

## Research Article

# Evaluation of Halogenated Coumarins for Antimosquito Properties

Venugopala K. Narayanaswamy,<sup>1,2</sup> Raquel M. Gleiser,<sup>3</sup> Kabange Kasumbwe,<sup>2</sup>  
Bandar E. Aldhubiab,<sup>1</sup> Mahesh V. Attimarad,<sup>1</sup> and Bharti Odhav<sup>2</sup>

<sup>1</sup>Department of Pharmaceutical Sciences, College of Clinical Pharmacy, King Faisal University, Al-Ahsa 31982, Saudi Arabia

<sup>2</sup>Department of Biotechnology and Food Technology, Durban University of Technology, Durban 4001, South Africa

<sup>3</sup>CREAN-IMBIV (CONICET-UNC), Facultad de Ciencias Agropecuarias, and FCEFYN, Universidad Nacional de Córdoba, Avenida Valparaíso s/n, 5016 Córdoba, Argentina

Correspondence should be addressed to Venugopala K. Narayanaswamy; [kvenugopala@kfu.edu.sa](mailto:kvenugopala@kfu.edu.sa) and Bharti Odhav; [odhavb@dut.ac.za](mailto:odhavb@dut.ac.za)

Received 26 August 2014; Accepted 2 December 2014; Published 25 December 2014

Academic Editor: Valdir Cechinel Filho

Copyright © 2014 Venugopala K. Narayanaswamy et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Mosquitoes are the major vectors of parasites and pathogens affecting humans and domestic animals. The widespread development of insecticide resistance and negative environmental effects of most synthetic compounds support an interest in finding and developing alternative products against mosquitoes. Natural coumarins and synthetic coumarin analogues are known for their several pharmacological properties, including being insecticidal. In the present study halogenated coumarins (3-mono/dibromo acetyl, 6-halogenated coumarin analogues) were screened for larvicidal, adulticidal, and repellent properties against *Anopheles arabiensis*, a zoophilic mosquito that is one of the dominant vectors of malaria in Africa. Five compounds exerted 100% larval mortality within 24 h of exposure. All coumarins and halogenated coumarins reversibly knocked down adult mosquitoes but did not kill them after 24 h of exposure. Repellent properties could not be evidenced. Five compounds were considered potential larvicidal agents for further research and development, while adulticidal activity was considered only mild to moderate.

## 1. Introduction

Mosquitoes are the major vectors of parasites and pathogens that cause malaria, filariasis, dengue fever, yellow fever, Japanese encephalitis, and other fevers affecting humans and domestic animals [1]. Vector control programs mainly use four classes of chemical insecticides: organochlorines, organophosphates, carbamates, and pyrethroids. Bacterial insecticides and insect growth regulators have also become more widely used in recent years. However, use of chemicals on a vast and increasing scale has led to the widespread development of resistance as a result of selection for certain genes [2], and some species have even become resistant to multiple insecticides [3]. Mosquito resistance to at least one insecticide used for malaria control has been identified in 64 countries [3]. Besides, synthetic organic insecticides, such as

those currently used to control mosquitoes, affect nontarget organisms and result in negative environmental effects [4]. Thus, there is an interest in the finding and development of alternative antimosquito products.

Coumarins (2*H*-1-benzopyran-2-one) are a class of phenolic substances found as secondary metabolites from plants, bacteria, and fungi, widely used as additives in food, perfumes, cosmetics, pharmaceuticals, optical brighteners, and dispersed fluorescent. Natural coumarins are known for their several pharmacological properties and have been recently reviewed by Venugopala et al. [5], also reported for synthetic coumarin analogues, such as analgesic, anti-inflammatory [6, 7], anticoagulant [8], antibacterial [9–11], antifungal [12], antiviral, anticancer [13], antihypertensive [14], antitubercular [15], antihyperglycemic [16], and antioxidant [17] properties. Warfarin is a synthetic coumarin analogue (known

