



Pesticide residues in *Rita rita* and *Cyprinus carpio* from river Ganga, India, and assessment of human health risk

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ABSTRACT

Present study was carried out to determine the concentration and bioaccumulation of pesticide residues in two commonly edible fishes: bagrid fish, *Rita rita* and common carp, *Cyprinus carpio* collected from river Ganga at Narora, India. The human health risk via consumption of these fishes was also assessed. The n-hexane extract of the muscle tissues was characterized by gas chromatography coupled to mass spectrometry and quantified by electron capture detector for pesticide residues. Bioaccumulation factor (BAF) in bagrid fish for detected pesticides was found to be higher than those in common carp. Estimated daily intake (EDI) values in our study were insignificantly higher than Average daily intake (ADI) values. Target hazard quotient (THQ) via consumption of selected fishes was found to be lower than the set 1.0, inferring non-carcinogenic risk. With regard to contaminants carcinogenic effects the total risk ratio (R) values of each pesticide was found lower than threshold risk limit except of heptachlor which indicates carcinogenic risk. The results justify pesticide pollution in river Ganga at Narora and thus more attention is required in order to help improve the health status of this ecosystem and reduce contamination of fishes.

1. Introduction

Pesticides are widely used in the world to control various pests in the crops. After application the pesticide residues tend to persist and enter the aquatic ecosystem organisms where they accumulate. Various studies have correlated the pesticide exposure with allergies, cancer, neuro abnormalities, endocrine dysfunctioning, abnormal physiology, developmental effects, headaches, stomachaches, vomiting, skin rash, coma etc., [1–6]. Regular consumption of food from pesticide contaminated source has both in-short duration (acute) and long-duration (chronic) effects. Acute pesticide poisoning has now become a rare evident however, long chronic toxicity caused by long duration exposure to low dose are commonly evident [6].

The river Ganga, the largest source of drinking water and irrigation in India also provides basic nutrition to the population living along the areas [7]. With the increase in population India has been undergoing rapid industrialization and economic development. Use of pesticides in agricultural sector has increased to hundred times to sustain more population in the country. Enormous quantities of pesticides are being applied along the Ganga river basin in agricultural fields [8–10]. Their residues finally find their way into the river by flash floods, leaching,

drainage and surface runoff. A large number of reports are available that show river Ganga is highly polluted [11–16]. At the international level, many reports are also available that show that the pesticide residues are present in water resources [17–20]. Presence of pesticide residues in the fish tissues [21–25] shows that these residues bio-accumulated along trophic level in the food chain.

In India, 60,000 MT of pesticides are being annually used of which maximum consumption occurs along river Ganga basin [26]. Besides the regular agricultural activities done along the Ganga basin, the dry beds of the river are used to grow vegetables and fruits, also add pesticides to the river during monsoon season.

Among all aquatic organisms fish is considered as suitable bio-indicator animal in monitoring environmental contamination. Fish uptakes pollutants directly through the water via gills, integuments, from the food intake and shows increased ability to bio-accumulate due to their lower mono-oxygenase (detoxifying enzyme) activity [27]. The pollutants present in the fish not only indicate persistence in the environment but also their transfer to other organisms through the food web. Fish as nutrition is an important source of not only proteins but also omega-3-polyunsaturated fatty acids which are recommended in cardiovascular diseases [28]. Besides fish fatty acids are used in the

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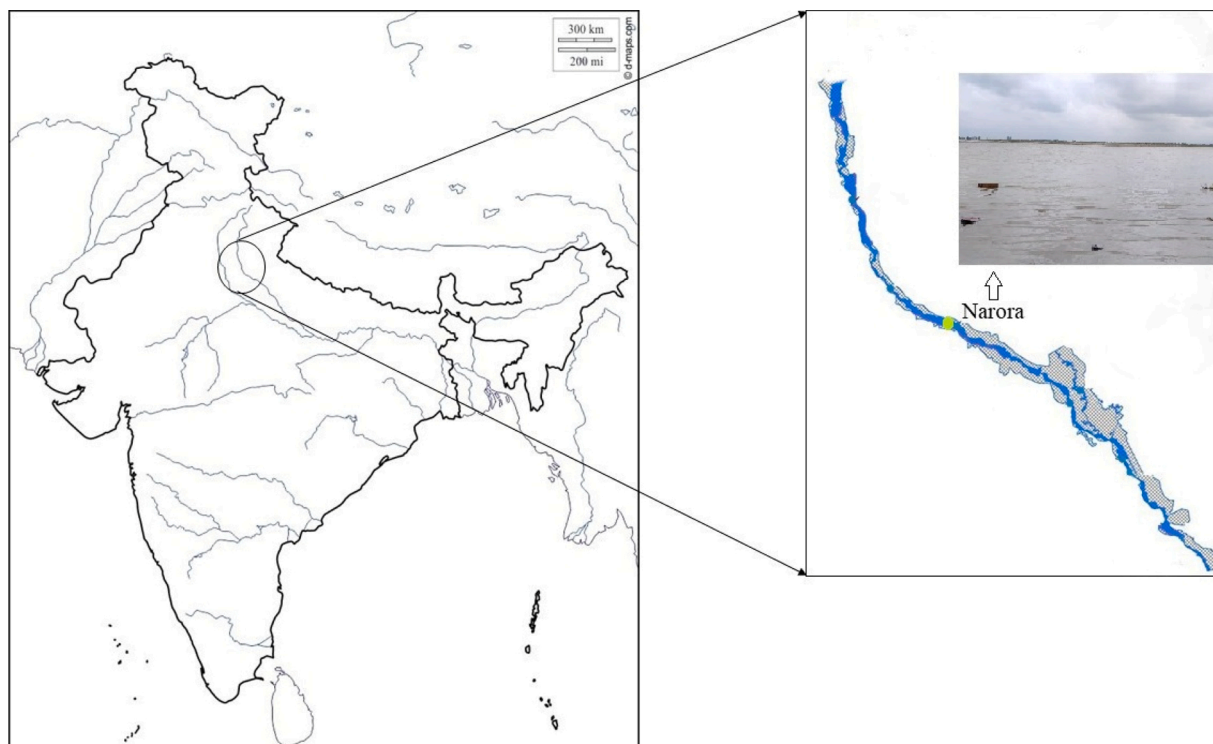


