

# Natural killer cell regulation - beyond the receptors

Carsten Watzl\*, Doris Urlaub, Frank Fasbender and Maren Claus

Address: IfADo - Leibniz Research Centre for Working Environment and Human Factors, Ardeystrasse 67, 44139 Dortmund, Germany

\* Corresponding author: Carsten Watzl (watzl@ifado.de)

*F1000Prime Reports* 2014, **6:87** (doi:10.12703/P6-87)

All F1000Prime Reports articles are distributed under the terms of the Creative Commons Attribution-Non Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/legalcode>), which permits non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

The electronic version of this article is the complete one and can be found at: <http://f1000.com/prime/reports/b/6/87>

## Abstract

Natural killer (NK) cells are lymphocytes that are important for early and effective immune responses against infections and cancer. In the last 40 years, many receptors, their corresponding ligands and signaling pathways that regulate NK cell functions have been identified. However, we now know that additional processes, such as NK cell education, differentiation and also the formation of NK cell memory, have a great impact on the reactivity of these cells. Here, we summarize the current knowledge about these modulatory processes.

## Introduction

In the mid-1970s, a novel immune cell type was described based on its ability to lyse allogeneic tumor cells without the need for prior sensitization. The term “natural cytotoxicity” was introduced to describe this feature and the cells mediating this effect were named NK cells [1–5]. In the last 40 years, much progress has been made in the understanding of the function and regulation of NK cells. We now know that NK cells contribute to effective innate immune responses and provide the first important line of defense against parasites, viruses and cancer [6–10]. NK cells derive from the common lymphocyte progenitor, but they are independent of a functional thymus and rely on germ-line-encoded surface receptors that do not undergo somatic recombination. One important step for the understanding of NK cell regulation was the realization that NK cells preferentially kill cells with low or no major histocompatibility complex (MHC) class I expression that led to the formulation of the “missing-self hypothesis” [11,12]. This concept was later supported through the identification of MHC class I-specific inhibitory receptors, such as Ly49 receptors in mice and killer cell immunoglobulin-like receptors (KIRs) in humans [13–19]. These inhibitory receptors possess immunoreceptor tyrosine-based inhibitory motifs (ITIMs) in their cytoplasmic tail that are phosphorylated upon binding to MHC class I. This leads to binding and activation of phosphatases, such as SHP1/2

and SH2 domain-containing inositol 5-phosphatase (SHIP), which in turn interfere with activating signaling pathways by dephosphorylation [20], effectively preventing NK cell activation.

NK cells are stimulated by a number of different activating receptors that can recognize a variety of ligands on potential target cells [21]. Engagement of these activating receptors can trigger NK cell functions via different signaling pathways [22–24]. Despite the diversity of these early signaling pathways, inhibitory receptors can effectively control NK cell activation [9,25]. It is, therefore, now generally accepted that NK cell activity is tightly regulated by an interplay between activating and inhibitory cell surface receptors. However, in recent years, it has become clear that this is not the only level at which the activity of NK cells is regulated. The fact that the triggering of the same receptor in individual NK cells does not necessarily lead to the same outcome already implies the presence of additional mechanisms for the regulation of NK cell functions. In the following article, we will describe three additional levels of NK cell regulation.

## NK cell education

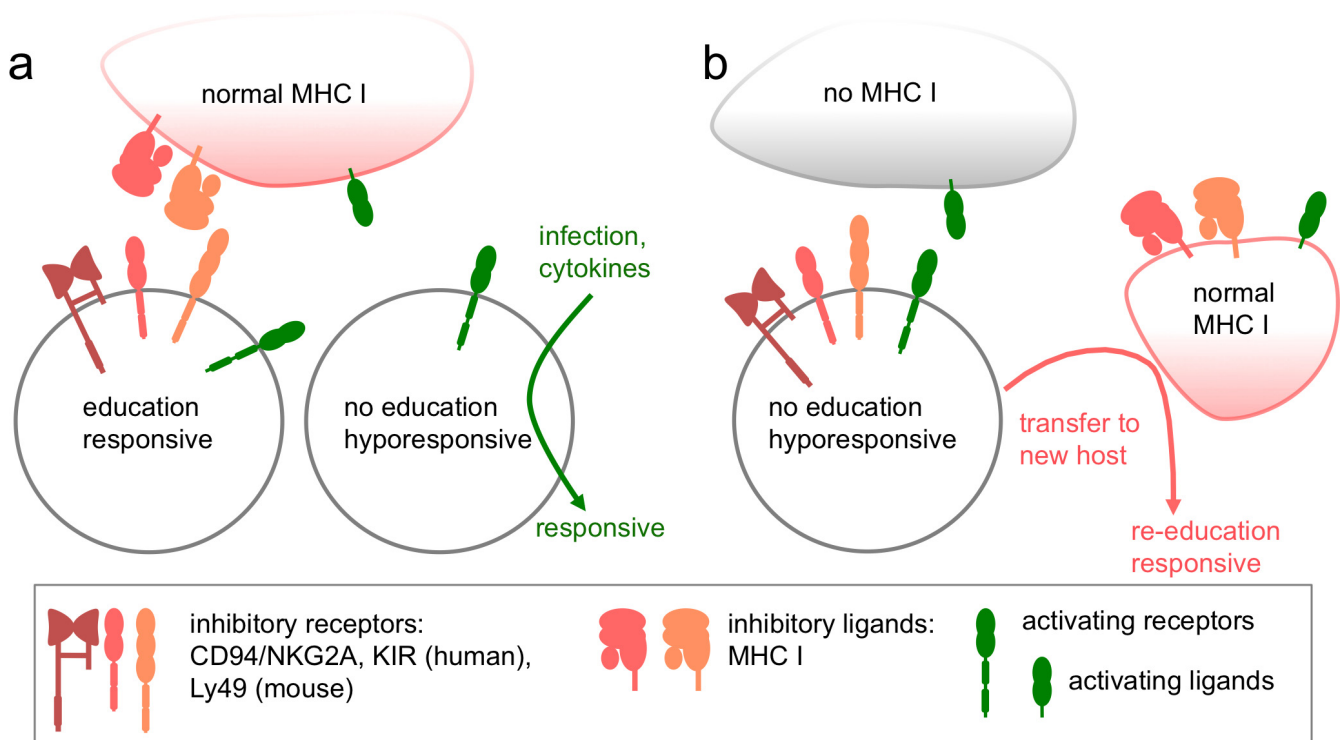
In accordance with the missing-self hypothesis, the “at least one” model was proposed [26]. This model assumed that NK cells need to express at least one inhibitory receptor that is specific for self-MHC class I in order to

prevent autoreactivity. This hypothesis was supported by data from human NK clones that were all found to express at least one self-specific inhibitory receptor [27]. However, it was also known that NK cells from MHC class I-deficient hosts were not autoreactive despite the lack of ligands for the inhibitory receptors [28,29]. This already suggested that additional mechanisms must exist to ensure that NK cells are not autoreactive in the absence of inhibitory signaling. Indeed, it was later discovered that a significant subset of NK cells present in healthy mice and humans lack self-specific inhibitory receptors [30–32]. These NK cells were not autoreactive and were found to be hyporesponsive when triggered through activating receptor stimulation. This adaptation of the reactivity of NK cells depending on the inhibitory receptor ligand matches is generally referred to as NK cell education [26] (Figure 1) and assures the self-tolerance of NK cells.

