

Review Article

The Impact of External Factors on the Epigenome: *In Utero* and over Lifetime

**Estela G. Torano,¹ María G. García,¹ Juan Luis Fernández-Morera,¹
Pilar Niño-García,² and Agustín F. Fernández¹**

¹*Cancer Epigenetics Laboratory, Instituto Universitario de Oncología del Principado de Asturias (IUOPA), HUCA, Universidad de Oviedo, 33006 Oviedo, Spain*

²*Servicio de Salud Laboral, Instituto Asturiano de Prevención de Riesgos Laborales, 33006 Oviedo, Spain*

Correspondence should be addressed to Agustín F. Fernández; affernandez@hca.es

Received 19 February 2016; Revised 12 April 2016; Accepted 26 April 2016

Academic Editor: Max Costa

Copyright © 2016 Estela G. Torano et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Epigenetic marks change during fetal development, adult life, and aging. Some changes play an important role in the establishment and regulation of gene programs, but others seem to occur without any apparent physiological role. An important future challenge in the field of epigenetics will be to describe how the environment affects both of these types of epigenetic change and to learn if interaction between them can determine healthy and disease phenotypes during lifetime. Here we discuss how chemical and physical environmental stressors, diet, life habits, and pharmacological treatments can affect the epigenome during lifetime and the possible impact of these epigenetic changes on pathophysiological processes.

1. Introduction

The meaning of the term epigenetics has evolved considerably over time. Conrad Hal Waddington coined the term in the 1940s to describe the “causal mechanisms” that give rise to phenotypes from genotypes in the developmental and differentiation processes [1]. Nowadays, the term is used to explain stable heritable chemical modifications to DNA and histones that affect gene expression without altering nucleotide sequence [2]. This new concept has allowed the consideration of a new perspective from which the complexity of many cellular processes such as genetic regulation, cellular development and differentiation, genomic imprinting, embryology, aging and cancer, and other diseases is understood. What is more, epigenetic alterations may occur due to chance or under environmental influence [3]. In the latter case, epigenetics moderates the genetic expression of a trait depending on the prevailing environmental conditions, a phenomenon which could confer an organism with the necessary plasticity to adapt to its environment and the capacity to induce

alternative phenotypes from the same genotype through the regulation of gene expression patterns [4].

Environmental epigenetics emerges from the idea that the interaction between the environment and the epigenome may alter the phenotype and might be related to disease susceptibility. And most importantly, these alterations could be transmitted down through generations [5, 6]. The epigenome is at risk of changes and alterations over time, and it will be dependent on internal, external, and/or stochastic factors [7]. In this review we will describe how external factors affect the epigenome and the consequences for health and disease during lifetime. We will discuss recent works on how epigenetic mechanisms, such as DNA methylation, histone posttranslational modifications, and noncoding RNAs, particularly microRNAs, are affected by environmental aspects, such as different chemical and physical environmental stressors, diet, unhealthy habits, and pharmacological treatments. The principal epigenetic mechanisms will be described and we will also discuss how epigenetic alterations caused by external factors could mediate the appearance of disease phenotypes.

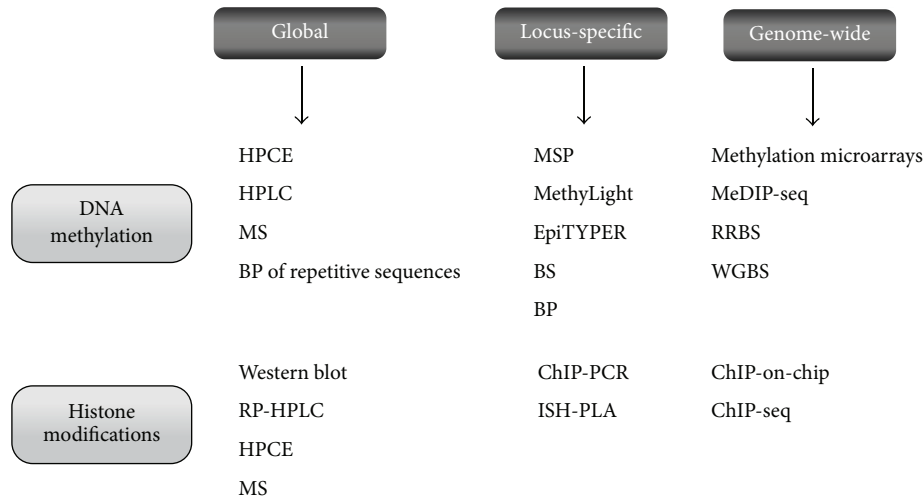


FIGURE 1: Summary of methods for DNA methylation and histone modification analysis. Different approaches, depending on whether global, locus-specific, or genome-wide analyses, are performed. HPCE: high-performance capillary electrophoresis; HPLC: high-performance liquid chromatography; MS: mass spectrometry; BP: bisulfite pyrosequencing; MSP: methylation-specific PCR; BS: bisulfite sequencing; MeDIP-seq: methylated DNA immunoprecipitation sequencing; RRBS: reduced representation bisulfite sequencing; WGBS: whole genome bisulfite sequencing; RP-HPLC: reversed-phase high-performance liquid chromatography (RP-HPLC); ChIP: chromatin immunoprecipitation; ISH-PLA: *in situ* hybridization and proximity ligation assay.

2. Epigenetic Mechanisms

Chromatin is a highly regulated complex of macromolecules in the nucleus formed by DNA, histone proteins, and RNA. The nucleosome is the basic repetitive unit of chromatin and consists of 147 base pairs around an octamer of the four core histones (H2A, H2B, H3, and H4) [8]. Differences in the degree of compaction determine its structure and complexity which in turn facilitate its differentiation into two functional states: euchromatin or heterochromatin, which are the transcriptionally active and inactive forms, respectively [9]. Epigenetic mechanisms such as DNA methylation and histone modifications participate in the remodeling of chromatin. These changes in chromatin structure are able to modify the accessibility of genes to transcriptional machinery, regulate gene expression during development and differentiation stages, and determine which genes are transcribed [10]. Aberrant profiles of these epigenetic processes may result in mismatches in important signaling pathways that alter various cell functions and may lead to the development of different diseases such as cancer [11].

DNA methylation is the best-known epigenetic mechanism in mammals, not only because it was the first discovered, but because it is also easier to measure [12, 13] (Figure 1). It is in fact one of the principal epigenetic events in the human genome and an important regulator of transcriptional activity, genomic imprinting, development, and tumorigenesis [14–16]. Methylation consists in the addition of a methyl group at the carbon 5 position of the cytosine ring to obtain 5-methylcytosine. It is a postreplication modification that appears nonuniformly in the human genome, which contains unmethylated areas intercalated by methylated regions [17, 18]. Methylation occurs predominantly in cytosines of CpG dinucleotides. In vertebrates, around 70–80% of all CpGs

are methylated, specifically in repressive heterochromatin regions and in repetitive sequences, such as retrotransposable elements [19, 20]. These CpGs are asymmetrically distributed into CpG-poor regions and CpG-dense regions called “CpG islands” which are located in the promoter regions in approximately 60% of genes and are usually nonmethylated [21, 22].

In general, CpG island methylation is related to gene silencing. DNA methylation promotes the binding of methyl binding proteins (MBPs), which mediate the recruitment of transcriptional repressors [23]. When this epigenetic change occurs in CpG islands located in promoter regions, the silencing can affect important cellular pathways involved in the development of multiple diseases such as cancer [24]. However, it is known that methylation is more dynamic in CpG shores, the genomic regions that are delimited as the 2 Kb regions flanking CpG islands, and in CpG shelf regions, that is, those beyond the CpG shores, 2–4 Kb beyond the CpG islands. Silencing by DNA methylation can also occur in these adjacent regions, where differences in methylation patterns have been found which are related to tissue specific differentiation and cancer [25–28]. Methylation can be catalyzed by two important groups of DNA methyltransferases (DNMTs) [29]: in the first group is DNMT1, which is essential in cell proliferation and ensures the maintenance of DNA-methylation patterns during DNA replication through the methylation of hemimethylated CpGs [30]. The second group includes DNMT3a and DNMT3b, which are required for *de novo* methylation and for establishing methylation patterns in early embryos and during development [31]. Their activity is also known to be necessary for the maintenance of methylation patterns in somatic cells [32].

DNA methylation is crucial for physiological development, playing a fundamental role in gene expression programs during cell fate differentiation [33, 34]. Several studies

have identified global DNA hypomethylation related to aging as well as tumoral processes [35–37]. This global decline of methylation levels has also been described as being associated with gene-specific hypermethylation [38, 39]. An illustration of these similar methylation patterns observed in both cancer and aging processes can be seen in relation to the promoter region of the estrogen receptor gene (ER), which is specifically hypermethylated in colon cancer and old individuals compared to normal patients and young people, respectively [39]. DNA methylation is also related to genomic imprinting, X chromosome inactivation mechanisms in females, and the silencing of foreign nucleic acids [40].

Histone modifications are another epigenetic mechanism that has been well studied and linked to development processes and aging [41, 42]. Chemical changes in histone protein residues are known as posttranslational modifications (PTMs). These variations usually affect protein functions or gene expression, ultimately impacting on biological processes [43]. PTMs involve reversible modifications commonly located in the N-terminal tails of histones, such as acetylation, methylation, phosphorylation, and ubiquitylation [44, 45]. These marks taken in combination form a “histone code,” which establishes a regulatory mechanism of chromatin dynamics which affects affinity in protein interactions, protein-DNA binding, and gene transcription [46, 47]. They also play an important role in DNA repair [48], DNA replication, and chromatin compaction [43]. These PTMs are “written” by different families of enzymes; for example, histone acetyltransferases (HATs) and deacetylases (HDACs) are involved in histone acetylation, while histone methyltransferases (HMTs) catalyze the transference of up to three methyl groups onto the lysine and arginine amino acids on histones, predominantly H3 and H4 [49]. The most common PTMs are in fact the acetylation and methylation of lysines on histone tails [46, 50]. Acetylation of lysine by its corresponding enzymes is linked to transcription activity [51]. However, histone methylation can be related to either gene activation or repression depending on which residues are affected; for example, H3K4me3 and H3K36me3 are active markers while H3K27me3 and H3K9me3 are both related to gene repression and heterochromatin [43, 46, 50, 52, 53].

Apart from the groups of enzymes above, there also exist proteins with specific domains that are able to identify these combinatorial PTMs. For example, proteins with bromodomains or chromodomains recognize and bind to acetylated lysines and methylated lysines, respectively [54, 55]. These effector proteins read and interpret histone marks [46, 56] and are involved in the regulation of transcriptional response. Greater knowledge of these protein functions has increased interest in their study as a promising new class of drug targets for a wide range of human diseases and for therapeutic development. For example, the development of BET (bromodomain and extraterminal) inhibitors selectively modulates the expression of genes involved in cell growth and invasion and antiapoptotic activity [57–59] associated with tumoral progression.

In some cases these epigenetic modifications are the result of environmental factors and may have important roles in the development of normal and pathological processes [60, 61].

In cancer disease, studies have identified PTMs associated with carcinogenesis. For example, loss of acetylation at Lys16 and trimethylation at Lys20 of histone H4 were found to be related to neoplastic processes [62]. Alterations in histone-modifying enzymes have been identified as the main cause of these changes in many cases [63–66] and as a result they have started to gain importance as a therapeutic target in recent years [67–69]. Histone PTMs are implicated not only in cancer, but also in a wide range of pathologies related to chronic diseases, such as diabetes and obesity [70], renal disease [71], and neuropathologies [72].

Although *ncRNAs* are not as well studied as DNA methylation or histone PTMs, they are attracting attention because of their important role for normal cell development and function, as well as in relation to disease. *ncRNAs* can be divided into short, intermediate, and heterogeneous group of long RNAs [73, 74]. *ncRNAs* such as micro (miRNAs) or long antisense noncoding RNAs (lncRNAs) have recently been acknowledged to play a role in epigenetics, and furthermore, they may be affected by certain environmental factors [75, 76]. An miRNA is a single-stranded RNA molecule, 21–22 nucleotides long, which has a completely different function compared to the most frequent single stranded RNA molecules such as messenger RNA (mRNA). In addition, miRNAs are not translated into protein and they are in part complementary to various mRNA molecules, to which they are able to bind to facilitate their elimination and subsequent gene expression [73]. lncRNAs are considered another epigenetic regulation mechanism of protein-coding gene expression, and they recruit histones and chromatin related proteins to specific sites [77, 78]. Loss of their activity may interfere with the transcription of various genes [79] and the aberrant functioning of lncRNA causes the deregulation of genes that are involved in a number of diseases [80]. When silencing affects tumor suppressor genes, it can contribute to cancer progress. One example is ANRIL, an lncRNA involved in the silencing of different tumor suppressor genes, such as NK4n/ARF/INK4a, p16/CDKN2A, and p15/CDKN2B, which are related to cell cycle and senescence [81, 82].

The epigenetic mechanisms described above are connected and integrated to regulate gene expression and cell fate through a complex scenario whereby the functions that they regulate interact [83]. For instance, the expression of miRNAs is usually controlled by DNA methylation, and epigenetic alterations can lead to disease phenotypes [84, 85].

3. Causes of Epigenetic Changes during Development, Adult Life and Aging

Epigenetic marks constantly change throughout life. Some of these changes are programmed and play important roles in the different stages of development, but the mechanisms involved are still not fully understood [86–88]. Intrinsic, or genetic, factors are of great importance in regulating certain epigenetic changes which occur over time. In support of this, the epigenomes of monozygotic twins are known to be more similar than those of dizygotic twins [89]. One important example of an intrinsic epigenetic mechanism which is programmed

during life is female puberty, which is initiated by the secretion of gonadotrophin-releasing hormone (GnRH) and the activation of the hypothalamic-pituitary-gonadal (HPG) axis [90]. Currently a few studies have related epigenetic mechanisms to female puberty regulation, supporting the notion that the activation of neuroendocrine pubertal components is mediated, at least in part, by epigenetic mechanisms [90].

