [24] and 2018 [25] for the United States and Europe, respectively) and as first-line TKI treatments (in 2017, ceritinib [26, 27] and alectinib [23, 28] and, in 2020, brigatinib [25, 29], for the United States and Europe, respectively).

At the time of the design and performance of the study, clinical trials had provided promising results for both brigatinib and alectinib as first-line TKIs in TKI-naïve ALK+ NSCLC patients, compared to crizotinib. The ALEX phase III study (<u>https://clinicaltrials.gov/</u>number, NCT02075840), showed that alectinib had a superior investigator-assessed PFS versus crizotinib (HR, 0.47; P < 0.001 [30]). At the second interim analysis of the ALTA-1L phase III trial (<u>https://clinicaltrials.gov/</u>

number, NCT02737501) the blinded independent review committee (BIRC)-assessed HR of PFS was 0.49 (log rank P < 0.0001 [31]. Moreover, both drugs present relevant intracranial efficacy: alectinib demonstrated superior efficacy versus crizotinib regardless of baseline CNS metastases [32] and brigatinib significantly delayed both CNS progression (without prior systemic progression) and systemic progression (without prior intracranial progression) compared with crizotinib [33]. Regarding ceritinib, direct comparison in first line has only been performed with chemotherapy [34, 35], although indirect comparison showed better results for ceritinib than crizotinib [36]. At the time of the beginning of the current study, two other ALK inhibitors were being tested in first line in comparison to crizotinib, although no results were available (ensartinib in eXalt3, NCT02767804, or lorlatinib in CROWN, NCT03052608).

Although no direct comparison between alectinib, brigatinib and ceritinib has been performed in a first-line setting, there are indirect comparisons in second line from which hypotheses can be drawn. Ceritinib, alectinib and brigatinib are effective in crizotinib-refractory ALK+ NSCLC patients [37-39], but no direct comparison between these drugs after crizotinib is available. An ongoing trial, ALTA-3 (https://clinicaltrials.gov/ number, NCT03596866), compares the efficacy of alectinib versus brigatinib in ALK+ NSCLC patients who had progressed on crizotinib; besides, and according to the current lack of direct comparisons, indirect analyses using available data have been performed to compare them. In fact, a matching-adjusted indirect comparison (MAIC) [40] between these drugs in crizotinib-refractory ALK+ NSCLC patients (using clinical data from the ALTA trial - date February 21, 2017 -, ASCEND-1 [41], ASCEND-2 [42], NP28761 [43] and NP28673 [44]) suggested that brigatinib may have prolonged PFS and OS versus ceritinib and prolonged PFS versus alectinib in patients after progression with crizotinib.

From a safety perspective, all ALKi are considered to be safe and tolerable in a similar fashion [45], although they show adverse events (AE), some of them common and others drug-specific. A systematic review [46] concluded that crizotinib was associated with more gastrointestinal and visual events, alectinib tended to have more hepatic and musculoskeletal AEs, ceritinib presented the highest incidence of clinically significant gastrointestinal AEs and laboratory abnormalities and brigatinib had a unique profile of increased early onset pulmonary AEs and hypertension associated with the 180 mg dose; these pulmonary AEs were found to be reduced when using the recommended initial dose of 90 mg [47]. This systematic review also suggested ceritinib to be less preferred by clinicians due to its safety profile. Regardless of their differences, most of the safety concerns associated with the mentioned ALKi can be minimized reducing administration dose [46].

According to first-line results with brigatinib and alectinib and indirect results of ceritinib, this last drug seems to have a lower efficacy both at systemic and cerebral levels when compared to brigatinib and alectinib [40]. As tolerability of all these ALKi is similar and alectinib has become the standard of care, a head to head clinical trial comparing brigatinib and alectinib as firstline therapy would be very interesting. However, since this head to head is not planned, results obtained with these second generation ALKi in the ALTA-2 study in second line, as well as in the MAIC analysis, will help to elucidate and refine the first-line therapy outline. Besides, *in silico* investigational approaches may be an alternative to compare the potential benefits of both drugs.

Concerning the mechanism of action of alectinib and brigatinib, both share ALK as a protein target, but they display completely different target profiles that could be determinant to define each drug mechanism. Beside ALK, brigatinib targets other tyrosine-protein kinases receptors such as EGFR [48–50], receptor-type tyrosine-protein kinase FLT3 [48, 51], tyrosine-protein kinase FER [52], ROS1 [51, 53], and insulin-like growth factor 1 receptor (IGF1R) [48, 49, 51, 54]. On the other hand, alectinib inhibits RET with comparable potency to ALK [55].

In silico tools are useful resources for predicting several (bio)chemical and (patho)physiological characteristics of likewise potential drugs [56]. These methods are used to improve *in vivo* and *in vitro* models and refine experimental programs of clinical and general biomedical studies involving lab work [57], and, in the long run, can reduce lab work and effectively succeed in 3R (reduce, reuse, recycle) [58]. Overall, these systems can be employed for the exploration of anticancer drug mechanisms of action and their efficacy in specific patient profiles.

In the present study, we created *in silico* systems biology-based mechanistic models of two first-line approved second generation ALKi, brigatinib and alectinib, in order to explore the potential differences between them with the aim of providing information or raising hypotheses towards the identification of strengths and weaknesses of the mechanisms of action of both drugs as first-line treatment for ALK+ NSCLC patients.

RESULTS

The main pathophysiological processes (namely "motives") described to be involved in ALK+ NSCLC were: (1) Cell growth and proliferation, (2) Sustained angiogenesis, (3) Evading apoptosis, (4) Tissue invasion and metastasis, (5) Immune evasion (Supplementary Table 1). Subsequently, each pathophysiological process was functionally characterized at protein level to determine its molecular effectors and used for focusing the analysis towards ALK+ NSCLC in a human biological network context (Figure 1 and Supplementary Table 2).

Brigatinib and alectinib protein target profiles were also carefully characterized and used in the posterior analyses (Figure 1 and Supplementary Table 3). Mechanistic systems biology models of brigatinib and alectinib obtained with TPMS technology were constructed with accuracy values of 94% to evaluate their mechanism of action and potential treatment efficacy in ALK+ NSCLC. Two distinct modelling approaches were used for that purpose: Artificial neural networks (ANN) [59], with the aim of detecting biological relationships; and samplingbased methods [60], in order to explain those relationships. A Sobol sensibility analysis was applied to brigatinib and alectinib mechanistic models in order to evaluate their robustness. The results of this analysis are available in the Supplementary Methods.

Effect of brigatinib and alectinib on cell growth, apoptosis and immune evasion through ALK and non-ALK inhibition

The relationships of each drug target with ALK+ NSCLC main pathophysiological motives were evaluated by the ANN and the results are shown in Table 1. The ANN analysis showed that, in general, alectinib presented a slightly lower correlation with ALK+ NSCLC pathophysiology than brigatinib (around 80% of the score obtained by brigatinib).

Evaluation of the relations between individual pathophysiological motives and drug targets suggested that both drugs affect cell growth and proliferation, apoptosis evasion and immune evasion through ALK inhibition. Regarding alectinib, its inhibition of RET might occur through modulation of the tumour immune response. On the other side, brigatinib non-ALK targets might affect the pathophysiological motives already affected by ALK inhibition (FLT3, IGF1R, and especially EGFR), as well as angiogenesis and invasiveness through FER and IGF1R inhibition.

Brigatinib and alectinib non-ALK targets affect differently cancer-related processes, including proliferation, apoptosis evasion, invasiveness and immune evasion

The comparison of the predicted mechanisms of action obtained by the mechanistic systems biology modeling using TPMS technology (Figure 2 and Supplementary Table 4) shows that both drugs act through ALK and some overlapping intracellular mechanisms (involving SHC1, GRB2, RASK). However, brigatinib seems to achieve a more diverse downstream effect, through PI3K, ERK and JAK/STAT.

A further evaluation of the impact of each drug on the activity of each protein present in the mechanisms of action, and on the pathophysiological motives previously defined was carried out. This analysis showed that brigatinib, compared to alectinib, has a stronger effect (TSignal) on most of the proteins and all the motives defining ALK+ NSCLC (Figure 3) except in immune evasion (Table 2), for which alectinib presents a greater effect.

Effect of brigatinib and alectinib on invasiveness and central nervous system metastasis

As shown in Table 2, brigatinib was predicted to have a potential stronger effect on metastasis effectors, which are related to invasiveness promotion and metastasis-site characteristics. In order to assess the possible role of each drug on brain metastasis, eight protein/gene effectors known to have a more important role in brain metastasis than in primary tumours were considered: FGFR1 [61–63]; Ki-67 [64, 65], ROBO1 [66]; S100A7 [67, 68]; S100B [69, 70]; SIRT1 [71]; SLIT2 [72]; and VEGFA [61, 67]. Out of these, six (Ki-67, ROBO1, S100A7, S100B, SLIT2, VEGFA) were found to be significantly more inhibited by brigatinib than alectinib (FDR < 0.05 and a change in TSignal > 20%). The current analysis also showed an association between brigatinib and prevention of brain metastasis, mainly through EGFR and IGF1R.