Fig. 1. Sketch map of the sampling site.

Table 1
Biometric data of the selected fishes collected from River Ganga.

Parameters	Name of fish	
	Bagrid fish (<i>Rita rita</i>) (n = 10)	Common carp (<i>Cyprinus carpio</i>) (n = 10)
Fish length (cm)	22.13 ± 4.13	27.33 ± 6.42
Fish weight (g)	350.71 ± 20	380.66 ± 17
Dietary habit ^a	Carnivorous	Omnivorous
Trophic level ^a	3.7	3.1
Lipid % (Muscle)	14.36 ± 1.19	8.49 ± 3.68

^a www. fish base org.

pharmaceutical and cosmetic preparations [29]. However, consumption of fish from the contaminated environment may causes accumulation of pollutants in the human body.

Pesticides risk assessment is given as a function of toxicological effects, that is usually expressed as the ratio of predicted environmental concentration and average daily consumption to average body weight. Around the globe various studies have been conducted to evaluate the health risk associated with consumption of pesticide contaminated fish [30–36] including number of studies from the country [37–39].

Therefore, for the present study we have chosen river Ganga at Narora as sampling station. The objective of the study was to estimate the concentration of pesticide residues in water and accumulation in two food fish species, bagrid catfish and common carp. Further the data were used to assess daily exposure and human health risk by the consumption of these fishes.

2. Materials and methods

2.1. Study site and the sampling

Fig. 1 displays the details of sampling site, ten fish samples of each

species and ten water samples were collected from Narora site (28°10'59"N 78°23'34"E) Uttar Pradesh of the river Ganga in August 2019. After collection the fishes were decapitated. Both the water and the fish samples were packed in ice-box and immediately brought to laboratory. The biometric data of both the fishes are given in Table 1.

2.1.1. Identification of fish

Fishes were identified following the keys given by Jhingran [40] and Fish Base org: -

Rita rita

Order: Siluriformes

Family: Bagridae

Fin formula: D 1 6; A ii 10-11; P 1 10; V I 6-7; Barbells 3 pairs

Cyprinus carpio

Order: Cypriniformes

Family: Cyprinidae

Fin formula: D 3-4/ 18-20; A 3-5; P₁ 1/15; P₂ 1/8

2.1.2. Sample preparation

In the laboratory, fishes were dissected and dorsal muscle tissues of the fish were taken out for pesticide residue analysis. Collected muscle samples 10 g of each sample was freeze-dried, grounded to fine powder, and stored at -20 °C before the process of extraction. The impurity particulates in the collected water samples were separated by filtration through 0.45-μm hydrophilic filters.

2.2. Sample extraction and clean up

2.2.1. Pesticides in water samples

Pesticides in water samples were analysed following the method of Muir and Sverko [41]. 2,4,5,6-tetrachloro-*m*-xylene (TCmX) as recovery surrogate was added in 1 L of filtered water sample. Liquid-liquid extraction with dichloromethane (35 mL) was performed. Na₂SO₄ column was used to remove water in the organic phase further *n*-hexane was used as an organic solvent. The column was packed from bottom to top with neutral silica, neutral alumina, and anhydrous sodium sulfate,