Besides the intrinsic epigenetic changes, there are some epigenetic alterations that take place apparently by chance, without evident biological function [7, 91]. The distinction between stochastic changes and environment-mediated changes is sometimes very difficult to establish as stochastic changes can potentially be modulated by both intrinsic and extrinsic (environmental) factors [7, 87]. Although this influence of environmental factors has been widely reported [92], the manner of the interplay between the environment and the epigenome remains largely unknown. Apart from intrinsic and stochastic factors that may activate epigenetic mechanisms, the main factors modulating these mechanisms are extrinsic factors like environmental situation. Many studies have provided support for the theory that environmental pressure during the developmental stages of early life (both prenatal and in childhood), such as nutritional status or exposure to toxic compounds, can affect epigenetic developmental programming. This has given rise to the term “developmental origin of health and disease” (DOHaD), which proposes a broad range of environmental phenomena in the early stages of life that may increase disease susceptibility during adult life [93]. Consequently, when studying the influence of the environment, it is necessary to take into account two dissimilar scenarios where the environment has an impact: embryonic development and during lifetime. The first scenario is the most susceptible to any external influence due to the high number of cell division events and the critical epigenetic changes that take place during cell differentiation [94]. Furthermore, the effect of any epigenetic change in an undifferentiated cell can be transmitted and amplified to future cell populations.

During embryonic development, environmental conditions can be modulated by two different principal factors: the lifestyle of the mother, which necessarily implies embryo exposures, and the anatomical/phenotypic circumstances of the mother, such as size of the uterus and placenta. It is a period where nutrient supply and chemical exposure have a critical influence on the epigenome [88] and any resulting epigenetic dysregulation could lead to disease development during adult life. As well as mother influence, studies in rats and mice have associated paternal obesity with a reduction in the implantation rate of the blastocyst [95–97].

In relation to the adult epigenome and how it is affected by the environment, despite an ample literature in the field, the molecular mechanisms implicated are still only poorly understood [94]. What is well known, though, is that the influence of external factors on the genome depends on the tissue type involved. For instance, it is easy to see that UV radiation could have a harmful effect on the skin, but rather less obvious to see that muscle can also be affected. In addition, when an alteration affects adult stem cells the consequences are likely to be more serious than when differentiated cells are involved

[88]. And most importantly, if the germline is affected, reproductive disorders might result and even the possibility of transgenerational inheritance of the epigenetic alterations [5].

For the purposes of this review we will only consider those external factors which can affect the epigenome. We will discuss how chemical and physical environmental stressors, diet, life habits, and pharmacological treatments can alter the epigenome during lifetime and how these alterations can determine healthy and disease phenotypes.

3.1. Chemical and Physical Environmental Stressors. Epigenetic marks can be affected by exposure to metals, air pollution, benzene, organic pollutants, and electromagnetic radiation [98]. Chemical and xenobiotic compounds in water or the atmosphere are other potential environmental stressors capable of changing epigenetic status. During embryonic development, the effect of exposure to environmental pollutants seems to have an even more crucial effect on the epigenome and increases the risk of developing disease in the F_1 , F_2 , and F_3 generations [99]. In this section we will describe several studies showing how several stressors such as metals and air pollutants can affect the epigenome, which in turn is related to the appearance of certain diseases.

It is known that environmental exposure to a variety of metals, such as arsenic, mercury, nickel, lead, and cadmium, has several impacts on human health. Many recent studies suggest that alterations in epigenetic mechanisms could play a key role in the molecular mechanisms involved in the metal exposure-related diseases. Arsenic (As) is considered the most widespread metal in the environment. It is present in rocks, soil, water, insecticides, and airborne particles, among other things [100, 101]. Chronic arsenic exposure is related to many health problems such as skin lesions, neuropathy, depression, cardiovascular diseases, and various kinds of cancers [102–106]. Experimental analyses have found DNA methylation changes after arsenic exposure, both global [107, 108] and gene-specific [109]. In addition, arsenic exposure is capable of inducing H3K4me3 and H3K9ac enrichment [110–113] and H3K27me3 decrease due to alterations in histone-modifying enzymes [113]. During human development, arsenic exposure has also been associated with changes in DNA methylation patterns of cord bloods during the prenatal period [114, 115] and with gene-specific DNA methylation changes in white blood cells [116] and in the placenta [117]. In adults, arsenic concentrations have been associated with LINE-1 DNA hypomethylation in different population-based studies [118, 119]. Furthermore, it has been found that it is able to induce DNA methylation changes in a gene-specific way. For instance, high arsenic exposure was found to be related to DNA hypermethylation of the tumor suppressor genes *p16* [120] and *RASSF1A* [121]. In Bangladeshi adults, a link was found between arsenic exposure and global PTM changes, positively correlated with H3K9me2 and inversely with H3K9ac [122]. The relationship between arsenic exposure and epigenetic alterations remains unclear, but an *in vitro* study has pointed to chronic arsenic exposure inducing loss of DNA methylation through SAM depletion [123].

Cadmium (Cd) is a chemical element which is widespread in the environment, in byproducts of industrial processes,

contaminated water, or soil, and it also has many common industrial uses, as a component in battery production, for example. Furthermore, cadmium exposure is mainly the result of diet, principally through cereals and vegetables, and also smoking. It causes many health problems, such as cancer, increased risk of bone fracture, kidney damage, and probably impaired early-life development [124, 125]. Epigenetic alterations could be implicated in cadmium toxicity mechanisms during embryonic development and lifetime. For instance, a set of genes related to transcriptional regulation control and apoptosis showed DNA methylation changes associated with maternal cadmium concentrations [126]. In adults, urinary cadmium concentrations of women were inversely associated with LINE-1 methylation and also negatively associated with *DNMT3B* expression [127].

Lead (Pb) is a poisonous heavy metal used in building construction, batteries, and consumer products, among other uses [128]. Exposure to lead is a great risk for human health, affecting a variety of fundamental molecular processes [129]. For instance, maternal exposure was shown to result in neurodevelopmental deficit [130] and reduced intelligence of the child [131], and *in vitro* Pb exposure of hESCs induced changes in the methylation status of genes involved in neurogenetic signaling pathways [132]. In humans, LINE-1 and Alu repeat DNA hypomethylation were correlated with Pb levels in umbilical cord blood [133, 134], and it was shown that early-life Pb exposure caused gender-specific changes in DNA methylation in dried blood spots [135]. In adults, male Pb levels were related to LINE-1 DNA hypomethylation [134, 136]. Another study, in women, has shown *COLIA2* promoter DNA hypomethylation with high exposure to lead [137].

Mercury (Hg) is a reactive metal whose physiological activity is unknown. Products containing mercury include batteries, fluorescent bulbs, medical products, dental amalgams, thermometers, and thermostats. However, humans are mainly exposed to mercury through fish and shellfish, which tend to concentrate mercury in their bodies. Some mercury-related health outcomes are immunotoxic effects, cardiovascular disease, cancer, and kidney disease [138–140]. *In vivo* studies in rats have revealed that prenatal exposure to mercury produces a reduction in neural cell proliferation, which is associated with DNA hypomethylation [141]. In humans, a recent study showed that *in utero* exposure to mercury, even at low levels, produced changes in DNA methylation [114]. Furthermore, studies have hypothesized that prenatal mercury exposure can change the proportion of immune cells in cord blood through DNA methylation changes [114, 142]. In adult women, there is an increase in the DNA methylation of the promoter region of the tumor suppressor gene *GSTM1* following high levels of mercury exposure [137]. Another study in male dental professionals found a correlation between SEPP1 DNA hypomethylation and hair mercury levels [143].

Nickel (Ni) is a metal that is widely occurring nowadays. It is used in jewelry, coins, batteries, and medical devices, among other things. The International Agency for Research on Cancer (IARC) has determined that some nickel compounds are carcinogenic to humans (mainly linked to respiratory cancers), but the state of knowledge of the molecular

mechanisms implicated is low and further research is required. *In vitro* experiments have shown that this compound is able to increase global levels of H3k4me3, H3K9me1, and H3K9me2 through demethylase inhibition [110, 144, 145]. In adults, nail concentrations of nickel were positively correlated with LINE-1 DNA methylation levels [118].

Besides alterations of DNA methylation or/and histone modifications, numerous changes in miRNA profiles have been associated with exposure to different metals, among them mercury, arsenic, and cadmium [146], and a negative correlation between mercury and lead levels and various miRNAs has been found in cervical swabs from pregnant women [147].

In addition to metals, air pollutants can also affect the epigenome. In adults, exposure to atmospheric pollutants, especially those which are traffic-related, has been associated with a reduction in lung function and with lung cancer, which could be due to changes in the DNA methylation of inflammation and immunity genes, as well as of repetitive elements [148, 149]. The effects of exposure to particulate matter (PM) on global and gene-specific methylation in workers in the steel industry, which has high levels of PM exposure, have also been investigated. Inducible nitric oxide synthase (iNOS) was found to have significantly decreased promoter methylation after PM exposure [150]. Moreover, exposure to black carbon, a marker of traffic particles, has also been found to be associated with aberrant global DNA methylation and to be related to cardiovascular disease [151]. In some cases, differential miRNA expression produced by environmental exposure, including air pollution, may be associated with human diseases. In adults, alterations of miR-9, miR-10b, miR-21, miR-128, miR-143, miR-155, miR-222, miR-223, and miR-338 associated with air pollution have been observed in various studies [152]. Both upregulation and downregulation of several miRNAs caused by diesel exhaust particle (DEP) exposure have been associated with human airway diseases [153].

Asbestos can also alter the epigenome. Exposure in adults induces malignant pleural mesothelioma (MPM), although the pathogenic mechanisms implicated in the tumor transformation are not well characterized [154]. Many studies have related asbestos exposure to promoter DNA methylation of many tumor suppressor genes such as *APC*, *CCND2*, *CDKN2A*, *CDKN2B*, *ESR1*, *HPPBP1*, *RASSF1*, *SLC6A20*, *SYK*, and *ZIC1* in MPM [155–157]. Along with asbestos, benzene is a major immunosuppressive agent. Bollati and collaborators found that low benzene exposure is able to induce peripheral blood DNA methylation changes, such as a decrease in LINE-1, AluI, and *MAGE-1* methylation and *p15* hypermethylation. These changes could increase the risk of developing acute myelogenous leukemia [158, 159].

Endocrine disruptors are chemical pollutants which at certain doses can affect the endocrine system and produce adverse developmental, reproductive, neurological, and immune effects. Many compounds act in this way, such as pesticides (DDT and methoxychlor), fungicides (vinclozolin), herbicides (atrazine), industrial chemicals (PCBs, dioxins), and plant hormones (phytoestrogens) [5, 160], although those most frequently affecting mammalian organisms are plastics, specifically bisphenol A (BPA) and the phthalates.

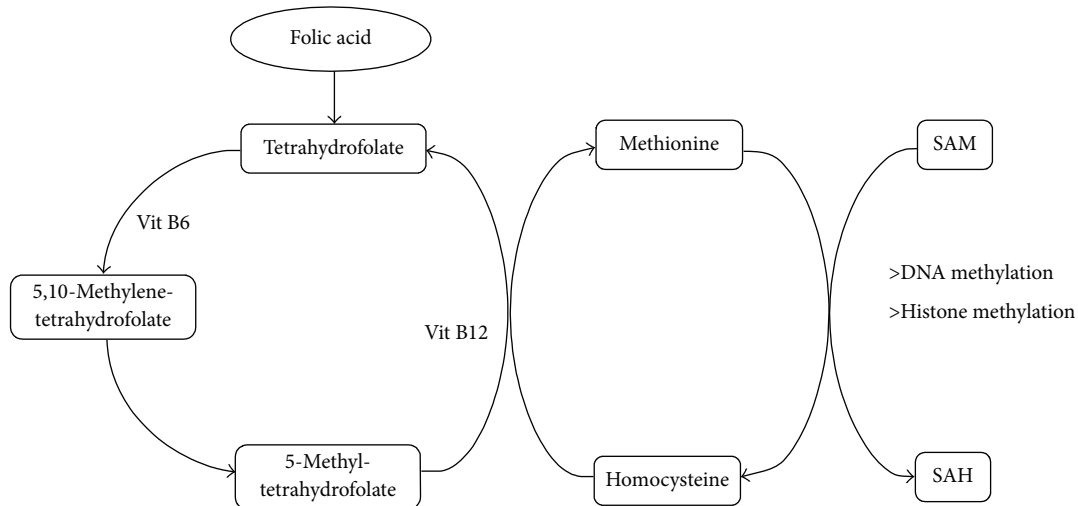


FIGURE 2: Scheme of methionine pathway. A metabolic pathway which represents the synthesis of SAM through folate intake. SAM: S-adenosylmethionine; SAH: S-adenosylhomocysteine.

The impacts of exposure to both these plastics on human health have been widely reviewed and reported by the National Toxicology Program-Center for the Evaluation of Risks to Human Reproduction [161, 162]. There is an ever increasing potential health risk associated with exposure to these two chemicals due to their extensive use in the manufacturing of polycarbonate plastics [163]. Effects of exposure include a decrease in female fertility and an increase in cancer susceptibility [164, 165]. BPA is a carbon-based synthetic compound employed to make certain consumer goods, such as CDs, DVDs, plastic bottles and containers, and the epoxy resins that line metal food and drink cans. Global hypomethylation has been found to result from BPA exposure in animal models such as *Agouti* mice [166, 167], in human spermatozoa [168], and on chromosome X in prepubescent girls [169]. *In vitro* study has related BPA treatment to an increase in promoter DNA methylation levels of *LAMP-3* (lysosomal-associated membrane protein 3), *Nsbp1* (nucleosome binding protein-1), and *Hpcall* (hippocalcin-like 1) genes [170, 171]. An *in vivo* study in rats has shown an increase in PGC-1 α DNA methylation related to induction of cardiomyopathy [172]. BPA has also been found capable of altering the expression and DNA methylation of many imprinted genes, including *Snrpn*, *Ube3a*, *Igf2*, *Kcnq1ot1*, *Cdkn1c*, and *Ascl2* [173, 174]. Apart from the DNA methylation changes produced by BPA, it also results in an increase in EZH2 activity and histone H3 trimethylation levels [170, 175].