Initially, two opposing mechanisms were discussed on how NK cells can become educated. In the “arming” or

“licensing” model, NK cells are assumed to be inactive by default and only acquire their full functionality through the engagement and the signaling of an inhibitory receptor [33,34]. In the “disarming” model, NK cells are active by default but are rendered hyporesponsive or anergic through the continuous stimulation via activating receptors recognizing endogenous ligands. They can only maintain their functionality if this chronic stimulation is counteracted by signals of inhibitory receptors [34]. While inhibitory receptors have opposite functions in both models, the outcome would be comparable – only NK cells with an inhibitory receptor for self-MHC class I can become functionally active. Additionally, the education of NK cells is not an all or nothing decision, but can be tuned in a quantitative way. The stronger the inhibitory interaction(s) of an NK cell is, the stronger it responds to activating receptor signals [35,36]. Therefore, the rheostat model has been proposed [37]. This model does not replace but rather supplements the arming or the disarming model, and describes NK

**Figure 1. NK cell education: adaption of the responsiveness depending on inhibitory receptor - ligand interactions**



(a) In normal major histocompatibility complex (MHC) class I-sufficient individuals (humans and mice), NK cells expressing inhibitory receptors recognizing those MHC class I molecules become educated. Those cells are responsive to activating receptor stimulation. The subset of NK cells that lacks inhibitory receptors for self MHC class I are non-educated and hyporesponsive when triggered through activating receptor stimulation. Under certain conditions, such as infections or cytokine stimulation, this subset can become responsive.

(b) In MHC class I-deficient individuals, NK cells are non-educated and hyporesponsive due to the lack of inhibitory ligands. After transfer to a new MHC class I-sufficient host, NK cells can become “re-educated” and responsive if they express the matching inhibitory receptors.

KIR, killer cell immunoglobulin-like receptor.

cell education as a dynamic process, rather than an on/off state.

With the data available today, both the arming and the disarming models are able to explain the observed phenotypes, but the mechanism remains unclear. The pros and cons of the different models have been extensively discussed [38,39]. It is not only inhibitory receptors that can dictate the reactivity of NK cells. The constitutive expression of a ligand for an activating NK cell receptor in mice results in the hyporesponsiveness of NK cells expressing the matching activating receptor [40,41]. This finding that chronic stimulation can render NK cells hyporesponsive would support the “disarming” model. Since many activating NK cell receptors are capable of recognizing endogenous ligands, NK cells probably have an individual activation threshold that is adjusted to their receptor expression and the available ligands [42]. Most importantly, these mechanisms must avoid autoreactivity while retaining maximal responsiveness of the NK cells toward infected or transformed cells.

While the functional mechanism of education is still unknown, we know from various studies that education is not final. The changes caused in educated and in hyporesponsive, uneducated cells are reversible, suggesting that there is plasticity in the education process. After transfer to an MHC-I0-sufficient environment, previously uneducated NK cells gained functional competence, whereas previously educated NK cells were rendered hyporesponsive in an MHC-I-deficient environment [43,44] (Figure 1). Similarly, non-educated human NK cells were shown to acquire functional inhibitory receptor expression upon stimulation with pro-inflammatory cytokines, which resulted in an educated phenotype [45]. Since this “re-education” is possible with mature cells and happens within a few days, it is likely uncoupled from the process of NK cell development in the bone marrow. However, findings from hematopoietic stem cell transplantations in humans suggest that the education of the donor NK cells remains even if the host MHC class I environment is different [46]. This would argue that at least in this setting, the cell responsible for the education of NK cells is of hematopoietic origin.

To complicate things even further, some inhibitory Ly49 receptors can also interact with their MHC class I ligand on the same cells *in cis* [47]. The importance of this *cis* binding for NK cell education is controversial. At least for some inhibitory receptors, it has been demonstrated that binding *in cis* contributes to this education [48,49], and the strength of the binding correlates with the potency of education [50]. However, in another experimental

setup, only the interaction *in trans* was effective for the education of NK cells [51]. Additionally, the fact that NK cells can adjust their reactivity through a change in their MHC class I environment supports the importance of the *trans* interactions for the re-education [43,44].

Why are NK cells that lack sufficient inhibitory receptors for self-MHC class I only rendered hyporesponsive and why are they not deleted as in the case of autoreactive T cells? The answer to this question could be that under certain circumstances, non-educated NK cells can be beneficial to the host. During an acute virus infection, the non-educated NK cells can become functional under the influence of pro-inflammatory cytokines and can even be more efficient than educated NK cells [52] (Figure 1). Similarly, non-educated NK cells can be more effective in mediating antibody-dependent cellular cytotoxicity in neuroblastoma patients treated with an anti-GD2 antibody [53]. Under these circumstances, the lack of inhibition may make the non-educated NK cells the better effector cells.

What is the molecular mechanism that determines the reactivity of educated NK cells? Interestingly, there are only a few transcriptional changes when comparing educated with non-educated NK cells [54]. One possible mechanism that could cause these functional differences without the need for changes in gene expression is the organization of receptors in the membrane. Nanoscopic analysis revealed that in educated NK cells, activating receptors were localized in nanodomains, whereas they were confined to an actin meshwork in non-educated cells [54]. In those nanodomains, the activating receptors could have the proper environment of signaling molecules needed for efficient NK cell activation [55]. This would be consistent with the finding that the triggering of activating receptors in educated NK cells results in an efficient activation of the integrin lymphocyte function-associated antigen 1 via inside-out signaling, thereby promoting the adhesion of educated NK cells to target cells [56].