Susceptibility of brigatinib and alectinib to bypass resistance mechanisms

The impact of resistance mechanisms, via protein mutations, on brigatinib's and alectinib's mechanisms of action TPMS models was evaluated over a total of 935 proteins and 2805 modifications (activation, inhibition, deletion) (Supplementary Table 5). Of those, 55 different modifications were identified as potential treatment resistances for brigatinib and 93 for alectinib. Among them, 37 were shared between the two drugs. The potential resistance mechanisms that affected alectinib to a greater extent than brigatinib were, mostly, related to the alternative pro-proliferative signaling mechanisms by which NSCLC cells could continue to proliferate. These mechanisms included proteins like MET, ERBB, FGFR, NTRK1 or PDGFR, among others. On the other hand, the potential resistance mechanisms that affected either both

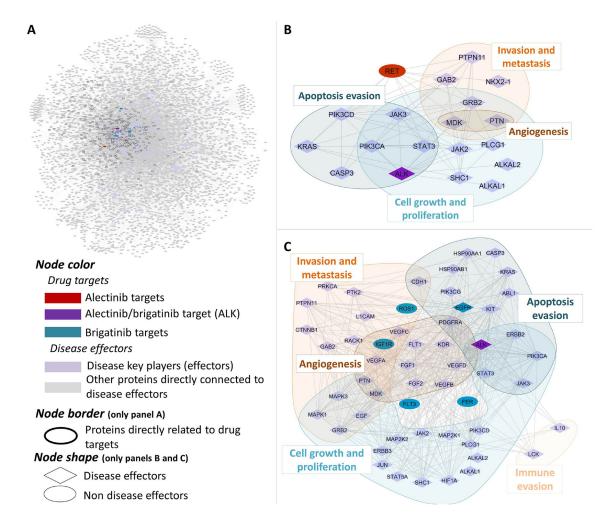


Figure 1: Human protein networks around ALK+ NSCLC molecular pathophysiology. General overview (A) and centered on main disease players (indicating their pathophysiological motives) and their relationship to alectinib (B) and brigatinib (C) drug targets.

drugs or brigatinib to a greater extent than alectinib were mainly potential downstream mediators such as SHC1, KRAS, PI3K or ERK.

Effect of concomitant treatments on the mechanism of action of brigatinib and alectinib

The potential interference with brigatinib or alectinib mechanisms of action was evaluated using a total of 654 drugs (Supplementary Table 6). The drugs that may impact the brigatinib mechanism may include angiotensinreceptor blockers, barbiturates and bisphosphonates. This can be mitigated by adjusting the brigatinib dose. For alectinib, drugs such as non-peptide inhibitors of the antidiuretic hormone can also interfere with it. However, concomitant use of brigatinib with strong/moderate CYP3A inhibitors/inducers can be managed [72].

DISCUSSION

The second generation ALKi brigatinib and alectinib have demonstrated efficacy in second line treatment in crizotinib-refractory ALK+ NSCLC patients, and in the first-line setting in ALEX (alectinib) and ALTA-1L (brigatinib) clinical trials. In the absence of a head to head trial between brigatinib and alectinib in the first-line setting, and beside the efficacy data and toxicity profile information obtained in independent trials, information from clinical trials in a second-line setting and indirect approaches may help to elucidate the best therapy against ALK+ NSCLC. In the present study, we applied *in silico* systems biology approaches to compare brigatinib and alectinib as first-line treatment for ALK+ NSCLC at a mechanistic level and thus highlighting the strengths and weaknesses of each ALK inhibitor. The present

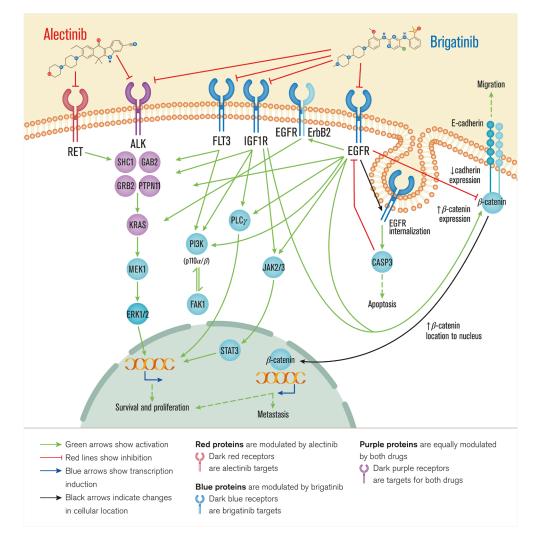


Figure 2: Overview of brigatinib's and alectinib's mechanisms of action. Receptor targets of each drug are depicted through the cell membrane and the following pathways and pathophysiological motives affected are depicted from the cell surface to the nucleus. Alectinib acts through ALK and RET, involved mainly in survival and proliferation, while brigatinib acts also through ALK and FLT3, IGF1R, and EGFR, signaling through overlapping intracellular mechanisms affecting cell survival and proliferation, metastasis, apoptosis and migration. Bibliographical validation information of interactions on the predicted mechanisms of action are shown in Supplementary data (Supplementary Table 1).

study indicates that both brigatinib and alectinib could be reasonable choices for first-line treatment, as also previously suggested by other authors [47]. The results obtained by our *in silico* model allow differentiating between the mechanisms of action of each drug, suggesting that both drugs may have similar efficacy as first-line treatment, and brigatinib may have higher impact in most studied pathways than alectinib. Other specific characteristics were highlighted for each drug.

According to previous publications [17, 47], brigatinib acts as a multi-kinase inhibitor with a broad-spectrum activity against ALK, FLT3, FER, ROS1, IGF1R, and EGFR targets, while alectinib acts on ALK and the proto-oncogene RET [55]. The analyses performed in this study to further determine the differences between brigatinib and alectinib's mechanisms of action point towards a potentially relevant role of RET, EFGR, IGF1R and FLT3 (besides ALK) in treating NSCLC. All these targets had been previously related to a greater or lesser extent to NSCLC development [73–77].

As predicted by TPMS analyses (Figure 2), brigatinib targets appear to show a more diverse range of effects compared to alectinib, mediated by ALK inhibition on NSCLC, such as: cell growth and proliferation (as for example STAT3 [78], PI3K [79], K-Ras [78, 80] or erbB2 [81] signaling); evading apoptosis (through EGFR-CASP3 interplay [82]); acting over sustained angiogenesis (IGF1R signaling [83, 84]); and tissue invasion and metastasis processes (modulating the E-cadherin- β -catenin axis [85–88]). These predicted results and the observed broad range of different effects of brigatinib could be explained by a wider cancer-related target profile of brigatinib. Moreover, it could also be associated with the relatively longer PFS observed with brigatinib in the crizotinib-refractory setting as compared to alectinib [89]. On the other hand, the analyses also suggest that alectinib might have a greater effect on immune evasion regulation through RET inhibition.

Central nervous system (CNS) is one of the most common sites of first progression in ALK+ NSCLC [89]. Even while receiving crizotinib (in around 25–50% of cases), efficacy end points are lower in relation to the CNS than overall [90–92]. In our study, brigatinib was predicted to have a potentially more robust impact on brain metastasis effectors than alectinib. Inhibition of EGFR might prevent CADH1 reduction mediated by PI3K/FAK1 and thus inhibit tissue invasion. Blocking EGFR and IGF1R pathways might also prevent β -catenin (CTNB1) upregulation, accumulation in the nucleus and transcription factor function. Intracranial responses to TKIs have also

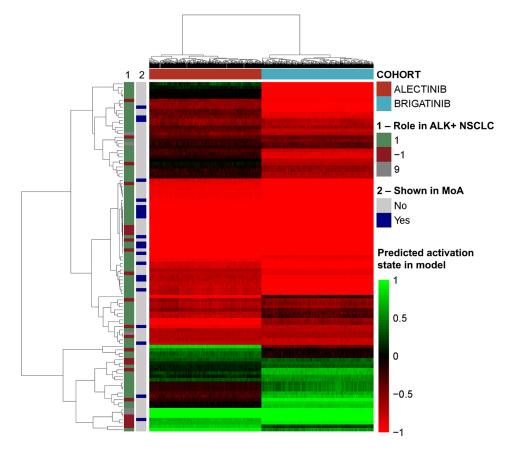


Figure 3: Heatmap of the effect induced by brigatinib and alectinib in each model solution over the effectors of the pathology. The vertical bars indicate the pathological effect of the effectors (1 if activated in the pathology, -1 if inhibited in the pathology, 9 if complex role) and whether the proteins appear in Figure 2 ("MoA").

been observed in previous studies. In the phase III trial ALTA-1L (https://clinicaltrials.gov/: NCT02737501) brigatinib was associated with a higher intracranial objective response rate (iORR) (78%) in individuals with ALK TKI-naive ALK+ NSCLC with baseline brain metastases compared to crizotinib (26%) [47]. Alectinib also showed superior intracranial activity versus crizotinib (81% and 50%, respectively) in the ALEX clinical trial, although progression in the brain with both agents has also been observed [93]. This intracranial efficacy is clearly explained by brain bioavailability in the case of alectinib, which shows a very good blood-brain barrier penetration [93], without being affected by MDR1/p-gp modulation [23, 94]. Brigatinib might be susceptible to MDR1/pgp modulation [95], although no major concerns were raised by the regulatory bodies [25]. Thus, some other factors might explain brigatinib's activity in the brain. The mechanistic study of the current analysis suggests that brigatinib might be able to reverse the activation of a greater percentage of metastasis effectors and, specifically, brain-related metastasis effectors, compared to alectinib. These effectors include the well-known proliferation marker Ki-67 [96] overexpressed in brain metastasis when compared to primary tumours [64, 65]; the ROBO1/SLIT2 axis, increased in brain metastasis [66] and involved in cell migration [97]; the pro-angiogenic VEGFA, related to increased brain metastatic potential [75, 98, 99]; and damage signal proteins (DAMPS) S100 proteins, involved in increased proliferation, anti-apoptotic, and migration capabilities [100, 101], which are increased in serum of brain metastatic patients and brain metastasis models [69, 70, 75, 100, 101]. Enhanced mechanistic impact over these - and other non brain-specific - metastasis effectors by brigatinib might explain its activity in the brain despite its lower blood-brain barrier penetration. The ongoing ALTA-3 trial will provide valuable information including intracranial progression after brigatinib versus alectinib in crizotinib-refractory patients that might help better understand their anti-metastatic mechanisms.