The effect of electromagnetic radiation on the epigenome has also been investigated. Prolonged exposure to ultraviolet (UV) light is associated with the development of various skin lesions and cutaneous malignancies [176, 177]. It is well known that solar UV radiation is involved in oxidative stress (3) [178], immune system alterations [179], and gene mutation and DNA damage (6) [180]. Some studies have also found changes at the epigenetic level associated with chronic exposure to ultraviolet radiation. One study found aberrant DNA hypermethylation linked to increased DNA methyltransferases activity and hypoacetylation of H3 and H4 in UV

exposed epidermal cells in mouse models [181]. These epigenetic modifications could be related to the transcriptional silencing of certain tumor suppressor genes and thus result in stimulating skin tumor formation. The identification of aberrant methylation in genes such as *Cip1/p21* and *p16 INK4a* [182], genes of the cadherin and laminin families [183], and the inactivation of the *RB1/p16* and *p53* pathways in cutaneous squamous cell carcinoma [184], as well as other studies, supports the notion that important epigenetic changes are mediated by chronic sun or UV radiation exposure. These and other findings have allowed the identification of genes with aberrant methylation that could be used as molecular markers which are commonly perturbed in malignant skin lesions (10) [185–187], and the development of new therapies to reverse those epigenetic changes associated with UV radiation. In addition to the clinical chemopreventive agents which exist for blocking the aberrant methylation or histone deacetylation caused in many tumorigenesis processes, there are also other natural compounds found in a variety of different foods which have a photoprotector effect. Several experimental studies in *in vivo* models have shown the beneficiary effect of proanthocyanidins from grape seeds [188] and polyphenols from green tea [189, 190], among others, in countering photocarcinogenesis.

3.2. Diet. Probably the most widely described example of diet affecting epigenetic marks is the study of the intake of folate and other methyl donors during prenatal stages. Vitamin B9, or folic acid, used in the synthesis of tetrahydrofolate, cannot be synthesized *de novo* by the human body; hence it needs to be supplied from the diet. Furthermore, vitamin B6 functions as a cofactor in the synthesis of 5-methyltetrahydrofolate, the methyl donor for the B12-dependent remethylation of homocysteine to S-adenosylmethionine (SAM), and the methyl donor group for DNA and histones which is necessary to maintain methylation levels (Figure 2). The methyl donor suppliers can affect the epigenome in a global manner and also in a locus-specific way [87, 191], and lack of folic acid

nutritional supplement contributes to the induction of cancer in animal models of disease [192]. The best example is the influence of maternal diet on the murine *Agouti* gene (A^{vy}). This gene is responsible for determining if mouse coat color is banded (*Agouti*) or solid (non-*Agouti*) and is regulated by the DNA methylation status of the intracisternal A-particle (IAP) retrotransposon located in the promoter region of the *Agouti* gene [193, 194]. When it is methylated, a mouse's coat has a normal appearance (solid color), but when the retrotransposon is unmethylated, the coat is banded or yellow and the mouse will have an increased risk of cancer and diabetes. The availability of methyl donors before and during pregnancy was found to increase the methylation status of the promoter *Agouti* gene in future offspring [195].

In humans, there are some studies showing the effects of diet and food availability on the epigenome and how these epigenetic changes could be involved in the appearance of several diseases in adulthood [191, 196]. A clear example of these relationships is reflected in the results of studies of the offspring of pregnant women during the Dutch famine of 1944-1945, during the last period of the Second World War [191, 197]. Another representative example showing the effects of nutrition comes from a study in Gambia showing differences in the epigenomes of children conceived during the nutrient-poor rainy season compared to those conceived in periods where the nutritional intake of the mother is better [196].

Although dietary folate deficiencies are most obvious when they take place during embryonic development, during adult life the amount of folate intake in the diet has been found to be related to epigenetic status in mammals [198, 199], including humans [200], and has also been related to methylation changes in colon cancer [201] and hyperhomocysteinemia [202]. Other dietary methyl donors are methionine, which is involved in the metabolic pathway of SAM and has been related to epigenetic-dependent hepatic disorders [203], and selenium, which is a dietary supplement capable of modifying the epigenetic status of prostate cancer cells and reducing both DNA and histone methylation levels and which has been suggested to improve cancer prevention through the activation of silenced genes [204]. In addition it has been described that deficiency in vitamin B12, another methyl donor, plays an important role in the adipocyte metabolism, and its deficiency leads to increased total cholesterol by limiting S-adenosylmethionine [205].

In relation to adults, caloric restriction (CR) is a dietary regimen based on a reduction in caloric intake which many studies have related to lifespan extension in various eukaryote organisms [206–212]. There are numerous studies supporting the notion that CR protects against many different diseases related to aging due to a reduction in oxidative stress and regulation of metabolic pathways [213–216]. The molecular mechanisms involved in this regulation are varied, and epigenetic marks could play an important role in the processes [217, 218]. For example, it is thought that CR might attenuate the epigenetic changes occurring during the progress of aging [219–221].

Bioactive dietary compounds such as polyphenols can alter developmental plasticity through the generation of

epigenetic changes and may also play a role in health and disease. In the last two decades much research has been focused on the mechanisms that could be responsible, at least in part, for the relationship between regular consumption of such bioactive compounds and the changes that may be produced in the epigenome and result in improvements in health and aging. This has given rise to the novel field of study known as epigenetic influence of nutrition [222]. Since numerous bioactive dietary compounds appear to have the potential to promote health or prevent diseases, such beneficial dietary supplements could be used to complement other therapies. Polyphenols, a structural class of organic chemicals characterized by the presence of large multiples of phenol structural units, are necessary in the human diet and can be found in fruits and vegetables [223]. Plant-origin polyphenols can be classified into different groups based on their chemical configuration and include flavonoids, stilbenes, phenolic acids, benzoquinones, acetophenones, lignins, and xanthenes [224]. Some authors estimate that more than 8000 distinct dietary polyphenols exist, including resveratrol (found in grapes), epigallocatechin-3-gallate (EGCG, in green tea), sulforaphane (SFN [1-isothiocyanato-4-(methylsulfinyl) butane], in broccoli), and curcumin (in turmeric) [225]. Although more studies are needed, the potential of green tea and broccoli in cancer chemoprevention seems to be mediated by epigenetic mechanisms, including DNMT and HDAC activity inhibition [226, 227].

3.3. Healthy and Unhealthy Habits. Although unhealthy lifestyle habits could be considered as a kind of environmental stressor, the fact that they depend on personal decisions has led us to describe them in a separate section.

It is well known that maternal tobacco smoke exposure (MTSE) is one of the most important risk factors during pregnancy for many diseases such as asthma, cancer, obesity, and type II diabetes [228–231]. MTSE is known to produce epigenetic changes that can affect birth-weight and fetal programming [232], specifically DNA methylation: there is global DNA hypomethylation and an increase in DNA promoter specific methylation in children exposed to prenatal smoking compared to children who were not [233]. In adults tobacco use has been related to an increase in promoter gene-specific DNA methylation, which in turn is linked to increased predisposition to diseases such as cancer [234–237]. To investigate in more depth the effect of tobacco smoking on DNA methylation, researchers have performed genome-wide DNA methylation analyses with the Illumina 450 K BeadChip. As a result, tobacco use has been related to changes in DNA methylation of CpG sites related to the development and function of the cellular, cardiovascular, detoxification, hematological, immune, tumorigenic, and reproduction systems [238–241]. Apart from DNA methylation, smoking also affects proper histone regulation across the polycomb repressive complex, coinciding with decreased H4k16ac and increased H3k27me3 [242]. Many substances contained in tobacco can also affect miRNAs. During development, a decrease in the expression of miRNAs, such as miR-16, miR-21, and miR-146a, have been related to nicotine and benzo(a)pyrene exposure in smoking mothers [243]. In adult smokers

miR-218 has been shown to be downregulated in bronchial epithelial cells [244]. Also, smoking has been associated with alterations in the expression of miRNAs such as miR-21, miR-34b, miR-125b, miR-146a, miR-223, and miR-340 [152]. Moreover, cigarette smoke condensate (CSC) causes aberrant overexpression of miR-31 in lung epithelium, and it could act as an oncomir promoting pulmonary carcinogenesis [245].

High alcohol consumption is also widely recognized to have many negative effects which lead to a deterioration in an individual's health. Alcohol can interfere with methionine metabolism through the inhibition of methionine synthase and, as a consequence, the long-term use of alcohol could lead to a decrease in the hepatocyte level of SAM [246]. In adult life, hypomethylation in LINE1 has been related to alcohol consumption in some tumors [247, 248]. Other studies have revealed that the use of alcohol alters DNA methylation patterns in hepatocarcinogenesis and neural stem cell differentiation [249, 250]. In addition, miR-125 and miR-126 downregulation has been observed in alcohol consumption related to hepatocellular carcinoma [251, 252].

Apart from DNA methylation and miRNA changes, ethanol induces gene activation through an increase in histone H3 and H4 acetylation and H3K4me3 [253–256], which may lead to immune system dysfunction [257]. Interestingly, epigenetic changes due to ethanol seem to be different depending on whether there is chronic or binge ethanol intake [253]. Furthermore, prenatal alcohol exposure also significantly affects the correct development of the fetus, including altering PcG/TrxG programming [258].

Substantial stress during early life can be a risk factor in the initial appearance of symptoms for individuals susceptible to bipolar disorder and other mental disorders [259]. Many studies have reported a relationship between early life stress and the aberrant DNA methylation of many genes such as the glucocorticoid receptor gene [260, 261] and the serotonin 1A receptor [262]. Also, stress is able to produce changes in histone modifications, such as increased levels of H3K4me3 and reduction of H3K9me3 levels in the dentate gyrus [263]. In this regard, there is an interesting study showing how prenatal maternal stress, generated by a natural disaster, was related to changes in DNA methylation patterns of blood cells, which could have an effect on the immune function of the offspring [264].

Physical exercise enhances or maintains physical fitness and is beneficial for human health in a number of ways. Little is known about the molecular mechanisms responsible, but several studies have shown that epigenetics is related to the effects of exercise on human health, since epigenetic changes in germ cells, skeletal muscle, and brain have been observed following a period of exercise [265–267].

3.4. Pharmacological Factors. Pharmacological treatment can also induce genome-wide epigenetic changes. Sodium valproate (VPA), a small fatty acid [268], has been widely studied. It is used to treat epilepsy, bipolar disorder, serious depression, migraine, and schizophrenia, as well as being used in cancer treatment and as a complementary treatment for latent HIV infection [269, 270]. Because of VPA's global HDAC inhibitor effect [271], it could possibly generate the

expression of some undesirable genes, and its side effects remain to be fully demonstrated.

Diethylstilbestrol (DES) is a “synthetic estrogen” which has been used for many years during pregnancy to prevent miscarriages and other pregnancy disorders but has been found to be associated with an increased risk of breast cancer and vaginal and cervical adenocarcinoma [272, 273]. It has been suggested that these side effects are mediated by epigenetic mechanisms, since it has been found that DES neonatal exposure in mice was related to decreased DNMT expression and alterations in DNA methylation in the mouse uterus [274]. In addition, it has been described that DES exposure in breast epithelial cells produces upregulation and downregulation of various miRNAs [275]. Moreover, miR-21 was consistently downregulated by DES exposure in MCF-7 breast cancer cell line [276].

Apart from these two drugs, there are many different medicines used on a daily basis that have recently been found to have epigenetic activity. For example, procaine, a local anesthetic, is now known to induce DNA demethylation [277]. And several antibiotics have also been implicated [278], such as pyrazinamide, a classic antituberculosis drug found to alter DNA methylation in the liver of treated rats, along with *LINE-1* hypomethylation and GSTP and *p16(INK4A)* promoter hypermethylation, all of which may be a side effect of its hepatic toxicity [279]. In addition, doxorubicin, an anthracycline antitumor antibiotic, has been found to inhibit DNMT1 and can induce apoptotic cell death [280]. miRNA profiles may also be altered by many different drugs used in therapy, for example, all-*trans*-retinoic acid in acute promyelocytic leukemia [281] and gemcitabine [282] and cisplatin [283] in ovarian cancer. Most importantly, perhaps, attention should be paid to the effects of medication on the germ cell epigenome and the potential transmission of epigenetic alterations to offspring. An example is where aberrant DNA methylation patterns were found in the sperm of patients treated with temozolomide, a chemotherapy drug used as a treatment for high-grade glioma [284].

4. Conclusions

The mammalian epigenome changes throughout embryonic development and with aging. Some of these changes are genetically programmed and others take place without any apparent function, though the molecular mechanisms involved are still to be elucidated. Moreover, what part of these changes is due to the interplay of environment with the epigenome and which is the result of individual genetics remain unknown. Further research needs to be focused on attempting to understand the causes of these changes in order to prevent the onset of diseases. In addition to all the environmental factors affecting the epigenome described in this review, we should take into account increased exposure to the nanomaterials and nanoparticles present in many everyday consumer goods, and consequently the study of the possible effects of this on the epigenome and human health will be a future, or rather is a current, challenge. The emergence of next-generation technologies will help us to answer some of these questions in the coming years.

Competing Interests

The authors declare that there is no conflict of interests.

Authors' Contributions

Estela G. Toraño and María G. García contributed equally to this review.

Acknowledgments

The authors thank Ronnie Lendrum for editorial assistance. Estela G. Toraño and María G. García received support from FICYT (BP11138 and BP13103). Juan Luis Fernández-Morera is supported by Sociedad Española de Diabetes. Agustín F. Fernández is sponsored by ISCIII-Subdirección General de Evaluación y Fomento de la Investigación (CP11/00131), and the Plan Nacional de I+D+I 2008/FEDER (PI11/01728). IUOPA is supported by the Obra Social Cajastur, Spain.

