

Comprehensive Review of Phytochemical Profiles and Health-Promoting Effects of Different Portions of Wampee (*Clausena lansium*)

Published as part of the ACS Omega virtual special issue "Phytochemistry".

Xin Huang,[#] Minghe Wang,[#] Saiyi Zhong, and Baojun Xu*



Cite This: *ACS Omega* 2023, 8, 26699–26714



Read Online

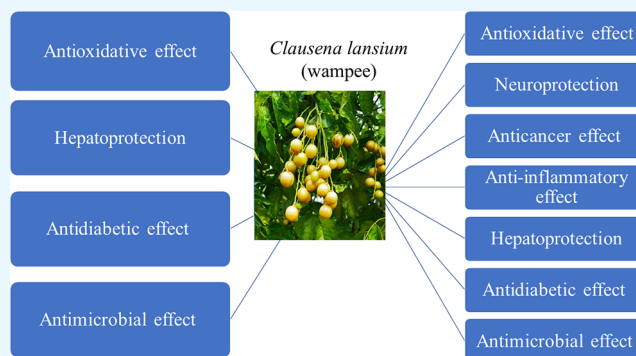
ACCESS |

Metrics & More

Article Recommendations

Supporting Information

ABSTRACT: *Clausena lansium*, commonly known as wampee, is a subtropical fruit from the Rutaceae family characterized by its high nutrient content and numerous bioactive substances. This low-fat fruit is abundant in fiber, vitamins, minerals, and essential amino acids. Wampee has been found to contain several bioactive compounds, including essential oils, phenolic compounds, and alkaloids. These bioactive constituents provide numerous health-enhancing properties, such as antioxidant, neuroprotective, anticarcinogenic, anti-inflammatory, hepatoprotective, antidiabetic, and antimicrobial effects. The relationship between these compounds and their impacts on health has been explored in various studies. While the disease-prevention efficacy of *C. lansium* has been established, additional research is necessary to elucidate the precise mechanisms and metabolic pathways involved. This paper presents a comprehensive review of wampee, focusing on its bioactive compounds, the beneficial effects derived from its consumption, and the evidence supporting the development of wampee-based functional foods in future studies.



1. INTRODUCTION

Clausena lansium (Lour.) Skeel, commonly referred to as wampee, is a subtropical fruit belonging to the Rutaceae family, whose origins are traced back to China.¹ It has been cultivated in China for more than 1500 years, and several varieties have been developed through selective breeding.² The fruit is commercially grown in China, Vietnam, Thailand, Malaysia, the Philippines, and India due to its high-value status. Wampee typically exhibits an oval shape and measures approximately 2.0 cm in diameter. Its yellow peel encases 1–5 seeds, as shown in Figure 1. It generally ripens from May to August in different cultivation areas due to geographical and climatic differences.³ The edible part of the wampee includes the pulp and peel. Besides being consumed as fresh fruit, wampee also can be processed into long-shelf-life food products, such as jams, beverages, preserved fruits, and wines. Dried wampee in Thailand is popular with locals.⁴ Processing is a possible way to expand the wampee market.

In addition, wampee could be employed in medicine rather than just being consumed as a typical food. In the view of Asian folk medicine, such as traditional Chinese medicine (TCM), the leaves, stem, bark, root, peel, and seeds of wampees can be used to treat various diseases. The water extraction of wampee leaves and peel can be used to treat

certain dermatological disorders, acute and chronic viral hepatitis, as well as asthma.⁵ The therapeutic properties of the wampee's root, bark, and stem were effective in addressing bronchitis and malarial fever, while the pulp and seeds were purportedly beneficial in alleviating symptoms of cough, dyspepsia, and specific gastrointestinal disorders.^{6,7} The wampee is renowned not only for its fruit but also for its substantial medicinal properties. Clausenolide-1-ethyl ether, a limonoid extracted from wampee, has been scientifically demonstrated to exhibit anti-HIV activity. Another compound, clausine-D, also derived from the wampee plant, possesses an antiplatelet effect that can aid in preventing blood clot formation. Additional pharmacological benefits attributed to the wampee include antibacterial, antifungal, and antimalarial activities. The myriad of therapeutic potentials this plant offers thus emphasizes its significance in medical research.⁸ In previous decades, with the help of advanced research

Received: April 26, 2023

Accepted: July 6, 2023

Published: July 19, 2023



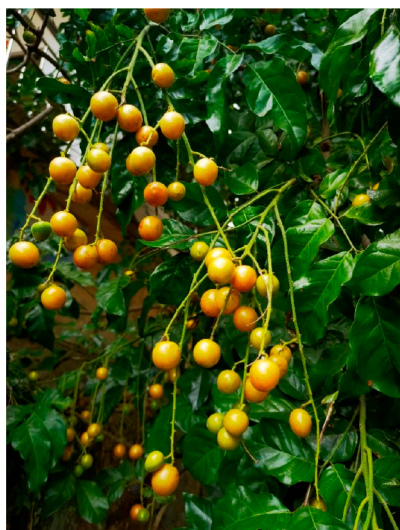


Figure 1. Wampee fruits on the tree.

techniques, the chemical composition and nutrient profiles of the wampee were explored. Relevant research has demonstrated that wampee is a rich source of various nutrients and phytochemicals, such as polyphenols, minerals, vitamins, and flavonoids, which possess health-promoting properties. These beneficial effects include antioxidative, anti-inflammatory, and antimicrobial activity.⁹

Several researchers have reported the bioactive compounds isolated from wampee, but no comprehensive information about the chemical profiles and bioactivities of wampee has been reported. This review aims to summarize the chemical constituents and corresponding bioactivities of different parts of the wampee. This comprehensive review is helpful for the better development of wampee fruit and related products.

2. CHEMICAL COMPOSITIONS

2.1. Proximate Compositions. The nutritional value of wampee is primarily attributed to its pulp, which serves as the primary edible portion. Supplemental Table 1 demonstrates the major macronutrient composition of wampee determined in several types of research. Wampee is a kind of tropical fruit with a high water content, and water occupies around 78.93 to 84.00% of the fresh weight (FW) of wampee pulp. The ash content in the wampee ranges from 0.9 to 2.6 g/100 g of FW. In every 100 g of the consumable pulp of the wampee fruit, the composition comprises 0.61 to 3.78 g of protein, 0.10 to 0.28 g of fat, and 9.9 to 14.1 g of carbohydrates by fresh weight. The fiber content is up to 4.58 g/100 g of wampee, establishing it as an outstanding source of dietary fiber. Additionally, the appropriate ratio between soluble sugar (10.16 g/100 g on average) and total acid (1.55 g/100 g on average) balances the sweet and sour taste to wampee. The low fat and sugar contents in wampee result in a low energy level of around 47 kcal/100 g. The variance of constituent content in other research may be attributed to the growing conditions, geographic factors, and analytical methods.^{4,10,11}

2.2. Vitamins and Minerals. Wampee could be a good supplement for vitamins and minerals in a daily diet. The most abundant vitamin in wampee is vitamin C (548 mg/kg), higher than other tropical fruits like lychee (364 mg/kg) and papaya (512 mg/kg).^{12,13} Other vitamins and precursors, including vitamin E (1.58 mg/kg), vitamin B1 (1.35 mg/kg), vitamin B2

(0.72 mg/kg), and niacin (0.33 mg/kg), as well as β -carotene (0.016 mg/kg), are also found in wampee. The wampee fruit also serves as a significant source of essential minerals. Each kilogram of wampee contains significant amounts of essential minerals such as 3500 mg potassium, 710 mg calcium, and 220 mg phosphorus, which are crucial for maintaining homeostasis in the body.¹¹ The contents of vitamins and minerals for fresh wampee are summarized in Supplemental Table 1.

2.3. Protein and Amino Acid. The protein content of wampee is 6.20%, while abundant amino acids were found in wampee pulp.¹⁴ A total of 17 amino acids were isolated and quantified from wampee and summarized in Supplemental Table 2.^{14–16} Among the identified amino acids in wampee pulp, alanine was the most abundant, with a concentration of 10.50 mg/g, followed by asparagine (6.10 mg/g), glutamine (4.10 mg/g), serine (3.00 mg/g), lysine (2.30 mg/g), and leucine (2.00 mg/g).

Amino acid assessment indicates that wampee contains nine essential amino acids (EAA), including histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, valine, and arginine, which make up around 28% of the total amount of amino acids. Therefore, the consumption of wampee may contribute to the sufficient daily intake of essential amino acids. Amino acids play a crucial role in physiological activities and food taste development. Based on their taste properties in foods, amino acids could be classified into three groups: bitter, sweet, and umami. The amino acid assessment reveals that the sweet amino acids (Thr, Ser, Gly, and Ala) are the dominant group of amino acids accounting for nearly 41% of the total amino acids. Additionally, the umami amino acids (Asp and Glu) make up about 28% of the total amino acids in wampee. As a result, it is reasonable that the amino acids in wampee contribute to developing its sweet and umami taste profile.

2.4. Carbohydrates. The carbohydrate content of wampee is primarily made up of free sugars and polysaccharides. Fructose, glucose, and sucrose are the most dominant free sugars in wampee. Some research reported a slight difference in soluble sugar content among different species of wampee fruit.¹⁷ The total soluble sugar content could be up to 87.35 mg/g, consisting of fructose (40.14 mg/g), glucose (28.39 mg/g), and sucrose (18.81 mg/g). The median total soluble sugar content in wampee is 100 mg/g, including 50.6 mg/g of sucrose.¹⁸

Besides the free sugars, the polysaccharides occupy a considerable portion of carbohydrates in the wampee. Polysaccharide is a group of bioactive compounds with different health benefits, such as antioxidative and anti-inflammatory activities, etc. Additionally, it involves immunoregulation and cell activities, thus playing a critical role in health.¹⁹ Polysaccharides in wampee are mainly composed of pectin, cellulose, and hemicellulose. Various methodologies, such as microwave-assisted extraction and ethanol sedimentation, have obtained crude polysaccharide extracts from wampee.

The composition of monosaccharides in the wampee may exhibit variations, which can be attributed to the difference in extraction methodologies and the specific portions of the wampee fruit utilized. For example, the polysaccharides obtained from the seed and peel of wampee were composed of mannose, galacturonic acid, galactose, arabinose, and glucose with a ratio of 11.70:55.37:9.58:17.81:2.03 and 2.76:48.28:15.83:27.63:2.74, respectively.^{20,21}

Table 1. Volatile and Phenolic Compounds of Wampee^a

Volatile compounds of wampee					Phenolic content of wampee (based on fresh weight)			
Compound	% Relative area				Portion	Phenolic compounds	Content (mg/100 g FW)	References
	Leaves	Pulp	Peel	Seed				
Sabinene	14.92	50.64	69.07	83.56	Leaves	Gallic acid	90.3	Kong et al., 2018 ³⁶
β -Bisabolene	9.88	ND	ND	0.15		Chlorogenic acid	221.8	
β -Caryophyllene	7.72	ND	ND	0.55		Caffeic acid	67.7	
α -Zingiberene	6.52	ND	ND	0.06		Rutin	1700.5	
3-Cyclohexen-1-ol	ND	15.17	0.28	0.51		Ferulic acid	82.4	
Cyclohexene	ND	6.5	0.17	0.39		Quercetin	2.87	
1,4-Cyclohexadiene	ND	6.19	0.32	ND	Catechin	1.91	Chang, Lu et al., 2018 ⁶⁷	
α -Phellandrene	1.38	5.03	10.63	3.08	Ferulic acid	2.94		
α -Pinene	1.99	2.08	9.41	4.26	Chlorogenic acid	1.39		
Myrcene	1.1	1.7	3.15	2.94	Vanillic acid	1.66		
Acetic acid	0.94	2.65	0.08	0.03	Vanillic acid	1.77		
Hexanal	1.55	0.47	0.04	ND	Ferulic acid	2.43		
Isosativene	0.38	ND	0.07	0.01	Rutin	15.87	Ye et al., 2019 ³²	
Butanal	8.61	ND	ND	ND	Syringin	15.85		
Reference	Chokeprasert et al., 2007 ²⁵				Catechin	262.7		
Phenolic content of wampee (based on dry weight)					Hesperetin	1.63		
Portion	Phenolic compounds	Content (mg/100 g DW)		References	<i>p</i> -Coumaric acid	9.87		
Leaves	Gallic acid	224.7		Chen et al., 2017 ³⁸	7-Hydroxycoumarin	12.03		
	(+)-Catechin	762.3			Ferulic acid	13.15		
	Vanillic acid	609.1			Rutin	98.94		
	Caffeic acid	1514.1			Syringin	9.8		
	Syringic acid	216.4			Rutin	656.8		
	Epicatechin	424.5			Chang et al., 2022 ⁴⁰	Benzoic acid	350.1	
<i>p</i> -Coumaric acid	12836.2		2-Methoxycinnamic acid	29.1				
Leaves	Ferulic acid	5417.2		Kaempferol		20.7		
	Rutin	9241.5		Hesperetin		20.9		
	Isoquercitrin	2760.9		Nobiletin		39.1		
	Quercitrin	2730.7		Tangeretin		48		
Quercetin	5641.3							
Peel	Proanthocyanidins	1.88		Chai et al., 2017 ³⁹				

^a“ND”: Not detected.