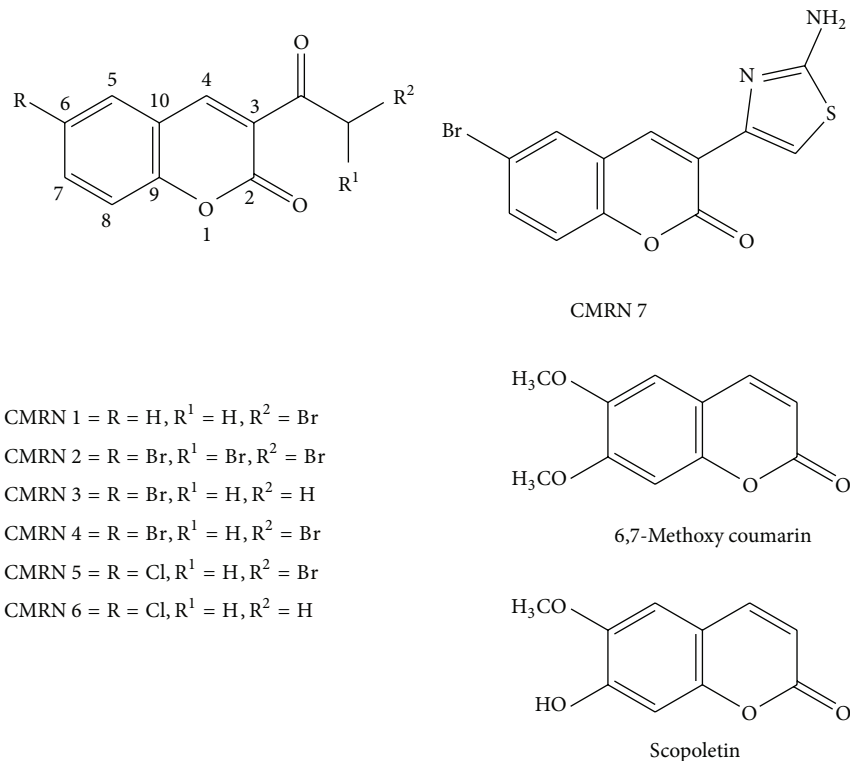


FIGURE 1: Synthetic halogenated coumarin compounds CMRN 1–CMRN 7, 6,7-dimethoxy coumarin, and scopoletin tested for antimosquito properties.

as Coumadin) that is used as an anticoagulant and is commercially available in the market with a trade name Coumadin. Essential oils and solvent extracts of plants containing coumarin have shown promising properties against mosquitoes. For example, coumarin extracted from southernwood (*Artemisia abrotanum* L.) and essential oil of carnation flowers (*Dianthus caryophyllus* L.) exerted a repellent effect against yellow fever mosquitoes (*Aedes aegypti* L.) and ticks (nymphs of *Ixodes ricinus* L.) [18]. Eight coumarin derivatives obtained from hexane extractions of the roots of *Esenbeckia grandiflora* Mart. were effective larvicides against *Ae. aegypti* [19]. The coumarin derivative pachyrrhizine, a compound from tubers of *Neorautanenia mitis*, showed larvicidal and adulticidal activities against *Anopheles gambiae* and *Culex quinquefasciatus* Say that were comparable to deltamethrin and alpha-cypermethrin, two standard mosquitoicides [20]. A component of *Tagetes lucida* Cav. hexane extract, 7-methoxy coumarin, showed larvicidal activity against *Ae. aegypti* [21]. The linear furanocoumarin imperatorin and the coumarin osthole extracted from *Cnidium monnieri* (L.) Cusson fruit were effective against larvae of *Culex pipiens pallens* Coquillett and *Ae. aegypti* and against *C. p. pallens* larvae resistant to various insecticides, suggesting that these coumarins and the pyrethroid and organophosphate insecticides do not share a common mode of action or elicit cross-resistance [22]. In this context, and in continuation of our search for novel chemical agents with antimosquito properties [23, 24], in the present study we undertake the screening of halogenated coumarins (3-mono/dibromo acetyl, 6-halogenated

coumarin analogues) for larvicidal and repellent properties against *Anopheles arabiensis*, one of the dominant vectors of malaria in Africa [25].