References

- [1] C. H. Waddington, "The epigenotype," *Endeavour*, vol. 1, pp. 18–20, 1942.
- [2] K. M. Godfrey, K. A. Lillycrop, G. C. Burdge, P. D. Gluckman, and M. A. Hanson, "Epigenetic mechanisms and the mismatch concept of the developmental origins of health and disease," *Pediatric Research*, vol. 61, pp. 5R–10R, 2007.
- [3] P. D. Gluckman, M. A. Hanson, and A. S. Beedle, "Non-genomic transgenerational inheritance of disease risk," *BioEssays*, vol. 29, no. 2, pp. 145–154, 2007.
- [4] R. Jaenisch and A. Bird, "Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals," *Nature Genetics*, vol. 33, pp. 245–254, 2003.
- [5] A. F. Fernández, E. G. Toraño, R. G. Urduñigo, A. G. Lana, I. A. Fernández, and M. F. Fraga, "The epigenetic basis of adaptation and responses to environmental change: perspective on human reproduction," *Advances in Experimental Medicine and Biology*, vol. 753, pp. 97–117, 2014.
- [6] M. K. Skinner, "Environmental epigenetics and a unified theory of the molecular aspects of evolution: a neo-Lamarckian concept that facilitates neo-Darwinian evolution," *Genome Biology and Evolution*, vol. 7, no. 5, pp. 1296–1302, 2015.
- [7] M. F. Fraga, "Genetic and epigenetic regulation of aging," *Current Opinion in Immunology*, vol. 21, no. 4, pp. 446–453, 2009.
- [8] E. I. Campos and D. Reinberg, "Histones: annotating chromatin," *Annual Review of Genetics*, vol. 43, pp. 559–599, 2009.
- [9] E. Li, "Chromatin modification and epigenetic reprogramming in mammalian development," *Nature Reviews Genetics*, vol. 3, no. 9, pp. 662–673, 2002.
- [10] B. E. Bernstein, A. Meissner, and E. S. Lander, "The mammalian epigenome," *Cell*, vol. 128, no. 4, pp. 669–681, 2007.
- [11] A. Portela and M. Esteller, "Epigenetic modifications and human disease," *Nature Biotechnology*, vol. 28, no. 10, pp. 1057–1068, 2010.
- [12] R. D. Hotchkiss, "The quantitative separation of purines, pyrimidines, and nucleosides by paper chromatography," *The Journal of Biological Chemistry*, vol. 175, no. 1, pp. 315–332, 1948.
- [13] E. Lara, V. Calvanese, A. F. Fernandez, and M. F. Fraga, "Techniques to study DNA methylation and histone modification," in *Epigenetic Aspects of Chronic Diseases*, pp. 21–39, Springer, London, UK, 2011.
- [14] P. M. Das and R. Singal, "DNA methylation and cancer," *Journal of Clinical Oncology*, vol. 22, no. 22, pp. 4632–4642, 2004.
- [15] M. Esteller, "Epigenetics in cancer," *The New England Journal of Medicine*, vol. 358, no. 11, pp. 1148–1159, 2008.
- [16] A. P. Feinberg and B. Tycko, "The history of cancer epigenetics," *Nature Reviews Cancer*, vol. 4, no. 2, pp. 143–153, 2004.
- [17] A. P. Bird, "CpG-rich islands and the function of DNA methylation," *Nature*, vol. 321, no. 6067, pp. 209–213, 1986.
- [18] M. Esteller, "Cancer epigenomics: DNA methylomes and histone-modification maps," *Nature Reviews Genetics*, vol. 8, no. 4, pp. 286–298, 2007.
- [19] A. P. Bird, "Gene number, noise reduction and biological complexity," *Trends in Genetics*, vol. 11, no. 3, pp. 94–100, 1995.
- [20] J. A. Yoder, N. S. Soman, G. L. Verdine, and T. H. Bestor, "DNA (cytosine-5)-methyltransferases in mouse cells and tissues. Studies with a mechanism-based probe," *Journal of Molecular Biology*, vol. 270, no. 3, pp. 385–395, 1997.
- [21] P. A. Jones and G. Liang, "Rethinking how DNA methylation patterns are maintained," *Nature Reviews Genetics*, vol. 10, no. 11, pp. 805–811, 2009.
- [22] J. G. Herman and S. B. Baylin, "Gene silencing in cancer in association with promoter hypermethylation," *The New England Journal of Medicine*, vol. 349, no. 21, pp. 2042–2054, 2003.
- [23] R. J. Klose and A. P. Bird, "Genomic DNA methylation: the mark and its mediators," *Trends in Biochemical Sciences*, vol. 31, no. 2, pp. 89–97, 2006.
- [24] P. A. Jones and S. B. Baylin, "The epigenomics of cancer," *Cell*, vol. 128, no. 4, pp. 683–692, 2007.
- [25] H. Han, C. C. Cortez, X. Yang, P. W. Nichols, P. A. Jones, and G. Liang, "DNA methylation directly silences genes with non-CpG island promoters and establishes a nucleosome occupied promoter," *Human Molecular Genetics*, vol. 20, no. 22, pp. 4299–4310, 2011.
- [26] X. Rao, J. Evans, H. Chae et al., "CpG island shore methylation regulates caveolin-1 expression in breast cancer," *Oncogene*, vol. 32, no. 38, pp. 4519–4528, 2013.
- [27] A. Doi, I.-H. Park, B. Wen et al., "Differential methylation of tissue- and cancer-specific CpG island shores distinguishes human induced pluripotent stem cells, embryonic stem cells and fibroblasts," *Nature Genetics*, vol. 41, no. 12, pp. 1350–1353, 2009.
- [28] R. A. Irizarry, C. Ladd-Acosta, B. Wen et al., "The human colon cancer methylome shows similar hypo- and hypermethylation at conserved tissue-specific CpG island shores," *Nature Genetics*, vol. 41, no. 2, pp. 178–186, 2009.
- [29] T. H. Bestor, "The DNA methyltransferases of mammals," *Human Molecular Genetics*, vol. 9, no. 16, pp. 2395–2402, 2000.
- [30] K. W. Kohn, M. I. Aladjem, J. N. Weinstein, and Y. Pommier, "Chromatin challenges during DNA replication: a systems representation," *Molecular Biology of the Cell*, vol. 19, no. 1, pp. 1–7, 2008.
- [31] M. Okano, D. W. Bell, D. A. Haber, and E. Li, "DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development," *Cell*, vol. 99, no. 3, pp. 247–257, 1999.

- [32] G.-D. Kim, J. Ni, N. Kelesoglu, R. J. Roberts, and S. Pradhan, "Co-operation and communication between the human maintenance and de novo DNA (cytosine-5) methyltransferases," *The EMBO Journal*, vol. 21, no. 15, pp. 4183–4195, 2002.
- [33] Y. Hirabayashi and Y. Gotoh, "Epigenetic control of neural precursor cell fate during development," *Nature Reviews Neuroscience*, vol. 11, no. 6, pp. 377–388, 2010.
- [34] V. Calvanese, A. F. Fernández, R. G. Urdinguio et al., "A promoter DNA demethylation landscape of human hematopoietic differentiation," *Nucleic Acids Research*, vol. 40, no. 1, pp. 116–131, 2012.
- [35] J. Catania and D. S. Fairweather, "DNA methylation and cellular ageing," *Mutation Research/DNAging*, vol. 256, no. 2–6, pp. 283–293, 1991.
- [36] P. A. Jones, "DNA methylation and cancer," *Cancer Research*, vol. 46, no. 2, pp. 461–466, 1986.
- [37] V. L. Wilson and P. A. Jones, "DNA methylation decreases in aging but not in immortal cells," *Science*, vol. 220, no. 4601, pp. 1055–1057, 1983.
- [38] C. Sidler, R. Woycicki, I. Kovalchuk, and O. Kovalchuk, "WI-38 senescence is associated with global and site-specific hypomethylation," *Aging*, vol. 6, no. 7, pp. 564–574, 2014.
- [39] J. P. Issa, "Aging, DNA methylation and cancer," *Critical Reviews in Oncology/Hematology*, vol. 32, no. 1, pp. 31–43, 1999.
- [40] K. D. Robertson and P. A. Jones, "DNA methylation: past, present and future directions," *Carcinogenesis*, vol. 21, no. 3, pp. 461–467, 2000.
- [41] S. Han and A. Brunet, "Histone methylation makes its mark on longevity," *Trends in Cell Biology*, vol. 22, no. 1, pp. 42–49, 2012.
- [42] S. Kim, C. G. Parks, Z. Xu et al., "Association between genetic variants in DNA and histone methylation and telomere length," *PLoS ONE*, vol. 7, no. 7, article e40504, 2012.
- [43] T. Kouzarides, "Chromatin modifications and their function," *Cell*, vol. 128, no. 4, pp. 693–705, 2007.
- [44] Y. I. Hassan and J. Zempleni, "A novel, enigmatic histone modification: biotinylation of histones by holocarboxylase synthetase," *Nutrition Reviews*, vol. 66, no. 12, pp. 721–725, 2008.
- [45] V. W. Zhou, A. Goren, and B. E. Bernstein, "Charting histone modifications and the functional organization of mammalian genomes," *Nature Reviews Genetics*, vol. 12, no. 1, pp. 7–18, 2011.
- [46] T. Jenuwein and C. D. Allis, "Translating the histone code," *Science*, vol. 293, no. 5532, pp. 1074–1080, 2001.
- [47] R. Karlič, H.-R. Chung, J. Lasserre, K. Vlahoviček, and M. Vingron, "Histone modification levels are predictive for gene expression," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 7, pp. 2926–2931, 2010.
- [48] D. Huertas, R. Sendra, and P. Muñoz, "Chromatin dynamics coupled to DNA repair," *Epigenetics*, vol. 4, no. 1, pp. 31–42, 2009.
- [49] S. Keating and A. El-Osta, "Transcriptional regulation by the Set7 lysine methyltransferase," *Epigenetics*, vol. 8, no. 4, 2013.
- [50] E. Birney, J. A. Stamatoyannopoulos, A. Dutta et al., "Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project," *Nature*, vol. 447, no. 7146, pp. 799–816, 2007.
- [51] T.-Y. Roh, S. Cuddapah, and K. Zhao, "Active chromatin domains are defined by acetylation islands revealed by genome-wide mapping," *Genes and Development*, vol. 19, no. 5, pp. 542–552, 2005.
- [52] J. C. Rice and C. D. Allis, "Histone methylation versus histone acetylation: new insights into epigenetic regulation," *Current Opinion in Cell Biology*, vol. 13, no. 3, pp. 263–273, 2001.
- [53] E. J. Richards and S. C. R. Elgin, "Epigenetic codes for heterochromatin formation and silencing: rounding up the usual suspects," *Cell*, vol. 108, no. 4, pp. 489–500, 2002.
- [54] A. Izzo and R. Schneider, "Chatting histone modifications in mammals," *Briefings in Functional Genomics*, vol. 9, no. 5–6, pp. 429–443, 2010.
- [55] M. Nikolov and W. Fischle, "Systematic analysis of histone modification readout," *Molecular BioSystems*, vol. 9, no. 2, pp. 182–194, 2013.
- [56] B. D. Strahl and C. D. Allis, "The language of covalent histone modifications," *Nature*, vol. 403, no. 6765, pp. 41–45, 2000.
- [57] M. A. Dawson, R. K. Prinjha, A. Dittmann et al., "Inhibition of BET recruitment to chromatin as an effective treatment for MLL-fusion leukaemia," *Nature*, vol. 478, no. 7370, pp. 529–533, 2011.
- [58] J. E. Delmore, G. C. Issa, M. E. Lemieux et al., "BET bromodomain inhibition as a therapeutic strategy to target c-Myc," *Cell*, vol. 146, no. 6, pp. 904–917, 2011.
- [59] J. Zuber, J. Shi, E. Wang et al., "RNAi screen identifies Brd4 as a therapeutic target in acute myeloid leukaemia," *Nature*, vol. 478, no. 7370, pp. 524–528, 2011.
- [60] R. L. Jirtle and M. K. Skinner, "Environmental epigenomics and disease susceptibility," *Nature Reviews Genetics*, vol. 8, no. 4, pp. 253–262, 2007.
- [61] F. Miao, Z. Chen, L. Zhang et al., "Profiles of epigenetic histone post-translational modifications at type 1 diabetes susceptible genes," *Journal of Biological Chemistry*, vol. 287, no. 20, pp. 16335–16345, 2012.
- [62] M. F. Fraga, E. Ballestar, A. Villar-Garea et al., "Loss of acetylation at Lys16 and trimethylation at Lys20 of histone H4 is a common hallmark of human cancer," *Nature Genetics*, vol. 37, no. 4, pp. 391–400, 2005.
- [63] P. Zhu, E. Martin, J. Mengwasser, P. Schlag, K.-P. Janssen, and M. Göttlicher, "Induction of HDAC2 expression upon loss of APC in colorectal tumorigenesis," *Cancer Cell*, vol. 5, no. 5, pp. 455–463, 2004.
- [64] S. Ropero, M. F. Fraga, E. Ballestar et al., "A truncating mutation of HDAC2 in human cancers confers resistance to histone deacetylase inhibition," *Nature Genetics*, vol. 38, no. 5, pp. 566–569, 2006.
- [65] P. Chi, C. D. Allis, and G. G. Wang, "Covalent histone modifications-miswritten, misinterpreted and mis-erased in human cancers," *Nature Reviews Cancer*, vol. 10, no. 7, pp. 457–469, 2010.
- [66] G. L. Dalgliesh, K. Furge, C. Greenman et al., "Systematic sequencing of renal carcinoma reveals inactivation of histone modifying genes," *Nature*, vol. 463, no. 7279, pp. 360–363, 2010.
- [67] R. J. Pierce, F. Dubois-Abdesselem, J. Lancelot, L. Andrade, and G. Oliveira, "Targeting schistosome histone modifying enzymes for drug development," *Current Pharmaceutical Design*, vol. 18, no. 24, pp. 3567–3578, 2012.
- [68] F. J. Dekker and H. J. Haisma, "Histone acetyl transferases as emerging drug targets," *Drug Discovery Today*, vol. 14, no. 19–20, pp. 942–948, 2009.
- [69] P. A. Cole, "Chemical probes for histone-modifying enzymes," *Nature Chemical Biology*, vol. 4, no. 10, pp. 590–597, 2008.
- [70] M. A. Reddy and R. Natarajan, "Epigenetic mechanisms in diabetic vascular complications," *Cardiovascular Research*, vol. 90, no. 3, pp. 421–429, 2011.
- [71] A. B. Sanz, M. D. Sanchez-Niño, A. M. Ramos et al., "NF- κ B in renal inflammation," *Journal of the American Society of Nephrology*, vol. 21, no. 8, pp. 1254–1262, 2010.