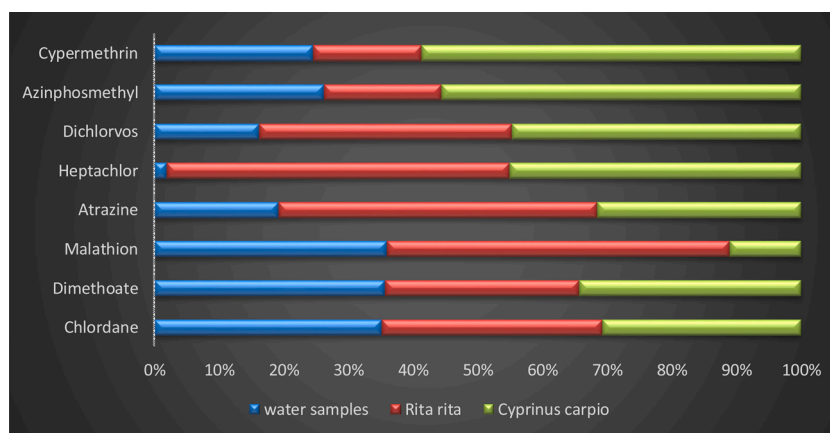


Fig. 2. Relative abundance of pesticides in water, muscle tissues of *Rita rita* and *Cyprinus carpio*.

to remove impurities in the extract. The extract was finally blown to dryness by purity nitrogen and the residues were redissolved with 20 μL of *n*-hexane.

2.2.2. Pesticides in fish samples

Soxhlet extraction method [42] was followed to extract pesticides in fish muscle. 10 g of muscle tissue after freeze dried were homogenized to fine powder with 35–40 g of activated anhydrous sodium sulfate. The prepared sample was packed and placed in extracting thimble of the Soxhlet apparatus. The mixture was extracted with 150 mL of acetone and *n*-hexane (20:80) v/v for 6 h. The extract was filtered and concentrated to 2 mL on water bath and transferred into (0.22 μm) membrane filter polyethylene (10 mL) syringe. The column was packed with neutral silica, acidic silica, florisil acidic alumina and sample mixture from bottom to top with other membrane filter placed on top. 15 mL of *n*-hexane was used to elute the packed column at a flow rate of 2 mL min^{-1} . High purity nitrogen at the gentle stream was used to dry eluent, followed by dry residues redissolution with 200 μL of *n*-hexane and addition of internal standard (pentachloronitrobenzene). Qualitative as well as quantitative analysis of analytes were done with GC–MS and GC-ECD. Fig. 2 depicts the relative abundance of pesticides detected in water samples and muscle tissues of the two fishes.

2.3. Determination of lipid content

Determination of lipid content was done gravimetrically [43]. 2 g of fine grounded muscle powder was dissolved in twenty millilitres of water, cyclohexane and 16 mL isopropyl (3:1) mixture. Ultrasonic extraction was done, mixture reached statically separated equilibrium, and the organic phase was collected. Extraction was repeated with 18 mL of cyclohexane and 6 mL isopropyl alcohol and combined with earlier then dried under a gentle nitrogen stream. The residue was weighed and recorded in mg and the percentage content was calculated per gram tissue sample.

2.4. Qualitative and quantitative analysis of pesticide residues by GC–MS

3 μL of sample was injected into a gas chromatography (Agilent 7890A, USA) instrument equipped with an electron capture detector (GC-ECD) (Agilent Technologies, USA) the analytical capillary column was DB-5 (30 m \times 0.25 mm i.d \times 0.25- μm film thickness, Agilent, USA). Nitrogen was used as carrier gas with the flow rate of (1 mL min^{-1}). Injector and detector temperatures were adjusted at 250 and 300 $^{\circ}\text{C}$. Started at 80 $^{\circ}\text{C}$ with 1 min hold, and the oven temperature was raised to 150 $^{\circ}\text{C}$ at 20 $^{\circ}\text{C}$ min^{-1} rate and finally to 300 $^{\circ}\text{C}$ (5 min hold) at the rate of 5 $^{\circ}\text{C}$ min^{-1} .

The instruments were calibrated with calibration standards during

analysis. Each sample was analyzed in duplicate. The recoveries of TCmX (surrogate standard) were $75 \pm 6\%$ in water samples and $68 \pm 6\%$ in fish samples. The recoveries of pesticides ranged from 73 to 100% in water samples and from 66 to 84% in fish samples. The method detection limits (MDLs) concentration of analytes were confirmed whose signal-to-noise (S/N) ratio was three and ranged from 0.05–100 $\mu\text{g L}^{-1}$ in water samples and from 0.01–100 $\mu\text{g g}^{-1}$ in fish samples. Concentration detected less than MDLs in samples was treated as not detected (nd).

2.5. Data analysis

The values of pesticides in both fishes were statistically analysed by Spearman correlation test by using SPSS statistical package (version 16.0; SPSS Inc., USA).

Bio-water accumulation factor (BAF) illustrates the partitioning of chemical between water and aquatic organisms, it provides the accumulation-scale of the contaminants in the organism.

BAF is calculated by the following equation:

$$\text{BAF} = C_f / c$$

Where C_f is the pollutant concentration in the fish ($\mu\text{g g}^{-1}$) normalized by lipid content of fish and c is concentration of pollutant in water ($\mu\text{g l}^{-1}$).