## 2. Materials and Methods

**2.1. Chemicals Tested.** The test compounds halogenated coumarin analogues (CMRN 1–CMRN 7) were synthesized as described previously [6, 7, 9–11, 26–28], using chemicals from Aldrich and Merck chemical company without further purification. Compounds 6,7-methoxy coumarin and scopoletin were obtained from Sigma-Aldrich. The products assessed were 3-(2-bromoacetyl)-2H-chromen-2-one (CMRN 1) [7, 11] and 6-bromo-3-(2,2-dibromoacetyl)-2H-chromen-2-one (CMRN 2) [28]; 3-acetyl-6-bromo-2H-chromen-2-one (CMRN 3) [6], 6-bromo-3-(2-bromoacetyl)-2H-chromen-2-one (CMRN 4) [9], and 3-(2-bromoacetyl)-6-chloro-2H-chromen-2-one (CMRN 5) [10]; 3-acetyl-6-chloro-2H-chromen-2-one (CMRN 6) [26]; 3-(2-aminothiazol-4-yl)-6-bromo-2H-chromen-2-one (CMRN 7) [27]; 6,7-methoxy coumarin; and scopoletin (Figure 1). The physicochemical characteristics of the halogenated coumarin analogues (CMRN 1–CMRN 7) are summarized in Table 1.

**2.2. Larvicidal Activity.** The *Anopheles arabiensis* used were from a colonized strain from Zimbabwe which had been reared according to the WHO (1975) guidelines [1] in an insectary simulating the temperature (27.5°C), humidity

TABLE 1: Physicochemical characteristics of 3-mono/dibromoacetyl-6-halogenated coumarin analogues CMRN 1–CMRN 7.

Code	M. F (M. Wt.)	Yield (%) <sup>a,b</sup>	m.p. (°C) reported	m.p. (°C) found	Mass (m/z)	c log P <sup>c</sup>
CMRN 1	C <sub>11</sub> H <sub>7</sub> BrO <sub>3</sub> (265)	96	120–122	121	266 (M + 1)	1.4023
CMRN 2	C <sub>11</sub> H <sub>5</sub> Br <sub>3</sub> O <sub>3</sub> (421)	95	146–148	147	422 (M + 1)	3.4753
CMRN 3	C <sub>11</sub> H <sub>7</sub> BrO <sub>3</sub> (265)	98	220–222	221	266 (M + 1)	2.0193
CMRN 4	C <sub>11</sub> H <sub>6</sub> Br <sub>2</sub> O <sub>3</sub> (343)	95	204–206	205	344 (M + 1)	2.2723
CMRN 5	C <sub>11</sub> H <sub>6</sub> BrClO <sub>3</sub> (299)	94	180–182	181	300 (M + 1)	2.1223
CMRN 6	C <sub>11</sub> H <sub>7</sub> ClO <sub>3</sub> (222)	95	218–220	219	223 (M + 1)	1.8693
CMRN 7	C <sub>12</sub> H <sub>7</sub> BrN <sub>2</sub> O <sub>2</sub> S (321)	87	210–212	211	322 (M + 1)	2.6992

<sup>a</sup>All of the products were characterized by spectral and physical data.

<sup>b</sup>Yields were on isolated basis.

<sup>c</sup>c log P was calculated using ChemBioDraw Ultra 13.0 v.

TABLE 2: Mortality of *Anopheles arabiensis* larvae exposed to coumarins and halogenated coumarins at 4 µg/mL and their negative (acetone) and positive (Temephos) controls and adult knock-down activity after 2-minute exposure to halogenated coumarins at 1000 µg/mL and their negative (acetone) and positive (DEET) controls against repellence assays (adjusted means and standard errors).