Furthermore, the difference in extraction method may cause the variation in monosaccharide composition, even from the same portion of the wampee. For example, galacturonic acid is one of the dominant monosaccharides in the pulp extract obtained through ethanol sedimentation, accounting for 62.16% of the total content. However, the ultrasonic microwave-assisted enzymatic method resulted in arabinose being the most abundant polysaccharide present in the pulp.^{22,23} The characteristics of polysaccharides extracted from a different portion of wampee have been summarized in Supplemental Table 3.

2.5. Volatile Compounds. Essential oils are natural bioactive products consisting of volatile compounds found in plants. The essential oils derived from wampee fruit exhibit potent health-promoting and pharmacological properties, including anti-inflammatory, antibacterial, and anticancer activities. These volatile components also contribute to the distinctive and appealing aroma of fruits. Thus, examining the chemical composition of essential oils and volatiles from wampee fruit is crucial, as they play a role in both its bioactivity and aroma. The volatile constituents of wampee essential oils can be classified into several major groups: terpenoids and their derivatives as well as alcohols and olefin compounds.

Various extraction methods could be applied to extract volatile compounds in plants, such as supercritical fluid

extraction, hydrodistillation, steam distillation, and hydrodiffusion, etc.²⁴ In addition, gas chromatography and mass spectrometry (GC-MS) are frequently utilized techniques to identify volatile compounds.

Volatile compounds were isolated and identified from fresh pulp, leaves, and seeds of wampee using GC-MS with a headspace sampler.²⁵ It is reported that the volatile compounds from wampee pulp are mainly made up of monoterpenes (76.0%) and alcohols (17.5%). In the wampee leaves, 86% of the entire volatile fraction was characterized. The primary components among these constituents were sesquiterpenes, accounting for 28%, and monoterpenes, comprising 22%. The major volatile components in the seed of wampee were found to be sabinene, β -phellandrene, and 4-terpineol by using GC-MS analysis.²⁶ Volatile oil was extracted from wampee pulp by supercritical fluid extraction.²⁷ The main compositions in wampee essential oil were 4-terpineol (26.94%), followed by γ -terpinene (14.39%), β -phellandrene (8.24%), sabinene (5.58%), and cymene (5.01%). Among these substances, β -phellandrene is an attractive resource for its potential antifungal activities.^{26,28} There are some differences between the characterized volatile compositions in the studies, possibly due to the differences in temperature, pressure, and polarity of different extraction methods.²⁹

Furthermore, the content of identical substances may exhibit considerable differences in various studies because of variations in climate, growing environment, and plucking. The volatile compositions in different parts of the wampee were also investigated, and it was found that monoterpene dominates the total volatile content.²⁵ Sabinene was found to have the highest amount in all parts of the wampee: leaves (14.92%), pulp (50.64%), peel (69.07%), and seed (83.56%). The volatile compounds in the wampee were summarized in Table 1.

2.6. Phenolic Compounds. Phenolic compounds are a massive group of phytochemicals, which are secondary metabolites synthesized through the shikimic acid and phenylpropanoid pathways in plants.³⁰ The typical chemical structure of phenolic compounds comprises an aromatic ring with one or more hydroxyl groups. Phenolic compounds can be divided into several classes, including phenolic acids (e.g., hydroxycinnamic acid, hydroxybenzoic acid), flavonoids (e.g., flavonols, flavanols, and anthocyanidins), tannins (e.g., hydrolyzable tannins, condensed tannins), stilbenes, coumarins, and lignans.³¹

The phenolic composition and concentrations of wampee may exhibit considerable discrepancies across various studies owing to the diversity in numerous aspects, such as geographical distribution, sample preparation, analytical approaches, and statistical analysis. Consequently, it is essential to consider these variables when evaluating the phenolic compound profiles derived from different studies.

The studies identifying phenolic compounds primarily focused on the pulp, peel, and leaf portion of wampee. Ye et al.³² extracted phytochemicals from wampee pulp using ethanol as the solvent and measured the phenolic content by the Folin–Ciocalteu method. The total phenolic content in wampee pulp was reported to be up to 907.8 mg gallic acid equivalent (GAE)/100 g FW, while the total flavonoid content in wampee pulp was up to 536.6 mg catechin equivalent (CE)/100 g FW. In another study, Prasad et al.³³ investigated the phenolic content in wampee peel using the Folin–Ciocalteu method and found that the total phenolic content in the ethanol extract fraction is 4.62 GAE mg/100 g dry weight (DW). The highest level of total phenolic content was observed in the ethyl acetate fraction, which was 33 mg GAE/100 g DW. In addition, Chang et al.³⁴ found that as the wampee leaves grow there is an accumulation of total phenolic content.

At least 20 phenolic compounds have been isolated and identified from different parts of wampee, including hydroxybenzoic acid (gallic acid, vanillic acid, and benzoic acid), hydroxycinnamic acid (ferulic acid, 2-methoxycinnamic acid, chlorogenic acid, caffeic acid, and *p*-coumaric acid), flavonol (rutin, kaempferol, quercetin, and isoquercitrin), flavanone (hesperetin), flavone (nobiletin, tangeretin), flavanol (catechin), and phenolic glycosides (syngin).^{32,35–40} In addition, coumarin is a class of phenolic compounds widely existing in wampee. All of these identified compounds were summarized in Table 1.

2.7. Alkaloids. Alkaloids represent a significant class of plant-derived secondary metabolites exhibiting pharmacological activity, and their biodynamic activities may benefit human health.⁴¹ Alkaloids are abundant in different parts of the wampee and could be classified into several types.

2.7.1. Carbazole Alkaloid. Carbazole alkaloid is an important subgroup of alkaloids. The fundamental chemical composition of carbazole is characterized by a central pyrrole

ring, which is coalesced with two benzene rings on either side.⁴² This class of compounds is considered as an anticancer agent because of their biological activities.⁴³

Serving as a significant source of carbazole alkaloids, various compounds have been extracted and identified from multiple sections of the wampee plant. Ten carbazole alkaloids named claulansines A–J were extracted from the stem of the wampee. Among these alkaloids, 10 μ M of claulansines A, F, H, I, and J effectively protected pheochromocytoma (PC12) cells from apoptosis.⁴⁴ Furthermore, claulansine F was found to be able to promote neuritogenesis in PC12 cells by activating the ERK signaling pathway, thus exerting its neuroprotective activity.

Moreover, a new carbazole alkaloid, claulansine K, was first isolated from wampee peels and exhibited *in vitro* α -glucosidase inhibitory activity.⁴⁵ One year later, Du et al.⁴⁶ characterized two new carbazole alkaloid claulansine S and T in the stem of the wampee. This research group extracted another seven carbazole alkaloids named claulansines L–R from the wampee stem in the same year. They subsequently discovered that claulansines N, P, Q, and S demonstrated hepatoprotective properties in human HepG2 hepatoma cells.⁴⁷ Subsequently, Liu et al.⁴⁸ extensively studied the alkaloid profiles in wampee fruits and isolated 16 carbazole alkaloids, while six were first reported. The research group also evaluated the neuroprotective activities of these alkaloids toward the human neuroblastoma SH-SY5Y cell line, proposing that wampee could potentially serve as a preventative and therapeutic agent for Parkinson's disease. Additionally, five new carbazole alkaloids were extracted from the stem of wampee, named claulansine A (Figure 2A), claulansine U, claulansine V, claulansine W, and (–)-(2′*R*)-claulamine A, respectively. These compounds presented potential protective effects on PC12 cells against serum deprivation and anticancer activities.⁴⁹ Recently, a new carbazole alkaloid, called claulansine X, has been identified in

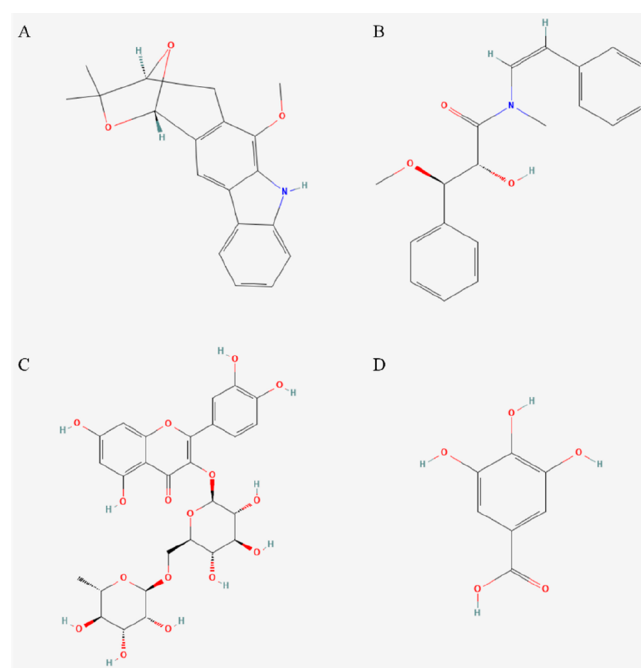


Figure 2. Structures of some important secondary metabolites: (A) claulansine A; (B) claulansamide C; (C) rutin; (D) gallic acid. The image was extracted from PubChem.

Table 2. Alkaloids in Wampee

Carbazole alkaloid in wampee						
No.	Name	Molecular formula	Source	Bioactivities	References	
1	3-Formyl-6-methoxycarbazole	C ₁₄ H ₁₁ NO ₂	Root	-	Li et al., 1991 ⁶	
2	Methyl 6-methoxycarbazole-3-carboxylate	C ₁₅ H ₁₃ NO ₃		-		
3	3-Formyl-1,6-dimethoxycarbazole	C ₁₅ H ₁₃ NO ₃		-		
4	Mafaicheenamines A	C ₁₉ H ₁₉ NO ₄	Twigs	Anticancer	Maneerat & Laphookhieo, 2010 ⁸⁸	
5	Mafaicheenamamine B	C ₁₉ H ₂₁ NO ₅				
6	Mafaicheenamines C	C ₁₉ H ₁₉ NO ₃				
7	Mafaicheenamines D	C ₁₉ H ₁₈ NO ₂	Root	Anticancer	Maneerat et al., 2012 ⁸⁹	
8	Mafaicheenamines E	C ₁₉ H ₁₈ NO ₃				
9	Claulansine A	C ₁₉ H ₁₉ NO ₃				
10	Claulansine B	C ₁₉ H ₁₉ NO ₄				
11	Claulansine C	C ₁₉ H ₂₀ NO ₅				
12	Claulansine D	C ₁₉ H ₁₉ NO ₅				
13	Claulansine E	C ₁₆ H ₁₃ NO ₃				
14	Claulansine F	C ₁₉ H ₁₇ NO ₃	Stem	Neuroprotective	Liu et al., 2012 ⁴⁴	
15	Claulansine G	C ₁₉ H ₁₇ NO ₂				
16	Claulansine H	C ₁₉ H ₁₉ NO ₄				
17	Claulansine I	C ₁₈ H ₁₈ NO ₂				
18	Claulansine J	C ₁₄ H ₁₁ NO ₄				
19	Claulansine K	C ₂₃ H ₂₃ NO ₈	Peel	-	Deng et al., 2014 ⁴⁵	
20	6-Methoxy-9H-carbazole-3-carboxylic acid	C ₁₄ H ₁₁ NO ₃	Stem	Anticancer	Jiang et al., 2014 ⁹⁰	
21	Claulansine L	C ₂₃ H ₂₃ NO ₈				
22	Claulansine M	C ₁₈ H ₁₅ NO ₂				
23	Claulansine N	C ₁₄ H ₁₂ NO ₃				
24	Claulansine O	C ₁₅ H ₁₄ NO ₃	Stem	Hepatoprotective	Du, Liu, Li, Ma et al., 2015 ⁵	
25	Claulansine P	C ₂₁ H ₂₃ NO ₄				
26	Claulansine Q	C ₁₅ H ₁₅ NO				
27	Claulansine R	C ₁₆ H ₁₇ NO ₂				
28	Claulansine S	C ₁₇ H ₁₉ NO				
29	Claulansine T	C ₁₈ H ₂₁ NO ₂	Stem	Hepatoprotective	Du, Liu, Li, Yang et al., 2015 ⁴⁶	
30	Claulansium A	C ₁₉ H ₁₇ NO ₄				
31	Claulansium B	C ₁₉ H ₂₁ NO ₃	Branches	Anticancer	Peng et al., 2018 ⁹¹	
32	Clausenalansine A	C ₁₈ H ₁₆ NO ₃				
33	Clausenalansine B	C ₁₄ H ₁₄ NO ₂				
34	Clausenalansine C	C ₁₅ H ₁₂ NO ₄				
35	Clausenalansine D	C ₁₅ H ₁₄ NO ₃	Fruit	Neuroprotective	Liu, Guo, Liu et al., 2019 ⁴⁸	
36	Clausenalansine E	C ₁₄ H ₁₄ NO ₂				
37	Clausenalansine F	C ₁₅ H ₁₆ NO ₂				
38	Claulansine U	C ₁₉ H ₁₇ NO ₄		Against serum deprivation injury		
39	Claulansine V	C ₁₉ H ₁₅ NO ₄		Against serum deprivation injury		
40	Claulansine W	C ₁₉ H ₁₉ NO ₃	Stem	Anticancer	Sun et al., 2020 ⁴⁹	
41	(-)-(2'R)-Claulamine A	C ₁₉ H ₁₇ NO ₃		/		
42	Claulansine X	C ₁₉ H ₂₀ NO ₄	Stem	Neuroprotective	Sun et al., 2021 ⁵⁰	
Amine alkaloid in wampee						
No.	Name	Molecular formula	Source	Bioactivities	References	
1	Clausenamide	C ₁₈ H ₁₉ NO ₃	Leaves	Hepatoprotective	Yang, Chen et al., 1987 ⁹²	
2	Neoclausenamide	C ₁₈ H ₁₉ NO ₃				
3	Dehydroclocloclausenamide	C ₁₈ H ₁₇ NO ₂				
4	Cyclocloclausenamide	C ₁₈ H ₁₇ NO ₂	Leaves	Hepatoprotective	Yang et al., 1988 ⁹³	
5	(E)-N-(4-Methoxyphenethyl)-2-methylbut-2-enamide	C ₁₄ H ₁₉ NO ₂	Leaves	-	Zhao et al., 2011 ⁹⁴	
6	Lansiumamide B	C ₁₈ H ₁₇ NO	Leaves	Anti-inflammatory	Matsui et al., 2013 ⁵⁵	
7	Lansiumamide C	C ₁₇ H ₁₈ NO				
8	Lansamide I	C ₁₈ H ₁₈ NO				
9	SB-204900	C ₁₈ H ₁₇ NO ₂				
10	Clausenalansamides C	C ₁₉ H ₂₁ NO ₃ Na	Leaves	-	Shen et al., 2017 ⁵⁴	
11	Clausenalansamides D	C ₁₈ H ₁₈ ClNO ₂ Na				
12	Clausenalansamides E	C ₁₈ H ₁₈ ClNO ₂ Na				
13	Clausenalansamides F	C ₁₉ H ₂₁ SNO ₄ Na				
14	Clausenalansamides G	C ₂₃ H ₂₉ NO ₈ Na				
15	Lansiumamide A	C ₁₇ H ₁₅ NO	Seeds	-	Lin, 1989 ⁵⁶	