In order to determine potential human health risk of tested fishes, the estimate daily intake (EDI), target hazard quotients (THQ) and Carcinogenic risk (R) were calculated.

The target hazard quotients (THQs), and carcinogenic risk ratio (R) were used in risk assessment. The $\text{THQ} > 1$ denotes that the daily exposure may cause human health hazard effects.

The calculations of EDI [44], THQ, and R are done using the formulae [45]:

$$\text{EDI} = \frac{C \times W_F}{W_{AB}}$$

Where,

C = Concentration of pollutant in food ($\mu\text{g g}^{-1}$), W_F = Average daily fish consumption in India is 55 g day^{-1} person $^{-1}$, and W_{AB} = Average adult body weight (70 kg) ([45], Jiang et al. [46],[1]).

$$\text{THQ} = \frac{E_F \times E_D \times F_{IR} \times C}{R_{FD} \times W_{AB} \times T_A} \times 10^{-3}$$

$$R = \frac{E_F \times E_D \times F_{IR} \times S_F \times C}{W_{AB} \times T_A} \times 10^{-3}$$

Where,

E_F = frequency of exposure (350 days year $^{-1}$), E_D = duration of

Table 2

The concentration of pesticide residues in water ($\mu\text{g/l}$) and fish tissues ($\mu\text{g/g}$ ww) from river Ganga, India.

Pesticide	LogK _{ow}	Surface water	Rita rita	
			Muscle tissues	Cyprinus carpio
Chlordane	6.16	0.104 ± 0.33 (0.54–0.301)	0.101 ± 0.25 (0.67–0.278)	0.091 ± 0.52 (nd-0.187)
Dimethoate	0.78	0.082 ± 0.15 (0.051–0.191)	0.069 ± 0.64 (0.063–0.079)	0.079 ± 0.64 (0.071–0.095)
Malathion	2.36	0.055 ± 0.80 (nd-0.109)	0.081 ± 0.32 (0.075–0.097)	0.017 ± 0.71 (nd-0.111)
Atrazine	2.61	0.051 ± 0.19 (nd-0.104)	0.131 ± 0.19 (0.076–0.189)	0.087 ± 0.23 (0.055–0.104)
Heptachlor	6.10	0.006 ± 0.20 (nd-0.0024)	0.167 ± 0.20 (0.143–0.201)	0.142 ± 0.20 (0.123–0.209)
Dichlorvos	1.43	0.059 ± 0.17 (nd-0.132)	0.142 ± 0.11 (0.111–0.267)	0.163 ± 0.13 (0.109–0.267)
Azinphosmethyl	2.75	0.065 ± 0.80 (0.055–0.102)	0.045 ± 0.10 (nd-0.201)	0.138 ± 0.19 (0.117–0.197)
Cypermethrin	6.60	0.076 ± 0.93 (0.061–0.097)	0.052 ± 0.47 (nd-0.177)	0.182 ± 0.52 (0.154–0.235)

Data shown as mean ± standard deviation; maximum and minimum concentrations are in parenthesis.

exposure (70 years), F_{IR} = Average daily fish consumption in India is 55 g day⁻¹ person⁻¹, R_{FD} = oral reference dose (mg kg⁻¹ day⁻¹), T_A = average life exposure time (365 days year⁻¹ × lifetime, assuming 70 years), and SF = oral cancer slope factor (mg kg⁻¹ day⁻¹)⁻¹.

Concentrations used in the present study of risk calculations are on a wet weight basis. Oral reference dose (R_{FD}) and oral cancer slope factor (SF) values were used from US EPA [45] for risk assessment.

3. Results and discussion

3.1. Concentration of pesticides in water and fish samples

In the present study eight pesticides viz chlordane, dimethoate, malathion, atrazine, heptachlor, dichlorvos, azinphosmethyl and cypermethrin were detected. Table 2 depicts the results of sample analysis as mean value ($\mu\text{g/l}$) concentration of pesticides. The observed concentration of chlordane and heptachlor were 0.104 $\mu\text{g/l}$ and 0.006 $\mu\text{g/l}$ respectively. The concentration of other pesticides was dimethoate 0.082 $\mu\text{g/l}$, cypermethrin 0.076 $\mu\text{g/l}$, azinphosmethyl 0.065 $\mu\text{g/l}$, dichlorvos 0.059 $\mu\text{g/l}$, malathion 0.055 $\mu\text{g/l}$ and atrazine 0.051 $\mu\text{g/l}$. The detected concentration of malathion in the present study was quite low from as reported earlier by (Sankaramakrishnan et al. [16] and Malik et al. [47]) from Kanpur and Lucknow sites of river Ganga. The newly introduced pesticides azinphosmethyl, dimethoate, atrazine, dichlorvos and cypermethrin were also detected in the present study as they are being continuously and abundantly used along the basin [8,48]. Rehana et al. [49] reported dimethoate in river Ganga from the same site, while Agnihotri et al. [50] reported heptachlor near Farrukhabad. Detection of malathion, heptachlor and chlordane shows their historical use and persistence in the ecosystem [51]. In accordance with the European Economic Commission (EEC Directive 80/778/EEC) [52] for drinking water the total pesticide level should not exceed 0.5 $\mu\text{g/l}$ and the individual pesticide not more than 0.1 $\mu\text{g/l}$. The detected concentrations of the above mentioned pesticides were within the range of EEC limit except for chlordane which exceeds the limit. The detected concentration of the pesticides could be attributed to the agricultural runoff resulting from extensive agricultural activities along the basin.