Compound code	Larval mortality	Knocked down
CMRN 1	98.9 ± 3.1 <sup>a</sup>	100.0 ± 10.9 <sup>a</sup>
CMRN 2	97.8 ± 3.1 <sup>a</sup>	96.7 ± 10.9 <sup>ab</sup>
CMRN 3	1.1 ± 3.1 <sup>b</sup>	91.1 ± 10.9 <sup>b</sup>
CMRN 4	98.9 ± 3.1 <sup>a</sup>	91.1 ± 10.9 <sup>b</sup>
CMRN 5	97.8 ± 3.1 <sup>a</sup>	97.8 ± 10.9 <sup>ab</sup>
CMRN 6	1.1 ± 3.1 <sup>b</sup>	95.6 ± 10.9 <sup>ab</sup>
CMRN 7	98.9 ± 3.1 <sup>a</sup>	81.1 ± 10.9 <sup>c</sup>
6,7-Methoxy coumarin	3.3 ± 3.1 <sup>b</sup>	96.7 ± 10.9 <sup>ab</sup>
Scopoletin	0.0 ± 3.1 <sup>b</sup>	96.7 ± 10.9 <sup>ab</sup>
Acetone (control)	0.0 ± 3.1 <sup>b</sup>	0.0 ± 10.9 <sup>d</sup>
Temephos	100.0 ± 3.1 <sup>a</sup>	
DEET		0.0 ± 10.9 <sup>d</sup>

<sup>a-d</sup>Within each column, compounds not sharing a letter differ significantly ( $P < 0.05$ ).

(70%), and lighting (12/12) of a malaria endemic environment. One mL of test compound (1 mg/mL) was added to distilled water (250 mL) producing a final concentration of 4 µg/mL. Thirty 3rd instar larvae were placed in the container. A negative control was set up using a solvent (acetone) and distilled water and a positive control included Temephos (Mostop; Agrivo), an effective emulsifiable organophosphate larvicidal used by the malarial control program. Each container was monitored for larval mortality at 24 h intervals for a period of three days and fed specially made cat food with reduced oil/fat content at regular intervals. Bioassays were triplicated. The percentage mortality was calculated relative to the initial number of exposed larvae. The larvicidal results are tabulated in Table 2.

2.3. *Insecticidal Activity.* Insecticidal activity assessment was conducted by exposing susceptible adult mosquitoes to a treated surface, in accordance with WHO protocol (1975) [1].

One mL of test compound solution (1 mg/mL) was sprayed onto a clean, dry, nonporous ceramic tile using a precalibrated Potter's Tower apparatus [8]. The tiles were then air-dried and the assay was initiated within 24 h of spraying, by fixing a cone over the sprayed tile and introducing thirty non-blood-fed, 2–5-day-old susceptible adult *A. arabiensis* mosquitoes into the cone. The effect of the test compounds was measured by determining the knockdown rate, which was based on temporary paralysis of the mosquitoes during a 60 min exposure period, and mortality 24 h postexposure. Deltamethrin (15 g/L; K-Othrine) was used as a positive control and acetone as a negative control. All bioassays were triplicate to ensure validity of results.

2.4. *Repellence Assays.* Repellent activity was assessed by topical application of the compound to skin and subsequent exposure of the treated areas of skin to unfed female mosquitoes. Ethical approval for the use of *Mastomys coucha* in these trials was approved from the Medical Research Council's Ethics Committee for Research on Animals. Adult *Mastomys* rodents were weighed individually and injected intraperitoneally with the correct concentration of sodium pentobarbital in comparison to the weight of the animal. The anesthetized rodents were then shaved on the ventral surface and a test compound (1 mL) was applied to each rodent's abdomen. Acetone was used as a solvent for the preparation of stock solution (1 mg/mL). Laboratory grade DEET (IUPAC: *N,N*-Diethyl-3-methylbenzamide) was used as the positive control and plain acetone was used as negative control.

Paper cups (500 mL) were modified by replacing the base of the cup with mosquito netting held in place with a rubber band and covering the mouth of the cup with transparent plastic film. Thirty unfed 4-day-old *A. arabiensis* females were introduced into the cup that was held in contact with the treated ventral surface of each rodent. Mosquito activity was observed through the transparent plastic film. After a period of 2 min, the numbers of mosquitoes probing were recorded. The cups holding the mosquitoes were removed and mosquitoes were then observed for 24 h. All tests were triplicated. The rodent was then returned to the animal facility and allowed to recover from anaesthetic. No adverse reactions to the applied components were observed on any of the *Mastomys* rodents during the 3 days they were monitored.