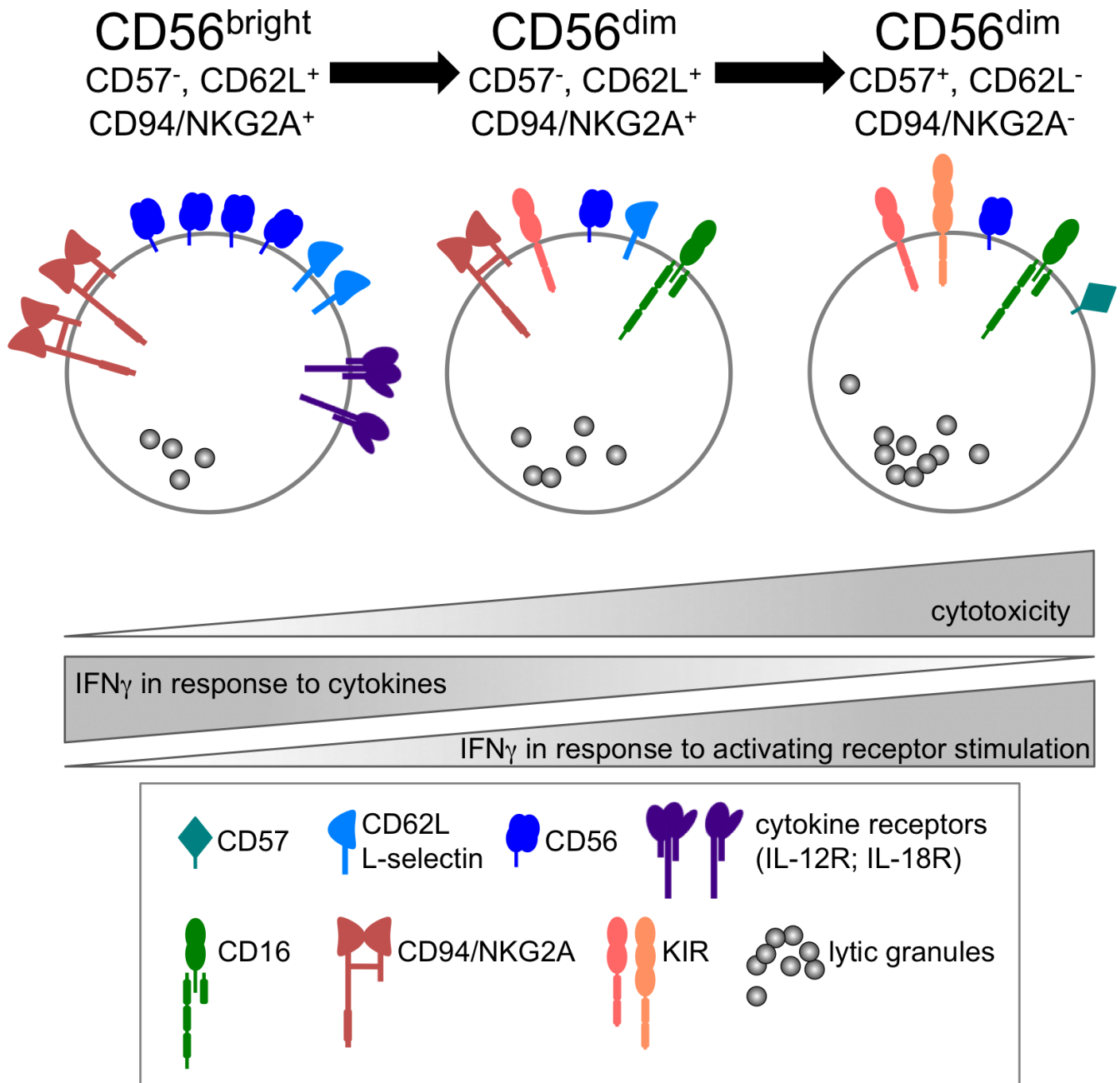
### **NK cell differentiation and subsets**

After the initial process of NK cell education, functionally competent NK cells can be found in the periphery. However, not all educated NK cells have the same functionality. Traditionally, human peripheral blood NK cells are divided in two functionally distinct subsets: CD56<sup>dim</sup> and CD56<sup>bright</sup> NK cells [57–59]. In recent years, it became clear that CD56<sup>dim</sup> NK cells can be further subdivided based on the expression of CD62L, CD57, or CD94/NKG2A [60–64]. Additionally, there is a developmental relationship between the different subpopulations, suggesting a differentiation of mature NK cells

starting from CD56<sup>bright</sup> via CD56<sup>dim</sup>, CD57<sup>-</sup>, CD62L<sup>+</sup>, CD94/NKG2A<sup>+</sup> to the more differentiated CD56<sup>dim</sup>, CD57<sup>+</sup>, CD62L<sup>-</sup>, CD94/NKG2A<sup>-</sup> NK cells [60–67] (Figure 2). Along this differentiation pathway, the functionality of NK cells change [68,69]. While CD56<sup>bright</sup> cells are not very cytotoxic, they are especially good at

producing IFN $\gamma$  after stimulation with pro-inflammatory cytokines, such as interleukin (IL)-12 and IL-18. This activity is gradually lost during the differentiation towards the more cytotoxic CD56<sup>dim</sup>, CD57<sup>+</sup> NK cells. In contrast, these most differentiated NK cells can produce more interferon gamma (IFN $\gamma$ ) when triggered via activating

Figure 2. Adaption of NK cell reactivity during differentiation



Functionally distinct subsets of human NK cells differ in their surface receptor expression and their reactivity towards activating receptors triggering or cytokine stimulation. See text for details.

IFN, interferon; IL, interleukin; KIR, killer cell immunoglobulin-like receptor.

surface receptors [70] and this IFN $\gamma$  competence has recently been linked to the epigenetic remodeling of the IFNG promoter [71]. Therefore, the functionality of NK cells changes during their differentiation (Figure 2), which may be important for the orchestration of successful NK-mediated immune responses. Recently, a study identified several thousand distinct subpopulations of NK cells in the peripheral blood of humans [72]. If this is reflected in additional differences in functionality, this will have to be addressed.

**Memories of an NK cell**

Recent studies have shown that NK cells can also acquire memory or memory-like functions, thereby challenging the classical distinction between innate and adaptive immunity [73]. As there are already excellent reviews about NK cell memory [73–76], we just want to briefly summarize the current knowledge about the different forms of NK cell memory that have been described so far (Table 1).

**Liver-restricted memory NK cells**

NK cells that exhibit a more potent secondary response were first described in a mouse model of delayed-type hypersensitivity (DTH) using hapten or viral antigens [77,78]. Recombination-activating gene (RAG)-deficient mice, lacking T and B cells, were sensitized with a hapten or a viral antigen and showed an NK cell-specific DTH response when challenged later with the same antigen. This antigen-specific type of NK cell memory is confined to CXCR6-positive liver NK cells [78,79]. However, it is currently unclear which receptors are responsible for the

antigen-specific response and how liver NK cells can mediate specific and localized immune reactions at the site of antigen re-challenge.

**CMV-specific memory NK cells**

In another form of antigen-specific NK cell memory, the receptor responsible for the effect is known. Cytomegalovirus (CMV) infections in mice have been shown to induce a rapid and clonal-like expansion of a NK cell subset expressing Ly49H, which recognizes the CMV-encoded protein m157 [80]. These NK cell memory subsets show enhanced immune responses upon secondary challenge with CMV. The activating receptor DNAX accessory molecule-1 (CD226) cooperates with Ly49H for the expansion of these memory NK cells by signaling through Fyn and protein kinase C $\eta$  (PKC $\eta$ ) [81]. Additionally, the expansion of CMV-specific memory NK cells is dependent on IL-12 and IL-15 and the subsequent signaling via signal transducer and activator of transcription 4 (STAT4) [82,83]. This induces the transcription factor zinc finger and BTB domain containing 32 (Zbtb32), which was shown to be essential for the proliferation and the protective capacity of the virus-specific NK cells [84]. Finally, the pro-apoptotic factor Bim is responsible for the contraction of the expanded Ly49H<sup>+</sup> NK cells population, resulting in mature, murine CMV-specific memory NK cells [85].