Table 2. continued

		Amine alkaloid in wampee			
No.	Name	Molecular formula	Source	Bioactivities	References
16	Lansiumamide B	C ₁₈ H ₁₇ NO			
17	Lansiumamide C	C ₁₈ H ₁₉ NO			
18	Lansumide I	C ₁₈ H ₁₇ NO			
19	Clausenalansamide A	C ₁₈ H ₁₉ NO ₃	Seeds	Nematicidal	Fan, Chen, Mei, Xu et al., 2018 ⁹⁵
20	Clausenalansamide B	C ₁₈ H ₁₉ NO ₂			
21	1'-Methoxyl-clausenalansamide B	C ₁₉ H ₂₁ NO ₃	Seeds	Nematicidal	Fan, Chen, Mei, Kong et al., 2018 ⁵⁷
22	3-Dehydroxy-3-methoxyl-secoclausenamamide	C ₁₈ H ₂₁ NO ₃			
23	2'-Dehydroxy-2'-acetoxyl-clausenalansamide B	C ₂₀ H ₂₁ NO ₃ Na			
24	Neoclausenamamide A	C ₂₀ H ₂₁ NO ₃ Na			
25	Neoclausenamamide B	C ₂₇ H ₂₅ NO ₄ Na			
26	Acetylclausenamamide	C ₂₀ H ₁₉ O ₃ N	Stem	Neuroprotective	Sun et al., 2021 ⁵⁰
27	Anhydroclausenamamide	C ₁₈ H ₁₈ O ₂ N			
28	1-Chloro-benzenepropanamide	C ₁₇ H ₁₉ O ₂ NCl			
29	Clauamide A	C ₁₆ H ₁₇ NO ₃	Stem	Hepatoprotective	Du, Liu, Li, Yang et al., 2015 ⁴⁶
30	(+)-(3S,4R,5S,6S)-Clauselansine C	C ₁₈ H ₁₇ NO ₂	Stem	Neuroprotective	Liu, Du et al., 2017 ⁶²

the ethanol extract of the wampee stem, exhibiting a moderate neuroprotective influence on the PC12 cell line.⁵⁰

In short, more than 40 carbazole alkaloids have been isolated and identified from different parts of the wampee and are summarized in Table 2. They are an essential and characteristic feature of a wampee.

Fan et al. recently reported that the pivotal regulatory enzymes, shikimate kinase and anthranilate synthase, upregulate carbazole alkaloid production in wampee through the carbazole alkaloid biosynthetic pathway.⁵¹ Alkaloids were discovered in different portions of the wampee, including the pulp, stem, branch, leaf, twig, root, and peel, and the stem contains the most abundant alkaloid profiles. Hence, the wampee stem can be considered an important source of carbazole alkaloids. Some of these carbazole alkaloids demonstrated antineoplastic, hepatoprotective, and neuroprotective properties in *in vitro* tests. In addition, Li et al. also suggested that carbazoles isolated from *Clausena* plants are a vital heterocyclic class of anticancer agents.⁵² However, further in-depth studies are needed to investigate the potential mechanisms of the bioactivities of these carbazole alkaloids to illustrate the health benefits of wampee.

2.7.2. Amine Alkaloid. Amide alkaloids are another important category of alkaloids abundant in various portions of the wampee. This group of compounds has attracted attention due to their excellent biological activities.

An amide alkaloid named clausenamamide was first isolated and characterized from wampee leaves.⁵³ In a later study, a greater variety of amide alkaloids were found in the leaves of wampee, such as clausenalansamides C (in Figure 2B) through G and clausenaline G,⁵⁴ lansiumamide C, lansumide I, lansiumamide B, and SB-204900.⁵⁵ Additionally, the wampee seed is another important source of amide alkaloids. Lin identified four cinnamamide derivatives in wampee seeds: lansiumamide A, lansumide-I, lansiumamide B, and lansiumamide C.⁵⁶ Later, Fan, Chen, Mei, and Kong et al. isolated amide alkaloids from wampee seeds, including 1'-methoxyl-clausenalansamide B, 3-dehydroxy-3-methoxyl-secoclausenamamide, 2'-dehydroxy-2'-acetoxyl-clausenalansamide B, neoclausenamamide-A, clausenalansamide A, clausenalansamide B, and neoclausenamamide-B, and confirmed their nematicidal activity.⁵⁷ Among the amide alkaloids derived from wampee seeds, lansiumamide B exhibits the broadest biological activities. These include an antiobesity

effect,⁵⁸ larvicidal activity,⁵⁹ antibacterial activity,⁶⁰ and fungicidal activity.⁶¹

Furthermore, continued research into the bioactive compounds in the wampee plant revealed the discovery of new amide alkaloids in the stem of the wampee. Du et al. reported clauamide A, isolated from wampee seeds, has potential hepatoprotective capacity.⁴⁶ Liu et al. extracted (+)-(3S,4R,5S,6S)-clauselansine C from wampee seeds and confirmed its neuroprotective activity.⁶² Recently, three more new amide alkaloids were found in wampee seeds, including acetylclausenamamide, anhydroclausenamamide, and 1-chloro-benzenepropanamide.⁵⁰

At least 30 amide alkaloids were extracted and characterized from different portions of the wampee. The amide alkaloids isolated from wampee were summarized in Table 2. The biological activities of amide alkaloids have been extensively investigated, revealing an array of potential therapeutic applications such as neuroprotective, anti-inflammatory, nematicidal, hepatoprotective, antidepressant, and antinociceptive effects. Hence, analyzing the amide alkaloid composition of wampee could potentially offer novel perspectives on its utilization, including the development of health-enhancing products. Also, developing the byproducts of wampee, like seeds, peels, leaves, and twigs, is a sustainable strategy that promotes the valorization of low-value products in the food industry.

3. HEALTH-PROMOTING EFFECTS OF WAMPEE

As mentioned in the previous section, wampee is a rich source of several bioactive compounds. The health-promoting effects of wampee reported in previous research include antioxidative, anti-inflammatory, hepatoprotective, antibacterial, and antineoplastic properties, etc. Most of the work was conducted using the *in vitro* model. However, these outcomes obtained from *in vitro* studies provide fundamental evidence for in-depth animal and clinical research.

3.1. Antioxidative Effects. Excessive production of reactive oxygen species (ROS) can result in an imbalanced redox reaction in tissues, leading to oxidative stress or even oxidative damage in the body.⁶³ ROS are produced both endogenously, as metabolic byproducts stemming from cellular oxygen metabolism within the organism, and exogenously, originating from environmental factors including ultraviolet

Table 3. Antioxidative Activity of Wampee^a

Portion Bioactive compound	Peel		Pulp	
	Phenolics		Phenolics	
	Ethyl acetate		Acetone	
Solvent	Ethyl acetate		Acetone	
TPC	330 ± 9.9 μg/g dry weight, expressed as gallic acid equivalents		7.92 mg/g GAE FW	9.07 mg/g GAE FW
TFC			7.16 mg/g CE FW	3.50 mg/g CE FW
DPPH	50 μg/mL (95% scavenging activity)		76.72 mg AsA equiv/100 g FW	208.1 mg AsA equiv/100 g FW
FRAP			264.3 mmol Fe ²⁺ /100 g FW	279.7 mmol Fe ²⁺ /100 g FW
ABTS				
Superoxide anion radical scavenging activity	50 μg/mL (88% scavenging activity)			
PSC			34.7 μmol Vit. C equiv/100 g FW	4927 μmol AsA equiv per 100 g FW
ORAC			5.44 mmol TE/100 g FW	11.03 mmol TE/100 g FW
Cell antioxidant <i>in vivo</i>				
Major finding	Ethyl acetate extract of wampee peel exhibits superior antioxidative capacity, which is mainly related to phenolic content.		Antioxidant capacity of wampee fruit is related to the total phenolic and total flavonoid contents.	It was apparent that wampee fruits had higher phenolic and flavonoid contents resulting in better antioxidant activities <i>in vitro</i> and cellular.
Reference	Prasad et al., 2010 ¹		Chang, Ye et al., 2018 ³⁴	Ye et al., 2019 ³²

^a“TPC”: Total phenolic content; “TFC”: Total flavonoid content; “DPPH”: 1,1-diphenyl-2-picrylhydrazyl; “FRAP”: Ferric reducing antioxidant power; “ABTS”: 2,2'-Azinobis(3-ethylbenzthiazoline-6-sulfonate); “PSC”: Peroxide radical scavenging capacity; “ORAC”: Oxygen radical absorbance capacity.

radiation, ionizing radiation, pollutants, heavy metals, and xenobiotics.⁶⁴ The most common ROS include hydroxyl radicals, hydrogen peroxide, and superoxide anions. Excessive ROS may cause irreversible cellular damage by inducing DNA/RNA damage, protein oxidation, and lipid peroxidation of polyunsaturated fatty acids. Consequently, increased oxidative stress heightens the risk of chronic diseases such as cancer, neurodegenerative disorders, and diabetes.⁶⁵ Hence, the dietary intake of antioxidants could be a feasible strategy to neutralize ROS and reduce the risk of related health problems.

To evaluate the antioxidative properties of wampee, a range of *in vitro* assays were used, including 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonate) (ABTS), oxygen radical absorbance capacity (ORAC), ferric reducing antioxidant power (FRAP), 2,2-diphenyl-1-picrylhydrazyl (DPPH), Trolox equivalent antioxidant capacity (TEAC), peroxide radical scavenging capacity (PSC), cellular antioxidant activity (CAA), self-oxidation of 1,2,3-phentriol, and lipid peroxidation assays.

The antioxidant capacity of wampee pulp can be primarily attributed to its rich phenolic compound content. Ye et al.³² examined the correlation between total phenolic content and antioxidant properties across five wampee pulp varieties. The total phenolic content was positively associated with both *in vitro* and *in vivo* antioxidant assays. The variety with the highest total phenolic content (907.8 mg of GAE/100 g) displayed the most potent antioxidant capacity in the ORAC, DPPH, and FRAP assays, with values of 11.03 mmol of Trolox equivalent/100 g, 208.1 mg ascorbic acid equivalent/100 g, and 279.7 mmol Fe²⁺/100 g, respectively. On the contrary, the variety with the lowest level of total phenolics (49.25 mg GAE/100 g) exhibited the weakest antioxidant capacity, with corresponding assay values of 1.08 mmol Trolox equivalent/100 g, 10.55 mg ascorbic acid equivalent/100 g, and 72.84 mmol Fe²⁺/100 g, respectively. Other research has similarly observed that wampee pulp with higher phenolic content displays enhanced free radical scavenging capabilities.^{34,40} In addition, Ye et al. analyzed the association between phenolic monomers and

antioxidant properties, finding that *in vitro* antioxidant capacity was primarily attributable to phenolic monomers such as ferulic acid, catechin, and syringic acid, while hesperetin contributed most to the *in vivo* antioxidant capacity of wampee pulp.³² Moreover, Zeng et al. reported that the ethanol extract of wampee fruit inhibited the expression of NF-κB, thereby protecting PC-12 cells from oxidative stress.⁶⁶

Wampee leaves exhibit remarkable antioxidant potential due to their rich phenolic composition. Chang et al. observed that wampee leaf buds with the highest total phenolic content (2046 mg GAE/100 g) yielded the highest ORAC value (415.8 μmol Trolox equivalent/g).⁶⁷ As the leaves developed, the total phenolic content consistently decreased, causing an initial reduction in antioxidant potential, followed by a subsequent increase. Interestingly, the total flavonoid content displayed an upward trend during leaf development. This observation can be attributed to the fact that despite the decline in total phenolic content affecting the antioxidant potential of wampee leaves the simultaneous production of flavonoid compounds may compensate for and sustain their antioxidant capacity. Thus, flavonoids represent a significant subgroup of total phenolics that may considerably contribute to the antioxidant properties of wampee leaves.