Repellence of the extracts was calculated using the following formula;

$$\begin{aligned} & \text{Percentage mosquitoes repelled} \\ & = \frac{\text{Number repelled}}{\text{Number introduced}} \times 100. \end{aligned} \quad (1)$$

**2.5. Statistical Analysis.** Generalized linear models assuming a Gaussian distribution were used to determine differences between treatments registered in larval mortality (larvicidal assays), adulticidal effects, and knockdown (in repellence assays). LSD Fisher test was used for post hoc analyses. In all cases, a value of  $P < 0.05$  was considered statistically significant.

### 3. Results and Discussion

The title compounds obtained were in good yields (87–98%) and characterization was completed by GCMS analysis. Purity of the compounds was confirmed by HPLC and it was more than 99%.  $c\text{Log}P$  of the compounds was calculated using ChemBioDraw Ultra 13.0 v and values were in the range of 1.8693–3.4753.

There was a significant effect of treatment on larval mortality ( $P < 0.001$ ) (Table 2). The highest activity was detected with CMRN 1, CMRN 2, CMRN 4, CMRN 5, and CMRN 7 showing close to 100% mortality after 24 h of exposure, which was the same as for the positive control Temephos. Compounds CMRN 3, CMRN 6, scopoletin, and 6,7-methoxy coumarin showed a statistically lower mortality that was equivalent to the negative control.

The promising larvicidal activity of CMRN 1, CMRN 2, CMRN 4, CMRN 5, and CMRN 7, which was comparable to the positive control Temephos, may be attributed to the presence of electron withdrawing halogen atoms (bromine and chlorine) on acetyl group at the third and sixth positions of the coumarin nucleus. Larvicidal and ovicidal activity of 4-methyl-7-hydroxy coumarin derivate against vectors *Aedes aegypti* and *Culex quinquefasciatus* have also been attributed to bromine atoms present at C-5 and C-8 positions [29]. Coumarin and mainly furanocoumarins can alter the detoxication capability of an organism, by reversibly or irreversibly inhibiting cytochrome P450 detoxication enzymes [30, 31].

On adulticidal assays, adult mosquito mortality of positive control K-Othrine showed 100% knockdown/mortality from the first 60 min of exposure, while the natural coumarin, synthetic compounds, and negative control did not knockdown mosquitoes throughout the 24 h observation period.

All components tested for repellence (except the controls) knocked down mosquitoes within the 2 min exposure time, and CMRN 1, CMRN 2, CMRN 5, CMRN 6, 6,7-methoxy coumarin, and scopoletin were the most potent, knocking approximately 100% of them ( $P < 0.001$ ) (Table 2). However, 24 h after exposure, all mosquitoes recovered. The few mosquitoes exposed to the coumarin analogues that were not knocked down did not attempt to bite; because of the low number of mosquitoes remaining active, no further statistical analyses were carried out.

An insect immobilization effect of coumarins has been reported for other insects [32, 33], and a slowly developing paralysis eventually leading to death has been a major feature of insect poisoning by coumarins such as surangin B [32]. Bioenergetic disruption of muscle has been determined as a prominent mechanism underlying the insecticidal action of surangin B [34]. Moreover, homology modeling and docking studies indicate that coumarin, as well as other terpene compounds, may act as acetylcholinesterase inhibitors and can block the octopamine receptor pathway and thus be neurotoxic against mosquitoes [35]. However, knockdown of mosquitoes resulting from short exposure to 3-mono/dibromoacetyl-6-halogenated coumarin analogues CMRN 1–CMRN 7 was fast (2 min or less) and reversible, and no adulticidal effects were detected from 60 min exposure. This may be partly explained by differences in experimental design, since topical application and injection were common procedures in previous reports on insecticidal properties [32, 33]. Besides mortality, coumarins may exert other negative effects on insect populations, such as decreasing their reproductive potential [36], which were not assessed in the present study.