- [72] M. Jakovcevski and S. Akbarian, "Epigenetic mechanisms in neurological disease," *Nature Medicine*, vol. 18, no. 8, pp. 1194–1204, 2012.
- [73] M. Esteller, "Non-coding RNAs in human disease," *Nature Reviews Genetics*, vol. 12, no. 12, pp. 861–874, 2011.
- [74] T. R. Mercer, M. E. Dinger, and J. S. Mattick, "Long non-coding RNAs: insights into functions," *Nature Reviews Genetics*, vol. 10, no. 3, pp. 155–159, 2009.
- [75] B. De Felice, F. Manfellotto, A. Palumbo et al., "Genome-wide microRNA expression profiling in placentas from pregnant women exposed to BPA," *BMC Medical Genomics*, vol. 8, no. 1, article 56, 2015.
- [76] S. Valadkhan, "lncRNAs in stress response," in *Long Non-Coding RNAs in Human Disease*, Current Topics in Microbiology and Immunology, Springer, 2015.
- [77] P. G. Hawkins and K. V. Morris, "RNA and transcriptional modulation of gene expression," *Cell Cycle*, vol. 7, no. 5, pp. 602–607, 2008.
- [78] N. Vadaie and K. V. Morris, "Long antisense non-coding RNAs and the epigenetic regulation of gene expression," *Biomolecular Concepts*, vol. 4, no. 4, pp. 411–415, 2013.
- [79] K. V. Morris, "Long antisense non-coding RNAs function to direct epigenetic complexes that regulate transcription in human cells," *Epigenetics*, vol. 4, no. 5, pp. 296–301, 2009.
- [80] A. Congrains, K. Kamide, M. Ohishi, and H. Rakugi, "ANRIL: molecular mechanisms and implications in human health," *International Journal of Molecular Sciences*, vol. 14, no. 1, pp. 1278–1292, 2013.
- [81] W. Yu, D. Gius, P. Onyango et al., "Epigenetic silencing of tumour suppressor gene p15 by its antisense RNA," *Nature*, vol. 451, no. 7175, pp. 202–206, 2008.
- [82] K. L. Yap, S. Li, A. M. Muñoz-Cabello et al., "Molecular interplay of the noncoding RNA ANRIL and methylated histone H3 Lysine 27 by polycomb CBX7 in transcriptional silencing of INK4a," *Molecular Cell*, vol. 38, no. 5, pp. 662–674, 2010.
- [83] K. Ikegami, J. Ohgane, S. Tanaka, S. Yagi, and K. Shiota, "Interplay between DNA methylation, histone modification and chromatin remodeling in stem cells and during development," *International Journal of Developmental Biology*, vol. 53, no. 2–3, pp. 203–214, 2009.
- [84] A. Lujambio, S. Ropero, E. Ballestar et al., "Genetic unmasking of an epigenetically silenced microRNA in human cancer cells," *Cancer Research*, vol. 67, no. 4, pp. 1424–1429, 2007.
- [85] H. Yin, P. Song, R. Su et al., "DNA Methylation mediated down-regulating of MicroRNA-33b and its role in gastric cancer," *Scientific Reports*, vol. 6, Article ID 18824, 2016.
- [86] W. Reik, "Stability and flexibility of epigenetic gene regulation in mammalian development," *Nature*, vol. 447, no. 7143, pp. 425–432, 2007.
- [87] R. Feil and M. F. Fraga, "Epigenetics and the environment: emerging patterns and implications," *Nature Reviews Genetics*, vol. 13, no. 2, pp. 97–109, 2012.
- [88] C. Huidobro, A. F. Fernandez, and M. F. Fraga, "Aging epigenetics: causes and consequences," *Molecular Aspects of Medicine*, vol. 34, no. 4, pp. 765–781, 2013.
- [89] Z. A. Kaminsky, T. Tang, S.-C. Wang et al., "DNA methylation profiles in monozygotic and dizygotic twins," *Nature Genetics*, vol. 41, no. 2, pp. 240–245, 2009.
- [90] P. A. Rzeczowska, H. Hou, M. D. Wilson, and M. R. Palmert, "Epigenetics: a new player in the regulation of mammalian puberty," *Neuroendocrinology*, vol. 99, pp. 139–155, 2014.
- [91] G. Vogt, M. Huber, M. Thiemann, G. van den Boogaart, O. J. Schmitz, and C. D. Schubart, "Production of different phenotypes from the same genotype in the same environment by developmental variation," *Journal of Experimental Biology*, vol. 211, no. 4, pp. 510–523, 2008.
- [92] A. P. Feinberg, "Phenotypic plasticity and the epigenetics of human disease," *Nature*, vol. 447, no. 7143, pp. 433–440, 2007.
- [93] P. D. Gluckman, M. A. Hanson, T. Buklijas, F. M. Low, and A. S. Beedle, "Epigenetic mechanisms that underpin metabolic and cardiovascular diseases," *Nature Reviews Endocrinology*, vol. 5, no. 7, pp. 401–408, 2009.
- [94] O. Aguilera, A. F. Fernández, A. Muñoz, and M. F. Fraga, "Epigenetics and environment: a complex relationship," *Journal of Applied Physiology*, vol. 109, no. 1, pp. 243–251, 2010.
- [95] M. Mitchell, H. W. Bakos, and M. Lane, "Paternal diet-induced obesity impairs embryo development and implantation in the mouse," *Fertility and Sterility*, vol. 95, no. 4, pp. 1349–1353, 2011.
- [96] N. K. Binder, M. Mitchell, and D. K. Gardner, "Parental diet-induced obesity leads to retarded early mouse embryo development and altered carbohydrate utilisation by the blastocyst," *Reproduction, Fertility and Development*, vol. 24, no. 6, pp. 804–812, 2012.
- [97] N. K. Binder, N. J. Hannan, and D. K. Gardner, "Paternal diet-induced obesity retards early mouse embryo development, mitochondrial activity and pregnancy health," *PLoS ONE*, vol. 7, no. 12, Article ID e52304, 2012.
- [98] A. Baccarelli and V. Bollati, "Epigenetics and environmental chemicals," *Current Opinion in Pediatrics*, vol. 21, no. 2, pp. 243–251, 2009.
- [99] F. Perera and J. Herbstman, "Prenatal environmental exposures, epigenetics, and disease," *Reproductive Toxicology*, vol. 31, no. 3, pp. 363–373, 2011.
- [100] D. K. Nordstrom, "Worldwide occurrences of arsenic in ground water," *Science*, vol. 296, no. 5576, pp. 2143–2145, 2002.
- [101] C. O. Abernathy, Y.-P. Liu, D. Longfellow et al., "Meeting on Arsenic: Health Effects, Mechanisms of Actions, and Research Issues, Hunt Valley, Maryland, 22–24 September 1997," *Environmental Health Perspectives*, vol. 107, no. 7, pp. 593–597, 1999.
- [102] M. Rahman, M. Tondel, I. A. Chowdhury, and O. Axelson, "Relations between exposure to arsenic, skin lesions, and glucosuria," *Occupational and Environmental Medicine*, vol. 56, no. 4, pp. 277–281, 1999.
- [103] R. R. Engel, C. Hopenhayn-Rich, O. Receveur, and A. H. Smith, "Vascular effects of chronic arsenic exposure: a review," *Epidemiologic Reviews*, vol. 16, no. 2, pp. 184–209, 1994.
- [104] J. I. Anetor, H. Wanibuchi, and S. Fukushima, "Arsenic exposure and its health effects and risk of cancer in developing countries: micronutrients as host defence," *Asian Pacific Journal of Cancer Prevention*, vol. 8, no. 1, pp. 13–23, 2007.
- [105] M. N. Bates, A. H. Smith, and C. Hopenhayn-Rich, "Arsenic ingestion and internal cancers: a review," *American Journal of Epidemiology*, vol. 135, no. 5, pp. 462–476, 1992.
- [106] J. Brinkel, M. H. Khan, and A. Kraemer, "A systematic review of arsenic exposure and its social and mental health effects with special reference to Bangladesh," *International Journal of Environmental Research and Public Health*, vol. 6, no. 5, pp. 1609–1619, 2009.
- [107] C. Q. Zhao, M. R. Young, B. A. Diwan, T. P. Coogan, and M. P. Waalkes, "Association of arsenic-induced malignant transformation with DNA hypomethylation and aberrant gene expression," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 20, pp. 10907–10912, 1997.

- [108] L. Benbrahim-Tallaa, R. A. Waterland, M. Styblo, W. E. Achanzar, M. M. Webber, and M. P. Waalkes, "Molecular events associated with arsenic-induced malignant transformation of human prostatic epithelial cells: aberrant genomic DNA methylation and K-ras oncogene activation," *Toxicology and Applied Pharmacology*, vol. 206, no. 3, pp. 288–298, 2005.
- [109] X. Cui, T. Wakai, Y. Shirai, K. Hatakeyama, and S. Hirano, "Chronic oral exposure to inorganic arsenate interferes with methylation status of p16INK4a and RASSF1A and induces lung cancer in A/J mice," *Toxicological Sciences*, vol. 91, no. 2, pp. 372–381, 2006.
- [110] Y. Chervona, A. Arita, and M. Costa, "Carcinogenic metals and the epigenome: understanding the effect of nickel, arsenic, and chromium," *Metallomics*, vol. 4, no. 7, pp. 619–627, 2012.
- [111] C. R. Tyler, A. K. Hafez, E. R. Solomon, and A. M. Allan, "Developmental exposure to 50 parts-per-billion arsenic influences histone modifications and associated epigenetic machinery in a region- and sex-specific manner in the adult mouse brain," *Toxicology and Applied Pharmacology*, vol. 288, no. 1, pp. 40–51, 2015.
- [112] C. R. Tyler, J. A. Weber, M. Labrecque, J. M. Hessinger, J. S. Edwards, and A. M. Allan, "ChIP-Seq analysis of the adult male mouse brain after developmental exposure to arsenic," *Data in Brief*, vol. 5, pp. 248–254, 2015.
- [113] X. Zhou, H. Sun, T. P. Ellen, H. Chen, and M. Costa, "Arsenite alters global histone H3 methylation," *Carcinogenesis*, vol. 29, no. 9, pp. 1831–1836, 2008.
- [114] A. Cardenas, D. C. Koestler, E. A. Houseman et al., "Differential DNA methylation in umbilical cord blood of infants exposed to mercury and arsenic in utero," *Epigenetics*, vol. 10, no. 6, pp. 508–515, 2015.
- [115] M. L. Kile, E. A. Houseman, A. A. Baccarelli et al., "Effect of prenatal arsenic exposure on DNA methylation and leukocyte subpopulations in cord blood," *Epigenetics*, vol. 9, no. 5, pp. 774–782, 2014.
- [116] M. Argos, L. Chen, F. Jasmine et al., "Gene-specific differential DNA methylation and chronic arsenic exposure in an epigenome-wide association study of adults in Bangladesh," *Environmental Health Perspectives*, vol. 123, no. 1, pp. 64–71, 2015.
- [117] B. B. Green, M. R. Karagas, T. Punshon et al., "Epigenome-wide assessment of DNA methylation in the placenta and arsenic exposure in the new hampshire birth cohort study (USA)," *Environmental Health Perspectives*, 2016.
- [118] S. M. Tajuddin, A. F. S. Amaral, A. F. Fernández et al., "Genetic and non-genetic predictors of LINE-1 methylation in leukocyte DNA," *Environmental Health Perspectives*, vol. 121, no. 6, pp. 650–656, 2013.
- [119] C. S. Wilhelm, K. T. Kelsey, R. Butler et al., "Implications of LINE1 methylation for bladder cancer risk in women," *Clinical Cancer Research*, vol. 16, no. 5, pp. 1682–1689, 2010.
- [120] G. Lu, H. Xu, D. Chang et al., "Arsenic exposure is associated with DNA hypermethylation of the tumor suppressor gene p16," *Journal of Occupational Medicine and Toxicology*, vol. 9, no. 1, article 42, 2014.
- [121] C. J. Marsit, M. R. Karagas, A. Schned, and K. T. Kelsey, "Carcinogen exposure and epigenetic silencing in bladder cancer," *Annals of the New York Academy of Sciences*, vol. 1076, pp. 810–821, 2006.
- [122] Y. Chervona, M. N. Hall, A. Arita et al., "Associations between arsenic exposure and global posttranslational histone modifications among adults in Bangladesh," *Cancer Epidemiology Biomarkers and Prevention*, vol. 21, no. 12, pp. 2252–2260, 2012.
- [123] J. F. Reichard, M. Schnekenburger, and A. Puga, "Long term low-dose arsenic exposure induces loss of DNA methylation," *Biochemical and Biophysical Research Communications*, vol. 352, no. 1, pp. 188–192, 2007.
- [124] L. Järup and A. Åkesson, "Current status of cadmium as an environmental health problem," *Toxicology and Applied Pharmacology*, vol. 238, no. 3, pp. 201–208, 2009.
- [125] M. Kippler, F. Tofail, R. Gardner et al., "Maternal cadmium exposure during pregnancy and size at birth: a prospective cohort study," *Environmental Health Perspectives*, vol. 120, no. 2, pp. 284–289, 2012.
- [126] A. P. Sanders, L. Smeester, D. Rojas et al., "Cadmium exposure and the epigenome: exposure-associated patterns of DNA methylation in leukocytes from mother-baby pairs," *Epigenetics*, vol. 9, no. 2, pp. 212–221, 2014.
- [127] M. B. Hossain, M. Vahter, G. Concha, and K. Broberg, "Low-level environmental cadmium exposure is associated with DNA hypomethylation in Argentinean women," *Environmental Health Perspectives*, vol. 120, no. 6, pp. 879–884, 2012.
- [128] R. Levin, M. J. Brown, M. E. Kashtock et al., "Lead exposures in U.S. Children, 2008: implications for prevention," *Environmental Health Perspectives*, vol. 116, no. 10, pp. 1285–1293, 2008.
- [129] M. A. Pokras and M. R. Kneeland, "Lead poisoning: using transdisciplinary approaches to solve an ancient problem," *EcoHealth*, vol. 5, no. 3, pp. 379–385, 2008.
- [130] C.-C. Lin, Y.-C. Chen, F.-C. Su et al., "In utero exposure to environmental lead and manganese and neurodevelopment at 2 years of age," *Environmental Research*, vol. 123, pp. 52–57, 2013.
- [131] D. C. Bellinger, "Lead neurotoxicity and socioeconomic status: conceptual and analytical issues," *NeuroToxicology*, vol. 29, no. 5, pp. 828–832, 2008.
- [132] M.-C. Senut, A. Sen, P. Cingolani, A. Shaik, S. J. Land, and D. M. Ruden, "Lead exposure disrupts global DNA methylation in human embryonic stem cells and alters their neuronal differentiation," *Toxicological Sciences*, vol. 139, no. 1, pp. 142–161, 2014.
- [133] J. R. Pilsner, H. Hu, A. Ettinger et al., "Influence of prenatal lead exposure on genomic methylation of cord blood DNA," *Environmental Health Perspectives*, vol. 117, no. 9, pp. 1466–1471, 2009.
- [134] R. O. Wright, J. Schwartz, R. J. Wright et al., "Biomarkers of lead exposure and DNA methylation within retrotransposons," *Environmental Health Perspectives*, vol. 118, no. 6, pp. 790–795, 2010.
- [135] A. Sen, N. Heredia, M.-C. Senut et al., "Early life lead exposure causes gender-specific changes in the DNA methylation profile of DNA extracted from dried blood spots," *Epigenomics*, vol. 7, no. 3, pp. 379–393, 2015.
- [136] C. Li, X. Yang, M. Xu, J. Zhang, and N. Sun, "Epigenetic marker (LINE-1 promoter) methylation level was associated with occupational lead exposure," *Clinical Toxicology*, vol. 51, no. 4, pp. 225–229, 2013.
- [137] C. W. Hanna, M. S. Bloom, W. P. Robinson et al., "DNA methylation changes in whole blood is associated with exposure to the environmental contaminants, mercury, lead, cadmium and bisphenol A, in women undergoing ovarian stimulation for IVF," *Human Reproduction*, vol. 27, no. 5, pp. 1401–1410, 2012.