CMV infection is also associated with the generation of memory NK cells in humans, where the expansion and long-term persistence of NKG2C<sup>+</sup> NK cells can be observed [86,87]. However, NKG2C does not seem to be involved

**Table 1. Characteristics of NK cell memory in different models Comparison of the four major types of NK cell memory. Lymphopenia-induced memory NK cells are not shown. See text for details**

| Memory type               | Liver                                                                       | CMV                                                                                                                                    | Fc $\epsilon$ R $\gamma$ Deficiency       | Cytokine-induced                                                 |
|---------------------------|-----------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------|-------------------------------------------|------------------------------------------------------------------|
| <b>Subpopulations</b>     | Thy1+CD11b+CD27-<br>Thy1+Ly49C/I+ [78,116]<br>CXCR6+ [78]<br>CD49a+DX5-[79] | Mouse: Ly49H+ [80]<br>Human:<br>NKG2C+ [86]<br>CD57+ [92]                                                                              | CD57dimFc $\epsilon$ R $\gamma$ - [100]   | CD25+ [107]                                                      |
| <b>Antigen</b>            | Haptens or viral antigens [78,117]                                          | m157 [80]                                                                                                                              | (m157?)                                   | -                                                                |
| <b>Proliferation</b>      | No [77]                                                                     | Yes [80]                                                                                                                               | Unknown                                   | Yes [96]                                                         |
| <b>Involved cytokines</b> | IL-12 [106]<br>CXCL16 [78]                                                  | IL-12, IL-15 [82]                                                                                                                      | IL-12?                                    | IL-12, IL-15, IL-18 [94]                                         |
| <b>Signaling</b>          | IL-12R [106]<br>CXCR6 [78]<br>NKG2D [77]                                    | IL-12R /STAT4 [82]<br>IL-15R<br>CD25 [108]<br>Ly49H/DAP12 [80]<br>DNAM-1 via Fyn, PKC [81]<br>Zbtb32 [84]<br>Bim [85]<br>miR-155 [110] | IL-12R?                                   | IL-12R /STAT4<br>IL-15R<br>IL-18R<br>CD25 [107]<br>miR-155 [112] |
| <b>Memory effect</b>      | DTH                                                                         | Enhanced cytotoxicity and IFN $\gamma$ production                                                                                      | Enhanced IFN $\gamma$ production and ADCC | Enhanced IFN $\gamma$ production                                 |

ADCC, antibody-dependent cellular cytotoxicity; DTH, delayed-type-hypersensitivity; IL, IFN $\Phi$ interferon; interleukin; PKC, protein kinase C; STAT4, signal transducer and activator of transcription 4.



in the direct recognition of CMV [88]. Interestingly, similar expansions of NKG2C<sup>+</sup> NK cells have been observed during and after other virus infections, such as hantavirus, HIV and hepatitis B [89–91], but they were always restricted to human CMV-seropositive individuals. Additionally, these NKG2C<sup>+</sup> NK cells are also positive for CD57 [92,93], demonstrating a terminal differentiation of these human CMV-dependent memory NK cells.

#### **Cytokine-induced memory-like NK cells**

*In vitro* exposure of NK cells to a combination of IL-12, IL-15 and IL-18 generates memory-like cells that show enhanced effector functions [94–96]. *In vivo*, inflammation or other immune responses could result in the exposure of NK cells to these cytokines. Dendritic cells are the main producers of IL-12 and IL-18, thereby regulating NK cells [97]. In a recent study, adoptive co-transfer of NK cells with dendritic cells without exogenous cytokines showed increased tumor infiltration relative to NK cell transfer alone [98]. Additionally, the improved effector functions of cytokine-exposed NK cells might be a valuable tool in enhancing the effectiveness of NK cell-based therapies against tumors [94,96,99].

Finally, there have been other reports describing NK cells with certain memory-like phenotypes. While they may be related to the types of memory described above, we list them here as separate examples of NK cell memory.

#### **FcεRγ-deficient memory NK cells**

A subpopulation of human NK cells has been described that is deficient for the FcεRγ signaling adaptor. In NK cells, FcεRγ is a signaling partner chain for CD16, an activating Fc receptor responsible for the recognition of antibody-coated cells. NK cells lacking FcεRγ display poor cytotoxicity but significantly enhanced IFNγ production upon CD16 stimulation [100]. FcεRγ<sup>-</sup> NK cells have a CD56<sup>dim</sup> phenotype and the existence of this subset is also associated with prior human CMV infection. However, these NK cells also demonstrate enhanced responses against other viruses [101].

#### **Lymphopenia-induced long-lived NK cells**

In a lymphopenic environment (e.g., Rag2<sup>-/-</sup> IL2rγ<sup>-/-</sup> mice), NK cells undergo a rapid but non-specific proliferation after adoptive transfer and, similar to memory T cells, show self-renewal at a steady state. These NK cells are able to respond robustly to viral infection more than 6 months after transfer [102–105].

#### **How does NK cell memory work?**

It is likely that the different forms of NK cell memory described above are not mutually exclusive and independent phenomena. Rather, there are some common

denominators that suggest that they are connected and possibly represent different forms of common “memory NK cells”. The signaling via pro-inflammatory cytokines seems to be important for the generation of NK cell memory. IL-12 in particular is essential for the generation of CMV-specific memory NK cells [82], for cytokine-induced memory NK cells, likely also for the generation of liver-restricted memory NK cells [106] and is possibly important for FcεRγ<sup>-</sup> memory NK cells. IL-12 induces the expression of a high-affinity IL-2 receptor via the up-regulation of the IL-2Rα chain (CD25) [107,108], which might serve as an early marker for memory NK cells. However, the molecular basis for the increased IFNγ production and the (in some cases) enhanced cytotoxicity of memory NK cells is still unclear. Analysis of gene expression data show specific differences between resting, activated and CMV-induced memory NK cells and suggests a common transcriptional program that is conserved in the memory differentiation of NK cells and CD8<sup>+</sup> T cells in response to infection [109]. Additionally, microRNAs (miRNA) have been shown to play a role in the regulation of NK cell functions, and the upregulation of miRNA-155 has been observed in CMV and cytokine-induced memory NK cells [110–112]. Finally, memory NK cells often display a more differentiated phenotype with the expression of CD57 and KLRG1 [92,93]. As described above, NK cells gain IFNγ-competence in response to activating receptor triggering when they mature, which is connected to a partial epigenetic remodeling of the IFNG promoter [71]. Extending these findings, recent data suggest that a broader epigenetic remodeling of the IFNG locus may be the basis for the enhanced IFNγ production of memory NK cells (C Romagnani, personal communication). Therefore, similar to what has been found for memory T cells [113,114], a global epigenetic reprogramming may also be responsible for the generation of memory NK cells [115]. However, in the current situation, it is very difficult to judge how much memory NK cells can contribute to immune responses against secondary infections with the same pathogen.

#### **Concluding remarks**

Some 45 years after the first description of “natural cytotoxicity”, we already know a lot about NK cells, their important contribution to early and effective immune responses and how their effector functions are regulated through different surface receptors and cytokines. However, we now know that processes, such as education, differentiation and finally also the formation of a memory pool, additionally impacts on the activity of NK cells. Uncovering the molecular details of these processes will greatly enhance our understanding of these important immune cells and will pave the way for more effective NK cell-based therapies.

## Abbreviations

ADCC, antibody-dependent cellular cytotoxicity; CMV, cytomegalovirus; DTH, delayed-type-hypersensitivity; IFN, interferon; IL, interleukin; ITIM, immunoreceptor tyrosine-based inhibitory motif; KIR, killer cell immunoglobulin-like receptor; MHC, major histocompatibility complex; NK, natural killer; RAG, recombination activating gene.

## Disclosures

The authors declare that they have no disclosures.

## Acknowledgments

The authors thank all the members of the Watzl lab for their support and for the helpful discussions. Our work is generously supported by the Deutsche Forschungsgemeinschaft DFG (WA 1552/5-1) and the SAW program of the Leibniz Association.