Prasad et al. examined the antioxidative properties of wampee peel extracts using four solvents: ethanol, hexane, ethyl acetate, butanol, and water.³³ The ethyl acetate extract demonstrated significant antioxidative potential, surpassing that of butylated hydroxytoluene (BHT) in *in vitro* tests. In the DPPH assay, antioxidative capacities were ranked as follows at a concentration of 50 μg/mL: ethyl acetate > BHT > water > ethanol > butanol > hexane. This outcome is likely due to the total phenolic content in the different fractions, with ethyl acetate, water, ethanol, butanol, and hexane fractions containing 330.0, 54.0, 46.2, 30.3, and 7.9 μg/g DW, respectively.³³ A similar study also reported higher total phenolic content in the ethyl acetate fraction.¹

Ethyl acetate is a polar solvent, whereas hexane is typically used to extract nonpolar substances, indicating that the

phenolic compounds in the wampee peel may possess high polarity. Moreover, the total phenolic content in the water fraction was significantly lower than that in the ethyl acetate fraction, suggesting that most antioxidative compounds in the wampee peel were not water-soluble. Thus, an appropriate solvent with both polar and hydrophobic properties can enhance the efficacy of isolating the predominant antioxidant constituents from the wampee peel.

In addition to phenolic substances, other compounds, such as polysaccharides extracted from wampee, have been identified as potential antioxidants.²⁰ The antioxidative capacity of acidic heteropolysaccharide extracted from wampee seed exhibited a concentration-dependent manner in both *in vitro* and *in vivo* tests. Polysaccharide extracts significantly reduced the malondialdehyde (MDA) level while increasing the SOD and glutathione (GSH) levels in high-fat diet-fed rats, demonstrating their potential ability to manage oxidative stress *in vivo*.²¹ However, different fractions of polysaccharide extracts might exhibit varying antioxidative properties. Peng et al.²⁰ attributed the variation to the phenols attached to polysaccharides, while Wu et al.²¹ ascribed it to differences in uronic acid content and structural characteristics.

Overall, various portions of the wampee exhibit significant antioxidant properties. Most research has primarily focused on the *in vitro* antioxidant capacity of wampee. Nonetheless, it is crucial to acknowledge that outcomes derived from *in vitro* tests may not simply be extrapolated to *in vivo* circumstances. The chemical and molecular targets of *in vitro* tests, such as ABTS and DPPH, are sterically hindered stable radicals that do not accurately represent short-lived radicals in the body.⁶⁸

In summary, phenolic compounds and polysaccharides in wampee are the primary constituents contributing to its antioxidative activity. Table 3 summarizes the antioxidant activity of wampee. Further comprehensive *in vivo* antioxidant tests are necessary to provide more evidence of the antioxidant properties of the wampee and determine the specific mechanisms.

3.2. Neuroprotection. Neurodegenerative diseases comprise a group of progressive disorders that impact neurons in the nervous system, leading to a wide array of symptoms, including cognitive dysfunction, impaired movement, memory loss, and, ultimately, death. Some of the most prevalent neurodegenerative diseases include Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), and amyotrophic lateral sclerosis (ALS).⁶⁹ The pathogenesis of these diseases, however, remains incompletely understood. Presently, there is a burgeoning interest in exploring natural products for their potential to target the hypothesized pathogenesis of neurodegenerative diseases. Adrenal pheochromocytoma (PC12) cells and human neuroblastoma SH-SY5Y cells are two widely used cell lines for establishing neuron injury models and investigating neuroprotective compounds.^{70,71} Prior studies have demonstrated that numerous bioactive compounds derived from wampee exhibit promising potential in protecting neuronal cells and may offer new insights into innovative treatments for neurodegenerative diseases.

Alkaloids comprise a large family of compounds with potential neuroprotective activity isolated from wampee. (–)-Clausenamide was one of the first compounds discovered with potential neuroprotective activity derived from wampee. Hu et al. demonstrated that (–)-clausenamide (1–100 μ M) significantly alleviates A β -induced neurotoxicity in differ-

entiated PC12 cells in a dose-dependent manner, as indicated by cell viability assays.⁷² This compound inhibited the production of reactive oxygen species (ROS) and the activation of caspase-3, which serve as markers for oxidative stress and apoptosis, respectively. Additionally, (–)-clausenamide activates the Nrf2/HO-1 antioxidant pathways, which play a crucial role in regulating oxidative stress and inflammation. Consequently, (–)-clausenamide may have potential as a natural neuroprotective agent for preventing and treating neurodegenerative diseases such as Alzheimer's disease. However, further research, including *in vivo* studies, is required to fully understand the mechanisms of action and potential therapeutic applications of this compound. Liu et al. developed a neuron injury model using a PC12 cell line and sodium nitroprusside (SNP) as an inducer.⁴⁴ They discovered that four alkaloids extracted from the wampee stem significantly increased cell viability against SNP-induced neurotoxicity. In a subsequent study, Liu et al. investigated the neuroprotective effects of alkaloids derived from the wampee stem.⁶² They observed that 5 out of 14 alkaloids moderately alleviate neurotoxicity induced by okadaic acid in PC12 cells. A total of 13 alkaloids were identified, while 5 of them could moderately alleviate the neurotoxicity induced by okadaic acid in PC12 cells.

Liu, Guo, and Liu et al. discovered that 16 carbazole alkaloids extracted from wampee pulp demonstrated significant protective capabilities against 6-OHDA-induced apoptosis in human neuroblastoma SH-SY5Y cells, similar to the positive control, curcumin.⁴⁸ Subsequently, Liu et al. isolated 12 geranylated carbazole alkaloids from the leaves and stems of the wampee, which exhibited similar protective activity with curcumin against 6-OHDA-induced neurotoxicity in SH-SY5Y cells.⁷³ More recently, Sun et al. reported that two carbazole alkaloids isolated from wampee stems showed only moderate protective effects on PC12 cells against serum deprivation injury.⁴⁹ Collectively, these studies underscore the neuroprotective potential of alkaloids derived from wampee, suggesting possible therapeutic applications for neurodegenerative diseases. However, as most of these investigations were preliminary, the mechanisms underlying the compounds' neuroprotective effects remain unclear, necessitating further research.

In addition to alkaloids, phenolic compounds have also demonstrated potential as neuroprotective agents. Li et al. extracted a flavonoid, Bu-7, from wampee leaves, which significantly enhanced the viability of PC12 cells following rotenone-induced injury.⁷⁴ Bu-7 inhibited the JNK and p38 signaling pathways via phosphorylation, consequently preventing rotenone-induced apoptosis in PC12 cells. Moreover, Bu-7 treatment attenuated mitochondrial membrane permeabilization and the collapse of the mitochondrial membrane potential (MMP), indicating the significant neuroprotective potential of Bu-7 and its possible application in treating Parkinson's disease. Later, Zeng et al. investigated the activity of wampee fruit ethanol extract in a hydrogen-peroxide-induced PC12 cell model.⁶⁶ The wampee fruit extracts significantly reversed cell apoptosis in PC12 cells in a dose-dependent manner, reduced intracellular ROS generation, and prevented MMP collapse. Furthermore, the extract treatment inhibited the NF- κ B pathway and downregulated caspase-3 expression, suggesting that the intracellular antioxidative capacity of wampee contributes to its protective effects against neurodegenerative diseases.

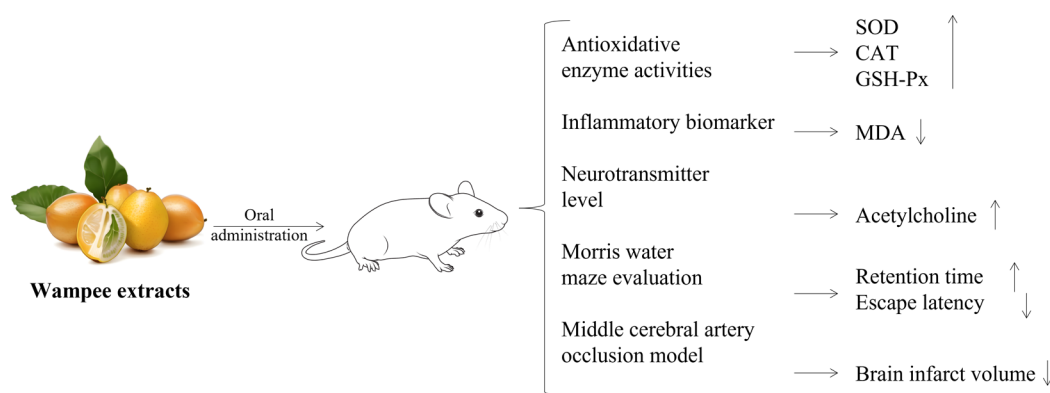


Figure 3. Experimental design of wampee neuroprotection effect and test index.

Further investigation of the neuroprotective effects of wampee using *in vivo* models is necessary. Tongun and Phachonpai developed a memory impairment model in rats induced by chronic restraint stress.⁷⁵ In the object recognition assessment, rats administered wampee peel extract (600 mg/kg) exhibited a higher discrimination index, while in the Morris water maze evaluation, they exhibited reduced escape latency and increased retention duration. The activity of the antioxidant enzyme in rat brain tissue, such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GSH-Px), was enhanced after wampee peel extract supplementation. The treatment group exhibited lower acetylcholinesterase (AChE) activity, indicating a higher essential neurotransmitter (ACh) level. Phachonpai and Tongun⁷⁶ reported similar findings and suggested that phenolic compounds might be the primary bioactive substances responsible for the antidementia activity of the wampee. In another study, Phachonpai and Tongun⁷⁷ induced focal cerebral ischemia in rats through middle cerebral artery occlusion (MCAO). Rats supplemented with the wampee peel extract displayed higher cognition test scores than the MCAO control group. Additionally, wampee peel extract treatment reduced inflammatory biomarkers and improved neuronal survival and cholinergic neurons in the hippocampal regions. The specific experiment design for the wampee neuroprotection effect was shown in Figure 3.

Both the *in vitro* and *in vivo* studies presented above demonstrate the considerable potential of wampee for addressing neurological issues. Alkaloids and phenolic compounds appear to be the primary categories of bioactive substances in wampee that contribute to its neuroprotective properties. However, further research is needed to elucidate the precise mechanisms of action of these bioactive compounds. Table 4 provides a summary of the neuroprotective effects of wampee.

3.3. Anti-Inflammation Effects. Inflammation is a multifaceted physiological response to tissue injury, infection, and other forms of cellular stress. Acute inflammation is an essential defense mechanism protecting the body from harmful stimuli. However, chronic inflammation contributes to the pathogenesis of numerous diseases such as arthritis, cardiovascular disease, and cancer. This persistent inflammatory state is characterized by the continuous activation of immune cells, tissues, and signaling pathways, leading to the production of pro-inflammatory cytokines, chemokines, and other mediators that may harm healthy tissues and exacerbate disease.⁷⁸

Wampee contains various bioactive compounds known for their anti-inflammatory properties. Inflammation can be induced by carrageenin, leading to the formation of edema in rat paws. A 200 mg/kg methanolic extract of wampee leaves and indomethacin treatments reduced rat paw edema by 55.5% and 64.0% within four hours, respectively, suggesting the potential of wampee extract as an anti-inflammatory agent.⁷ Similarly, Huang et al. discovered that wampee leaf extract diminished TNF- α production by suppressing the TLR4/MYD88/TRAF6 signaling pathways in LPS-treated RAW 264.7 cells.⁷⁹ Matsui et al. identified two cinnamamides, lansiumamide B and SB-204900, that inhibited histamine release and decreased IL-6, COX-2, and TNF- α formation, thus exhibiting anti-inflammatory activity in rat basophilic leukemia cells.⁵⁵ The mechanism of the wampee anti-inflammation effect is shown in Figure 4. Wampee stem-derived alkaloids demonstrated anti-inflammatory potential comparable to curcumin in LPS-induced murine microglial BV2 cells.⁸⁰ The alkaloid 3-formyl-6-methoxycarbazole, isolated from wampee roots, showed anti-inflammatory activity similar to dexamethasone, significantly suppressing TNF- α at a concentration of 50 $\mu\text{g/mL}$ in LPS-induced monocyte cells.⁸¹ Shen et al. reported that alkaloids from the wampee stem not only inhibited TNF- α and NO production but also suppressed fMLP/CB-induced superoxide anion generation in LPS-induced RAW 264.7 cells.⁸² Likewise, coumarin and alkaloid compounds from wampee leaves, such as wampetin and imperatorin, effectively reduced superoxide anion generation in RAW 264.7 cells with IC₅₀ values of 6.8 and 1.7 μM , respectively.⁵⁴

In summary, both *in vivo* and *in vitro* studies have demonstrated the potent anti-inflammatory properties of wampee. It has been posited that the notable anti-inflammatory compound present in the wampee could be attributed to alkaloids, which exhibit their effects by inhibiting inflammatory signaling cascades and reducing the production of reactive species. Table 5 summarizes the anti-inflammation effect of wampee.