The acquisition of resistance to TKI therapy still seems inevitable. However, next generation TKIs are able to more strongly inhibit ALK – both in its wild type form and presenting secondary mutations - suggesting a better control over these progression mechanisms. In fact, brigatinib presents a high selectivity for ALK and low propensity for pharmacological failure [17, 19], showing higher potency than alectinib towards ALK-rearrangement fusions [14, 102, 103]. Brigatinib selectivity over ALK has been also proven in patients with ALK fusion proteins with and without secondary mutations [104, 105]. Little is known about the capacity of ALK TKIs to prevent bypass resistance mechanisms. The evaluation of the impact of developing non-ALK-related resistances on the efficacy of the drugs performed in the current study suggests that alectinib might be more susceptible to bypass resistance mechanisms. The results of our in silico analysis also suggest that brigatinib might block or prevent the development of upstream bypass resistance mechanisms more effectively than alectinib, which could translate into resistance-free treatment for a longer period of time. This would probably occur due to a mechanism of action that reaches a larger number of intracellular effectors involved in ALK-independent resistance mechanisms, including JAK/STAT, MEK/ERK, PI3K or PLCy [105, 106]. According to our in silico results, brigatinib is predicted to modulate these pathways that are involved in different NSCLC-related pathophysiological processes, more strongly than alectinib. Thus, given the broader impact of brigatinib on ALK secondary mutations compared to other ALK TKIs [17, 19] and the results of the current analysis regarding bypass mechanisms, it could be hypothesized that brigatinib would prevent the generation of a wider spectrum of resistance mechanisms compared to alectinib. This low resistance predisposition of brigatinib could be related to the efficacy results in terms of PFS observed in the indirect comparison (MAIC) between brigatinib and alectinib/ceritinib by Reckamp [40]. However, further pre-clinical and clinical studies are needed to validate these hypotheses, and ALTA-3, comparing brigatinib to alectinib in ALK+ NSCLC patients who had progressed on crizotinib, might provide interesting conclusions in this regard.

There are two dimensions in which drugs can affect each other: through metabolic and mechanistic interactions. According to the recommendations of the technical specifications [23, 25], whereas both drugs interact with CYP3A – among other enzymes and transporters–, only brigatinib has strict interactions with the usage of inductors, inhibitors and substrates of CYP3A family cytochromes [25, 96]. The current study evaluated the mechanistic interaction between drugs commonly used in cancer patients, regarding the interference of the signal induced by the targets of co-treatments.

According to the current knowledge and the data herein presented, brigatinib might be more prone to present relevant metabolic and mechanistic interactions with other drugs than alectinib, which might be a safer option in poly-treated patients. Use of more than one drug (e.g., to treat cancer or treatment-derived complications, or pre-existing conditions) is common in cancer patients, and polypharmacy (5 or more concomitant drugs) has been shown to occur at a higher frequency in cancer survivors than in non-cancer age- and sex-matched controls [107]. Polypharmacy is especially common among the elderly or in end-of-life settings [108]. Thus, drug interactions must be carefully taken into account when considering different treatment options. However, as NSCLC adenocarcinoma patients tend to be younger and tend to be non-smokers compared to other cancer patients [102, 109], potential drug interference due to polypharmacy might not represent a determinant factor for treatment selection in clinical practice.

8 8								
Pathophysiological processes (motives)	Brigatinib's targets							Alectinib's targets
	ALK	FLT3	FER	ROS1	IGF1R	EGFR	ALK	RET
NSCLC ALK ⁺	++	+	+	+	+	+++	++	+
Cell growth and proliferation	+++	++	++	+	+++	+++	+++	+
Evading apoptosis	+++	++	+	+	++	++++	+++	+
Sustained angiogenesis	+	+	++	+	+++	+	+	+
Tissue invasion and metastasis	+	+	++	+	++	++	+	+
Immune evasion	+++	+++	+	+	+	++	+++	++++

Table 1: Effect of brigatinib's and alectinib's drug targets in ALK+ NSCLC

Relationship of brigatinib's and alectinib's targets and pathophysiological motives calculated by artificial neural networks (ANN). High (++++) corresponds to *p*-value < 0.05, Medium-high (+++) corresponds to *p*-values < 0.15 and Medium (++) corresponds to *p*-value < 0.25, while Low (+) correspond to *p*-value > 0.25.

As previously stated, ALKi activity is affected by several factors, including tumour intrinsic characteristics (e.g., ALK fusion gene variants or presence of other primary gene co-mutations) and extrinsic factors (e.g., impact of prior treatments such as presence of ALK secondary mutations, or development of by-pass resistances), and also drug-dependent characteristics (e.g., blood-brain barrier crossing).

The current study aimed to explore mechanistic differences between brigatinib and alectinib that could affect efficacy of both drugs in an *in silico* approach. However, beside efficacy data, drug toxicity profile is an important determinant of treatment selection. According to previous publications, we considered that although all ALKi present common and specific adverse events, alectinib and brigatinib are similarly well tolerated and can be managed by reducing dose or interrupting treatment [46].

In order to better contextualize the hypotheses raised from the mechanistic analyses, other parameters need to be considered and have been herein discussed (ALK secondary mutations, safety concerns), and must be taken into account in the clinical practice. Besides, in silico modelling approaches can be used as predictive tools and hypothesis generators, limited by the information about diseases and drugs. For example, unknown targets or not yet described pathophysiological processes might have a role in the mechanisms of action of the evaluated drugs. Nevertheless, the models were built by considering the whole human protein network and a wide range of drug-pathology relationships (Supplementary Table 7) [60], not only limited to NSCLC or oncologic indications, and they present cross-validation accuracies above 80% in the case of ANN models and above 90% in sampling methods-based models. Thus, even if modelling approaches based on systems biology are limited by the amount of available information and some assumptions have to be made, in silico techniques are helpful for understanding fundamental processes in cancer [110, 111]. These approaches allow us to explore investigational or marketed drugs with reduced experimental cost and in different settings. This proves to be especially important if clinical investigations are not going to be done soon or are complex to be conducted, as in the case of the brigatinib versus alectinib head to head study in a firstline setting. Similarly, a comparison to other second and third generation ALKi that have recently shown benefit with respect to crizotinib in the first line setting (ensartinib in eXalt3, NCT02767804, or lorlatinib in CROWN, NCT03052608) could provide further insights into the mechanisms behind ALK+NSCLC treatment. Thus, systems biology and artificial intelligence approaches can contribute to exploring unanswered questions and this may guide the development of ALK TKIs and the identification of the optimal treatment sequence in ALK+ NSCLC patients. Further in silico studies with the aim of identifying the best treatment sequence after brigatinib are ongoing.

MATERIALS AND METHODS

Molecular characterization of ALK+ NSCLC pathophysiology and drugs

To carefully characterize the pathophysiology of ALK+ NSCLC, we conducted an extensive and detailed full-length review of relevant review articles over the last 5 years in the PubMed database (from December 3rd 2013 to December 3rd 2018) using the following search string: (("ALK+-positive" [TITLE] or "ALK+" [TITLE]) and ("Non-Small Cell Lung Cancer" [TITLE] or "NSCLC" [TITLE]) AND ("MOLECULAR" [TITLE/ABSTRACT] "PATHOGENESIS" [TITLE/ABSTRACT] or or "PATHOPHYSIOLOGY" [TITLE/ABSTRACT]) and Review[ptyp]) and (("Non-Small Cell Lung Cancer" [TITLE] or "NSCLC" [TITLE]) AND ("MOLECULAR" [TITLE/ABSTRACT] or "PATHOGENESIS" [TITLE/ ABSTRACT] or "PATHOPHYSIOLOGY" [TITLE/ ABSTRACT]) and Review[ptyp]). The search was also expanded using article reference lists. The main

Motive	% Effectors m	ore reversed	_	
	Brigatinib MoA	Alectinib MoA	Drug with highest Tsignal	FDR T-Test
Cell growth and proliferation	87%	10%	Brigatinib	< 0.05
Evading apoptosis	69%	27%	Brigatinib	< 0.05
Sustained angiogenesis	65%	29%	Brigatinib	< 0.05
Tissue invasion and metastasis	63%	37%	Brigatinib	< 0.05
Immune evasion	25%	75%	Alectinib	< 0.05

Percentage of effectors reversed indicate the proportion of proteins in each motive with a significant difference (FDR < 0.05) and stronger modulation considering the total number of effectors affected in the mechanism of action (MoA) for each drug.

pathophysiological processes (motives) described to be involved in ALK+ NSCLC were identified (Supplementary Table 1). Subsequently, each motive was further functionally characterized at protein level to determine its molecular effectors. A total of 174 proteins were identified (Supplementary Table 2).

For drug protein target profile definition (brigatinib and alectinib), a dedicated review of databases (DrugBank [112], STITCH [113], SuperTarget [114]) and of scientific literature was performed (Supplementary Table 3).

TPMS technology: systems biology-based model creation

Therapeutic Performance Mapping System (TPMS) (Anaxomics Biotech, Barcelona, Spain) is a top-down systems biology approach based on artificial intelligence and pattern recognition models. This methodology integrates available biological, pharmacological and medical information to generate mathematical models that simulate the mechanisms of action of drugs in a pathophysiological human context (Figure 4). TPMS models are trained using a compendium of biological and clinical data characteristics of the human physiology (Supplementary Table 7).