## References

- Herberman RB, Nunn ME, Holden HT, Lavrin DH: **Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic and allogeneic tumors. II. Characterization of effector cells.** *Int J Cancer* 1975, **16**:230-9.
- Herberman RB, Nunn ME, Lavrin DH: **Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic acid allogeneic tumors. I. Distribution of reactivity and specificity.** *Int J Cancer* 1975, **16**:216-29.
- Kiessling R, Klein E, Pross H, Wigzell H: **"Natural" killer cells in the mouse. II. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Characteristics of the killer cell.** *Eur J Immunol* 1975, **5**:117-21.
- Kiessling R, Klein E, Wigzell H: **"Natural" killer cells in the mouse. I. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Specificity and distribution according to genotype.** *Eur J Immunol* 1975, **5**:112-7.
- Peter HH, Kalden JR, Seeland P, Diehl V, Eckert G: **Humoral and cellular immune reactions 'in vitro' against allogeneic and autologous human melanoma cells.** *Clin Exp Immunol* 1975, **20**:193-207.
- Jost S, Altfeld M: **Control of human viral infections by natural killer cells.** *Annu Rev Immunol* 2013, **31**:163-94.
- Long EO, Kim HS, Liu D, Peterson ME, Rajagopalan S: **Controlling natural killer cell responses: integration of signals for activation and inhibition.** *Annu Rev Immunol* 2013, **31**:227-58.
- Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S: **Functions of natural killer cells.** *Nat Immunol* 2008, **9**:503-10.
- Watzl C, Urlaub D: **Molecular mechanisms of natural killer cell regulation.** *Front Biosci (Landmark Ed)* 2012, **17**:1418-32.
- Orange JS: **Natural killer cell deficiency.** *J Allergy Clin Immunol* 2013, **132**:515-25; quiz 526.
- Kärre K, Ljunggren HG, Piontek G, Kiessling R: **Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategy.** *Nature* 1986, **319**:675-8.
- Ljunggren HG, Kärre K: **Host resistance directed selectively against H-2-deficient lymphoma variants. Analysis of the mechanism.** *J Exp Med* 1985, **162**:1745-59.
- Karlhofer FM, Ribaudo RK, Yokoyama WM: **MHC class I alloantigen specificity of Ly-49+ IL-2-activated natural killer cells.** *Nature* 1992, **358**:66-70.
- Yokoyama WM, Seaman WE: **The Ly-49 and NKR-PI gene families encoding lectin-like receptors on natural killer cells: the NK gene complex.** *Annu Rev Immunol* 1993, **11**:613-35.
- Wagtmann N, Biassoni R, Cantoni C, Verdiani S, Malnati MS, Vitale M, Bottino C, Moretta L, Moretta A, Long EO: **Molecular clones of the p58 NK cell receptor reveal immunoglobulin-related molecules with diversity in both the extra- and intracellular domains.** *Immunity* 1995, **2**:439-49.
- Wagtmann N, Rajagopalan S, Winter CC, Peruzzi M, Long EO: **Killer cell inhibitory receptors specific for HLA-C and HLA-B identified by direct binding and by functional transfer.** *Immunity* 1995, **3**:801-9.
- Colonna M, Navarro F, Bellón T, Llano M, García P, Samaridis J, Angman L, Cella M, López-Botet M: **A common inhibitory receptor for major histocompatibility complex class I molecules on human lymphoid and myelomonocytic cells.** *J Exp Med* 1997, **186**:1809-18.
- D'Andrea A, Chang C, Franz-Bacon K, McClanahan T, Phillips JH, Lanier LL: **Molecular cloning of NKBI. A natural killer cell receptor for HLA-B allotypes.** *J Immunol* 1995, **155**:2306-10.
- Colonna M, Samaridis J: **Cloning of immunoglobulin-superfamily members associated with HLA-C and HLA-B recognition by human natural killer cells.** *Science* 1995, **268**:405-8.
- Long EO: **Regulation of immune responses through inhibitory receptors.** *Annu Rev Immunol* 1999, **17**:875-904.
- Moretta A, Bottino C, Vitale M, Pende D, Cantoni C, Mingari MC, Biassoni R, Moretta L: **Activating receptors and coreceptors involved in human natural killer cell-mediated cytotoxicity.** *Annu Rev Immunol* 2001, **19**:197-223.
- Lanier LL: **Up on the tightrope: natural killer cell activation and inhibition.** *Nat Immunol* 2008, **9**:495-502.
- Vivier E, Nunès JA, Vély F: **Natural killer cell signaling pathways.** *Science* 2004, **306**:1517-9.
- Gasser S, Raulet DH: **Activation and self-tolerance of natural killer cells.** *Immunol Rev* 2006, **214**:130-42.
- Watzl C, Long EO: **Signal transduction during activation and inhibition of natural killer cells.** *Curr Protoc Immunol* 2010, **Chapter 11**:Unit 11.9B.
- Raulet DH, Held W, Correa I, Dorfman JR, Wu MF, Corral L: **Specificity, tolerance and developmental regulation of natural killer cells defined by expression of class I-specific Ly49 receptors.** *Immunol Rev* 1997, **155**:41-52.
- Valiante NM, Uhrberg M, Shilling HG, Lienert-Weidenbach K, Arnett KL, D'Andrea A, Phillips JH, Lanier LL, Parham P: **Functionally and structurally distinct NK cell receptor repertoires in the peripheral blood of two human donors.** *Immunity* 1997, **7**:739-51.
- Höglund P, Ohlén C, Carbone E, Franksson L, Ljunggren HG, Latour A, Koller B, Kärre K: **Recognition of beta 2-microglobulin-negative (beta 2m-) T-cell blasts by natural killer cells from normal but not from beta 2m- mice: nonresponsiveness controlled by beta 2m- bone marrow in chimeric mice.** *Proc Natl Acad Sci USA* 1991, **88**:10332-6.
- Zimmer J, Donato L, Hanau D, Cazenave JP, Tongio MM, Moretta A, de la Salle H: **Activity and phenotype of natural killer cells in peptide transporter (TAP)-deficient patients (type I bare lymphocyte syndrome).** *J Exp Med* 1998, **187**:117-22.
- Fernandez NC, Treiner E, Vance RE, Jamieson AM, Lemieux S, Raulet DH: **A subset of natural killer cells achieves self-tolerance without expressing inhibitory receptors specific for self-MHC molecules.** *Blood* 2005, **105**:4416-23.



**Human NK cell education by inhibitory receptors for MHC class I.** *Immunity* 2006, **25**:331-42.



33. Yokoyama WM, Kim S: **Licensing of natural killer cells by self-major histocompatibility complex class I.** *Immunol Rev* 2006, **214**:143-54.
34. Raulet DH, Vance RE: **Self-tolerance of natural killer cells.** *Nat Rev Immunol* 2006, **6**:520-31.
35. Brodin P, Lakshmikanth T, Johansson S, Kärre K, Höglund P: **The strength of inhibitory input during education quantitatively tunes the functional responsiveness of individual natural killer cells.** *Blood* 2009, **113**:2434-41.



36. Joncker NT, Fernandez NC, Treiner E, Vivier E, Raulet DH: **NK cell responsiveness is tuned commensurate with the number of inhibitory receptors for self-MHC class I: the rheostat model.** *J Immunol* 2009, **182**:4572-80.