3.4. Hepatoprotection. In recent years, increasing interest has emerged in exploring the potential hepatoprotective activity of the wampee and its constituents. Du et al. isolated several novel carbazole alkaloids from the wampee stem and assessed their hepatoprotective activity against *N*-acetyl-*p*-aminophenol (APAP)-induced toxicity in HepG2 cells, a human hepatocellular carcinoma cell line.⁴⁷ The study discovered that five of these alkaloids demonstrated hepatoprotective activity comparable to bicyclol, a drug with

Table 4. Neuroprotective Effects of Wampee

No.	Sources	Model	Study design	Result	References
1	(-)-Clausenamide isolated from wampee stem	A β 25–35-induced cell apoptosis in PC12 cells	Cell viability, Western blot, MMP, flow cytometry	(-)-Clausenamide (10 μ M) reduces the collapse of MMP by 42.4%. (-)-Clausenamide (10 μ M) treatment increased the viability of PC12 to 90% from 70% (-)-Clausenamide inactivated the p38 MAPK pathway and inhibited the expression of P53 and caspase 3 The cell viability of negative control group was 76%, while the treatment of four alkaloids (10 μ M) with neuroprotective potential restored the value to 91.0, 88.2, 84.8, and 83.7%, respectively. The cell viability of negative control group was 70.5%, while the treatment of five alkaloids (10 μ M) with neuroprotective potential restored the value to 91.2, 89.7, 83.5, 83.4, and 83.3%, respectively. The EC50 value of 16 carbazole alkaloids ranged from 0.36 to 10.69 μ M, while the EC50 value for curcumin (positive control) was 5.82 μ M.	Hu et al., 2010 ⁷² Liu et al., 2012 ⁴⁴ Liu, Du et al., 2017 ⁶⁵ Liu, Guo, Liu et al., 2019 ⁴⁸
2	Alkaloids isolated from wampee stem	SNP-induced toxicity in PC12 cells	Cell viability		Liu et al., 2012 ⁴⁴
3	Alkaloids isolated from wampee stem	Okadaic acid-induced injury in PC12 cells	Cell viability		Liu, Du et al., 2017 ⁶⁵
4	Alkaloids isolated from wampee pulp	6-Hydroxydopamine-induced cell apoptosis in human neuroblastoma SH-SY5Y cells	Cell viability		Liu, Guo, Liu et al., 2019 ⁴⁸
5	Alkaloids isolated from wampee stem and leaves	6-Hydroxydopamine-induced cell apoptosis in human neuroblastoma SH-SY5Y cells	Cell viability		Liu, Guo, Wang et al., 2019 ⁷³
6	Alkaloids isolated from wampee stem	Serum deprivation injury in PC12 cells	Cell viability		Sun et al., 2020 ⁴⁹
7	Flavonoid isolated from wampee leaves	Rotenone-induced injury in PC12 cells	Cell viability, Western blot, flow cytometry, measurement of mitochondrial membrane potential	The cell viabilities of negative control, positive control, and alkaloid treatment groups (10 μ M) were 47.4, 83.8, 67.8, and 63.3%, respectively The cell viability of cells treated with rotenone was 69.2%, and this increased significantly by 14.4%, 3.1%, and 18.1% after treated with 0.1, 1, and 10 μ M Bu-7, respectively, Bu-7 attenuated the rotenone-induced mitochondrial potential reduction in cells. Bu-7 inhibited activation of the JNK and p38 signaling pathways and suppressed caspase 3 activity in the rotenone-treated cells. Bu-7 exerted its activities in a bell-shaped dose–response relationship. The cell viability of negative control group was 55%, but the wampee fruit extract (1, 10, 20 μ M) increased this value to 64.12, 72.13, and 76.26%, respectively.	Li et al., 2011 ⁷⁴ Zeng et al., 2014 ⁶⁶
8	Ethanol extract of wampee pulp	H ₂ O ₂ -induced neurotoxicity in PC12 cells	Cell viability, Western blot, measurement of mitochondrial membrane potential		Wampee pulp extract downregulated caspase-3 and pnf- κ B p65 and suppressed NF- κ B p65 entry to the nucleus from the cytoplasm.
9	Ethanol extract of wampee peel	Memory impairment induced by chronic restraint stress in rats	Object recognition test, Morris water maze test, ache activity lipid peroxidation, SOD activity determination	High dose of wampee peel extract (600 mg/kg) significantly increased the discrimination index of rats. The rats received wampee peel extract treatment displayed shorter time of escape of latency and longer retention time. Wampee peel extract decreased the lipid peroxidation index. It improved the antioxidant enzymes activities while inhibiting pain. Medium dose of wampee peel extract treatment (400 mg/kg) significantly reduced the brain infarct volume of rats; decreased the MDA level in the brain; and enhanced the neuronal survival and cholinergic neurons in hippocampal regions.	Tongun & Phachonpai, 2020 ⁷⁵ Phachonpai & Tongun, 2021b ⁷⁷
10	Ethanol extract of wampee peel	Rats received permanent right middle cerebral artery occlusion	Morris water maze test, passive avoidance test, determination of cerebral infarct volume, Nissl staining and survival neuron densities counts, determination of the cholinergic neuron densities, measurement of lipid peroxidation	High dose of wampee peel extract treatment (600 mg/kg) increased the discrimination index of rats. The rats that received wampee peel extract treatment displayed shorter time of escape of latency and longer retention time. Wampee peel extract alleviated oxidative stress. It improved the antioxidant enzyme activities while inhibiting pain.	Phachonpai & Tongun, 2021a ⁶⁶
11	Ethanol extract of wampee peel	Memory impairment induced by chronic restraint stress in rats	Object recognition test, Morris water maze test, superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GSH-Px), and ache activities and the malondialdehyde (MDA) determination		Phachonpai & Tongun, 2021a ⁶⁶

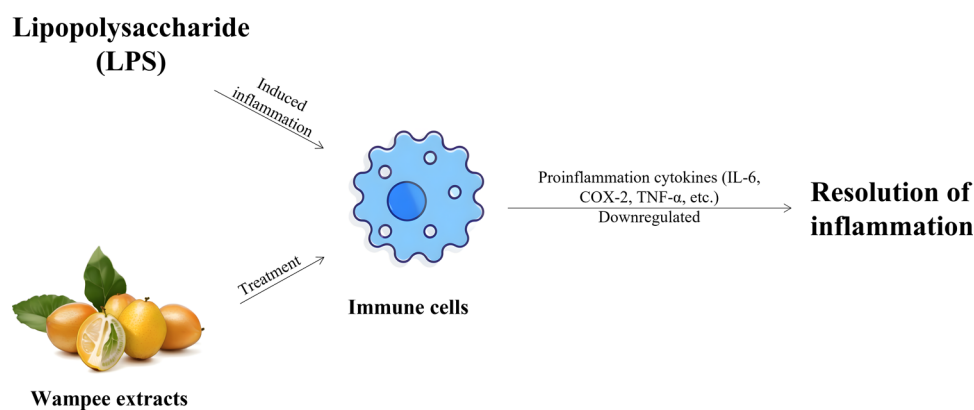


Figure 4. Mechanism of wampee anti-inflammation effect.

established hepatoprotective effects. Similarly, Liu et al. identified a monoterpene from the stem of the wampee that exhibited similar activity to bicyclol in protecting HepG2 cells from APAP-induced toxicity.⁸³

The hepatoprotective properties of wamees have been investigated in animal models. Carbon tetrachloride (CCl_4) is a widely used agent to induce liver injury in mice by generating the trichloromethyl-oxygen radical through hepatic cytochrome P450 activation, leading to lipid peroxidation. Liu et al. demonstrated that administering eight alkaloids isolated from wampee leaves at a dosage of 250 mg/kg significantly decreased serum alanine transaminase (ALT) levels in CCl_4 -intoxicated mice.⁸⁴ Moreover, the two alkaloids protected against acetaminophen- and thioacetamide-induced liver damage. It is hypothesized that these alkaloids might exhibit hepatoprotective effects by inhibiting the interaction between CCl_4 and hepatic cytochrome P450, consequently diminishing the production of subsequent free radicals.

Additionally, carbon tetrachloride (CCl_4)-induced liver injury results in the rapid release of alanine transaminase (ALT), aspartate transaminase (AST), and alkaline phosphatase (ALP) into the bloodstream. Elevated serum levels of these enzymes were indicative of liver damage. Adebajo et al. discovered that administering a 200 mg/kg dosage of methanolic wampee leaf extract improved the activities of ALT, AST, and ALP in the livers of CCl_4 -intoxicated mice while simultaneously downregulating these enzymes' activity in plasma.⁷ The mechanism of the wampee hepatoprotective effect is also illustrated in Figure 5. Consequently, these findings indicate that wampee extract could potentially mitigate hepatocellular damage induced by CCl_4 *in vivo*. Table 5 summarizes the hepatoprotective effect of wampee.

3.5. Antidiabetic Effects. Numerous extracts from wampee have demonstrated antidiabetic properties. Adebajo et al.⁷ discovered that wampee stem extract effectively reduced glucose levels in rats. This extract treatment increased insulin secretion in both *in vivo* and *in vitro* experiments. Additionally, the wampee peel extract (3.4 mL/kg/day) significantly mitigated the physiological abnormalities associated with diabetes, while decreasing swelling and lipid accumulation in rat organs. Furthermore, an 8-week oral administration of lansiumamide B (20 mg/kg), an alkaloid isolated from the seeds of wampee, significantly reduced the body weight of HFD-fed mice by diminishing the visceral and subcutaneous fat tissues. The serum cholesterol, high-density lipoprotein (HDL), and low-density lipoprotein (LDL) levels in high-fat

diet fed mice were reduced by 30.4%, 22.1%, and 47.7%, respectively.⁵⁸

Kong et al. investigated the impact of polyphenol extracts from the wampee leaf on streptozotocin (STZ)-induced type 2 diabetic rats.³⁶ The extract exhibited potent antihyperlipidemic properties, which effectively reduced the concentrations of total cholesterol (TC), triglycerides (TG), low-density lipoprotein cholesterol (LDL-C), and high-density lipoprotein cholesterol (HDL-C) within a four-week duration. The test index was illustrated in Figure 6. The bioactive constituents in wampee peel extract, including rutin (in Figure 2C), gallic acid (in Figure 2D), ferulic acid, chlorogenic acid, and caffeic acid, were identified to exhibit potential antidiabetic properties.³⁶

α -Glucosidase is an enzyme responsible for breaking down complex carbohydrates into simple sugars in the small intestine. This process is crucial for the absorption of carbohydrates into the bloodstream. However, in individuals with diabetes mellitus, α -glucosidase activity can result in elevated blood sugar levels after a meal, a phenomenon termed postprandial hyperglycemia. Condensed tannins and proanthocyanidins isolated from wampee have demonstrated inhibitory effects against α -glucosidase through competitive inhibition and conformational alteration in a dose-dependent manner.⁸⁵ Moreover, Shen et al. found that a high-fat diet altered gut microbiota composition in mice and increased the expression of inflammatory cytokines, such as tumor necrosis factor- α (TNF- α), while oral administration of wampee extract could reverse these changes.⁸⁶ Consequently, wampee may attenuate hyperglycemia and hyperlipidemia by inhibiting α -glucosidase and positively modulating the abundance of gut microbiota associated with metabolic parameters, thereby exhibiting antidiabetic activity.

These findings suggest the potential of wampee in preventing diabetes and provide scientific evidence for developing wampee-based functional food. Table 5 summarizes the antidiabetic activities of the wampee.