Pesticide concentration detected in fish samples is given in Table 2. Mean concentrations (wet weight, ww) ranged from 0.167 $\mu\text{g g}^{-1}$ for heptachlor to 0.045 $\mu\text{g g}^{-1}$ for azinphosmethyl in bagrid *R. rita*. Mean concentration in *C. carpio* ranged from 0.182 $\mu\text{g g}^{-1}$ for dichlorvos to 0.017 $\mu\text{g g}^{-1}$ for malathion. The concentration of other pesticides were dichlorvos 0.142 $\mu\text{g/l}$, atrazine 0.131 $\mu\text{g/l}$, chlordane 0.101 $\mu\text{g/l}$, malathion 0.081 $\mu\text{g/l}$, cypermethrin 0.076 $\mu\text{g/l}$ and dimethoate 0.069 $\mu\text{g/l}$

Table 3

Health hazard index for pesticide in fish *Rita rita* and *Cyprinus carpio*.

Name of pesticide	ADI ($\mu\text{g/kg/d}$) (US EPA)	EDI ($\mu\text{g/kg/d}$)	
		<i>Rita rita</i>	<i>Cyprinus carpio</i>
Chlordane	0.016	0.079	0.071
Dimethoate	0.01	0.054	0.062
Malathion	0.007	0.063	0.013
Atrazine	0.005	0.102	0.068
Heptachlor	0.002	0.131	0.111
Dichlorvos	0.002	0.111	0.128
Azinphosmethyl	0.001	0.035	0.108
Cypermethrin	0.02	0.040	0.143

in *R. rita*. In *C. carpio* the concentration of these pesticides were dichlorvos 0.163 $\mu\text{g/l}$, heptachlor 0.142 $\mu\text{g/l}$, azinphosmethyl 0.138 $\mu\text{g/l}$, atrazine 0.087 $\mu\text{g/l}$, chlordane 0.091 $\mu\text{g/l}$ and dimethoate 0.079 $\mu\text{g/l}$. These results are in partial agreement with those obtained by [53], who found atrazine and chlorpyrifos residues in the muscle tissue of Tilapia fish collected from Rosetta Nile branch, Egypt. Dimethoate and malathion in the present study were reported lower than the earlier findings of Akhtar et al. [54]. Similarly, heptachlor and chlordane were reported lower than the value as reported earlier by Samanta [55] in fish tissue from the river Ganga at West Bengal site. Due to continuous exposure, contaminants accumulate and get concentrated in the muscle tissues compared to water. Various parameters influence the bioaccumulation of pesticides in fish, including water solubility, degree of ionization, stability, and size or shape of the chemical, and lipid content of the species [56]. The differences between pesticide concentrations in fishes from this study can be attributed to these factors and to differences in exposure. From the data presented in Table 2, it may be concluded that pesticide concentrations were lower in the omnivorous species (*C. carpio*) than in carnivorous species (*R. rita*).

3.2. Bioaccumulation factors

The bioaccumulation factor is the ratio of given chemical contaminant found inside the tissue of the fish to that found in the surrounding water. Pesticides being lipophilic, the detected concentration inside the tissue is normalized with lipid content. The present study shows, the BAF of malathion was found higher in both the selected fishes *R. rita* and *C. carpio*. Chlordane, dimethoate, atrazine, azinphosmethyl and cypermethrin in *R. rita* showed more accumulation than in *C. carpio*. Higher BAF might be accredited to poor water solubility and relative high log K_{ow} values (Table 2) [57,58]. From the studies conducted, it is known that bioaccumulation of chemical contaminant in the fish is due to the cumulative effect of many physiological and environmental conditions such as fish species, fish age, total lipid content including the environmental concentration of the contaminant [59,60]. BAF is always accessed by the hypothesis that the fish remains in the steady state in the surrounding environment, however it is not possible in the natural conditions. Further river Ganga is subjected to dynamic conditions because of large anthropogenic activities. Therefore, lipid content is not the only factor responsible for varied BAF difference but the total conditions of contexture of water.

3.3. Exposure assessment and risk characterization

Fish consumption is one of the most common sources of pesticide entry into human body [61]. Pesticide residues have a high potential negative effect on consumers [62,63]. Estimated daily intake (EDIs) was calculated to assess potential pesticide exposure to humans using mean values of each in the fish tissue as given in Table 2. Table 3 shows comparison between calculated EDIs and the acceptable daily intake (ADI) value issued by regulatory agency US EPA [45]. For the risk characterization all the chemicals were grouped into adversity groups based on chemical specific adversity and use of Rfd. From the

Table 4
Non-carcinogenic (THQ) and carcinogenic risks (R) of pesticide residues.