Because no (active) mosquitoes were observed attempting to bite the rodent treated with coumarin analogues, further assessment of the repellent properties of these compounds after different treatment times and concentrations merits further testing.

### 4. Conclusions

The present study evaluates 3-mono/dibromoacetyl-6-halogenated coumarin analogues CMRN 1–CMRN 7, scopoletin, and 6,7-methoxy coumarin for larvicidal and repellent effects against the malaria vector *A. arabiensis*. Compounds CMRN 1, CMRN 2, CMRN 4, CMRN 5, and CMRN 7 were considered potential larvicidal agents for further research and development, because these compounds exerted close to 100% mortality within 24 h of exposure. Adulticidal activity on the other hand was considered negligible and repellence should be further explored. Of particular interest are the bromo- and chloroanalogues of CMRN 2, CMRN 4, and CMRN 5 that have potential to be used to prevent and control malaria by controlling the vector *A. arabiensis*.

### Conflict of Interests

The authors declare that they have no conflict of interests.

### Acknowledgments

The authors are grateful to College of Clinical Pharmacy, King Faisal University, and Durban University of Technology, Durban, for facilities. The funding institutions had no involvement in the study design, in the collection, analysis, and interpretation of data, in the writing of the paper, or in the decision to submit the paper for publication. Raquel M. Gleiser is a Career Member of CONICET.