3.6. Antimicrobial Effects. Wampee has potential antimicrobial activity. Both He et al.⁸⁷ and Ma et al.²⁶ found that the essential oils (EOs) of *Clausena lansium* played an important role in inhibiting the growth of five *Candida* species. The EOs, extracted from chicken heart wampee (CHW) pericarp and abundant in β -phellandrene and β -sesquiphellandrene, showed the most significant antimicrobial efficacy. For example, He et al.⁸⁷ reported a 23.1 mm inhibition zone diameter for β -phellandrene, while Ma et al.²⁶ observed comparable outcomes. Following β -phellandrene treatment, the inhibition zone diameters for the five *Candida* species

Table S. Health Benefit Effects of Wampee

Health effect	Sources	Cell model	Result	Reference
Anti-inflammation effect	Alkaloids isolated from wampee stem	Murine microglial BV2 cells	Carbazole alkaloids displayed considerable anti-inflammatory potential against LPS-stimulated NO production in cell model, with IC ₅₀ values of around 5 μM	Liu et al., 2015 ⁸⁰
	Alkaloids and coumarins isolated from wampee stem	Human neutrophils and RAW264.7 cell lines	Imperatorin, isochlorogenic acid, and osthole were the most potent inhibitors for fMLP/CB-induced superoxide anion generation with IC ₅₀ values of 0.20, 0.81, and 0.0086 μM, respectively. The extracted alkaloid, such as wampetin, decreases up to 98.9% NO of control which is higher than positive control (indomethacin, 30.9%).	Shen et al., 2012 ⁸²
Anti-inflammation effect	Alkaloids and coumarins isolated from wampee leaves	Human neutrophils	Wampetin also downregulated the expression of TNF-α. Imperatorin and wampetin exhibited potent inhibitory ability toward fMLP/CB-induced superoxide anion generation with IC ₅₀ values of 1.7 and 6.8 μM, respectively.	Shen et al., 2017 ⁵⁴
	Alkaloid isolated from wampee root	RAW264.7 cell lines	Wampee extract inactivated TNF-α in a dose-dependent manner, through suppressing TLR4/MyD88/TRAF6 pathways, while up to 100 μg/mL wampee extract inhibited 80.6% TNF-α expression.	Huang et al., 2019 ⁷⁹
Hepatoprotective	Methanol extract of wampee stem	Monocyte cell line U937, human gingival fibroblast (HGF) cell line	3-Formyl-6-methoxycarbazole (50 μg/mL) displayed comparable inhibitory ability to dexamethasone (positive control) in suppressing TNF-α.	Rodanant et al., 2015 ⁸¹
	Methanol extract of wampee stem	Rats fed with high-fat diet	100 mg/kg wampee extract had comparable inflammatory effect to indomethacin that significantly alleviated the rat paw edema induced by inflammation.	Adebajo et al., 2009 ⁷
Antidiabetes	Methanol extract of wampee stem	Rats fed with high-fat diet	The low dose (100 mg/kg) and high dose (200 mg/kg) could decrease sleep duration time 5.3% and 8.4%, respectively. The methanol extract increased AST, ALT and ALP activity in serum and decreased these enzymes' activity in plasma.	Adebajo et al., 2009 ⁷
	Leaves of wampee including eight compounds	CCl ₄ -induced hepatotoxicity in mice	These eight compounds (250 mg/kg) could decrease the elevated SGPT of CCl ₄ -intoxicated mice. The cyclo-clausenamide was the most potent one among these compounds. The inhibition rate reached 76.7% after the cyclo-clausenamide treatment.	Liu et al., 1996 ⁸⁴
Antimicrobial	Glucopyranoside isolated from wampee stem	HepG2 (human hepatocellular liver carcinoma cell line) cells	The HepG2 cell survival rate was increased around 10% after the extracted compound (10 μM) treatment.	Liu et al., 2017 ⁶²
	Polyphenol extract from wampee leaves	Streptozotocin-induced type 2 diabetic rats	Higher body weight. Lower food intake. Less excretion.	Kong et al., 2018 ³⁶
Antidiabetes	Methanol extract of wampee stem	Glucose-induced INS-1 cell line, rat	Lower level of fasting blood glucose. Lower organ coefficient. Less liver damage and lipid accumulation.	Adebajo et al., 2009 ⁷
	Proanthocyanidins isolated from wampee pulp	Chemical reaction model	The rat treated with methanol extract of wampee stem (100 mg/kg) showed 15.8% lower level of glucose than control group. The extract also increased the insulin secretion in both <i>in vitro</i> (38.5% higher than control) and <i>in vivo</i> (1.74 times over control) tests.	Adebajo et al., 2009 ⁷
Antimicrobial	Essential oils extracted from pericarps	<i>Candida</i> strains	Proanthocyanidins isolated from wampee (0.26 μg/mL) suppressed the enzyme activity to 50%.	Yang et al., 2020 ⁸³
	Essential oils and three main compounds extracted from seed	<i>Candida</i> strains	The essential oil extracted from chicken heart wampee which was rich in β-phellandrene and β-sesquiphellandrene had the most powerful effect. The inhibition zone diameter of <i>C. glabrata</i> was 23.1 mm after the essential oils (10 μL/disc) treatment.	He et al., 2019 ⁸⁷
Antimicrobial	Three carbazole alkaloids extracted from twigs roots	<i>Porphyromonas gingivalis</i>	The 4-terpineol was the most potent compound among the essential oils and its main compound. The inhibition zone diameters of five <i>Candida</i> species was 26.7–35.6 mm after the 4-terpineol (10 μL/disc) treatment.	Ma et al., 2021 ⁸⁶
			Inhibition zone diameters of <i>Porphyromonas gingivalis</i> was 19.06–20.94 mm after the lansine (50 μg/mL) treatment. The lansine was the most powerful compound among extracted carbazole alkaloids.	Rodanant et al., 2015 ⁸¹

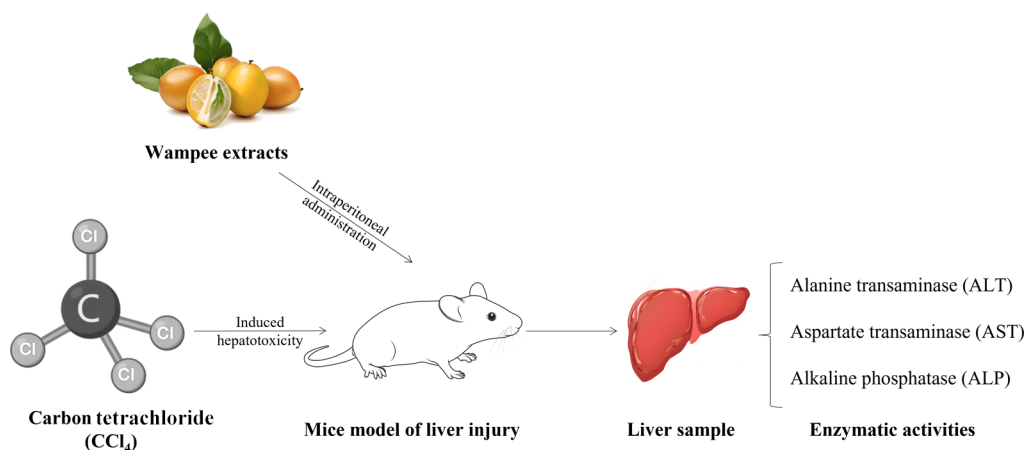


Figure 5. Mechanism of the wampee hepatoprotective effect.

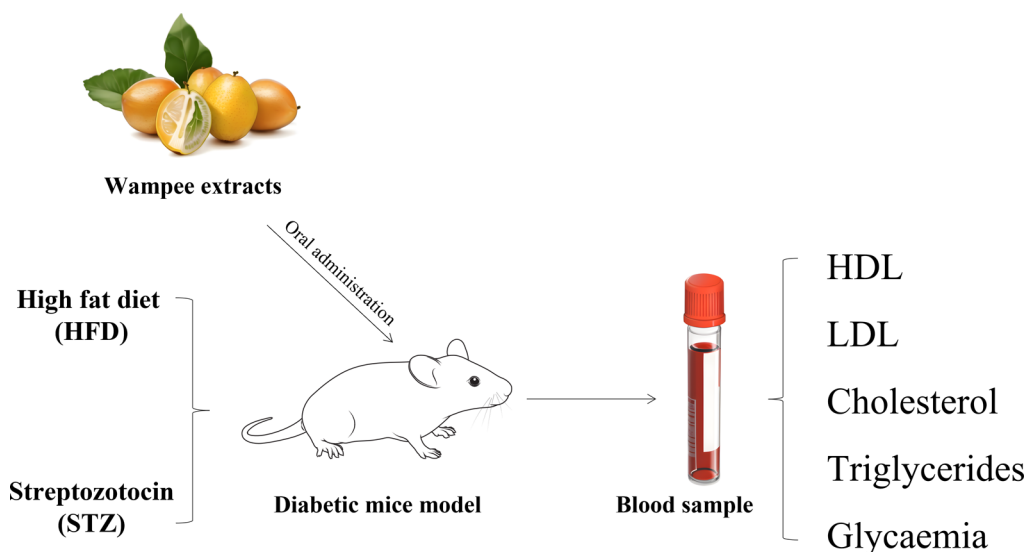


Figure 6. Wampee extract was used for diabetes treatment.

examined ranged from 15.6 mm to 22.1 mm. In addition to β -phellandrene, the principal constituents of the EOs, sabinene, and 4-terpineol, they also demonstrated inhibitory effects on the growth of the bacterial strain. The compound 4-terpineol demonstrated the most significant efficacy, with inhibition zone diameters ranging from 26.7 to 35.6 mm when tested against five distinct *Candida* species.²⁶ The extracts from the twigs and roots of *Clausena lansium* also showed antimicrobial activity. The *P. gingivalis* growth was inhibited after three carbazole alkaloid treatments. Among the investigated carbazole alkaloids, lansine exhibited the most potent efficacy, as evidenced by its inhibition zone diameters ranging from 19.06 to 20.94 mm.⁸¹ Based on the current studies, different bioactive compounds isolated from wampee exhibited potential antimicrobial activity.

Nonetheless, no research has been conducted to investigate the precise mechanisms underlying the effects of these compounds. Therefore, further investigation is required to understand the antimicrobial effect of wampee. These compounds could be extracted from wampee and made into an antimicrobial drug for further application. Table 5 summarized the antimicrobial effects of wampee.

4. CONCLUSIONS

The wampee fruit has been the subject of numerous studies due to its abundant nutrient content. This comprehensive review thoroughly studies the health implications and chemical constituents of wampee. In summary, wampee is an excellent source of dietary fiber with low-fat and low-calorie content and is enriched with significant levels of vitamin C, potassium, calcium, and nine essential amino acids. It is a good source of low-calorie food. Numerous bioactive constituents in wampee contribute to an array of health-promoting effects, such as antioxidative properties and neuroprotection and anticancer, anti-inflammatory, hepatoprotective, antidiabetic, and antimicrobial activities. The beneficial promoting effect and its corresponding bioactive substances have been studied and identified from wampee. However, the specific mechanism of these substances has not been explained clearly. The metabolic pathways and toxicology research of the bioactive substances extracted from wampee still require further study.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsomega.3c02759>.

Supplemental Tables 1–3 as mentioned in the text (PDF)

AUTHOR INFORMATION

Corresponding Author

Baojun Xu – Food Science and Technology Program,
Department of Life Sciences, BNU-HKBU United
International College, Zhuhai, Guangdong 519087, China;
orcid.org/0000-0003-0739-3735; Phone: +86-756-
3620636; Email: baojunxu@uic.edu.cn

Authors

Xin Huang – Food Science and Technology Program,
Department of Life Sciences, BNU-HKBU United
International College, Zhuhai, Guangdong 519087, China
Minghe Wang – Food Science and Technology Program,
Department of Life Sciences, BNU-HKBU United
International College, Zhuhai, Guangdong 519087, China
Saiyi Zhong – College of Food Science and Technology,
Guangdong Ocean University, Guangdong Provincial Science
and Technology Innovation Center for Subtropical Fruit and
Vegetable Processing, Zhanjiang 524088, China

Complete contact information is available at:
<https://pubs.acs.org/10.1021/acsomega.3c02759>

Author Contributions

[#]These authors contributed equally.

Funding

This study is supported by a research grant (project code: UICR0200007–23) from BNU-HKBU United International College.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We are grateful to all the members of the Food Science and Technology Program at BNU-HKBU United International College for their scientific assistance.