37. Brodin P, Kärre K, Höglund P: **NK cell education: not an on-off switch but a tunable rheostat.** *Trends Immunol* 2009, **30**:143-9.
38. Elliott JM, Yokoyama WM: **Unifying concepts of MHC-dependent natural killer cell education.** *Trends Immunol* 2011, **32**:364-72.
39. Höglund P, Brodin P: **Current perspectives of natural killer cell education by MHC class I molecules.** *Nat Rev Immunol* 2010, **10**:724-34.
40. Sun JC, Lanier LL: **Tolerance of NK cells encountering their viral ligand during development.** *J Exp Med* 2008, **205**:1819-28.



41. Tripathy SK, Keyel PA, Yang L, Pingel JT, Cheng TP, Schneeberger A, Yokoyama WM: **Continuous engagement of a self-specific activation receptor induces NK cell tolerance.** *J Exp Med* 2008, **205**:1829-41.



42. Orr MT, Lanier LL: **Natural killer cell education and tolerance.** *Cell* 2010, **142**:847-56.
43. Elliott JM, Wahle JA, Yokoyama WM: **MHC class I-deficient natural killer cells acquire a licensed phenotype after transfer into an MHC class I-sufficient environment.** *J Exp Med* 2010, **207**:2073-9.



44. Joncker NT, Shifrin N, Delebecque F, Raulet DH: **Mature natural killer cells reset their responsiveness when exposed to an altered MHC environment.** *J Exp Med* 2010, **207**:2065-72.



45. Juelke K, Killig M, Thiel A, Dong J, Romagnani C: **Education of hyporesponsive NK cells by cytokines.** *Eur J Immunol* 2009, **39**:2548-55.
46. Haas P, Loiseau P, Tamouza R, Cayuela J, Moins-Teisserenc H, Busson M, Henry G, Falk CS, Charron D, Socié G, Toubert A, Dulphy N: **NK-cell education is shaped by donor HLA genotype after unrelated allogeneic hematopoietic stem cell transplantation.** *Blood* 2011, **117**:1021-9.



47. Held W, Mariuzza RA: **Cis interactions of immunoreceptors with MHC and non-MHC ligands.** *Nat Rev Immunol* 2008, **8**:269-78.
48. Bessoles S, Angelov GS, Back J, Leclercq G, Vivier E, Held W: **Education of murine NK cells requires both cis and trans**

**recognition of MHC class I molecules.** *J Immunol* 2013, **191**:5044-51.



49. Chalifour A, Scarpellino L, Back J, Brodin P, Devèvre E, Gros F, Lévy F, Leclercq G, Höglund P, Beermann F, Held W: **A Role for cis Interaction between the Inhibitory Ly49A receptor and MHC class I for natural killer cell education.** *Immunity* 2009, **30**:337-47.



50. Jonsson AH, Yang L, Kim S, Taffner SM, Yokoyama WM: **Effects of MHC class I alleles on licensing of Ly49A+ NK cells.** *J Immunol* 2010, **184**:3424-32.
51. Ebihara T, Jonsson AH, Yokoyama WM: **Natural killer cell licensing in mice with inducible expression of MHC class I.** *Proc Natl Acad Sci USA* 2013, **110**:E4232-7.



52. Orr MT, Murphy WJ, Lanier LL: **'Unlicensed' natural killer cells dominate the response to cytomegalovirus infection.** *Nat Immunol* 2010, **11**:321-7.



53. Tarek N, Le Luduec J, Gallagher MM, Zheng J, Venstrom JM, Chamberlain E, Modak S, Heller G, Dupont B, Cheung NV, Hsu KC: **Unlicensed NK cells target neuroblastoma following anti-GD2 antibody treatment.** *J Clin Invest* 2012, **122**:3260-70.



54. Guia S, Jaeger BN, Piatek S, Maifert S, Trombik T, Fenis A, Chevrier N, Walzer T, Kerdiles YM, Marguet D, Vivier E, Ugolini S: **Confinement of activating receptors at the plasma membrane controls natural killer cell tolerance.** *Sci Signal* 2011, **4**:ra21.



55. Watzl C, Long EO: **Natural killer cell inhibitory receptors block actin cytoskeleton-dependent recruitment of 2B4 (CD244) to lipid rafts.** *J Exp Med* 2003, **197**:77-85.
56. Thomas LM, Peterson ME, Long EO: **Cutting edge: NK cell licensing modulates adhesion to target cells.** *J Immunol* 2013, **191**:3981-5.



57. Lanier LL, Le AM, Civin CI, Loken MR, Phillips JH: **The relationship of CD16 (Leu-11) and Leu-19 (NKH-1) antigen expression on human peripheral blood NK cells and cytotoxic T lymphocytes.** *J Immunol* 1986, **136**:4480-6.

58. Cooper MA, Fehniger TA, Turner SC, Chen KS, Ghaehri BA, Ghayur T, Carson WE, Caligiuri MA: **Human natural killer cells: a unique innate immunoregulatory role for the CD56(bright) subset.** *Blood* 2001, **97**:3146-51.

59. Jacobs R, Hintzen G, Kemper A, Beul K, Kempf S, Behrens G, Sykora KW, Schmidt RE: **CD56bright cells differ in their KIR repertoire and cytotoxic features from CD56dim NK cells.** *Eur J Immunol* 2001, **31**:3121-7.

60. Yu J, Mao HC, Wei M, Hughes T, Zhang J, Park I, Liu S, McClory S, Marcucci G, Trotta R, Caligiuri MA: **CD94 surface density identifies a functional intermediary between the CD56bright and CD56dim human NK-cell subsets.** *Blood* 2010, **115**:274-81.



61. Juelke K, Killig M, Luetke-Eversloh M, Parente E, Gruen J, Morandi B, Ferlazzo G, Thiel A, Schmitt-Knosalla I, Romagnani C: **CD62L expression identifies a unique subset of polyfunctional CD56dim NK cells.** *Blood* 2010, **116**:1299-307.