## References

- [1] M. W. Service, "Management of vectors," in *Pest and Vector Management in Tropics*, A. Youdeowei and M. Service, Eds., pp. 265–280, Longman, London, UK, 1983.
- [2] A. Baleta, "Insecticide resistance threatens malaria control in Africa," *The Lancet*, vol. 374, no. 9701, pp. 1581–1582, 2009.
- [3] WHOPEs, "Pesticides and their application for the control of vectors and pests of public health importance," in *WHO Pesticide Evaluation Scheme*, p. 114, WHOPEs, 6th edition, 2006.
- [4] D. Pimentel and H. Lehman, *The Pesticide Question: Environment, Economics and Ethics*, Chapman and Hall, London, UK, 1993.
- [5] K. N. Venugopala, V. Rashmi, and B. Odhav, "Review on natural coumarin lead compounds for their pharmacological activity," *BioMed Research International*, vol. 2013, Article ID 963248, 14 pages, 2013.
- [6] K. N. Venugopala and B. S. Jayashree, "Synthesis of carboxamides of 2'-amino-4'-(6-bromo-3-coumarinyl) thiazole as analgesic and antiinflammatory agents," *Indian Journal of Heterocyclic Chemistry*, vol. 12, no. 4, pp. 307–310, 2003.
- [7] K. N. Venugopala, B. S. Jayashree, and M. Attimarad, "Synthesis and evaluation of some substituted 2-arylamino coumarinyl thiazoles as potential NSAIDs," *Asian Journal of Chemistry*, vol. 16, no. 2, pp. 872–876, 2004.
- [8] C. Potter, "An improved laboratory apparatus for applying direct sprays and surface films, with data on the electrostatic charge on atomized spray fluids," *Annals of Applied Biology*, vol. 39, no. 1, pp. 1–28, 1952.
- [9] K. N. Venugopala and B. Jayashree, "Microwave-induced synthesis of schiff bases of aminothiazolyl bromocoumarins as antibacterials," *Indian Journal of Pharmaceutical Sciences*, vol. 70, no. 1, pp. 88–91, 2008.
- [10] G. K. Rao, K. N. Venugopala, and P. N. Sanjay Pai, "Microwave-assisted synthesis of some 6-chloro-3-[2-(substituted anilino)-1,3-thiazol-4-yl]-2H-1-benzopyran-2-ones as antibacterial agents," *Indian Journal of Heterocyclic Chemistry*, vol. 17, no. 4, pp. 397–400, 2008.
- [11] K. N. Venugopala, G. K. Rao, P. N. S. Pai, and G. L. Ganesh, "Synthesis and characterization of carboxamides of 2'-amino-4'-(3-(2H-1-benzopyran-2-one))thiazole as antimicrobial agents," *Asian Journal of Chemistry*, vol. 20, no. 3, pp. 1697–1701, 2008.
- [12] Y. Shi and C.-H. Zhou, "Synthesis and evaluation of a class of new coumarin triazole derivatives as potential antimicrobial agents," *Bioorganic and Medicinal Chemistry Letters*, vol. 21, no. 3, pp. 956–960, 2011.
- [13] K. Paul, S. Bindal, and V. Luxami, "Synthesis of new conjugated coumarin-benzimidazole hybrids and their anticancer activity," *Bioorganic and Medicinal Chemistry Letters*, vol. 23, no. 12, pp. 3667–3672, 2013.
- [14] K. M. Amin, F. M. Awadalla, A. A. M. Eissa, S. M. Abou-Seri, and G. S. Hassan, "Design, synthesis and vasorelaxant evaluation of novel coumarin-pyrimidine hybrids," *Bioorganic and Medicinal Chemistry*, vol. 19, no. 20, pp. 6087–6097, 2011.
- [15] A. Arshad, H. Osman, M. C. Bagley, C. K. Lam, S. Mohamad, and A. S. M. Zahariluddin, "Synthesis and antimicrobial properties of some new thiazolyl coumarin derivatives," *European Journal of Medicinal Chemistry*, vol. 46, no. 9, pp. 3788–3794, 2011.
- [16] B. C. Raju, A. K. Tiwari, J. A. Kumar et al., " $\alpha$ -glucosidase inhibitory antihyperglycemic activity of substituted chromenone derivatives," *Bioorganic and Medicinal Chemistry*, vol. 18, no. 1, pp. 358–365, 2010.
- [17] G. Melagraki, A. Afantitis, O. Igglessi-Markopoulou et al., "Synthesis and evaluation of the antioxidant and anti-inflammatory activity of novel coumarin-3-aminoamides and their alpha-lipoic acid adducts," *European Journal of Medicinal Chemistry*, vol. 44, no. 7, pp. 3020–3026, 2009.
- [18] H. Tunón, W. Thorsell, A. Mikiver, and I. Malander, "Arthropod repellency, especially tick (*Ixodes ricinus*), exerted by extract from *Artemisia abrotanum* and essential oil from flowers of *Dianthus caryophyllum*," *Fitoterapia*, vol. 77, no. 4, pp. 257–261, 2006.
- [19] P. E. S. De Oliveira, L. M. Conserva, A. C. Brito, and R. P. L. Lemos, "Coumarin derivatives from *Esenbeckia grandiflora* and its larvicidal activity against *Aedes aegypti*," *Pharmaceutical Biology*, vol. 43, no. 1, pp. 53–57, 2005.
- [20] C. C. Joseph, M. M. Ndoile, R. C. Malima, and M. H. H. Nkunya, "Larvicidal and mosquitocidal extracts, a coumarin, isoflavonoids and pterocarpanes from *Neorautanenia mitis*," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 98, no. 8, pp. 451–455, 2004.
- [21] A. Oranday, G. Martinez, A. Nuñez, C. Rivas, and A. E. Flores, "Coumarin isolated from *Tagetes lucida* Cav. exhibits larvicidal activity in *Aedes aegypti* (L.)," *Southwestern Entomologist*, vol. 33, no. 4, pp. 315–317, 2008.
- [22] Z. Wang, J.-R. Kim, M. Wang, S. Shu, and Y.-J. Ahn, "Larvicidal activity of *Cnidium monnieri* fruit coumarins and structurally related compounds against insecticide-susceptible and insecticide-resistant *Culex pipiens pallens* and *Aedes aegypti*," *Pest Management Science*, vol. 68, no. 7, pp. 1041–1047, 2012.
- [23] K. N. Venugopala, M. Krishnappa, S. K. Nayak et al., "Synthesis and antimosquito properties of 2,6-substituted benzo[d]thiazole and 2,4-substituted benzo[d]thiazole analogues against *Anopheles arabiensis*," *European Journal of Medicinal Chemistry*, vol. 65, pp. 295–303, 2013.
- [24] V. K. Narayanaswamy, R. M. Gleiser, R. K. Chalannavar, and B. Odhav, "Antimosquito properties of 2-substituted phenyl/benzylamino-6-(4-chlorophenyl)-5-methoxycarbonyl-4-methyl-3,6-dihydropyrimidin-1-ium chlorides against *Anopheles arabiensis*," *Medicinal Chemistry*, vol. 10, no. 2, pp. 211–219, 2014.
- [25] M. E. Sinka, M. J. Bangs, S. Manguin et al., "The dominant *Anopheles* vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic précis," *Parasites and Vectors*, vol. 3, no. 1, article 117, 2010.
- [26] D. Chopra, K. N. Venugopala, B. S. Jayashree, and T. N. G. Row, "3-acetyl-6-chloro-2H-chromen-2-one," *Acta Crystallographica Section E: Structure Reports Online*, vol. 62, no. 6, pp. o2310–o2312, 2006.
- [27] D. Chopra, A. R. Choudhury, K. N. Venugopala et al., "3-(2-Amino-1,3-thiazol-4-yl)-6-bromo-2H-chromen-2-one," *Acta Crystallographica Section E: Structure Reports*, vol. 65, no. 12, pp. 3047–3048, 2009.
- [28] D. Chopra, K. N. Venugopala, and G. K. Rao, "6-Bromo-3-(dibromo-acet-yl)-2H-chromen-2-one," *Acta Crystallographica Section E*, vol. 63, no. 12, p. 4872, 2007.
- [29] M. Deshmukh, P. Pawar, M. Joseph, U. Phalgune, R. Kashalkar, and N. R. Deshpanded, "Efficacy of 4-methyl-7-hydroxy coumarin derivatives against *Aedes aegypti* and *Culex quinquefasciatus*," *Indian Journal of Experimental Biology*, vol. 46, no. 11, pp. 788–792, 2008.