- [138] Y.-S. Hong, Y.-M. Kim, and K.-E. Lee, "Methylmercury exposure and health effects," *Journal of Preventive Medicine and Public Health*, vol. 45, no. 6, pp. 353–363, 2012.
- [139] M. R. Karagas, A. L. Choi, E. Oken et al., "Evidence on the human health effects of low-level methylmercury exposure," *Environmental Health Perspectives*, vol. 120, no. 6, pp. 799–806, 2012.
- [140] J.-D. Park and W. Zheng, "Human exposure and health effects of inorganic and elemental mercury," *Journal of Preventive Medicine and Public Health*, vol. 45, no. 6, pp. 344–352, 2012.
- [141] R. Bose, N. Onishchenko, K. Edoff, A. M. Janson Lang, and S. Ceccatelli, "Inherited effects of low-dose exposure to methylmercury in neural stem cells," *Toxicological Sciences*, vol. 130, no. 2, pp. 383–390, 2012.
- [142] K. M. Bakulski, H. J. Lee, J. I. Feinberg et al., "Prenatal mercury concentration is associated with changes in DNA methylation at *TCEANC2* in newborns," *International Journal of Epidemiology*, vol. 44, no. 4, pp. 1249–1262, 2015.
- [143] J. M. Goodrich, N. Basu, A. Franzblau, and D. C. Dolinoy, "Mercury biomarkers and DNA methylation among michigan dental professionals," *Environmental and Molecular Mutagenesis*, vol. 54, no. 3, pp. 195–203, 2013.
- [144] H. Chen, Q. Ke, T. Kluz, Y. Yan, and M. Costa, "Nickel ions increase histone H3 lysine 9 dimethylation and induce transgene silencing," *Molecular and Cellular Biology*, vol. 26, no. 10, pp. 3728–3737, 2006.
- [145] X. Zhou, Q. Li, A. Arita, H. Sun, and M. Costa, "Effects of nickel, chromate, and arsenite on histone 3 lysine methylation," *Toxicology and Applied Pharmacology*, vol. 236, no. 1, pp. 78–84, 2009.
- [146] H.-W. Ryu, D. H. Lee, H.-R. Won, K. H. Kim, Y. J. Seong, and S. H. Kwon, "Influence of toxicologically relevant metals on human epigenetic regulation," *Toxicological Research*, vol. 31, no. 1, pp. 1–9, 2015.
- [147] A. P. Sanders, H. H. Burris, A. C. Just et al., "Altered miRNA expression in the cervix during pregnancy associated with lead and mercury exposure," *Epigenomics*, vol. 7, no. 6, pp. 885–896, 2015.
- [148] J. Lepeule, M.-A. C. Bind, A. A. Baccarelli et al., "Epigenetic influences on associations between air pollutants and lung function in elderly men: the normative aging study," *Environmental Health Perspectives*, vol. 122, no. 6, pp. 566–572, 2014.
- [149] L. Hou, X. Zhang, Y. Zheng et al., "Altered methylation in tandem repeat element and elemental component levels in inhalable air particles," *Environmental and Molecular Mutagenesis*, vol. 55, no. 3, pp. 256–265, 2014.
- [150] L. Tarantini, M. Bonzini, P. Apostoli et al., "Effects of particulate matter on genomic DNA methylation content and iNOS promoter methylation," *Environmental Health Perspectives*, vol. 117, no. 2, pp. 217–222, 2009.
- [151] A. Baccarelli and S. Ghosh, "Environmental exposures, epigenetics and cardiovascular disease," *Current Opinion in Clinical Nutrition and Metabolic Care*, vol. 15, no. 4, pp. 323–329, 2012.
- [152] K. Vrijens, V. Bollati, and T. S. Nawrot, "MicroRNAs as potential signatures of environmental exposure or effect: a systematic review," *Environmental Health Perspectives*, vol. 123, no. 5, pp. 399–411, 2015.
- [153] M. J. Jardim, R. C. Fry, I. Jaspers, L. Dailey, and D. Diaz-Sanchez, "Disruption of MicroRNA expression in human airway cells by diesel exhaust particles is linked to tumorigenesis-associated pathways," *Environmental Health Perspectives*, vol. 117, no. 11, pp. 1745–1751, 2009.
- [154] B. C. Christensen, E. A. Houseman, C. J. Marsit et al., "Aging and environmental exposures alter tissue-specific DNA methylation dependent upon CPG island context," *PLoS Genetics*, vol. 5, no. 8, Article ID e1000602, 2009.
- [155] J. A. Tsou, L. Y. C. Shen, K. D. Siegmund et al., "Distinct DNA methylation profiles in malignant mesothelioma, lung adenocarcinoma, and non-tumor lung," *Lung Cancer*, vol. 47, no. 2, pp. 193–204, 2005.
- [156] B. C. Christensen, J. J. Godleski, C. J. Marsit et al., "Asbestos exposure predicts cell cycle control gene promoter methylation in pleural mesothelioma," *Carcinogenesis*, vol. 29, no. 8, pp. 1555–1559, 2008.
- [157] Y. Y. Cheng, M. B. Kirschner, N. C. Cheng et al., "ZIC1 is silenced and has tumor suppressor function in malignant pleural mesothelioma," *Journal of Thoracic Oncology*, vol. 8, no. 10, pp. 1317–1328, 2013.
- [158] V. Bollati, A. Baccarelli, L. Hou et al., "Changes in DNA methylation patterns in subjects exposed to low-dose benzene," *Cancer Research*, vol. 67, no. 3, pp. 876–880, 2007.
- [159] A. M. Melnick, "Epigenetics in AML," *Best Practice and Research: Clinical Haematology*, vol. 23, no. 4, pp. 463–468, 2010.
- [160] M. K. Skinner, M. Manikkam, and C. Guerrero-Bosagna, "Epigenetic transgenerational actions of endocrine disruptors," *Reproductive Toxicology*, vol. 31, no. 3, pp. 337–343, 2011.
- [161] M. D. Shelby, "NTP-CERHR monograph on the potential human reproductive and developmental effects of di (2-ethylhexyl) phthalate (DEHP)," *NTP CERHR MON*, no. 18, 2006.
- [162] M. D. Shelby, "NTP-CERHR monograph on the potential human reproductive and developmental effects of bisphenol A," *NTP CERHR MON*, vol. 22, no. 5, pp. 7–64, 2008.
- [163] R. U. Halden, "Plastics and health risks," *Annual Review of Public Health*, vol. 31, pp. 179–194, 2010.
- [164] R. R. Newbold, W. N. Jefferson, and E. Padilla-Banks, "Prenatal Exposure to Bisphenol A at environmentally relevant doses adversely affects the murine female reproductive tract later in life," *Environmental Health Perspectives*, vol. 117, no. 6, pp. 879–885, 2009.
- [165] M. Durando, L. Kass, J. Piva et al., "Prenatal bisphenol A exposure induces preneoplastic lesions in the mammary gland in Wistar rats," *Environmental Health Perspectives*, vol. 115, no. 1, pp. 80–86, 2007.
- [166] O. S. Anderson, M. S. Nahar, C. Faulk et al., "Epigenetic responses following maternal dietary exposure to physiologically relevant levels of bisphenol A," *Environmental and Molecular Mutagenesis*, vol. 53, no. 5, pp. 334–342, 2012.
- [167] D. C. Dolinoy, D. Huang, and R. L. Jirtle, "Maternal nutrient supplementation counteracts bisphenol A-induced DNA hypomethylation in early development," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 32, pp. 13056–13061, 2007.
- [168] M. Miao, X. Zhou, Y. Li et al., "LINE-1 hypomethylation in spermatozoa is associated with Bisphenol A exposure," *Andrology*, vol. 2, no. 1, pp. 138–144, 2014.
- [169] J. H. Kim, L. S. Rozek, A. S. Soliman et al., "Bisphenol A-associated epigenomic changes in prepubescent girls: a cross-sectional study in Gharbiah, Egypt," *Environmental Health: A Global Access Science Source*, vol. 12, article 33, 2013.
- [170] Y.-I. Weng, P.-Y. Hsu, S. Liyanarachchi et al., "Epigenetic influences of low-dose bisphenol A in primary human breast epithelial cells," *Toxicology and Applied Pharmacology*, vol. 248, no. 2, pp. 111–121, 2010.