REFERENCES

- Prasad, K. N.; Xie, H.; Hao, J.; Yang, B.; Qiu, S.; Wei, X.; Chen, F.; Jiang, Y. Antioxidant and anticancer activities of 8-hydroxy-psoralen isolated from wampee [*Clausena lansium* (Lour.) Skeels] peel. *Food Chem.* **2010**, *118* (1), 62–66.
- Zhao, Z.; Gao, A.; Huang, J.; Luo, R.; Yu, K. Screening of sweet wampee [*Clausena lansium* (Lour.) Skeels] progenies in the early growth stage based on chloroplast genome analysis. *Genet. Resour. Crop Evol.* **2021**, *68*, 1747–1750.
- Rodrigues, S.; de Brito, E. S.; de Oliveira Silva, E. Wampee—*Clausena lansium*. In *Exotic Fruits*; Rodrigues, S., de Oliveira Silva, E., de Brito, E. S., Eds.; Academic Press: 2018; pp 439–441.
- Lim, T., *Clausena lansium*. In *Edible Medicinal and Non-medicinal Plants*; Springer, 2012; pp 871–883.
- Du, Y.; Liu, H.; Li, C.; Ma, J.; Zhang, D.; Li, L.; Sun, H.; Bao, X.; Zhang, D. Bioactive carbazole alkaloids from the stems of *Clausena lansium*. *Fitoterapia* **2015**, *103*, 122–128.
- Wen-Shyong, L.; McChesney, J. D.; El-Feraly, F. S. Carbazole alkaloids from *Clausena lansium*. *Phytochemistry* **1991**, *30* (1), 343–346.
- Adebajo, A.C.; Iwalewa, E.O.; Obuotor, E.M.; Ibikunle, G.F.; Omisore, N.O.; Adewunmi, C.O.; Obaparusi, O.O.; Klaes, M.; Adetogun, G.E.; Schmidt, T.J.; Verspohl, E.J. Pharmacological properties of the extract and some isolated compounds of *Clausena lansium* stem bark: anti-trichomonal, antidiabetic, anti-inflammatory, hepatoprotective and antioxidant effects. *J. Ethnopharmacol.* **2009**, *122* (1), 10–19.
- Ismail, A. A.; Ahmad, B. A.; Mohamed, A.; RasedeeAbdullah; Siddiq, I. A.; Mohamed, Y. I.; Zeenelabdin, A. A review of traditional uses, phytochemical and pharmacological aspects of selected members of *Clausena* genus (Rutaceae). *J. Med. Plants Res.* **2012**, *6* (38), 5107–5118.
- Peng, W.; Fu, X.; Li, Y.; Xiong, Z.; Shi, X.; Zhang, F.; Huo, G.; Li, B. Phytochemical study of stem and leaf of *Clausena lansium*. *Molecules* **2019**, *24* (17), 3124.
- Yu, H.; Zhao, Z.; Gao, A.; Luo, R. Quality evaluation of 23 species of *Clausena lansium* (Lour.) Skeels germplasm resources based on principal component analysis and cluster analysis. *Chin J. Trop Crop.* **2022**, *7*, 1357–1364.
- Li, S.; Chen, W.; Xu, Y.; Xiao, G.; Wu, J.; Chen, Z. Nutrition composition of seedless wampee. *Food Sci. Technol.* **2005**, *06*, 96–98.
- Wall, M. M. Ascorbic acid and mineral composition of longan (*Dimocarpus longan*), lychee (*Litchi chinensis*) and rambutan (*Nephelium lappaceum*) cultivars grown in Hawaii. *J. Food Compos Anal.* **2006**, *19* (6), 655–663.
- Wall, M. M. Ascorbic acid, vitamin A, and mineral composition of banana (*Musa sp.*) and papaya (*Carica papaya*) cultivars grown in Hawaii. *J. Food Compos Anal.* **2006**, *19* (5), 434–445.
- Huang, W.-Y.; Cai, Y.-Z.; Corke, H.; Sun, M. Survey of antioxidant capacity and nutritional quality of selected edible and medicinal fruit plants in Hong Kong. *J. Food Compos Anal.* **2010**, *23* (6), 510–517.
- Zhang, Y.; Huang, Y.; Huang, J.; Li, R.; Liu, J. Study on amino acids from the fruit of *Clausena lansium*. *J. Chin. Med. Mater.* **2006**, *29* (9), 921–924.
- Dai, H.; Lai, Z.; Huang, B.; Li, J.; Xiao, W.; Wang, X. Determination of amino acids in flesh of waxberry and wampee by RP-HPLC. *J. Zhongkai Univ. Agric. Technol.* **2008**, *21* (003), 7–11.
- Sun, D.; Lu, X.; Liang, J.; Hu, Y.; Xie, J. Fruit quality and constituent of sugars and organic acids in wampee cultivars. *Chin. J. Trop. Crops.* **2012**, *33* (8), 1418–1421.
- Lu, Y.; Lin, Z.; Zeng, Y.; Qiu, J.; Pan, Q. Diversity of wampee (*Clausena Lansium* (Lour.) Skeels) germplasms in China revealed based on fruit quality traits. *Mol. Plant Breed.* **2015**, *13* (2), 317–330.
- Yang, W.; Zhao, P.; Li, X.; Guo, L.; Gao, W. The potential roles of natural plant polysaccharides in inflammatory bowel disease: A review. *Carbohydr. Polym.* **2022**, *277*, No. 118821.
- Peng, J.; Bu, Z.; Ren, H.; He, Q.; Yu, Y.; Xu, Y.; Wu, J.; Cheng, L.; Li, L. Physicochemical, structural, and functional properties of wampee (*Clausena lansium* (Lour.) Skeels) fruit peel pectin extracted with different organic acids. *Food Chem.* **2022**, *386*, No. 132834.
- Wu, H.; Min, T.; Li, X.; Li, L.; Lai, F.; Tang, Y.; Yang, X. Physicochemical properties and antioxidant activities of acidic polysaccharides from wampee seeds. *Int. J. Biol. Macromol.* **2013**, *59*, 90–95.
- Song, C.; Huang, F.; Liu, L.; Zhou, Q.; Zhang, D.; Fang, Q.; Lei, H.; Niu, H. Characterization and prebiotic properties of pectin polysaccharide from *Clausena lansium* (Lour.) Skeels fruit. *Int. J. Biol. Macromol.* **2022**, *194*, 412–421.
- Wen, P.; Pei, Z.; Zhu, T.; Yu, Z.; Geng, Y.; Chen, C.; Xue, C. Preparation technology optimization of soluble dietary fiber and its structure characterization and composition of monosaccharide from *Clausena lansium* sarcocarp. *Sci. Technol. Food Ind.* **2020**, *41* (21), 29–36.
- Tongnuanchan, P.; Benjakul, S. Essential oils: Extraction, bioactivities, and their uses for food preservation. *J. Food Sci.* **2014**, *79* (7), R1231–R1249.
- Chokeprasert, P.; Charles, A. L.; Sue, K.-H.; Huang, T.-C. Volatile components of the leaves, fruits and seeds of wampee [*Clausena lansium* (Lour.) Skeels]. *J. Food Compos Anal.* **2007**, *20* (1), 52–56.
- Ma, Y.; Wang, Y.; Zhou, X.; Yang, H.; Zhang, H.; Chen, W.; Zhang, H.; Zhang, Y.; He, X. The influence of the chemical

composition of essential oils of *Clausena lansium* seeds on the growth of *Candida* strains. *Sci. Rep.* **2021**, *11* (1), 1–10.

(27) Huang, Y.; Zhang, Y.; Huang, J.; Li, R.; Liu, J. Study on chemical constituents of volatile oil and trace elements from fruits of *Clausena lansium*. *Chin. J. Chin. Mater. Med.* **2006**, *31* (11), 898–900.

(28) Tampieri, M. P.; Galuppi, R.; Macchioni, F.; Carelle, M. S.; Falcioni, L.; Cioni, P. L.; Morelli, I. The inhibition of *Candida albicans* by selected essential oils and their major components. *Mycopathologia* **2005**, *159* (3), 339–345.

(29) Yousefi, M.; Rahimi-Nasrabadi, M.; Pourmortazavi, S. M.; Wysokowski, M.; Jesionowski, T.; Ehrlich, H.; Mirsadeghi, S. Supercritical fluid extraction of essential oils. *Trends Anal. Chem.* **2019**, *118*, 182–193.

(30) Laura, A.; Moreno-Escamilla, J. O.; Rodrigo-García, J.; Alvarez-Parrilla, E., Phenolic compounds. In *Postharvest physiology and biochemistry of fruits and vegetables*; Elsevier: 2019; pp 253–271.

(31) Saranraj, P.; Behera, S. S.; Ray, R. C., Traditional foods from tropical root and tuber crops: Innovations and challenges. In *Innovations in traditional foods*; Elsevier: 2019; pp 159–191.

(32) Ye, Y.; Chang, X.; Brennan, M. A.; Brennan, C. S.; Guo, X. Comparison of phytochemical profiles, cellular antioxidant and anti-proliferative activities in five varieties of wampee (*Clausena lansium*) fruits. *Int. J. Food Sci. Technol.* **2019**, *54* (7), 2487–2493.

(33) Prasad, K. N.; Hao, J.; Yi, C.; Zhang, D.; Qiu, S.; Jiang, Y.; Zhang, M.; Chen, F. Antioxidant and anticancer activities of wampee (*Clausena lansium* (Lour.) Skeels) peel. *J. Biotechnol. Biomed.* **2009**, No. 612805.

(34) Chang, X.; Ye, Y.; Pan, J.; Lin, Z.; Qiu, J.; Guo, X.; Lu, Y. Comparative assessment of phytochemical profiles and antioxidant activities in selected five varieties of wampee (*Clausena lansium*) fruits. *Int. J. Food Sci. Technol.* **2018**, *53* (12), 2680–2686.

(35) Li, Q.; Chang, X.; Guo, R.; Wang, Q.; Guo, X. Dynamic effects of fermentation on phytochemical composition and antioxidant properties of wampee (*Clausena lansium* (Lour.) Skeel) leaves. *Food Sci. Nutr.* **2019**, *7* (1), 76–85.

(36) Kong, F.; Su, Z.; Guo, X.; Zeng, F.; Bi, Y. Antidiabetic and lipid-lowering effects of the polyphenol extracts from the leaves of *Clausena lansium* (Lour.) Skeels on streptozotocin-induced type 2 diabetic rats. *J. Food Sci.* **2018**, *83* (1), 212–220.

(37) Chang, X.; Lu, Y.; Lin, Z.; Qiu, J.; Guo, X.; Pan, J.; Mehmood, A. A. Impact of leaf development stages on polyphenolics profile and antioxidant activity in *Clausena lansium* (Lour.) Skeels. *Biomed Res. Int.* **2018**, No. 7093691.

(38) Chen, G.-L.; Zhang, X.; Chen, S.-G.; Han, M.-D.; Gao, Y.-Q. Antioxidant activities and contents of free, esterified and insoluble-bound phenolics in 14 subtropical fruit leaves collected from the south of China. *J. Funct. Foods* **2017**, *30*, 290–302.

(39) Chai, W.-M.; Lin, M.-Z.; Feng, H.-L.; Zou, Z.-R.; Wang, Y.-X. Proanthocyanidins purified from fruit pericarp of *Clausena lansium* (Lour.) Skeels as efficient tyrosinase inhibitors: Structure evaluation, inhibitory activity and molecular mechanism. *Food Funct.* **2017**, *8* (3), 1043–1051.

(40) Chang, X.; Ye, Y.; Pan, J.; Lin, Z.; Qiu, J.; Peng, C.; Guo, X.; Lu, Y. Comparative analysis of phytochemical profiles and antioxidant activities between sweet and sour wampee (*Clausena lansium*) Fruits. *Foods* **2022**, *11* (9), 1230.

(41) Debnath, B.; Singh, W. S.; Das, M.; Goswami, S.; Singh, M. K.; Maiti, D.; Manna, K. Role of plant alkaloids on human health: A review of biological activities. *Mater. Today Chem.* **2018**, *9*, 56–72.

(42) Yang, J.-H.; Wang, X.-Y.; Zhou, Y.-P.; Lu, R.; Chen, C.-H.; Zhang, M.-H.; Cheng, Y.-Y.; Morris-Natschke, S. L.; Lee, K.-H.; Wang, Y.-S. Carbazole alkaloids from *Clausena anisum-olens*: Isolation, characterization, and anti-HIV evaluation. *Molecules* **2020**, *25* (1), 99.

(43) Shaikh, M. S.; Karpoomath, R.; Thapliyal, N.; Rane, R. A.; Palkar, M. B.; Faya, A. M.; Patel, H. M.; Alwan, W. S.; Jain, K.; Hampannavar, G. A. Current perspective of natural alkaloid carbazole and its derivatives as antitumor agents. *Anti-Cancer Agents Med. Chem.* **2015**, *15* (8), 1049–1065.

(44) Liu, H.; Li, C.-J.; Yang, J.-Z.; Ning, N.; Si, Y.-K.; Li, L.; Chen, N.-H.; Zhao, Q.; Zhang, D.-M. Carbazole alkaloids from the stems of *Clausena lansium*. *J. Nat. Prod.* **2012**, *75* (4), 677–682.

(45) Deng, H.; Mei, W.; Wang, H.; Guo, Z.; Dong, W.; Wang, H.; Li, S.; Dai, H. Carbazole alkaloids from the peels of *Clausena lansium*. *J. Asian Nat. Prod. Res.* **2014**, *16* (10), 1024–1028.

(46) Du, Y.; Liu, H.; Li, C.; Yang, J.; Ma, J.; Zhang, D.; Sun, H.; Zhang, D. Carbazole and amide alkaloids from the stems of *Clausena lansium*. *J. Asian Nat. Prod. Res.* **2015**, *17* (11), 1048–1053.

(47) Du, Y.-Q.; Liu, H.; Li, C.-J.; Ma, J.; Zhang, D.; Li, L.; Sun, H.; Bao, X.-Q.; Zhang, D.-M. Bioactive carbazole alkaloids from the stems of *Clausena lansium*. *Fitoterapia* **2015**, *103*, 122–128.

(48) Liu, Y.; Guo, J.; Liu, Y.; Hu, S.; Yan, G.; Qiang, L.; Fu, Y. Carbazole alkaloids with potential neuroprotective activities from the fruits of *Clausena lansium*. *J. Agric. Food Chem.* **2019**, *67* (20), 5764–5771.

(49) Sun, X.; Ma, J.; Li, C.; Zang, Y.; Huang, J.; Wang, X.; Chen, N.; Chen, X.; Zhang, D. Carbazole alkaloids with bioactivities from the stems of *Clausena lansium*. *Phytochem. Lett.* **2020**, *38*, 28–32.

(50) Sun, X.; Li, C.; Ma, J.; Zang, Y.; Huang, J.; Chen, N.; Wang, X.; Zhang, D. New amide alkaloids and carbazole alkaloid from the stems of *Clausena lansium*. *Fitoterapia* **2021**, *154*, No. 104999.

(51) Fan, Y.; Sahu, S. K.; Yang, T.; Mu, W.; Wei, J.; Cheng, L.; Yang, J.; Liu, J.; Zhao, Y.; Lisby, M.; Liu, H. The *Clausena lansium* (Wampee) genome reveal new insights into the carbazole alkaloids biosynthesis pathway. *Genomics* **2021**, *113* (6), 3696–3704.

(52) Li, H.; Feng, Z.; Wang, Y.; Lin, L. Anticancer carbazole alkaloids and coumarins from *Clausena* plants: A review. *Chin. J. Nat. Med.* **2017**, *15* (12), 881–888.