Pesticide	Rfd (mg/kg/day) ⁻¹ (US EPA)	SF (mg/kg/day) ⁻¹ (US EPA)	THQ		R	
			<i>Rita rita</i>	<i>Cyprinus carpio</i>	<i>Rita rita</i>	<i>Cyprinus carpio</i>
Chlordane	0.00006	0.35	1.36E06	1.31E04	2.87E05	4.39E05
Dimethoate	0.0002		2.59E01	2.97E01		
Malathion	0.02	0.0038	3.05E03	6.40E04	2.31E07	4.86E08
Atrazine	0.035	0.22	2.81E03	1.87E03	2.17E05	1.44E05
Heptachlor	0.0005	4.5	2.51E01	4.49E06	5.66E04	4.81E04
Dichlorvos	0.0005	0.29	2.13E01	2.45E01	3.10E05	3.56E05
Azinphosmethyl	0.2		3.39E04	1.03E04		
Cypermethrin	0.01		3.91E03	1.37E02		

assumption, the daily exposure to pesticides via consumption of these fish from the selected site of river Ganga, the potential risks in human are certain (Table 4). However, the target health quotient (THQs) values for both the fishes were less than 1.0, which indicate lower risk of eating fish from the study area. The calculated R ($< 1 \times 10^{-4}$) value associated with consumption of pesticide contaminated fish indicates negligible cancer risk US EPA [45]. However, the calculated R values of heptachlor were found to be higher than 1×10^{-4} for both the fishes indicating the heptachlor associated risk of cancer. Moreover, human behaviour including dietary habits varies greatly based on various criteria such as locality and socioeconomic status. Furthermore when considering specific single chemicals or assessment groups, such as pesticides in our case, we cannot determine just a part of the overall risk and cannot provide an integrated assessment of the multiple risks triggered by exposure to different toxic stimuli [64–67]. In this study grouping of the chemicals on adversity groups was based on common adversities instead of chemical structure or common mode of action approach that might lead to the underestimation of the real risk from multiple exposure to chemicals. It is certified by many studies that combined exposure from many sources, even at low levels can lead in time to unexpected toxic mixture effects [68,69]. Previous studies also suggested that various pesticides potentially pose health risk to the populations [70,71,72].

4. Conclusion

The present study provides a broad overview of pesticide status in water and two different edible fishes of the river Ganga. Surface water contains pesticide residue in quite detectable limit with mean concentration of chlordane 0.104 µg/l and heptachlor of 0.006 µg/l. Bagrid catfish *R. rita* tends to accumulate more residues than common carp *C. carpio*. Bioaccumulation factor was calculated for pesticides and found to be higher for malathion than other pesticides in both fishes. Lower than 1.0 target health quotient (THQ) implies lower non-carcinogenic risk via consumption of these fishes. Nevertheless, for heptachlor, exposure to carcinogenic risk are high. In summary, due to enormous use of pesticides in the agricultural field basin along the river, potential health risk associated with fish consumption cannot be ignored. The total residue limit should not exceed more than 0.1 µg/g [US EPA]. Further the fish is an important dietary source of proteins and possesses therapeutic values thus, effective measures need to be taken that help reduce total pesticide consumption along the basin and contamination of the fish.

Author's statement

Data will be available on request to the authors.