- [171] W. Y. Tang, L. M. Morey, Y. Y. Cheung, L. Birch, G. S. Prins, and S. M. Ho, "Neonatal exposure to estradiol/bisphenol A alters promoter methylation and expression of Nbspl and Hpcall genes and transcriptional programs of Dnmt3a/b and Mbd2/4 in the rat prostate gland throughout life," *Endocrinology*, vol. 153, no. 1, pp. 42–55, 2012.
- [172] Y. Jiang, W. Xia, J. Yang et al., "BPA-induced DNA hypermethylation of the master mitochondrial gene PGC-1 α contributes to cardiomyopathy in male rats," *Toxicology*, vol. 329, pp. 21–31, 2015.
- [173] Z. Mao, W. Xia, H. Chang, W. Huo, Y. Li, and S. Xu, "Paternal BPA exposure in early life alters Igf2 epigenetic status in sperm and induces pancreatic impairment in rat offspring," *Toxicology Letters*, vol. 238, no. 3, pp. 30–38, 2015.
- [174] M. Susiarjo, I. Sasson, C. Mesaros, and M. S. Bartolomei, "Bisphenol A exposure disrupts genomic imprinting in the mouse," *PLoS Genetics*, vol. 9, no. 4, Article ID e1003401, 2013.
- [175] L. F. Doherty, J. G. Bromer, Y. Zhou, T. S. Aldad, and H. S. Taylor, "In utero exposure to diethylstilbestrol (DES) or bisphenol-A (BPA) increases EZH2 expression in the mammary gland: an epigenetic mechanism linking endocrine disruptors to breast cancer," *Hormones and Cancer*, vol. 1, no. 3, pp. 146–155, 2010.
- [176] C. Jhappan, F. P. Noonan, and G. Merlino, "Ultraviolet radiation and cutaneous malignant melanoma," *Oncogene*, vol. 22, no. 20, pp. 3099–3112, 2003.
- [177] D. L. Narayanan, R. N. Saladi, and J. L. Fox, "Ultraviolet radiation and skin cancer," *International Journal of Dermatology*, vol. 49, no. 9, pp. 978–986, 2010.
- [178] X. Zhang, B. S. Rosenstein, Y. Wang, M. Lebowohl, and H. Wei, "Identification of possible reactive oxygen species involved in ultraviolet radiation-induced oxidative DNA damage," *Free Radical Biology and Medicine*, vol. 23, no. 7, pp. 980–985, 1997.
- [179] S. K. Katiyar, "UV-induced immune suppression and photocarcinogenesis: chemoprevention by dietary botanical agents," *Cancer Letters*, vol. 255, no. 1, pp. 1–11, 2007.
- [180] M. Ichihashi, M. Ueda, A. Budiyanto et al., "UV-induced skin damage," *Toxicology*, vol. 189, no. 1–2, pp. 21–39, 2003.
- [181] V. Nandakumar, M. Vaid, T. O. Tollefsbol, and S. K. Katiyar, "Aberrant DNA hypermethylation patterns lead to transcriptional silencing of tumor suppressor genes in UVB-exposed skin and UVB-induced skin tumors of mice," *Carcinogenesis*, vol. 32, no. 4, pp. 597–604, 2011.
- [182] S. K. Katiyar, T. Singh, R. Prasad, Q. Sun, and M. Vaid, "Epigenetic alterations in ultraviolet radiation-induced skin carcinogenesis: interaction of bioactive dietary components on epigenetic targets," *Photochemistry and Photobiology*, vol. 88, no. 5, pp. 1066–1074, 2012.
- [183] U. G. Sathyanarayana, A. Y. Moore, L. Li et al., "Sun exposure related methylation in malignant and non-malignant skin lesions," *Cancer Letters*, vol. 245, no. 1–2, pp. 112–120, 2007.
- [184] K. Murao, Y. Kubo, N. Ohtani, E. Hara, and S. Arase, "Epigenetic abnormalities in cutaneous squamous cell carcinomas: frequent inactivation of the RB1/p16 and p53 pathways," *British Journal of Dermatology*, vol. 155, no. 5, pp. 999–1005, 2006.
- [185] R. van Doorn, N. A. Gruis, R. Willemze, P. A. Van Der Velden, and C. P. Tensen, "Aberrant DNA methylation in cutaneous malignancies," *Seminars in Oncology*, vol. 32, no. 5, pp. 479–487, 2005.
- [186] A. Marini, A. Mirmohammadsadegh, S. Nambiar, A. Gustrau, T. Ruzicka, and U. R. Hengge, "Epigenetic inactivation of tumor suppressor genes in serum of patients with cutaneous melanoma," *Journal of Investigative Dermatology*, vol. 126, no. 2, pp. 422–431, 2006.
- [187] J. J. Lee, G. F. Murphy, and C. G. Lian, "Melanoma epigenetics: novel mechanisms, markers, and medicines," *Laboratory Investigation*, vol. 94, no. 8, pp. 822–838, 2014.
- [188] A. Mittal, C. A. Elmets, and S. K. Katiyar, "Dietary feeding of proanthocyanidins from grape seeds prevents photocarcinogenesis in SKH-1 hairless mice: relationship to decreased fat and lipid peroxidation," *Carcinogenesis*, vol. 24, no. 8, pp. 1379–1388, 2003.
- [189] S. K. Katiyar and C. A. Elmets, "Green tea polyphenolic antioxidants and skin photoprotection (Review)," *International Journal of Oncology*, vol. 18, no. 6, pp. 1307–1313, 2001.
- [190] S. K. Katiyar and K. Mukhtar, "Tea in chemoprevention of cancer," *International Journal of Oncology*, vol. 8, no. 2, pp. 221–238, 1996.
- [191] B. T. Heijmans, E. W. Tobi, A. D. Stein et al., "Persistent epigenetic differences associated with prenatal exposure to famine in humans," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 44, pp. 17046–17049, 2008.
- [192] I. P. Pogribny, S. A. Ross, C. Wise et al., "Irreversible global DNA hypomethylation as a key step in hepatocarcinogenesis induced by dietary methyl deficiency," *Mutation Research—Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 593, no. 1–2, pp. 80–87, 2006.
- [193] D. C. Dolinoy, C. Weinhouse, T. R. Jones, L. S. Rozek, and R. L. Jirtle, "Variable histone modifications at the Avy metastable epiallele," *Epigenetics*, vol. 5, no. 7, pp. 637–644, 2010.
- [194] G. L. Wolff, R. L. Kodell, S. R. Moore, and C. A. Cooney, "Maternal epigenetics and methyl supplements affect agouti gene expression in Avy/a mice," *The FASEB Journal*, vol. 12, no. 11, pp. 949–957, 1998.
- [195] R. A. Waterland, D. C. Dolinoy, J.-R. Lin, C. A. Smith, X. Shi, and K. G. Tahiliani, "Maternal methyl supplements increase offspring DNA methylation at Axin fused," *Genesis*, vol. 44, no. 9, pp. 401–406, 2006.
- [196] R. A. Waterland, R. Kellermayer, E. Laritsky et al., "Season of conception in rural gambia affects DNA methylation at putative human metastable epialleles," *PLoS Genetics*, vol. 6, no. 12, Article ID e1001252, 2010.
- [197] E. W. Tobi, J. J. Goeman, R. Monajemi et al., "DNA methylation signatures link prenatal famine exposure to growth and metabolism," *Nature Communications*, vol. 5, article no. 5592, 2014.
- [198] M. K. Keyes, H. Jang, J. B. Mason et al., "Older age and dietary folate are determinants of genomic and p16-specific DNA methylation in mouse colon," *Journal of Nutrition*, vol. 137, no. 7, pp. 1713–1717, 2007.
- [199] J. Kotsopoulos, K.-J. Sohn, and Y.-I. Kim, "Postweaning dietary folate deficiency provided through childhood to puberty permanently increases genomic DNA methylation in adult rat liver," *Journal of Nutrition*, vol. 138, no. 4, pp. 703–709, 2008.
- [200] S. Bae, C. M. Ulrich, L. B. Bailey et al., "Impact of folic acid fortification on global DNA methylation and one-carbon biomarkers in the Women's Health Initiative Observational Study cohort," *Epigenetics*, vol. 9, no. 3, pp. 396–403, 2014.
- [201] F. Coppedè, F. Migheli, A. Lopomo et al., "Gene promoter methylation in colorectal cancer and healthy adjacent mucosa specimens: correlation with physiological and pathological characteristics, and with biomarkers of one-carbon metabolism," *Epigenetics*, vol. 9, no. 4, pp. 621–633, 2014.

- [202] A. Kalani, P. K. Kamat, S. Givvimani et al., "Nutri-epigenetics ameliorates blood-brain barrier damage and neurodegeneration in hyperhomocysteinemia: role of folic acid," *Journal of Molecular Neuroscience*, vol. 52, no. 2, pp. 202–215, 2014.
- [203] M. A. Avila, E. R. García-Trevijano, M. L. Martínez-Chantar et al., "S-Adenosylmethionine revisited: its essential role in the regulation of liver function," *Alcohol*, vol. 27, no. 3, pp. 163–167, 2002.
- [204] N. Xiang, R. Zhao, G. Song, and W. Zhong, "Selenite reactivates silenced genes by modifying DNA methylation and histones in prostate cancer cells," *Carcinogenesis*, vol. 29, no. 11, pp. 2175–2181, 2008.
- [205] A. Adaikalakoteswari, S. Finer, P. D. Voyias et al., "Vitamin B12 insufficiency induces cholesterol biosynthesis by limiting s-adenosylmethionine and modulating the methylation of SREBF1 and LDLR genes," *Clinical Epigenetics*, vol. 7, no. 1, article 14, 2015.
- [206] B. H. Natelson, J. E. Ottenweller, R. J. Servatius, S. Drastal, M. T. Bergen, and W. N. Tapp, "Effect of stress and food restriction on blood pressure and lifespan of dahl salt-sensitive rats," *Journal of Hypertension*, vol. 10, no. 12, pp. 1457–1462, 1992.
- [207] T. D. Pugh, T. D. Oberley, and R. Weindruch, "Dietary intervention at middle age: caloric restriction but not dehydroepiandrosterone sulfate increases lifespan and lifetime cancer incidence in mice," *Cancer Research*, vol. 59, no. 7, pp. 1642–1648, 1999.
- [208] R. J. Colman, R. M. Anderson, S. C. Johnson et al., "Caloric restriction delays disease onset and mortality in rhesus monkeys," *Science*, vol. 325, no. 5937, pp. 201–204, 2009.
- [209] S.-J. Lin, M. Kaerberlein, A. A. Andalis et al., "Calorie restriction extends *Saccharomyces cerevisiae* lifespan by increasing respiration," *Nature*, vol. 418, no. 6895, pp. 344–348, 2002.
- [210] S. N. Austad, "Life extension by dietary restriction in the bowl and doily spider, *Frontinella pyramitela*," *Experimental Gerontology*, vol. 24, no. 1, pp. 83–92, 1989.
- [211] D. K. Ingram, R. Weindruch, E. L. Spangler, J. R. Freeman, and R. L. Walford, "Dietary restriction benefits learning and motor performance of aged mice," *The Journal of Gerontology*, vol. 42, no. 1, pp. 78–81, 1987.
- [212] T. Chapman and L. Partridge, "Female fitness in *Drosophila melanogaster*: an interaction between the effect of nutrition and of encounter rate with males," *Proceedings of the Royal Society B: Biological Sciences*, vol. 263, no. 1371, pp. 755–759, 1996.
- [213] B. J. Merry, "Molecular mechanisms linking calorie restriction and longevity," *International Journal of Biochemistry and Cell Biology*, vol. 34, no. 11, pp. 1340–1354, 2002.
- [214] R. S. Sohal and R. Weindruch, "Oxidative stress, caloric restriction, and aging," *Science*, vol. 273, no. 5271, pp. 59–63, 1996.
- [215] W. M. Teeuwisse, R. L. Widya, M. Paulides et al., "Short-term caloric restriction normalizes hypothalamic neuronal responsiveness to glucose ingestion in patients with type 2 diabetes," *Diabetes*, vol. 61, no. 12, pp. 3255–3259, 2012.
- [216] S. R. Rose, S. Burstein, G. A. Burghen, P. Pitukcheewanont, S. Shope, and V. Hodnickak, "Caloric restriction for 24 hours increases mean night growth hormone," *Journal of Pediatric Endocrinology and Metabolism*, vol. 12, no. 2, pp. 175–183, 1999.
- [217] M. Fang, D. Chen, and C. S. Yang, "Dietary polyphenols may affect DNA methylation," *Journal of Nutrition*, vol. 137, no. 1, supplement, pp. 223S–228S, 2007.
- [218] A. F. Fernández and M. F. Fraga, "The effects of the dietary polyphenol resveratrol on human healthy aging and lifespan," *Epigenetics*, vol. 6, no. 7, pp. 870–874, 2011.
- [219] U. Muñoz-Najar and J. M. Sedivy, "Epigenetic control of aging," *Antioxidants and Redox Signaling*, vol. 14, no. 2, pp. 241–259, 2011.
- [220] L. Chouliaras, D. L. A. van den Hove, G. Kenis et al., "Caloric restriction attenuates age-related changes of DNA methyltransferase 3a in mouse hippocampus," *Brain, Behavior, and Immunity*, vol. 25, no. 4, pp. 616–623, 2011.
- [221] Y. Li, L. Liu, and T. O. Tollefsbol, "Glucose restriction can extend normal cell lifespan and impair precancerous cell growth through epigenetic control of hTERT and p16 expression," *FASEB Journal*, vol. 24, no. 5, pp. 1442–1453, 2010.
- [222] O. S. Anderson, K. E. Sant, and D. C. Dolinoy, "Nutrition and epigenetics: an interplay of dietary methyl donors, one-carbon metabolism and DNA methylation," *Journal of Nutritional Biochemistry*, vol. 23, no. 8, pp. 853–859, 2012.
- [223] A. Link, F. Balaguer, and A. Goel, "Cancer chemoprevention by dietary polyphenols: promising role for epigenetics," *Biochemical Pharmacology*, vol. 80, no. 12, pp. 1771–1792, 2010.
- [224] L. Bravo, "Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance," *Nutrition Reviews*, vol. 56, no. 11, pp. 317–333, 1998.
- [225] V. Cheyner, "Polyphenols in foods are more complex than often thought," *The American Journal of Clinical Nutrition*, vol. 81, no. 1, supplement, pp. 223S–229S, 2005.
- [226] S. M. Henning, P. Wang, C. L. Carpenter, and D. Heber, "Epigenetic effects of green tea polyphenols in cancer," *Epigenomics*, vol. 5, no. 6, pp. 729–741, 2013.
- [227] R. H. Dashwood and E. Ho, "Dietary histone deacetylase inhibitors: from cells to mice to man," *Seminars in Cancer Biology*, vol. 17, no. 5, pp. 363–369, 2007.
- [228] G. Filippini, M. Farinotti, and M. Ferrarini, "Active and passive smoking during pregnancy and risk of central nervous system tumours in children," *Paediatric and Perinatal Epidemiology*, vol. 14, no. 1, pp. 78–84, 2000.
- [229] M. A. Norman, E. A. Holly, D. K. Ahn, S. Preston-Martin, B. A. Mueller, and P. M. Bracci, "Prenatal exposure to tobacco smoke and childhood brain tumors: results from the United States West Coast childhood brain tumor study," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 5, no. 2, pp. 127–133, 1996.
- [230] K. Mattsson, K. Källén, M. P. Longnecker, A. Rignell-Hydbom, and L. Rylander, "Maternal smoking during pregnancy and daughters' risk of gestational diabetes and obesity," *Diabetologia*, vol. 56, no. 8, pp. 1689–1695, 2013.
- [231] R. M. Whyatt, F. P. Perera, W. Jedrychowski, R. M. Santella, S. Garte, and D. A. Bell, "Association between polycyclic aromatic hydrocarbon-DNA adduct levels in maternal and newborn white blood cells and glutathione S-transferase P1 and CYP1A1 polymorphisms," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 9, no. 2, pp. 207–212, 2000.
- [232] M. A. Suter, A. M. Anders, and K. M. Aagaard, "Maternal smoking as a model for environmental epigenetic changes affecting birthweight and fetal programming," *Molecular Human Reproduction*, vol. 19, no. 1, pp. 1–6, 2013.
- [233] C. V. Breton, H.-M. Byun, M. Wenten, F. Pan, A. Yang, and F. D. Gilliland, "Prenatal tobacco smoke exposure affects global and gene-specific DNA methylation," *American Journal of Respiratory and Critical Care Medicine*, vol. 180, no. 5, pp. 462–467, 2009.
- [234] D.-H. Kim, H. H. Nelson, J. K. Wiencke et al., "p16^{INK4a} and histology-specific methylation of CpG islands by exposure to tobacco smoke in non-small cell lung cancer," *Cancer Research*, vol. 61, no. 8, pp. 3419–3424, 2001.