Mechanism of action models

In order to obtain the mechanism of action (MoA) of brigatinib and alectinib, drug-ALK+ NSCLC mathematical models were generated following the same methodology as described in Jorba [60] and applied in previous studies [63, 115, 116]. As input, TPMS takes the activation (+1) and inactivation (-1) of the drug target

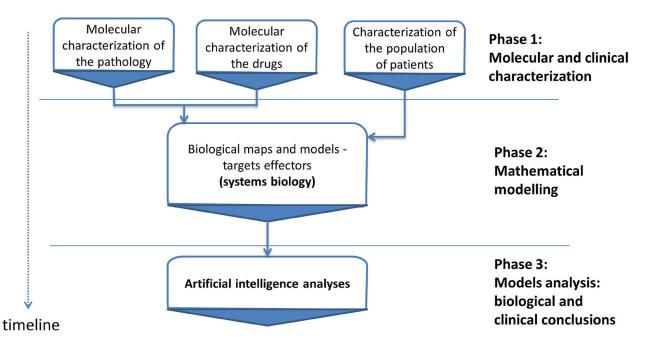


Figure 4: Study workflow. Overview of the *in silico* study approach showing the main phases employed to simulate the mechanisms of action (MoA) of brigatinib and alectinib with respect to ALK+NSCLC molecular characterization. TPMS is a validated top-down systems biology approach that integrates all available biological, pharmacological and medical knowledge (protein network, truth table and specific data compilation) by means of pattern recognition models and artificial intelligence to create mathematical models that simulate *in silico* the behavior of human physiology.

proteins (Supplementary Table 3), and as output the protein states of the pathology of interest (Supplementary Table 2). It then optimizes the paths between both protein sets and computes the activation and inactivation values of the full human interactome. The resulting subnetwork of proteins with non-null outputs and their values will define the MoA of the drug. The impact of each drug over the activity of the proteins effectors of the pathophysiological disease was quantified using the Tsignal (i.e., the average signal values of the protein effectors), as described in Jorba [60]. More detailed information on the modelling methodology can be found in Supplementary Methods.

Sobol sensibility analysis

In order to analyze the impact of the noise in the final MoAs affecting the biological conclusions reached, a Sobol sensibility analysis was performed over the constructed TPMS mathematical models [117]. Detailed of the analysis implementation can be found in Supplementary Methods.

Drug-(patho)physiology motive relation finding

Artificial neural networks (ANN) were used to identify relations between proteins (e.g., drug targets) and clinical elements of the network [118], which is an approach previously used and validated by several publications [53, 119–121]. This strategy was used to perform an efficacy evaluation of brigatinib and alectinib from each of their targets towards ALK+ NSCLC pathophysiological motives and its corresponding proteins. Detailed information on the modelling methodology can be found in Supplementary Methods.

Evaluation of the impact of potential resistances over the mechanisms of action

In order to identify possible cancer resistances, the TPMS models were evaluated for possible mutations to identify the key nodes or proteins with higher impact on the effector proteins TSignal. Because both brigatinib and alectinib mechanisms of action had a vast amount of proteins or nodes (> 5000), the universe of possible key nodes was reduced to the list of proteins around ALK+ NSCLC effectors and around the drugs' target proteins. To evaluate the addition of a mutation in the system, the impact of a protein activation, inhibition and deletion over the mechanisms of action of brigatinib and alectinib was tested. To do so, the resulting TSignal of the altered models was computed and compared to the original one. Finally, the *p*-value of the difference between the Tsignals over ALK+ NSCLC with and without the mutation was calculated, and the ones with *p*-values ≤ 0.022 were selected (Supplementary Table 5).

Evaluation of drug interferences over the mechanisms of action

To identify possible co-treatment interferences, a list of pharmacological treatments potentially co-administered with brigatinib or alectinib was created and evaluated. To do so, we generated a list of all treatments for common conditions (either in the general population and in the ALK+ NSCLC population) and treatments for brigatinib/ alectinib-associated adverse drug reactions, according to DrugBank database [112]. After that, the mechanisms of action of brigatinib and alectinib was perturbed by activating the co-treatment protein targets, each drug one by one, and the TSignal was computed. Finally, the differences in the TSignal between the original and the perturbed system and the corresponding *p*-values were calculated, and the ones with *p*-values < 0.1 and 0.05 were selected (Supplementary Table 6).

CONCLUSIONS

An in silico head to head based on the mechanism of action evaluation between brigatinib and alectinib has been performed highlighting the advantages of using one before the other from an efficacy point of view. Brigatinib appears to have a wider mechanism of action, presenting targets that potentially act more strongly in most of the ALK+ NSCLC pathophysiological pathways, including invasiveness to the CNS. On the other side, alectinibinduced RET inhibition might contribute to reducing the tumour immune evasion mechanisms. In general, both drugs are known to be well-tolerated and, although shown and predicted to have a similar efficacy for the treatment of ALK+ NSCLC in a first-line setting, the differences in their target profiles might allow for identification, in subsequent studies, of different patient profiles that might benefit from either of them, beside considering potential safety concerns in specific patient subpopulations. Future clinical studies will be needed to confirm these findings. The used approach can be applied for the evaluation of other next-generation ALKi, even if not vet approved, or exploring other questions, such as optimal treatment sequence.

Author contributions

AFN, AL and EC contributed to setting up fundamental questions regarding ALK+ NSCLC treatment. EC, CSV and MC contributed to the study design. AFN, AL, CSV, CM and AN contributed to data acquisition. CSV, MC, GJ, JMM and BO contributed to data analysis. EC contributed to clinical interpretation of data. AFN, AL, CM, AN and CSV contributed to writing the manuscript. EC and MC contributed to the critical revision of the manuscript.

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CONFLICTS OF INTEREST

EC: Speaker or advisory board: Takeda, Astra Zeneca, Roche, MSD, Novartis, Boheringer Ingelheim, Pfizer, BMS; Travel expenses: Roche, Pfizer, Takeda, BMS. AL, CM, AN and AFN are full time employees of Takeda Farmacéutica España S.A. CSV, GJ, MC and JMM are full time employees of Anaxomics Biotech. BO declares no competing interests.

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REFERENCES

- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin. 2015; 65:87–108. <u>https://doi.org/10.3322/caac.21262</u>. [PubMed]
- Wong MC, Lao XQ, Ho KF, Goggins WB, Tse SL. Incidence and mortality of lung cancer: global trends and association with socioeconomic status. Sci Rep. 2017; 7:14300. <u>https://doi.org/10.1038/s41598-017-14513-7</u>. [PubMed]
- SEER. National Cancer Institute. Surveillance, Epidemiology, and End Results (SEER) Program. Cancer Statistics Review, 1975–2016. <u>https://seer.cancer.gov/</u> csr/1975_2016/.
- Griffin R, Ramirez RA. Molecular targets in non-small cell lung cancer. Ochsner J. 2017; 17:388–92. <u>https://doi. org/10.1043/TOJ-17-0033</u>. [PubMed]
- Morris TA, Khoo C, Solomon BJ. Targeting ROS1 Rearrangements in Non-small Cell Lung Cancer: Crizotinib and Newer Generation Tyrosine Kinase Inhibitors. Drugs. 2019; 79:1277–86. <u>https://doi.org/10.1007/s40265-019-01164-3</u>. [PubMed]
- Rotow J, Bivona TG. Understanding and targeting resistance mechanisms in NSCLC. Nat Rev Cancer. 2017; 17:637–58. <u>https://doi.org/10.1038/nrc.2017.84</u>. [PubMed]
- Griesinger F, Roeper J, Pöttgen C, Willborn KC, Eberhardt WE. Brain metastases in ALK-positive NSCLC - time to

adjust current treatment algorithms. Oncotarget. 2018; 9:35181–94. <u>https://doi.org/10.18632/oncotarget.26073</u>. [PubMed]

- Johung KL, Yeh N, Desai NB, Williams TM, Lautenschlaeger T, Arvold ND, Ning MS, Attia A, Lovly CM, Goldberg S, Beal K, Yu JB, Kavanagh BD, et al. Extended survival and prognostic factors for patients with ALK-rearranged non-small-cell lung cancer and brain metastasis. J Clin Oncol. 2016; 34:123–29. <u>https://doi. org/10.1200/JCO.2015.62.0138. [PubMed]</u>
- Yoshida T, Oya Y, Tanaka K, Shimizu J, Horio Y, Kuroda H, Sakao Y, Hida T, Yatabe Y. Clinical impact of crizotinib on central nervous system progression in ALK-positive nonsmall lung cancer. Lung Cancer. 2016; 97:43–47. <u>https:// doi.org/10.1016/j.lungcan.2016.04.006</u>. [PubMed]
- Sullivan I, Planchard D. ALK inhibitors in non-small cell lung cancer: the latest evidence and developments. Ther Adv Med Oncol. 2016; 8:32–47. <u>https://doi.org/10.1177/1758834015617355</u>. [PubMed]
- 11. European Medicines Agency (EMA). Xalkori. 2019. <u>https://www.ema.europa.eu/en/medicines/human/EPAR/xalkori.</u>
- Li J, Huang Y, Wu M, Wu C, Li X, Bao J. Structure and energy based quantitative missense variant effect analysis provides insights into drug resistance mechanisms of anaplastic lymphoma kinase mutations. Sci Rep. 2018; 8:10664. <u>https:// doi.org/10.1038/s41598-018-28752-9</u>. [PubMed]
- Lin JJ, Zhu VW, Yoda S, Yeap BY, Schrock AB, Dagogo-Jack I, Jessop NA, Jiang GY, Le LP, Gowen K, Stephens PJ, Ross JS, Ali SM, et al. Impact of EML4-ALK Variant on Resistance Mechanisms and Clinical Outcomes in ALK-Positive Lung Cancer. J Clin Oncol. 2018; 36:1199–206. <u>https://doi.org/10.1200/JCO.2017.76.2294</u>. [PubMed]
- Gainor JF, Dardaei L, Yoda S, Friboulet L, Leshchiner I, Katayama R, Dagogo-Jack I, Gadgeel S, Schultz K, Singh M, Chin E, Parks M, Lee D, et al. Molecular Mechanisms of Resistance to First- and Second-Generation ALK Inhibitors in ALK-Rearranged Lung Cancer. Cancer Discov. 2016; 6:1118–33. <u>https://doi.org/doi:10.1158/2159-8290.CD-16-0596</u>. [PubMed]
- Food and Drug Administration (FDA). Highlights of prescribing information: Xalkori. 2019. <u>http://www.fda. gov/MedicalDevices/ProductsandMedicalProcedures/</u> <u>InVitroDiagnostics/ucm301431.htm</u>.
- 16. Solomon BJ, Mok T, Kim DW, Wu YL, Nakagawa K, Mekhail T, Felip E, Cappuzzo F, Paolini J, Usari T, Iyer S, Reisman A, Wilner KD, et al, and PROFILE 1014 Investigators. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. N Engl J Med. 2014; 371:2167–77. <u>https://doi.org/10.1056/NEJMoa1408440</u>. [PubMed]
- Ali R, Arshad J, Palacio S, Mudad R. Brigatinib for ALKpositive metastatic non-small-cell lung cancer: design, development and place in therapy. Drug Des Devel Ther. 2019; 13:569–580. <u>https://doi.org/10.2147/DDDT.S147499</u>. [PubMed]