Data availability statement

The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- [1] J. Jurewicz, W. Hanke, Prenatal and childhood exposure to pesticides and neurobehavioral development: review of epidemiological studies, *Int. J. Occup. Med. Environ. Health* 21 (2) (2008) 121–132.
- [2] S.M. Engel, J. Wetmur, J. Chen, C. Zhu, D.B. Barr, R.L. Canfield, M.S. Wolff, Prenatal exposure to organophosphates, paraoxonase 1, and cognitive development in childhood, *Environ. Health Perspect.* 119 (8) (2011) 1182–1188.
- [3] M.F. Bouchard, J. Chevri er, K.G. Harley, K. Kogut, M. Vedar, N. Calderon, C. Trujillo, C. Johnson, A. Bradman, D.B. Barr, B. Eskenazi, Prenatal exposure to organophosphate pesticides and IQ in 7-year-old children, *Environ. Health Perspect.* 119 (8) (2011) 1189–1195.
- [4] S.A. Mansour, Pesticide exposure—Egyptian scene, *Toxicology* 198 (1–3) (2004) 91–115.
- [5] Z.U. Shah, S. Parveen, A review on pesticides pollution in aquatic ecosystem and probable adverse effects on fish, *Pollut. Res.* 39 (2) (2020) 309–321.
- [6] V.P. Kalyabina, E.N. Esimbekova, K.V. Kopylova, V.A. Kratasyuk, Pesticides: formulations, distribution pathways and effects on human health—a review, *Toxicol. Rep.* 8 (2021) 1179–1192.
- [7] P. Sharma, P.K. Meher, A. Kumar, Y.P. Gautam, K.P. Mishra, Changes in water quality index of Ganges river at different locations in Allahabad, *Sustain. Water Qual. Ecol.* 3 (2014) 67–76.
- [8] NGRBA, National Ganga River Basin Authority (Ministry of Environment and Forests, Government of India), Volume I, Environmental and Social Management Framework (ESMF), 2011, pp. 7–10. Environmental and Social Analysis January 2011, <http://moef.nic.in/downloads/public-information/Draft%20ESA%20Volume%20I.pdf>.
- [9] K.M. Holvoet, P. Seuntjens, P.A. Vanrolleghem, Monitoring and modeling pesticide fate in surface waters at the catchment scale, *Ecol. Modell.* 209 (1) (2007) 53–64.
- [10] S. Takatori, M. Okihashi, Y. Okamoto, Y. Kitagawa, S. Kakimoto, H. Murata, T. Sumimoto, Y. Tanaka, A rapid and easy multiresidue method for the determination of pesticide residues in vegetables, fruits, and cereals using liquid chromatography/tandem mass spectrometry, *J. AOAC Int.* 91 (4) (2008) 871–883.
- [11] S.N. Khuman, P. Chakraborty, Air-water exchange of pesticidal persistent organic pollutants in the lower stretch of the transboundary river Ganga, India, *Chemosphere* 233 (2019) 966–974.
- [12] L. Singh, S.K. Choudhary, P.K. Singh, Pesticide concentration in water and sediment of River Ganga at selected sites in middle Ganga plain, *Int. J. Environ. Sci.* 3 (1) (2012) 260–274.
- [13] Mutiyar, Mittal, Status of organochlorine pesticides in Ganga river basin: anthropogenic or glacial? *Drink. Water Eng. Sci. Discuss* 5 (2013) 1–30.
- [14] R. Pandey, D. Raghuvanshi, D.N. Shukla, Water quality of river Ganga along ghats in Allahabad city, UP, India, *Adv. Appl. Sci. Res.* 5 (4) (2014) 181–186.
- [15] A. Kumari, R.K. Sinha, K. Gopal, Concentration of organochlorine pesticide residues in Ganga water in Bihar, India, *Environ. Ecol.* 19 (2) (2001) 351–356.
- [16] N. Sankaramakrishnan, A.K. Sharma, R. Sanghi, Organochlorine and organophosphorous pesticide residues in ground water and surface waters of Kanpur, Uttar Pradesh, India, *Environ. Int.* 31 (1) (2005) 113–120.