62. Lopez-Vergès S, Milush JM, Pandey S, York VA, Arakawa-Hoyt J, Pircher H, Norris PJ, Nixon DF, Lanier LL: **CD57 defines a functionally distinct population of mature NK cells in the human CD56dimCD16+ NK-cell subset.** *Blood* 2010, **116**: 3865-74.
63. Björkström NK, Riese P, Heuts F, Andersson S, Fauriat C, Ivarsson MA, Björklund AT, Flodström-Tullberg M, Michaëlsson J, Rottenberg ME, Guzmán CA, Ljunggren H, Malmberg K: **Expression patterns of NKG2A, KIR, and CD57 define a process of CD56dim NK-cell differentiation uncoupled from NK-cell education.** *Blood* 2010, **116**:3853-64.
64. Béziat V, Descours B, Parizot C, Debré P, Vieillard V: **NK cell terminal differentiation: correlated stepwise decrease of NKG2A and acquisition of KIRs.** *PLoS ONE* 2010, **5**:e11966.
65. Chan A, Hong D, Atzberger A, Kollnberger S, Filer AD, Buckley CD, McMichael A, Enver T, Bowness P: **CD56bright human NK cells differentiate into CD56dim cells: role of contact with peripheral fibroblasts.** *J Immunol* 2007, **179**:89-94.
66. Huntington ND, Legrand N, Alves NL, Jaron B, Weijer K, Plet A, Corcuff E, Mortier E, Jacques Y, Spits H, Di Santo, James P: **IL-15 trans-presentation promotes human NK cell development and differentiation in vivo.** *J Exp Med* 2009, **206**:25-34.
67. Romagnani C, Juelke K, Falco M, Morandi B, D'Agostino A, Costa R, Ratto G, Forte G, Carrega P, Lui G, Conte R, Strowig T, Moretta A, Münz C, Thiel A, Moretta L, Ferlazzo G: **CD56brightCD16- killer Ig-like receptor- NK cells display longer telomeres and acquire features of CD56dim NK cells upon activation.** *J Immunol* 2007, **178**:4947-55.
68. Nagler A, Lanier LL, Cwirla S, Phillips JH: **Comparative studies of human FcR111-positive and negative natural killer cells.** *J Immunol* 1989, **143**:3183-91.
69. Ellis TM, Fisher RI: **Functional heterogeneity of Leu 19<sup>+</sup> "bright" and Leu 19<sup>+</sup> "dim" lymphokine-activated killer cells.** *J Immunol* 1989, **142**:2949-54.
70. Fauriat C, Long EO, Ljunggren H, Bryceson YT: **Regulation of human NK-cell cytokine and chemokine production by target cell recognition.** *Blood* 2010, **115**:2167-76.
- F1000Prime RECOMMENDED**
71. Luetke-Eversloh M, Cicek BB, Siracusa F, Thom JT, Hamann A, Frischbutter S, Baumgrass R, Chang H, Thiel A, Dong J, Romagnani C: **NK cells gain higher IFN- $\gamma$  competence during terminal differentiation.** *Eur J Immunol* 2014, **44**:2074-84.
- F1000Prime RECOMMENDED**
72. Horowitz A, Strauss-Albee DM, Leipold M, Kubo J, Nemat-Gorgani N, Dogan OC, Dekker CL, Mackey S, Maecker H, Swan GE, Davis MM, Norman PJ, Guethlein LA, Desai M, Parham P, Blish CA: **Genetic and environmental determinants of human NK cell diversity revealed by mass cytometry.** *Sci Transl Med* 2013, **5**:208ra145.
- F1000Prime RECOMMENDED**
73. Vivier E, Raulet DH, Moretta A, Caligiuri MA, Zitvogel L, Lanier LL, Yokoyama WM, Ugolini S: **Innate or adaptive immunity? The example of natural killer cells.** *Science* 2011, **331**:44-9.
74. Min-Oo G, Kamimura Y, Hendricks DW, Nabekura T, Lanier LL: **Natural killer cells: walking three paths down memory lane.** *Trends Immunol* 2013, **34**:251-8.
75. Rölle A, Pollmann J, Cerwenka A: **Memory of infections: an emerging role for natural killer cells.** *PLoS Pathog* 2013, **9**: e1003548.
76. Sun JC, Ugolini S, Vivier E: **Immunological memory within the innate immune system.** *EMBO J* 2014, **33**:1295-303.
77. O'Leary JG, Goodarzi M, Drayton DL, von Andrian, Ulrich H: **T cell- and B cell-independent adaptive immunity mediated by natural killer cells.** *Nat Immunol* 2006, **7**:507-16.
- F1000Prime RECOMMENDED**
78. Paust S, Gill HS, Wang B, Flynn MP, Moseman EA, Senman B, Szczepanik M, Telenti A, Askenase PW, Compans RW, von Andrian, Ulrich H: **Critical role for the chemokine receptor CXCR6 in NK cell-mediated antigen-specific memory of haptens and viruses.** *Nat Immunol* 2010, **11**:1127-35.
- F1000Prime RECOMMENDED**
79. Peng H, Jiang X, Chen Y, Sojka DK, Wei H, Gao X, Sun R, Yokoyama WM, Tian Z: **Liver-resident NK cells confer adaptive immunity in skin-contact inflammation.** *J Clin Invest* 2013, **123**:1444-56.
80. Sun JC, Beilke JN, Lanier LL: **Adaptive immune features of natural killer cells.** *Nature* 2009, **457**:557-61.
- F1000Prime RECOMMENDED**
81. Nabekura T, Kanaya M, Shibuya A, Fu G, Gascoigne, Nicholas RJ, Lanier LL: **Costimulatory molecule DNAM-1 is essential for optimal differentiation of memory natural killer cells during mouse cytomegalovirus infection.** *Immunity* 2014, **40**:225-34.
- F1000Prime RECOMMENDED**
82. Sun JC, Madera S, Bezman NA, Beilke JN, Kaplan MH, Lanier LL: **Proinflammatory cytokine signaling required for the generation of natural killer cell memory.** *J Exp Med* 2012, **209**:947-54.
- F1000Prime RECOMMENDED**
83. Firth MA, Madera S, Beaulieu AM, Gasteiger G, Castillo EF, Schluns KS, Kubo M, Rothman PB, Vivier E, Sun JC: **Nfil3-independent lineage maintenance and antiviral response of natural killer cells.** *J Exp Med* 2013, **210**:2981-90.
- F1000Prime RECOMMENDED**
84. Beaulieu AM, Zawislak CL, Nakayama T, Sun JC: **The transcription factor Zbtb32 controls the proliferative burst of virus-specific natural killer cells responding to infection.** *Nat Immunol* 2014, **15**:546-53.
- F1000Prime RECOMMENDED**
85. Min-Oo G, Bezman NA, Madera S, Sun JC, Lanier LL: **Proapoptotic Bim regulates antigen-specific NK cell contraction and the generation of the memory NK cell pool after cytomegalovirus infection.** *J Exp Med* 2014, **211**:1289-96.
- F1000Prime RECOMMENDED**
86. Gumá M, Angulo A, Vilches C, Gómez-Lozano N, Malats N, López-Botet M: **Imprint of human cytomegalovirus infection on the NK cell receptor repertoire.** *Blood* 2004, **104**:3664-71.
87. Gumá M, Budt M, Sáez A, Brckalo T, Hengel H, Angulo A, López-Botet M: **Expansion of CD94/NKG2C+ NK cells in response to human cytomegalovirus-infected fibroblasts.** *Blood* 2006, **107**:3624-31.
88. Della Chiesa M, Falco M, Bertaina A, Muccio L, Alicata C, Frassoni F, Locatelli F, Moretta L, Moretta A: **Human cytomegalovirus infection promotes rapid maturation of NK cells expressing activating killer Ig-like receptor in patients transplanted with NKG2C-/- umbilical cord blood.** *J Immunol* 2014, **192**:1471-9.
- F1000Prime RECOMMENDED**
89. Björkström NK, Lindgren T, Stoltz M, Fauriat C, Braun M, Evander M, Michaëlsson J, Malmberg K, Klingström J, Ahlm C, Ljunggren H: **Rapid expansion and long-term persistence of elevated NK cell**

numbers in humans infected with hantavirus. *J Exp Med* 2011, **208**:13-21.