- Vavalà T, Novello S. Alectinib in the treatment of ALKpositive non-small cell lung cancer: an update on its properties, efficacy, safety and place in therapy. Ther Adv Med Oncol. 2018; 10:1758835918789364. <u>https://doi.org/10.1177/1758835918789364</u>. [PubMed]
- Zhang S, Anjum R, Squillace R, Nadworny S, Zhou T, Keats J, Ning Y, Wardwell SD, Miller D, Song Y, Eichinger L, Moran L, Huang WS, et al. The potent ALK inhibitor brigatinib (AP26113) overcomes mechanisms of resistance to first- and second-generation ALK inhibitors in preclinical models. Clin Cancer Res. 2016; 22:5527–38. <u>https://doi. org/10.1158/1078-0432.CCR-16-0569</u>. [PubMed]
- 20. US Food and Drug Administration. FDA approves Zykadia for late-stage lung cancer: breakthrough therapy drug approved four months ahead of review completion goal date. 2014. <u>https://www.fda.gov/newsevents/newsroom/</u> pressannouncements/ucm395299.htm.
- 21. European Medicines Agency (EMA). 2019. <u>https://www.ema.europa.eu/en/medicines/human/EPAR/zykadia</u>.
- 22. US Food and Drug Administration (FDA). Highlights of prescribing information: alecensa. 2019. <u>http://www.fda.gov/CompanionDiagnostics</u>.
- 23. European Medicines Agency (EMA). Alecensa. 2019. <u>https://www.ema.europa.eu/en/medicines/human/EPAR/</u> <u>alecensa</u>.
- 24. Food and Drug Administration (FDA). Highlights of prescribing information: alunbrig. 2019. <u>https://www.alunbrig.com</u>.
- 25. European Medicines Agency (EMA). Alunbrig. Available 2019. <u>https://www.ema.europa.eu/en/medicines/human/</u> <u>EPAR/alunbrig</u>.
- 26. Food and Drug Administration (FDA). FDA broadens ceritinib indication to previously untreated ALK-positive metastatic NSCLC. 2017. <u>https://www.fda.gov/drugs/resources-information-approved-drugs/fda-broadens-ceritinib-indication-previously-untreated-alk-positive-metastatic-nsclc</u>.
- 27. European Medicines Agency (EMA). Zykadia. 2019. https://www.ema.europa.eu/en/documents/smop/chmppost-authorisation-summary-positive-opinion-zykadiaii-12_en.pdf.
- Food and Drug Administration (FDA). Alectinib approved for (ALK) positive metastatic non-small cell lung cancer (NSCLC). 2017. <u>https://www.fda.gov/drugs/resourcesinformation-approved-drugs/alectinib-approved-alkpositive-metastatic-non-small-cell-lung-cancer-nsclc.</u>
- 29. Food and Drug Administration (FDA). FDA approves brigatinib for ALK-positive metastatic NSCLC. <u>https://www.fda.gov/drugs/drug-approvals-and-databases/fda-approves-brigatinib-alk-positive-metastatic-nsclc</u>.
- 30. Peters S, Camidge DR, Shaw AT, Gadgeel S, Ahn JS, Kim DW, Ou SI, Pérol M, Dziadziuszko R, Rosell R, Zeaiter A, Mitry E, Golding S, et al, and ALEX Trial Investigators. Alectinib versus crizotinib in untreated ALK-positive non-

small-cell lung cancer. N Engl J Med. 2017; 377:829–38. https://doi.org/10.1056/NEJMoa1704795. [PubMed]

- 31. Camidge DR, Kim HR, Ahn MJ, Yang JC, Han JY, Hochmair MJ, Lee KH, Delmonte A, Garcia Campelo MR, Kim DW, Griesinger F, Felip E, Califano R, et al. Brigatinib vs crizotinib in patients with ALK inhibitor-naive advanced ALK+ NSCLC: updated results from the phase III ALTA-1L trial. Ann Oncol. 2019; 30:ix195–96. <u>https://doi. org/10.1093/annonc/mdz446</u>.
- 32. Camidge DR, Dziadziuszko R, Peters S, Mok T, Noe J, Nowicka M, Gadgeel SM, Cheema P, Pavlakis N, de Marinis F, Cho BC, Zhang L, Moro-Sibilot D, et al. Updated efficacy and safety data and impact of the EML4-ALK fusion variant on the efficacy of alectinib in untreated ALK-positive advanced non-small cell lung cancer in the Global Phase III ALEX Study. J Thorac Oncol. 2019; 14:1233–43. https://doi.org/10.1016/j.jtho.2019.03.007. [PubMed]
- Camidge DR, Kim DW, Tiseo M, Langer CJ, Ahn MJ, Shaw AT, Huber RM, Hochmair MJ, Lee DH, Bazhenova LA, Gold KA, Ou SI, West HL, et al. Exploratory Analysis of Brigatinib Activity in Patients With Anaplastic Lymphoma Kinase-Positive Non–Small-Cell Lung Cancer and Brain Metastases in Two Clinical Trials. J Clin Oncol. 2018; 36:2693–701. <u>https://doi.org/10.1200/JCO.2017.77.5841</u>. [PubMed]
- 34. Felip E, Orlov S, Park KS, Yu CJ, Tsai CM, Nishio M, Cobo Dols M, McKeage MJ, Su WC, Mok T, Scagliotti GV, Spigel D, Branle F, et al. ASCEND-3: A single-arm, openlabel, multicenter phase II study of ceritinib in ALKi-naïve adult patients (pts) with ALK-rearranged (ALK+) non-small cell lung cancer (NSCLC). J Clin Oncol. 2015; 33:S8060. https://doi.org/10.1200/jco.2015.33.15_suppl.8060.
- 35. Soria JC, Tan DS, Chiari R, Wu YL, Paz- Ares L, Wolf J, Geater SL, Orlov S, Cortinovis D, Yu CJ, Hochmair M, Cortot AB, Tsai CM, et al. First-line ceritinib versus platinum-based chemotherapy in advanced ALK-rearranged non-small-cell lung cancer (ASCEND-4): a randomised, open-label, phase 3 study. Lancet. 2017; 389:917–29. https://doi.org/10.1016/S0140-6736(17)30123-X. [PubMed]
- Li J, Knoll S, Bocharova I, Tang W, Signorovitch J. Comparative efficacy of first-line ceritinib and crizotinib in advanced or metastatic anaplastic lymphoma kinasepositive non-small cell lung cancer: an adjusted indirect comparison with external controls. Curr Med Res Opin. 2019; 35:105–11. <u>https://doi.org/10.1080/03007995.2018.</u> <u>1541443. [PubMed]</u>
- 37. Shaw AT, Kim TM, Crinò L, Gridelli C, Kiura K, Liu G, Novello S, Bearz A, Gautschi O, Mok T, Nishio M, Scagliotti G, Spigel DR, et al. Ceritinib versus chemotherapy in patients with ALK-rearranged non-small-cell lung cancer previously given chemotherapy and crizotinib (ASCEND-5): a randomised, controlled, open-label, phase 3 trial. Lancet Oncol. 2017; 18:874–86. https://doi.org/10.1016/S1470-2045(17)30339-X. [PubMed]