- [235] H. Kim, M. K. Young, S. K. Jin et al., "Tumor-specific methylation in bronchial lavage for the early detection of non-small-cell lung cancer," *Journal of Clinical Oncology*, vol. 22, no. 12, pp. 2363–2370, 2004.
- [236] Y. Liu, Q. Lan, J. M. Siegfried, J. D. Luketich, and P. Keohavong, "Aberrant promoter methylation of p16 and MGMT genes in lung tumors from smoking and never-smoking lung cancer patients," *Neoplasia*, vol. 8, no. 1, pp. 46–51, 2006.
- [237] L. Ottini, P. Rizzolo, E. Siniscalchi et al., "Gene promoter methylation and DNA repair capacity in monozygotic twins with discordant smoking habits," *Mutation Research—Genetic Toxicology and Environmental Mutagenesis*, vol. 779, pp. 57–64, 2015.
- [238] S. Zeilinger, B. Kühnel, N. Klopp et al., "Tobacco smoking leads to extensive genome-wide changes in DNA methylation," *PLoS ONE*, vol. 8, no. 5, Article ID e63812, 2013.
- [239] M. M. Monick, S. R. H. Beach, J. Plume et al., "Coordinated changes in AHRH methylation in lymphoblasts and pulmonary macrophages from smokers," *American Journal of Medical Genetics—Part B: Neuropsychiatric Genetics*, vol. 159, no. 2, pp. 141–151, 2012.
- [240] B. R. Joubert, S. E. Håberg, R. M. Nilsen et al., "450K epigenome-wide scan identifies differential DNA methylation in newborns related to maternal smoking during pregnancy," *Environmental Health Perspectives*, vol. 120, no. 10, pp. 1425–1431, 2012.
- [241] W. Besingi and Å. Johansson, "Smoke-related DNA methylation changes in the etiology of human disease," *Human Molecular Genetics*, vol. 23, no. 9, pp. 2290–2297, 2014.
- [242] M. Hussain, M. Rao, A. E. Humphries et al., "Tobacco smoke induces polycomb-mediated repression of Dickkopf-1 in lung cancer cells," *Cancer Research*, vol. 69, no. 8, pp. 3570–3578, 2009.
- [243] M. A. Maccani, M. Avissar-Whiting, C. E. Banister, B. McGonnigal, J. F. Padbury, and C. J. Marsit, "Maternal cigarette smoking during pregnancy is associated with downregulation of miR-16, miR-21 and miR-146a in the placenta," *Epigenetics*, vol. 5, no. 7, pp. 583–589, 2010.
- [244] F. Schembri, S. Sridhar, C. Perdomo et al., "MicroRNAs as modulators of smoking-induced gene expression changes in human airway epithelium," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 7, pp. 2319–2324, 2009.
- [245] S. Xi, M. Yang, Y. Tao et al., "Cigarette smoke induces C/EBP- β -mediated activation of mir-31 in normal human respiratory epithelia and lung cancer cells," *PLoS ONE*, vol. 5, no. 10, article e13764, 2010.
- [246] K. K. Kharbanda, "Alcoholic liver disease and methionine metabolism," *Seminars in Liver Disease*, vol. 29, no. 2, pp. 155–165, 2009.
- [247] E. S. Schernhammer, E. Giovannucci, T. Kawasaki, B. Rosner, C. S. Fuchs, and S. Ogino, "Dietary folate, alcohol and B vitamins in relation to LINE-1 hypomethylation in colon cancer," *Gut*, vol. 59, no. 6, pp. 794–799, 2010.
- [248] I. M. Smith, W. K. Mydlarz, S. K. Mithani, and J. A. Califano, "DNA global hypomethylation in squamous cell head and neck cancer associated with smoking, alcohol consumption and stage," *International Journal of Cancer*, vol. 121, no. 8, pp. 1724–1728, 2007.
- [249] R. A. Hlady, R. L. Tiedemann, W. Puszyk et al., "Epigenetic signatures of alcohol abuse and hepatitis infection during human hepatocarcinogenesis," *Oncotarget*, vol. 5, no. 19, pp. 9425–9443, 2014.
- [250] F. C. Zhou, Y. Balaraman, M. Teng, Y. Liu, R. P. Singh, and K. P. Nephew, "Alcohol alters DNA methylation patterns and inhibits neural stem cell differentiation," *Alcoholism: Clinical and Experimental Research*, vol. 35, no. 4, pp. 735–746, 2011.
- [251] S. Bala and G. Szabo, "MicroRNA signature in alcoholic liver disease," *International Journal of Hepatology*, vol. 2012, Article ID 498232, 6 pages, 2012.
- [252] Y. Ladeiro, G. Couchy, C. Balabaud et al., "MicroRNA profiling in hepatocellular tumors is associated with clinical features and oncogene/tumor suppressor gene mutations," *Hepatology*, vol. 47, no. 6, pp. 1955–1963, 2008.
- [253] A. R. Aroor, R. J. Restrepo, K. K. Kharbanda, and S. D. Shukla, "Epigenetic histone modifications in a clinically relevant rat model of chronic ethanol-binge-mediated liver injury," *Hepatology International*, vol. 8, supplement 2, pp. 421–430, 2014.
- [254] M. Choudhury, P.-H. Park, D. Jackson, and S. D. Shukla, "Evidence for the role of oxidative stress in the acetylation of histone H3 by ethanol in rat hepatocytes," *Alcohol*, vol. 44, no. 6, pp. 531–540, 2010.
- [255] A. Ghezzi, H. R. Krishnan, L. Lew, F. J. Prado III, D. S. Ong, and N. S. Atkinson, "Alcohol-induced histone acetylation reveals a gene network involved in alcohol tolerance," *PLoS Genetics*, vol. 9, no. 12, Article ID e1003986, 2013.
- [256] A. Page, P. P. Paoli, S. J. Hill et al., "Alcohol directly stimulates epigenetic modifications in hepatic stellate cells," *Journal of Hepatology*, vol. 62, no. 2, pp. 388–397, 2015.
- [257] B. J. Curtis, A. Zahs, and E. J. Kovacs, "Epigenetic targets for reversing immune defects caused by alcohol exposure," *Alcohol Research: Current Reviews*, vol. 35, no. 1, pp. 97–113, 2013.
- [258] K. J. Veazey, D. Muller, and M. C. Golding, "Prenatal alcohol exposure and cellular differentiation: a role for Polycomb and Trithorax group proteins in FAS phenotypes?" *Alcohol Research: Current Reviews*, vol. 35, no. 1, pp. 77–85, 2012.
- [259] G. S. Leverich, L. L. Altshuler, M. A. Frye et al., "Factors associated with suicide attempts in 648 patients with bipolar disorder in the Stanley Foundation bipolar network," *Journal of Clinical Psychiatry*, vol. 64, no. 5, pp. 506–515, 2003.
- [260] P. O. McGowan, A. Sasaki, A. C. D'Alessio et al., "Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse," *Nature Neuroscience*, vol. 12, no. 3, pp. 342–348, 2009.
- [261] L. J. van der Knaap, H. Riese, J. J. Hudziak et al., "Glucocorticoid receptor gene (NR3C1) methylation following stressful events between birth and adolescence. the TRAILS study," *Translational Psychiatry*, vol. 4, article e381, 2014.
- [262] B. Le François, J. Soo, A. M. Millar et al., "Chronic mild stress and antidepressant treatment alter 5-HT1A receptor expression by modifying DNA methylation of a conserved Sp4 site," *Neurobiology of Disease*, vol. 82, pp. 332–341, 2015.
- [263] R. G. Hunter, K. J. McCarthy, T. A. Milne, D. W. Pfaff, and B. S. McEwen, "Regulation of hippocampal H3 histone methylation by acute and chronic stress," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 49, pp. 20912–20917, 2009.
- [264] L. Cao-Lei, R. Massart, M. J. Suderman et al., "DNA methylation signatures triggered by prenatal maternal stress exposure to a natural disaster: project ice storm," *PLoS ONE*, vol. 9, no. 9, Article ID e107653, 2014.

- [265] J. Denham, B. J. O'Brien, J. T. Harvey, and F. J. Char-char, "Genome-wide sperm DNA methylation changes after 3 months of exercise training in humans," *Epigenomics*, vol. 7, no. 5, pp. 717–731, 2015.
- [266] M. D. Nitert, T. Dayeh, P. Volkov et al., "Impact of an exercise intervention on DNA methylation in skeletal muscle from first-degree relatives of patients with type 2 diabetes," *Diabetes*, vol. 61, no. 12, pp. 3322–3332, 2012.
- [267] J. L. Abel and E. F. Rissman, "Running-induced epigenetic and gene expression changes in the adolescent brain," *International Journal of Developmental Neuroscience*, vol. 31, no. 6, pp. 382–390, 2013.
- [268] V. Santini, A. Gozzini, and G. Ferrari, "Histone deacetylase inhibitors: molecular and biological activity as a premise to clinical application," *Current Drug Metabolism*, vol. 8, no. 4, pp. 383–394, 2007.
- [269] A. Villar-Garea and M. Esteller, "Histone deacetylase inhibitors: understanding a new wave of anticancer agents," *International Journal of Cancer*, vol. 112, no. 2, pp. 171–178, 2004.
- [270] K. Huber, G. Doyon, J. Plaks, E. Fyne, J. W. Mellors, and N. Sluis-Cremer, "Inhibitors of histone deacetylases: correlation between isoform specificity and reactivation of HIV type 1 (HIV-1) from latently infected cells," *Journal of Biological Chemistry*, vol. 286, no. 25, pp. 22211–22218, 2011.
- [271] M. Göttlicher, S. Minucci, P. Zhu et al., "Valproic acid defines a novel class of HDAC inhibitors inducing differentiation of transformed cells," *The EMBO Journal*, vol. 20, no. 24, pp. 6969–6978, 2002.
- [272] J. L. Fleming, T. H.-M. Huang, and A. E. Toland, "The role of parental and grandparental epigenetic alterations in familial cancer risk," *Cancer Research*, vol. 68, no. 22, pp. 9116–9121, 2008.
- [273] M. Veurink, M. Koster, and L. T. W. de Jong-van den Berg, "The history of DES, lessons to be learned," *Pharmacy World and Science*, vol. 27, no. 3, pp. 139–143, 2005.
- [274] K. Sato, H. Fukata, Y. Kogo, J. Ohgane, K. Shiota, and C. Mori, "Neonatal exposure to diethylstilbestrol alters expression of DNA methyltransferases and methylation of genomic DNA in the mouse uterus," *Endocrine Journal*, vol. 56, no. 1, pp. 131–139, 2009.
- [275] P.-Y. Hsu, D. E. Deatherage, B. A. T. Rodriguez et al., "Xenoestrogen-induced epigenetic repression of microRNA-9-3 in breast epithelial cells," *Cancer Research*, vol. 69, no. 14, pp. 5936–5945, 2009.
- [276] S. L. Tilghman, M. R. Bratton, H. C. Segar et al., "Endocrine disruptor regulation of microRNA expression in breast carcinoma cells," *PLoS ONE*, vol. 7, no. 3, article e32754, 2012.
- [277] A. Villar-Garea, M. F. Fraga, J. Espada, and M. Esteller, "Procaine is a DNA-demethylating agent with growth-inhibitory effects in human cancer cells," *Cancer Research*, vol. 63, no. 16, pp. 4984–4989, 2003.
- [278] S. Badal, Y. F. Her, and L. J. Maher III, "Non-antibiotic effects of fluoroquinolones in mammalian cells," *The Journal of Biological Chemistry*, vol. 290, no. 36, pp. 22287–22297, 2015.
- [279] V. M. Kovalenko, T. V. Bagnyukova, O. V. Sergienko et al., "Epigenetic changes in the rat livers induced by pyrazinamide treatment," *Toxicology and Applied Pharmacology*, vol. 225, no. 3, pp. 293–299, 2007.
- [280] T. Yokochi and K. D. Robertson, "Doxorubicin inhibits DNMT1, resulting in conditional apoptosis," *Molecular Pharmacology*, vol. 66, no. 6, pp. 1415–1420, 2004.
- [281] R. Garzon, F. Pichiorri, T. Palumbo et al., "MicroRNA gene expression during retinoic acid-induced differentiation of human acute promyelocytic leukemia," *Oncogene*, vol. 26, no. 28, pp. 4148–4157, 2007.
- [282] T. Boren, Y. Xiong, A. Hakam et al., "MicroRNAs and their target messenger RNAs associated with ovarian cancer response to chemotherapy," *Gynecologic Oncology*, vol. 113, no. 2, pp. 249–255, 2009.
- [283] A. Sorrentino, C.-G. Liu, A. Addario, C. Peschle, G. Scambia, and C. Ferlini, "Role of microRNAs in drug-resistant ovarian cancer cells," *Gynecologic Oncology*, vol. 111, no. 3, pp. 478–486, 2008.
- [284] I. Berthaut, D. Montjean, L. Dessolle et al., "Effect of temozolomide on male gametes: an epigenetic risk to the offspring?" *Journal of Assisted Reproduction and Genetics*, vol. 30, no. 6, pp. 827–833, 2013.