- [30] P. Letteron, V. Descatoire, D. Larrey, M. Tinel, J. Geneve, and D. Pessayre, "Inactivation and induction of cytochrome P-450 by various psoralen derivatives in rats," *Journal of Pharmacology and Experimental Therapeutics*, vol. 238, no. 2, pp. 685–692, 1986.
- [31] J. J. Neal, "Inhibition of insect cytochromes P450 by furano-coumarins," *Pesticide Biochemistry and Physiology*, vol. 50, no. 1, pp. 43–50, 1994.
- [32] R. A. Nicholson and A. G. Zhang, "Surangin B: insecticidal properties and mechanism underlying its transmitter-releasing action in nerve-terminal fractions isolated from mammalian brain," *Pesticide Biochemistry and Physiology*, vol. 53, no. 3, pp. 152–163, 1995.
- [33] M. D. Moreira, M. C. Picanço, L. C. D. A. Barbosa et al., "Plant compounds insecticide activity against Coleoptera pests of stored products," *Pesquisa Agropecuária Brasileira*, vol. 42, no. 7, pp. 909–915, 2007.
- [34] J. Zheng, D. Leong, G. Lees, and R. A. Nicholson, "Studies on the interaction of surangin B with insect mitochondria, insect synaptosomes, and rat cortical neurones in primary culture," *Pesticide Biochemistry and Physiology*, vol. 61, no. 1, pp. 1–13, 1998.
- [35] B. Khanikor, P. Parida, R. N. S. Yadav, and D. Bora, "Comparative mode of action of some terpene compounds against octopamine receptor and acetyl cholinesterase of mosquito and human system by the help of homology modeling and docking studies," *Journal of Applied Pharmaceutical Science*, vol. 3, no. 2, pp. 6–12, 2013.
- [36] R. Kaur and P. J. Rup, "Evaluation of regulatory influence of four plant growth regulators on the reproductive potential and longevity of melon fruit fly (*Bactrocera cucurbitae*)," *Phytoparasitica*, vol. 30, no. 3, pp. 224–230, 2002.