- Novello S, Mazières J, Oh IJ, de Castro J, Migliorino MR, Helland Å, Dziadziuszko R, Griesinger F, Kotb A, Zeaiter A, Cardona A, Balas B, Johannsdottir HK, et al. Alectinib versus chemotherapy in crizotinib-pretreated anaplastic lymphoma kinase (ALK)-positive non-small-cell lung cancer: results from the phase III ALUR study. Ann Oncol. 2018; 29:1409– 16. https://doi.org/10.1093/annonc/mdy121. [PubMed]
- 39. Huber RM, Hansen KH, Paz-Ares Rodríguez L, West HL, Reckamp KL, Leighl NB, Tiseo M, Smit EF, Kim DW, Gettinger SN, Hochmair MJ, Kim SW, Langer CJ, et al. Brigatinib in Crizotinib-Refractory ALK+ NSCLC: 2-Year Follow-up on Systemic and Intracranial Outcomes in the Phase 2 ALTA Trial. J Thorac Oncol. 2020; 15:404–15. https://doi.org/10.1016/j.jtho.2019.11.004. [PubMed]
- Reckamp K, Lin HM, Huang J, Proskorovsky I, Reichmann W, Krotneva S, Kerstein D, Huang H, Lee J. Comparative efficacy of brigatinib versus ceritinib and alectinib in patients with crizotinib-refractory anaplastic lymphoma kinase-positive non-small cell lung cancer. Curr Med Res Opin. 2019; 35:569–76. <u>https://doi.org/10.1080/03007995</u> .2018.1520696. [PubMed]
- 41. Kim DW, Mehra R, Tan DS, Felip E, Chow LQ, Camidge DR, Vansteenkiste J, Sharma S, De Pas T, Riely GJ, Solomon BJ, Wolf J, Thomas M, et al. Activity and safety of ceritinib in patients with ALK-rearranged non-small-cell lung cancer (ASCEND-1): updated results from the multicentre, open-label, phase 1 trial. Lancet Oncol. 2016; 17:452–63. https://doi.org/10.1016/s1470-2045(15)00614-2.
- 42. Mok T, Spigel D, Felip E, de Marinis F, Ahn MJ, Groen HJM, Wakelee HA, Hida T, Crino L, Nishio M, Scagliotti GV, Branle F, Emeremni C, et al. ASCEND-2: A single-arm, open-label, multicenter phase II study of ceritinib in adult patients (pts) with ALK-rearranged (ALK+) non-small cell lung cancer (NSCLC) previously treated with chemotherapy and crizotinib (CRZ). Journal of Clinical Oncology. 2015; 33:8059. https://doi.org/10.1200/jco.2015.33.15_suppl.8059.
- Camidge DR, Gadgeel S, Ou SH, Gandhi L, Riely G, Cetnar J, West H, Socinski M, Chiappori A, Mekhail T, Chao B, Borghaei H, Gold K, et al. MA07.02 Updated efficacy and safety data from the phase 2 NP28761 study of alectinib in ALK-positive non-small-cell lung cancer. J Thorac Oncol. 2017; 12:S378. https://doi.org/10.1016/j.jtho.2016.11.426.
- 44. Barlesi F, Dingemans AM, Yang JC, Ou SI, Ahn JS, De Petris L, Hughes B, Lena H, Bordogna W, Golding S, Morcos PN, Balas B, Zeaiter A, Kim D. Updated efficacy and safety from the global phase II NP28673 study of alectinib in patients (pts) with previously treated ALK b non-small-cell lung cancer (NSCLC). Ann Oncol. 2016; 27:1263P. <u>https://doi.org/10.1093/annonc/mdw383.63</u>.
- Breadner D, Blanchette P, Shanmuganathan S, Boldt RG, Raphael J. Efficacy and safety of ALK inhibitors in ALKrearranged non-small cell lung cancer: A systematic review and meta-analysis. Lung Cancer. 2020; 144:57–63. <u>https:// doi.org/10.1016/j.lungcan.2020.04.011</u>. [PubMed]

- 46. Kassem L, Shohdy KS, Lasheen S, Abdel-Rahman O, Ali A, Abdel-Malek RR. Safety issues with the ALK inhibitors in the treatment of NSCLC: A systematic review. Crit Rev Oncol Hematol. 2019; 134:56–64. <u>https://doi.org/10.1016/j. critrevonc.2018.11.004</u>. [PubMed]
- Sabari JK, Santini FC, Schram AM, Bergagnini I, Chen R, Mrad C, Lai WV, Arbour KC, Drilon A. The activity, safety, and evolving role of brigatinib in patients with ALKrearranged non-small cell lung cancers. Onco Targets Ther. 2017; 10:1983–92. <u>https://doi.org/10.2147/OTT.S109295</u>. [PubMed]
- Amanam I, Gupta R, Mambetsariev I, Salgia R. The brigatinib experience: a new generation of therapy for ALK-positive non-small-cell lung cancer. Future Oncol. 2018; 14:1897–908. <u>https://doi.org/10.2217/fon-2017-0545</u>. [PubMed]
- Jain RK, Chen H. Spotlight on brigatinib and its potential in the treatment of patients with metastatic ALK-positive non-small cell lung cancer who are resistant or intolerant to crizotinib. Lung Cancer (Auckl). 2017; 8:169–77. <u>https:// doi.org/10.2147/LCTT.S126507</u>. [PubMed]
- Uchibori K, Inase N, Araki M, Kamada M, Sato S, Okuno Y, Fujita N, Katayama R. Brigatinib combined with anti-EGFR antibody overcomes osimertinib resistance in EGFRmutated non-small-cell lung cancer. Nat Commun. 2017; 8:14768. <u>https://doi.org/10.1038/ncomms14768</u>. [PubMed]
- 51. Huang WS, Liu S, Zou D, Thomas M, Wang Y, Zhou T, Romero J, Kohlmann A, Li F, Qi J, Cai L, Dwight TA, Xu Y, et al. Discovery of Brigatinib (AP26113), a Phosphine Oxide-Containing, Potent, Orally Active Inhibitor of Anaplastic Lymphoma Kinase. J Med Chem. 2016; 59:4948–64. <u>https://doi.org/10.1021/acs.jmedchem.6b00306. [PubMed]</u>
- 52. Stanicka J, Rieger L, O'Shea S, Cox O, Coleman M, O'Flanagan C, Addario B, McCabe N, Kennedy R, O'Connor R. FES-related tyrosine kinase activates the insulin-like growth factor-1 receptor at sites of cell adhesion. Oncogene. 2018; 37:3131–50. <u>https://doi. org/10.1038/s41388-017-0113-z</u>. [PubMed]
- Rashdan S, Gerber DE. A crowded, but still varied, space: brigatinib in anaplastic lymphoma kinase-rearranged nonsmall cell lung cancer. Transl Cancer Res. 2017; 6:S78–82. <u>https://doi.org/10.21037/tcr.2017.02.12</u>. [PubMed]
- Mezquita L, Planchard D. The role of brigatinib in crizotinib-resistant non-small cell lung cancer. Cancer Manag Res. 2018; 10:123–30. <u>https://doi.org/10.2147/</u> <u>CMAR.S129963. [PubMed]</u>
- 55. Kodama T, Tsukaguchi T, Satoh Y, Yoshida M, Watanabe Y, Kondoh O, Sakamoto H. Alectinib shows potent antitumor activity against RET-rearranged non-small cell lung cancer. Mol Cancer Ther. 2014; 13:2910–18. <u>https://doi. org/10.1158/1535-7163.MCT-14-0274</u>. [PubMed]
- 56. Workman P, Aboagye EO, Balkwill F, Balmain A, Bruder G, Chaplin DJ, Double JA, Everitt J, Farningham DA,

Glennie MJ, Kelland LR, Robinson V, Stratford IJ, et al, and Committee of the National Cancer Research Institute. Guidelines for the welfare and use of animals in cancer research. Br J Cancer. 2010; 102:1555–77. <u>https://doi.org/10.1038/sj.bjc.6605642</u>. [PubMed]

- Trisilowati, Mallet DG. *In Silico* Experimental Modeling of Cancer Treatment. ISRN Oncol. 2012; 2012:828701. <u>https://doi.org/10.5402/2012/828701. [PubMed]</u>
- Jean-Quartier C, Jeanquartier F, Jurisica I, Holzinger A. In silico cancer research towards 3R. BMC Cancer. 2018; 18:408. <u>https://doi.org/10.1186/s12885-018-4302-0</u>. [PubMed]
- Romeo-Guitart D, Forés J, Herrando-Grabulosa M, Valls R, Leiva-Rodríguez T, Galea E, González-Pérez F, Navarro X, Petegnief V, Bosch A, Coma M, Mas JM, Casas C. Neuroprotective Drug for Nerve Trauma Revealed Using Artificial Intelligence. Sci Rep. 2018; 8:1879. <u>https://doi. org/10.1038/s41598-018-19767-3</u>. [PubMed]
- Jorba G, Aguirre-Plans J, Junet V, Segú-Vergés C, Ruiz JL, Pujol A, Fernández-Fuentes N, Mas JM, Oliva B. *In-silico* simulated prototype-patients using TPMS technology to study a potential adverse effect of sacubitril and valsartan. PLoS One. 2020; 15:e0228926. <u>https://doi.org/10.1371/</u> journal.pone.0228926. [PubMed]
- Ni W, Chen W, Lu Y. Emerging findings into molecular mechanism of brain metastasis. Cancer Med. 2018; 7:3820– 33. <u>https://doi.org/10.1002/cam4.1667</u>. [PubMed]
- 62. Popper HH. Progression and metastasis of lung cancer. Cancer Metastasis Rev. 2016; 35:75–91. <u>https://doi.org/10.1007/s10555-016-9618-0. [PubMed]</u>
- Preusser M, Berghoff AS, Berger W, Ilhan-Mutlu A, Dinhof C, Widhalm G, Dieckmann K, Wöhrer A, Hackl M, von Deimling A, Streubel B, Birner P. High rate of FGFR1 amplifications in brain metastases of squamous and nonsquamous lung cancer. Lung Cancer. 2014; 83:83–89. https://doi.org/10.1016/j.lungcan.2013.10.004. [PubMed]
- Wang X, Jiao W, Zhao Y, Zhang L, Yao R, Wang Y, Wang M, Luo Y, Zhao J. CUG-binding protein 1 (CUGBP1) expression and prognosis of brain metastases from non-small cell lung cancer. Thorac Cancer. 2016; 7:32–38. https://doi.org/10.1111/1759-7714.12268. [PubMed]
- 65. Matheus RS, Bernardi FC, Gallo CP, da Silva AP, Rodrigues OR, Capelozzi M, Lopes A, Fenezelian S, Saldiva PH, Capelozzi VL. Nuclear markers (star volume, mitotic index, AgNOR and Ki-67) of the primary tumor and its metastasis in non-small cell lung carcinomas. Pathol Res Pract. 2004; 200:13–23. <u>https://doi.org/10.1016/j.prp.2003.12.007</u>. [PubMed]
- 66. Tseng RC, Lee SH, Hsu HS, Chen BH, Tsai WC, Tzao C, Wang YC. SLIT2 attenuation during lung cancer progression deregulates β-catenin and E-cadherin and associates with poor prognosis. Cancer Res. 2010; 70:543–51. <u>https://doi.org/10.1158/0008-5472.CAN-09-2084</u>. [PubMed]