(53) Yang, M.; Cao, Y.; Li, W.; Yang, Y.; Chen, Y.; Huang, L. Isolation and structural elucidation of Clausenamides from the leaves of *Clausena lansium* (Lour.) Skeels. *Acta Pharm. Sin.* **1987**, *22* (1), 33–40.

(54) Shen, D.; Kuo, P.; Huang, S.; Hwang, T.; Chan, Y.; Shieh, P.; Ngan, N.; Thang, T.; Wu, T. Constituents from the leaves of *Clausena lansium* and their anti-inflammatory activity. *J. Nat. Med.* **2017**, *71* (1), 96–104.

(55) Matsui, T.; Ito, C.; Furukawa, H.; Okada, T.; Itoigawa, M. Lansiumamide B and SB-204900 isolated from *Clausena lansium* inhibit histamine and TNF- α release from RBL-2H3 cells. *Inflamm. Res.* **2013**, *62* (3), 333–341.

(56) Lin, J.-H. Cinnamide derivatives from *Clausena lansium*. *Phytochemistry* **1989**, *28* (2), 621–622.

(57) Fan, Y.; Chen, H.; Mei, W.; Kong, F.; Li, F.; Chen, P.; Cai, C.; Huang, M.; Dai, H. Nematicidal amide alkaloids from the seeds of *Clausena lansium*. *Fitoterapia* **2018**, *128*, 20–25.

(58) Huang, L.; Li, D.; Xu, Y.-S.; Feng, Z.-L.; Meng, F.-C.; Zhang, Q.-W.; Gan, L.-S.; Lin, L.-G. Clausoxamine, an alkaloid possessing a 1,3-oxazine-4-one ring from the seeds of *Clausena lansium* and the anti-obesity effect of lansiumamide B. *RSC Adv.* **2017**, *7* (74), 46900–46905.

(59) Han, Y.; Li, L.-c.; Hao, W.-b.; Tang, M.; Wan, S.-q. Larvicidal activity of lansiumamide B from the seeds of *Clausena lansium* against *Aedes albopictus* (Diptera: Culicidae). *Parasitol. Res.* **2013**, *112* (2), 511–516.

(60) Li, L.; Feng, X.; Tang, M.; Hao, W.; Han, Y.; Zhang, G.; Wan, S. Antibacterial activity of Lansiumamide B to tobacco bacterial wilt (*Ralstonia solanacearum*). *Microbiol. Res.* **2014**, *169* (7–8), 522–526.

(61) Xu, H.; Chen, T.; Huang, L.; Shen, Q.; Lian, Z.; Shi, Y.; Ouyang, M.-A.; Song, L. Synthesis and fungicidal activity of lansiumamide A and B and their derivatives. *Molecules* **2018**, *23* (7), 1499.

(62) Liu, J.; Du, Y.; Li, C.; Li, L.; Chen, F.; Yang, J.; Chen, N.; Zhang, D. Alkaloids from the stems of *Clausena lansium* and their neuroprotective activity. *RSC Adv.* **2017**, *7* (56), 35417–35425.

(63) Rotariu, D.; Babes, E. E.; Tit, D. M.; Moisi, M.; Bustea, C.; Stoicescu, M.; Radu, A.-F.; Vesa, C. M.; Behl, T.; Bungau, A. F.; Bungau, S. G. Oxidative stress—complex pathological issues

concerning the hallmark of cardiovascular and metabolic disorders. *Biomed. Pharmacother.* **2022**, *152*, No. 113238.

(64) Pizzino, G.; Irrera, N.; Cucinotta, M.; Pallio, G.; Mannino, F.; Arcoraci, V.; Squadrito, F.; Altavilla, D.; Bitto, A. Oxidative stress: harms and benefits for human health. *Oxid. Med. Cell. Longev.* **2017**, No. 8416763.

(65) Srinivas, U. S.; Tan, B. W.; Vellayappan, B. A.; Jayasekharan, A. D. ROS and the DNA damage response in cancer. *Redox Biol.* **2019**, *25*, No. 101084.

(66) Zeng, X.; Zhou, X.; Cui, L.; Liu, D.; Wu, K.; Li, W.; Huang, R. The fruits of wampee inhibit H₂O₂-induced apoptosis in PC12 cells via the NF- κ B pathway and regulation of cellular redox status. *Molecules* **2014**, *19* (6), 7368–7387.

(67) Chang, X.; Lu, Y.; Lin, Z.; Qiu, J.; Guo, X.; Pan, J.; Abbasi, A. M. Impact of leaf development stages on polyphenolics profile and antioxidant activity in *Clausena lansium* (Lour.) Skeels. *Biomed Res. Int.* **2018**, No. 7093691.

(68) Schaich, K. M.; Tian, X.; Xie, J. Hurdles and pitfalls in measuring antioxidant efficacy: A critical evaluation of ABTS, DPPH, and ORAC assays. *J. Funct. Foods* **2015**, *14*, 111–125.

(69) Maragakis, N. J.; Rothstein, J. D. Mechanisms of disease: Astrocytes in neurodegenerative disease. *Nat. Clin. Pract. Neurol.* **2006**, *2* (12), 679–689.

(70) Xicoy, H.; Wieringa, B.; Martens, G. J. The SH-SY5Y cell line in Parkinson's disease research: A systematic review. *Mol. Neurodegener.* **2017**, *12* (1), 1–11.

(71) Wiatrak, B.; Kubis-Kubiak, A.; Piwowar, A.; Barg, E. PC12 cell line: Cell types, coating of culture vessels, differentiation and other culture conditions. *Cells* **2020**, *9* (4), 958.

(72) Hu, J.-F.; Chu, S.-F.; Ning, N.; Yuan, Y.-H.; Xue, W.; Chen, N.-H.; Zhang, J.-T. Protective effect of (–) Clausenamide against A β -induced neurotoxicity in differentiated PC12 cells. *Neurosci. Lett.* **2010**, *483* (1), 78–82.

(73) Liu, Y.; Guo, J.; Wang, X.; Liu, Y.; Zhang, W.; Wang, T.; Qiang, L.; Fu, Y. Geranylated carbazole alkaloids with potential neuroprotective activities from the stems and leaves of *Clausena lansium*. *Bioorg. Chem.* **2019**, *92*, No. 103278.

(74) Li, B.; Yuan, Y.; Hu, J.; Zhao, Q.; Zhang, D.; Chen, N. Protective effect of Bu-7, a flavonoid extracted from *Clausena lansium*, against rotenone injury in PC12 cells. *Acta Pharmacol. Sin.* **2011**, *32* (11), 1321–1326.

(75) Tongun, T.; Phachonpai, W. Cognitive booster of wampee peel extract on chronic restraint stress-induced memory dysfunction in rats. *J. Appl. Pharm. Sci.* **2020**, *10* (7), 019–026.

(76) Phachonpai, W.; Tongun, T. Cognition enhancing effects of *Clausena lansium* (Lour.) peel extract attenuate chronic restraint stress-induced memory deficit in rats. *Heliyon* **2021**, *7* (5), No. e07003.

(77) Phachonpai, W.; Tongun, T. Neuroprotective and cognitive enhancing effects of *Clausena lansium* (Lour.) skeels peels extract in ischemic rat brains. *J. Appl. Pharm. Sci.* **2021**, *11* (9), 001–008.

(78) Shin, S. A.; Joo, B. J.; Lee, J. S.; Ryu, G.; Han, M.; Kim, W. Y.; Park, H. H.; Lee, J. H.; Lee, C. S. Phytochemicals as anti-inflammatory agents in animal models of prevalent inflammatory diseases. *Molecules* **2020**, *25* (24), 5932.

(79) Huang, G.; Li, J.; Li, W.; Liu, T.; Jiang, G.; Wu, T.; Zou, Y.; Huang, J.; Tao, L.; Zhu, Z.; et al. Suppression of inflammation by ethanol extract of *Clausena lansium* via modulation of TLR4/MYD88/TRAF6 signaling pathway in RAW 264.7 macrophages. *Eur. J. Inflamm.* **2019**, *17*, No. 2058739219841973.

(80) Liu, J.; Li, C.-J.; Ni, L.; Yang, J.-Z.; Li, L.; Zang, C.-x.; Bao, X.-Q.; Zhang, D.; Zhang, D.-M. Anti-inflammatory alkaloid glycoside and quinoline alkaloid derivatives from the stems of *Clausena lansium*. *RSC Adv.* **2015**, *5* (98), 80553–80560.

(81) Rodanant, P.; Surarit, R.; Laphookhieo, S.; Kuvatanasuchati, J. In vitro evaluation of the antibacterial and anti-inflammation activities of *Clausena lansium* (Lour.) Skeels. *Songklanakarinn J. Sci. Technol.* **2015**, *37* (1), 43–48.

(82) Shen, D.-Y.; Chao, C.-H.; Chan, H.-H.; Huang, G.-J.; Hwang, T.-L.; Lai, C.-Y.; Lee, K.-H.; Thang, T. D.; Wu, T.-S. Bioactive constituents of *Clausena lansium* and a method for discrimination of aldose enantiomers. *Phytochemistry* **2012**, *82*, 110–117.

(83) Liu, J.; Li, C.-J.; Du, Y.-Q.; Li, L.; Sun, H.; Chen, N.-H.; Zhang, D.-M. Bioactive compounds from the stems of *Clausena lansium*. *Molecules* **2017**, *22* (12), 2226.

(84) Liu, G. T.; Li, W. X.; Chen, Y. Y.; Wei, H. L. Hepatoprotective action of nine constituents isolated from the leaves of *Clausena lansium* in mice. *Drug Dev. Res.* **1996**, *39* (2), 174–178.

(85) Ou-Yang, C.; Chai, W.; Xu, X.; Song, S.; Wei, Q.; Huang, Q.; Zou, Z. Inhibitory potential of proanthocyanidins from the fruit pulp of *Clausena lansium* (Lour.) Skeels against α -glucosidase and non-enzymatic glycation: Activity and mechanism. *Process Biochem.* **2020**, *91*, 364–373.

(86) Shen, S.; Liao, Q.; Huang, L.; Li, D.; Zhang, Q.; Wang, Y.; Lee, S. M.-Y.; Lin, L. Water soluble fraction from ethanolic extract of *Clausena lansium* seeds alleviates obesity and insulin resistance, and changes the composition of gut microbiota in high-fat diet-fed mice. *J. Funct. Foods* **2018**, *47*, 192–199.

(87) He, X.; Zhang, L.; Chen, J.; Sui, J.; Yi, G.; Wu, J.; Ma, Y. Correlation between chemical composition and antifungal activity of *Clausena lansium* essential oil against *Candida spp.* *Molecules* **2019**, *24* (7), 1394.

(88) Maneerat, W.; Laphookhieo, S. Antitumoral alkaloids from *Clausena lansium*. *Heterocycles* **2010**, *81* (5), 1261.

(89) Maneerat, W.; Ritthiwigrom, T.; Cheenpracha, S.; Laphookhieo, S. Carbazole alkaloids and coumarins from *Clausena lansium* roots. *Phytochem. Lett.* **2012**, *5* (1), 26–28.

(90) Jiang, H.-Y.; Zhang, W.-J.; You, C.-X.; Yang, K.; Fan, L.; Feng, J.-B.; Chen, J.; Yang, Y.-J.; Wang, C.-F.; Deng, Z.-W.; Yin, H.-B.; Du, S.-S. Two new cytotoxic constituents from the *Clausena lansium* (Lour.) Skeels. *Phytochem. Lett.* **2014**, *9*, 92–95.

(91) Peng, W.; Zheng, L.; Ji, C.; Shi, X.; Xiong, Z.; Shangguan, X. Carbazole alkaloids isolated from the branch and leaf extracts of *Clausena lansium*. *Chin. J. Nat. Med.* **2018**, *16* (7), 509–512.

(92) Ming-He, Y.; Yan-Yong, C.; Liang, H. Studies on the chemical constituents of *Clausena Lansium* (Lour.) Skeels II: the isolation and structural elucidation of neo-clausenamide and dehydrocycloclausenamide. *Acta Chimica Sinica English Edition* **1987**, *5* (3), 267–272.

(93) Ming-He, Y.; Yan-Yong, C.; Liang, H. Three novel cyclic amides from *Clausena lansium*. *Phytochemistry* **1988**, *27* (2), 445–450.

(94) Zhao, Q.; Yang, J.-Z.; Li, C.-J.; Chen, N.-H.; Zhang, D.-M. A new megastigmane glucoside and a new amide alkaloid from the leaves of *Clausena lansium* (Lour.) Skeels. *J. Asian Nat. Prod. Res.* **2011**, *13* (04), 361–366.

(95) Fan, Y.; Chen, H.; Mei, W.; Xu, S.; Zhou, L.; Huang, M. Amide alkaloids from the seeds of *Clausena lansium* and their nematocidal activities. *J. Trop. Subtrop. Bot.* **2018**, *26* (1), 85–91.

(96) Ma, Y.; Wang, Y.; Zhou, X.; Yang, H.; Zhang, H.; Chen, W.; Zhang, H.; Zhang, Y.; He, X. The influence of the chemical composition of essential oils of *Clausena lansium* seeds on the growth of *Candida* strains. *Sci. Rep.* **2021**, *11* (1), 1–10.