90. Brunetta E, Fogli M, Varchetta S, Bozzo L, Hudspeth KL, Marcenaro E, Moretta A, Mavilio D: **Chronic HIV-1 viremia reverses NKG2A/NKG2C ratio on natural killer cells in patients with human cytomegalovirus co-infection.** *AIDS* 2010, **24**:27-34.
91. Béziat V, Dalgard O, Asselah T, Halfon P, Bedossa P, Boudifa A, Hervier B, Theodorou I, Martinot M, Debré P, Björkström NK, Malmberg K, Marcellin P, Vieillard V: **CMV drives clonal expansion of NKG2C+ NK cells expressing self-specific KIRs in chronic hepatitis patients.** *Eur J Immunol* 2012, **42**:447-57.
92. Lopez-Vergès S, Milush JM, Schwartz BS, Pando MJ, Jarjoura J, York VA, Houchins JP, Miller S, Kang S, Norris PJ, Nixon DF, Lanier LL: **Expansion of a unique CD57<sup>+</sup>NKG2Chi natural killer cell subset during acute human cytomegalovirus infection.** *Proc Natl Acad Sci USA* 2011, **108**:14725-32.
93. Hendricks DW, Balfour HH, Dunmire SK, Schmeling DO, Hogquist KA, Lanier LL: **Cutting edge: NKG2C(hi)CD57+ NK cells respond specifically to acute infection with cytomegalovirus and not Epstein-Barr virus.** *J Immunol* 2014, **192**:4492-6.
94. Cooper MA, Elliott JM, Keyel PA, Yang L, Carrero JA, Yokoyama WM: **Cytokine-induced memory-like natural killer cells.** *Proc Natl Acad Sci USA* 2009, **106**:1915-9.
95. Romee R, Schneider SE, Leong JW, Chase JM, Keppel CR, Sullivan RP, Cooper MA, Fehniger TA: **Cytokine activation induces human memory-like NK cells.** *Blood* 2012, **120**:4751-60.
96. Ni J, Miller M, Stojanovic A, Garbi N, Cerwenka A: **Sustained effector function of IL-12/15/18-preactivated NK cells against established tumors.** *J Exp Med* 2012, **209**:2351-65.
97. Chijioke O, Münz C: **Dendritic cell derived cytokines in human natural killer cell differentiation and activation.** *Front Immunol* 2013, **4**:365.
98. Cui Y, Yang X, Zhu W, Li J, Wu X, Pang Y: **Immune response, clinical outcome and safety of dendritic cell vaccine in combination with cytokine-induced killer cell therapy in cancer patients.** *Oncol Lett* 2013, **6**:537-41.
99. Lehmann D, Spanholtz J, Sturtzel C, Tordoir M, Schlechta B, Groenewegen D, Hofer E: **IL-12 directs further maturation of ex vivo differentiated NK cells with improved therapeutic potential.** *PLoS ONE* 2014, **9**:e87131.
100. Hwang I, Zhang T, Scott JM, Kim AR, Lee T, Kakarla T, Kim A, Sunwoo JB, Kim S: **Identification of human NK cells that are deficient for signaling adaptor FcR $\gamma$  and specialized for antibody-dependent immune functions.** *Int Immunol* 2012, **24**:793-802.
101. Zhang T, Scott JM, Hwang I, Kim S: **Cutting edge: antibody-dependent memory-like NK cells distinguished by FcR $\gamma$  deficiency.** *J Immunol* 2013, **190**:1402-6.
102. Sun JC, Beilke JN, Bezman NA, Lanier LL: **Homeostatic proliferation generates long-lived natural killer cells that respond against viral infection.** *J Exp Med* 2011, **208**:357-68.
103. Pric M, Blazar BR, Farrar MA, Jameson SC: **In vivo survival and homeostatic proliferation of natural killer cells.** *J Exp Med* 2003, **197**:967-76.
104. Jamieson AM, Isnard P, Dorfman JR, Coles MC, Raulet DH: **Turnover and proliferation of NK cells in steady state and lymphopenic conditions.** *J Immunol* 2004, **172**:864-70.
105. Ranson T, Vosshenrich, Christian AJ, Corcuff E, Richard O, Müller W, Di Santo, James P: **IL-15 is an essential mediator of peripheral NK-cell homeostasis.** *Blood* 2003, **101**:4887-93.
106. Majewska-Szczepanik M, Paust S, von Andrian, Ulrich H, Askenase PV, Szczepanik M: **Natural killer cell-mediated contact sensitivity develops rapidly and depends on interferon- $\alpha$ , interferon- $\gamma$  and interleukin-12.** *Immunology* 2013, **140**:98-110.
107. Leong JW, Chase JM, Romee R, Schneider SE, Sullivan RP, Cooper MA, Fehniger TA: **Preactivation with IL-12, IL-15, and IL-18 induces CD25 and a functional high-affinity IL-2 receptor on human cytokine-induced memory-like natural killer cells.** *Biol Blood Marrow Transplant* 2014, **20**:463-73.
108. Lee S, Fragoso MF, Biron CA: **Cutting edge: a novel mechanism bridging innate and adaptive immunity: IL-12 induction of CD25 to form high-affinity IL-2 receptors on NK cells.** *J Immunol* 2012, **189**:2712-6.
109. Bezman NA, Kim CC, Sun JC, Min-Oo G, Hendricks DW, Kamimura Y, Best JA, Goldrath AW, Lanier LL: **Molecular definition of the identity and activation of natural killer cells.** *Nat Immunol* 2012, **13**:1000-9.
110. Zawislak CL, Beaulieu AM, Loeb GB, Karo J, Canner D, Bezman NA, Lanier LL, Rudensky AY, Sun JC: **Stage-specific regulation of natural killer cell homeostasis and response against viral infection by microRNA-155.** *Proc Natl Acad Sci USA* 2013, **110**:6967-72.
111. Sullivan RP, Leong JW, Fehniger TA: **MicroRNA regulation of natural killer cells.** *Front Immunol* 2013, **4**:44.
112. Trotta R, Chen L, Ciarlariello D, Josyula S, Mao C, Costinean S, Yu L, Butchar JP, Tridandapani S, Croce CM, Caligiuri MA: **miR-155 regulates IFN- $\gamma$  production in natural killer cells.** *Blood* 2012, **119**:3478-85.
113. Zediak VP, Wherry EJ, Berger SL: **The contribution of epigenetic memory to immunologic memory.** *Curr Opin Genet Dev* 2011, **21**:154-9.
114. Weng N, Araki Y, Subedi K: **The molecular basis of the memory T cell response: differential gene expression and its epigenetic regulation.** *Nat Rev Immunol* 2012, **12**:306-15.
115. Cichocki F, Miller JS, Anderson SK, Bryceson YT: **Epigenetic regulation of NK cell differentiation and effector functions.** *Front Immunol* 2013, **4**:55.
116. Gillard GO, Bivas-Benita M, Hovav A, Grandpre LE, Panas MW, Seaman MS, Haynes BF, Letvin NL: **Thy1+ NK [corrected] cells from vaccinia virus-primed mice confer protection against vaccinia virus challenge in the absence of adaptive lymphocytes.** *PLoS Pathog* 2011, **7**:e1002141.
117. Tong L, Assenmacher M, Zänker KS, Jahn P: **Virus-specific peptide dependent NK cell cytotoxicity.** *Inflamm Allergy Drug Targets* 2014, **13**:128-33.