- Hu L, Zhang J, Zhu H, Min J, Feng Y, Zhang H. Biological characteristics of a specific brain metastatic cell line derived from human lung adenocarcinoma. Med Oncol. 2010; 27:708–14. <u>https://doi.org/10.1007/s12032-009-9273-1</u>. [PubMed]
- Wolf R, Howard OM, Dong HF, Voscopoulos C, Boeshans K, Winston J, Divi R, Gunsior M, Goldsmith P, Ahvazi B, Chavakis T, Oppenheim JJ, Yuspa SH. Chemotactic activity of S100A7 (Psoriasin) is mediated by the receptor for advanced glycation end products and potentiates inflammation with highly homologous but functionally distinct S100A15. J Immunol. 2008; 181:1499–506. <u>https://doi.org/10.4049/jimmunol.181.2.1499</u>. [PubMed]
- Bechmann T, Madsen JS, Brandslund I, Lund ED, Ormstrup T, Jakobsen EH, Jylling AM, Steffensen KD, Jakobsen A. Predicting brain metastases of breast cancer based on serum S100B and serum HER2. Oncol Lett. 2013; 6:1265–70. https://doi.org/10.3892/ol.2013.1536. [PubMed]
- 70. Chen L, Hu X, Wu H, Jia Y, Liu J, Mu X, Wu H, Zhao Y. Over-expression of S100B protein as a serum marker of brain metastasis in non-small cell lung cancer and its prognostic value. Pathol Res Pract. 2019; 215:427–32. https://doi.org/10.1016/j.prp.2018.11.011. [PubMed]
- Han L, Liang XH, Chen LX, Bao SM, Yan ZQ. SIRT1 is highly expressed in brain metastasis tissues of non-small cell lung cancer (NSCLC) and in positive regulation of NSCLC cell migration. Int J Clin Exp Pathol. 2013; 6:2357–65. [PubMed]
- 72. Tugnait M, Gupta N, Hanley MJ, Sonnichsen D, Kerstein D, Dorer DJ, Venkatakrishnan K, Narasimhan N. Effects of Strong CYP2C8 or CYP3A Inhibition and CYP3A Induction on the Pharmacokinetics of Brigatinib, an Oral Anaplastic Lymphoma Kinase Inhibitor, in Healthy Volunteers. Clin Pharmacol Drug Dev. 2020; 9:214–23. https://doi.org/10.1002/cpdd.723. [PubMed]
- 73. Takeuchi K, Soda M, Togashi Y, Suzuki R, Sakata S, Hatano S, Asaka R, Hamanaka W, Ninomiya H, Uehara H, Lim Choi Y, Satoh Y, Okumura S, et al. RET, ROS1 and ALK fusions in lung cancer. Nat Med. 2012; 18:378–81. <u>https://doi.org/10.1038/nm.2658. [PubMed]</u>
- 74. Bethune G, Bethune D, Ridgway N, Xu Z. Epidermal growth factor receptor (EGFR) in lung cancer: an overview and update. J Thorac Dis. 2010; 2:48–51. [PubMed]
- 75. Zhao J, Zou M, Lv J, Han Y, Wang G, Wang G. Effective treatment of pulmonary adenocarcinoma harboring triple EGFR mutations of L858R,T790M, and cis-C797S by osimertinib, bevacizumab, and brigatinib combination therapy: a case report. Onco Targets Ther. 2018; 11:5545– 50. https://doi.org/10.2147/OTT.S170358. [PubMed]
- 76. van der Veeken J, Oliveira S, Schiffelers RM, Storm G, van Bergen En Henegouwen PM, Roovers RC. Crosstalk between epidermal growth factor receptor- and insulin-like growth factor-1 receptor signaling: implications for cancer therapy. Curr Cancer Drug Targets. 2009; 9:748–60. <u>https:// doi.org/10.2174/156800909789271495</u>. [PubMed]

- 77. Lim SH, Kim SY, Kim K, Jang H, Ahn S, Kim KM, Kim NK, Park W, Lee SJ, Kim ST, Park SH, Park JO, Park YS, et al. The implication of FLT3 amplification for FLT targeted therapeutics in solid tumors. Oncotarget. 2017; 8:3237–45. https://doi.org/10.18632/oncotarget.13700. [PubMed]
- Ricciuti B, De Giglio A, Mecca C, Arcuri C, Marini S, Metro G, Baglivo S, Sidoni A, Bellezza G, Crinò L, Chiari R. Precision medicine against ALK-positive non-small cell lung cancer: beyond crizotinib. Med Oncol. 2018; 35:72. <u>https://doi.org/10.1007/s12032-018-1133-4</u>. [PubMed]
- Marsit CJ, Zheng S, Aldape K, Hinds PW, Nelson HH, Wiencke JK, Kelsey KT. PTEN expression in non-small-cell lung cancer: evaluating its relation to tumor characteristics, allelic loss, and epigenetic alteration. Hum Pathol. 2005; 36:768–76. <u>https://doi.org/10.1016/j.humpath.2005.05.006</u>. [PubMed]
- Mou H, Moore J, Malonia SK, Li Y, Ozata DM, Hough S, Song CQ, Smith JL, Fischer A, Weng Z, Green MR, Xue W. Genetic disruption of oncogenic Kras sensitizes lung cancer cells to Fas receptor-mediated apoptosis. Proc Natl Acad Sci U S A. 2017; 114:3648–53. <u>https://doi.org/10.1073/ pnas.1620861114</u>. [PubMed]
- Chuang JC, Stehr H, Liang Y, Das M, Huang J, Diehn M, Wakelee HA, Neal JW. ERBB2-Mutated Metastatic Non-Small Cell Lung Cancer: Response and Resistance to Targeted Therapies. J Thorac Oncol. 2017; 12:833–42. https://doi.org/10.1016/j.jtho.2017.01.023. [PubMed]
- Song JY, Lee SW, Hong JP, Chang SE, Choe H, Choi J. Epidermal growth factor competes with EGF receptor inhibitors to induce cell death in EGFR-overexpressing tumor cells. Cancer Lett. 2009; 283:135–42. <u>https://doi. org/10.1016/j.canlet.2009.03.034</u>. [PubMed]
- Heidegger I, Kern J, Ofer P, Klocker H, Massoner P. Oncogenic functions of IGF1R and INSR in prostate cancer include enhanced tumor growth, cell migration and angiogenesis. Oncotarget. 2014; 5:2723–35. <u>https://doi. org/10.18632/oncotarget.1884</u>. [PubMed]
- Nurwidya F, Andarini S, Takahashi F, Syahruddin E, Takahashi K. Implications of Insulin-like Growth Factor 1 Receptor Activation in Lung Cancer. Malays J Med Sci. 2016; 23:9–21. [PubMed]
- Liu X, Su L, Liu X. Loss of CDH1 up-regulates epidermal growth factor receptor via phosphorylation of YBX1 in nonsmall cell lung cancer cells. FEBS Lett. 2013; 587:3995– 4000. https://doi.org/10.1016/j.febslet.2013.10.036. [PubMed]
- 86. Wang X, Wang H, Li G, Song Y, Wang S, Zhu F, Guo C, Zhang L, Shi Y. Activated macrophages down-regulate expression of E-cadherin in hepatocellular carcinoma cells via NF-κB/Slug pathway. Tumour Biol. 2014; 35:8893–901. https://doi.org/10.1007/s13277-014-2159-7. [PubMed]
- Warsito D, Sjöström S, Andersson S, Larsson O, Sehat B. Nuclear IGF1R is a transcriptional co-activator of LEF1/TCF. EMBO Rep. 2012; 13:244–50. <u>https://doi.org/10.1038/embor.2011.251</u>. [PubMed]

- Jamwal G, Singh G, Dar MS, Singh P, Bano N, Syed SH, Sandhu P, Akhter Y, Monga SP, Dar MJ. Identification of a unique loss-of-function mutation in IGF1R and a crosstalk between IGF1R and Wnt/β-catenin signaling pathways. Biochim Biophys Acta-Mol Cell Res. 2018; 1865:920–31. https://doi.org/10.1016/j.bbamcr.2018.03.013. [PubMed]
- Gainor JF, Ou SH, Logan J, Borges LF, Shaw AT. The central nervous system as a sanctuary site in ALK-positive non-small-cell lung cancer. J Thorac Oncol. 2013; 8:1570– 73. <u>https://doi.org/10.1097/JTO.000000000000029</u>. [PubMed]
- Costa DB, Kobayashi S, Pandya SS, Yeo WL, Shen Z, Tan W, Wilner KD. CSF concentration of the anaplastic lymphoma kinase inhibitor crizotinib. J Clin Oncol. 2011; 29:e443–45. <u>https://doi.org/10.1200/jco.2010.34.1313</u>. [PubMed]
- 91. Rangachari D, Yamaguchi N, VanderLaan PA, Folch E, Mahadevan A, Floyd SR, Uhlmann EJ, Wong ET, Dahlberg SE, Huberman MS, Costa DB. Brain metastases in patients with EGFR-mutated or ALK-rearranged non-small-cell lung cancers. Lung Cancer. 2015; 88:108–11. <u>https://doi. org/10.1016/j.lungcan.2015.01.020</u>. [PubMed]
- 92. Zhang I, Zaorsky NG, Palmer JD, Mehra R, Lu B. Targeting brain metastases in ALK-rearranged non-small-cell lung cancer. Lancet Oncol. 2015; 16:e510–21. <u>https://doi. org/10.1016/S1470-2045(15)00013-3</u>. [PubMed]
- 93. Gadgeel SM, Shaw AT, Govindan R, Gandhi L, Socinski MA, Camidge DR, De Petris L, Kim DW, Chiappori A, Moro-Sibilot DL, Duruisseaux M, Crino L, De Pas T, et al. Pooled analysis of CNS response to alectinib in two studies of pretreated patients with ALK-positive non-small-cell lung cancer. J Clin Oncol. 2016; 34:4079–85. <u>https://doi.org/10.1200/JCO.2016.68.4639</u>. [PubMed]
- Addeo A, Tabbò F, Robinson T, Buffoni L, Novello S. Precision medicine in ALK rearranged NSCLC: A rapidly evolving scenario. Crit Rev Oncol Hematol. 2018; 122:150– 56. <u>https://doi.org/10.1016/j.critrevonc.2017.12.015</u>. [PubMed]
- Li D, Yang W, Arthur C, Liu JS, Cruz-Niera C, Yang MQ. Systems biology analysis reveals new insights into invasive lung cancer. BMC Syst Biol. 2018; 12:117. <u>https://doi. org/10.1186/s12918-018-0637-z</u>. [PubMed]
- 96. Tan J, Li M, Zhong W, Hu C, Gu Q, Xie Y. Tyrosine kinase inhibitors show different anti-brain metastases efficacy in NSCLC: A direct comparative analysis of icotinib, gefitinib, and erlotinib in a nude mouse model. Oncotarget. 2017; 8:98771–81. <u>https://doi.org/10.18632/oncotarget.21936</u>. [PubMed]
- 97. Kong R, Yi F, Wen P, Liu J, Chen X, Ren J, Li X, Shang Y, Nie Y, Wu K, Fan D, Zhu L, Feng W, Wu JY. Myo9b is a key player in SLIT/ROBO-mediated lung tumor suppression. J Clin Invest. 2015; 125:4407–20. <u>https://doi.org/10.1172/JCI81673. [PubMed]</u>
- Chen LJ, Li XY, Zhao YQ, Liu WJ, Wu HJ, Liu J, Mu XQ, Wu HB. Down-regulated microRNA-375 expression

as a predictive biomarker in non-small cell lung cancer brain metastasis and its prognostic significance. Pathol Res Pract. 2017; 213:882–88. <u>https://doi.org/10.1016/j.</u> prp.2017.06.012. [PubMed]

- 99. Sun D, Li X, Ma M, Liu J, Xu Y, Ye L, Hou H, Wang C, Li X, Jiang Y. The predictive value and potential mechanisms of miRNA-328 and miRNA-378 for brain metastases in operable and advanced non-small-cell lung cancer. Jpn J Clin Oncol. 2015; 45:464–73. <u>https://doi.org/10.1093/jjco/hyv009</u>. [PubMed]
- 100. Pang X, Min J, Liu L, Liu Y, Ma N, Zhang H. S100B protein as a possible participant in the brain metastasis of NSCLC. Med Oncol. 2012; 29:2626–32. <u>https://doi.org/10.1007/ s12032-012-0169-0. [PubMed]</u>
- 101. Jiang W, Jia Q, Liu L, Zhao X, Tan A, Ma N, Zhang H. S100B promotes the proliferation, migration and invasion of specific brain metastatic lung adenocarcinoma cell line. Cell Biochem Funct. 2011; 29:582–88. <u>https://doi. org/10.1002/cbf.1791</u>. [PubMed]
- 102. Fontana D, Ceccon M, Gambacorti- Passerini C, Mologni L. Activity of second-generation ALK inhibitors against crizotinib-resistant mutants in an NPM-ALK model compared to EML4-ALK. Cancer Med. 2015; 4:953–65. https://doi.org/10.1002/cam4.413. [PubMed]
- 103. Ceccon M, Mologni L, Giudici G, Piazza R, Pirola A, Fontana D, Gambacorti-Passerini C. Treatment Efficacy and Resistance Mechanisms Using the Second-Generation ALK Inhibitor AP26113 in Human NPM-ALK-Positive Anaplastic Large Cell Lymphoma. Mol Cancer Res. 2015; 13:775–83. <u>https://doi.org/10.1158/1541-7786.MCR-14-0157</u>. [PubMed]
- 104. Bazhenova L, Hodgson JG, Langer CJ, Simon GR, Gettinger SN, Ou SH, Reckamp KL, West HJ, Chiappori A, Koh HA, Molina JR, Shaw AT, Patel JD, et al. Activity of brigatinib (BRG) in crizotinib (CRZ)-resistant ALK+ NSCLC patients (pts) according to ALK plasma mutation status. J Clin Oncol. 2017; 35:9065. <u>https://doi.org/10.1200/ JCO.2017.35.15_suppl.9065</u>.
- 105. Katayama R. Drug resistance in anaplastic lymphoma kinase-rearranged lung cancer. Cancer Sci. 2018; 109:572– 80. <u>https://doi.org/10.1111/cas.13504.</u> [PubMed]
- 106. Rothenstein JM, Chooback N. ALK inhibitors, resistance development, clinical trials. Curr Oncol. 2018; 25:S59–67. <u>https://doi.org/10.3747/co.25.3760</u>. [PubMed]
- 107. Murphy CC, Fullington HM, Alvarez CA, Betts AC, Lee SJ, Haggstrom DA, Halm EA. Polypharmacy and patterns of prescription medication use among cancer survivors. Cancer. 2018; 124:2850–57. <u>https://doi.org/10.1002/ cncr.31389</u>. [PubMed]
- 108. LeBlanc TW, McNeil MJ, Kamal AH, Currow DC, Abernethy AP. Polypharmacy in patients with advanced cancer and the role of medication discontinuation. Lancet Oncol. 2015; 16:e333–41. <u>https://doi.org/10.1016/S1470-2045(15)00080-7</u>. [PubMed]

- 109. Rodig SJ, Mino-Kenudson M, Dacic S, Yeap BY, Shaw A, Barletta JA, Stubbs H, Law K, Lindeman N, Mark E, Janne PA, Lynch T, Johnson BE, et al. Unique clinicopathologic features characterize ALK-rearranged lung adenocarcinoma in the western population. Clin Cancer Res. 2009; 15:5216–23. https://doi.org/10.1158/1078-0432.CCR-09-0802. [PubMed]
- 110. Gao T, Qian J. EAGLE: an algorithm that utilizes a small number of genomic features to predict tissue/cell typespecific enhancer-gene interactions. PLoS Comput Biol. 2019; 15:e1007436. <u>https://doi.org/10.1371/journal.pcbi.1007436</u>. [PubMed]
- 111. Kuperstein I, Bonnet E, Nguyen HA, Cohen D, Viara E, Grieco L, Fourquet S, Calzone L, Russo C, Kondratova M, Dutreix M, Barillot E, Zinovyev A. Atlas of Cancer Signalling Network: a systems biology resource for integrative analysis of cancer data with Google Maps. Oncogenesis. 2015; 4:e160–14. <u>https://doi.org/10.1038/ oncsis.2015.19</u>. [PubMed]
- 112. Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, Sajed T, Johnson D, Li C, Sayeeda Z, Assempour N, Iynkkaran I, Liu Y, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. Nucleic Acids Res. 2018; 46:D1074–82. <u>https://doi.org/10.1093/nar/gkx1037</u>. [PubMed]
- 113. Szklarczyk D, Santos A, von Mering C, Jensen LJ, Bork P, Kuhn M. STITCH 5: augmenting protein-chemical interaction networks with tissue and affinity data. Nucleic Acids Res. 2016; 44:D380–84. <u>https://doi.org/10.1093/nar/ gkv1277. [PubMed]</u>
- 114. Hecker N, Ahmed J, von Eichborn J, Dunkel M, Macha K, Eckert A, Gilson MK, Bourne PE, Preissner R. SuperTarget goes quantitative: update on drug-target interactions. Nucleic Acids Res. 2012; 40:D1113–17. <u>https://doi. org/10.1093/nar/gkr912</u>. [PubMed]
- 115. Lorén V, Garcia-Jaraquemada A, Naves JE, Carmona X, Mañosa M, Aransay AM, Lavin JL, Sánchez I, Cabré E, Manyé J, Domènech E. ANP32E, a Protein Involved in Steroid-Refractoriness in Ulcerative Colitis, Identified by a Systems Biology Approach. J Crohns Colitis. 2019; 13:351– 61. <u>https://doi.org/10.1093/ecco-jcc/jiy171</u>. [PubMed]
- 116. Iborra-Egea O, Gálvez-Montón C, Roura S, Perea-Gil I, Prat-Vidal C, Soler-Botija C, Bayes-Genis A. Mechanisms of action of sacubitril/valsartan on cardiac remodeling: a systems biology approach. NPJ Syst Biol Appl. 2017; 3:12. <u>https://doi.org/10.1038/s41540-017-0013-4</u>. [PubMed]
- 117. Zhang XY, Trame MN, Lesko LJ, Schmidt S. Sobol sensitivity analysis: A tool to guide the development and evaluation of systems pharmacology models. CPT Pharmacometrics Syst Pharmacol. 2015; 4:69–79. <u>https:// doi.org/10.1002/psp4.6</u>. [PubMed]
- 118. Bishop CM. Pattern recognition and machine learning. Springer Science and Business Media. 2006.
- 119. Fiuza-Luces C, Santos-Lozano A, Llavero F, Campo R, Nogales-Gadea G, Díez-Bermejo J, Baladrón C, González-

Murillo Á, Arenas J, Martín MA, Andreu AL, Pinós T, Gálvez BG, et al. Muscle molecular adaptations to endurance exercise training are conditioned by glycogen availability: a proteomics-based analysis in the McArdle mouse model. J Physiol. 2018; 596:1035–61. <u>https://doi.org/10.1113/JP275292</u>. [PubMed]

- 120. Herrando-Grabulosa M, Mulet R, Pujol A, Mas JM, Navarro X, Aloy P, Coma M, Casas C. Novel Neuroprotective Multicomponent Therapy for Amyotrophic Lateral Sclerosis Designed by Networked Systems. PLoS One. 2016; 11:e0147626. <u>https://doi.org/10.1371/journal.pone.0147626</u>. [PubMed]
- 121. Herreros-Villanueva M, Perez-Palacios R, Castillo S, Segu C, Sardon T, Mas JM, Martin AC, Arroyo R. Biological Relationships between miRNAs used for Colorectal Cancer Screening. J Mol Biomark Diagn. 2018; 09. <u>https://doi.org/10.4172/2155-9929.1000398</u>.