- [17] S. Afful, A.K. Anim, Y. Serfor-Armah, Spectrum of organochlorine pesticide residues in fish samples from the Densu Basin, *Res. J. Environ. Earth Sci.* 2 (3) (2010) 133–138.
- [18] J.R. Fianko, A. Donko, S.T. Lowor, P.O. Yeboah, E.T. Glover, T. Adom, A. Faanu, Health risk associated with pesticide contamination of fish from the Densu River Basin in Ghana, *J. Environ. Prot.* 2 (2011) 115–123.
- [19] A. Ccancapa, A. Masiá, V. Andreu, Y. Picó, Spatio-temporal patterns of pesticide residues in the Turia and Júcar Rivers (Spain), *Sci. Total Environ.* 540 (2016) 200–210.
- [20] M. Kapsi, C. Tsoutsis, A. Paschalidou, T. Albanis, Environmental monitoring and risk assessment of pesticide residues in surface waters of the Louros River (NW Greece), *Sci. Total Environ.* 650 (2019) 2188–2198.
- [21] R. Zhou, L. Zhu, Q. Kong, Persistent chlorinated pesticides in fish species from Qiantang River in East China, *Chemosphere* 68 (5) (2007) 838–847.
- [22] Y. Guo, X.Z. Meng, H.L. Tang, E.Y. Zeng, Tissue distribution of organochlorine pesticides in fish collected from the Pearl River Delta, China: implications for fishery input source and bioaccumulation, *Environ. Pollut.* 155 (1) (2008) 150–156.
- [23] V. Belenguer, F. Martínez-Capel, A. Masiá, Y. Picó, Patterns of presence and concentration of pesticides in fish and waters of the Júcar River (Eastern Spain), *J. Hazard. Mater.* 265 (2014) 271–279.
- [24] D.Q. Wang, Y.X. Yu, X.Y. Zhang, S.H. Zhang, Y.P. Pang, X.L. Zhang, Z.Q. Yu, M. H. Wu, J.M. Fu, Polycyclic aromatic hydrocarbons and organochlorine pesticides in fish from Taihu Lake: their levels, sources, and biomagnification, *Ecotoxicol. Environ. Saf.* 82 (2012) 63–70.
- [25] U. Arisekar, R.J. Shakila, G. Jeyasekaran, R. Shalini, P. Kumar, A.M. Malani, V. Rani, Accumulation of organochlorine and pyrethroid pesticide residues in fish, water, and sediments in the Thamirabarani river system of southern peninsular India, *Environ. Nanotechnol. Monit. Manag.* 11 (2019) 100194.
- [26] B. Kumar, V.K. Verma, A.K. Naskar, P. Chakraborty, S. Kumar, D. Mukherjee, Human health risk from hexachlorocyclohexane and dichlorodiphenyltrichloroethane pesticides, through consumption of vegetables: estimation of daily intake and hazard quotients, *J. Xenobiotics* 3 (1) (2013) e6.
- [27] M. Davodi, A. Esmaili-Sari, N. Bahramifarr, Concentration of polychlorinated biphenyls and organochlorine pesticides in some edible fish species from the Shadegan Marshes (Iran), *Ecotoxicol. Environ. Saf.* 74 (3) (2011) 294–300.
- [28] F. Jabeen, A.S. Chaudhry, Chemical compositions and fatty acid profiles of three freshwater fish species, *Food Chem.* 125 (3) (2011) 991–996.
- [29] Z.R. Nengroo, A. Rauf, Fatty acid composition and antioxidant activities of five medicinal plants from Kashmir, *Ind. Crops Prod.* 140 (2019) 111596.
- [30] S. Afful, A.K. Anim, Y. Serfor-Armah, Spectrum of organochlorine pesticide residues in fish samples from the Densu Basin, *Res. J. Environ. Earth Sci.* 2 (3) (2010) 133–138.
- [31] J.R. Fianko, A. Donko, S.T. Lowor, P.O. Yeboah, E.T. Glover, T. Adom, A. Faanu, Health risk associated with pesticide contamination of fish from the Densu River Basin in Ghana, *J. Environ. Prot.* 2 (2011) 115–123.
- [32] A. Ccancapa, A. Masiá, V. Andreu, Y. Picó, Spatio-temporal patterns of pesticide residues in the Turia and Júcar Rivers (Spain), *Sci. Total Environ.* 540 (2016) 200–210.
- [33] M. Kapsi, C. Tsoutsis, A. Paschalidou, T. Albanis, Environmental monitoring and risk assessment of pesticide residues in surface waters of the Louros River (NW Greece), *Sci. Total Environ.* 650 (2019) 2188–2198.
- [34] Y. Fang, Z. Nie, Y. Yang, Q. Die, F. Liu, J. He, Q. Huang, Human health risk assessment of pesticide residues in market-sold vegetables and fish in a northern metropolis of China, *Environ. Sci. Pollut. Res. - Int.* 22 (8) (2015) 6135–6143.
- [35] I.A. Granados-Galván, D.G. Rodríguez-Meza, A. Luna-González, H.A. González-Ocampo, Human health risk assessment of pesticide residues in snappers (Lutjanus) fish from the Navachiste Lagoon complex, Mexico, *Mar. Pollut. Bull.* 97 (1–2) (2015) 178–187.
- [36] T. Robinson, U. Ali, A. Mahmood, M.J.I. Chaudhry, J. Li, G. Zhang, K.C. Jones, R. N. Malik, Concentrations and patterns of organochlorines (OCs) in various fish species from the Indus River, Pakistan: a human health risk assessment, *Sci. Total Environ.* 541 (2016) 1232–1242.
- [37] S.N. Sruthi, M.S. Shyleshchandra, S.P. Mathew, E.V. Ramasamy, Contamination from organochlorine pesticides (OCPs) in agricultural soils of Kuttanad agroecosystem in India and related potential health risk, *Environ. Sci. Pollut. Res. - Int.* 24 (1) (2017) 969–978.
- [38] M.W. Aktar, M. Paramasivam, D. Sengupta, S. Purkait, M. Ganguly, S. Banerjee, Impact assessment of pesticide residues in fish of Ganga river around Kolkata in West Bengal, *Environ. Monit. Assess.* 157 (1) (2009) 97–104.
- [39] U. Arisekar, R.J. Shakila, R. Shalini, G. Jeyasekaran, Pesticides contamination in the Thamirabarani, a perennial river in peninsular India: the first report on ecotoxicological and human health risk assessment, *Chemosphere* 267 (2021) 129251.
- [40] A.G. Jhingran, The Fish Genetic Resources of India. Bureau of Fish Genetic Resources, Allahabad and Maya Press Pvt. Ltd., Allahabad, India, 1984, p. 83.
- [41] D. Muir, E. Sverko, Analytical methods for PCBs and organochlorine pesticides in environmental monitoring and surveillance: a critical appraisal, *Anal. Bioanal. Chem.* 386 (4) (2006) 769–789.
- [42] A. Therdeppitak, K. Yammeng, Determination of organochlorine pesticides in commercial fish by gas chromatography with electron capture detector and confirmation by gas chromatography mass spectrometry, *Health* (1) (2003) 2.
- [43] F. Smedes, Determination of total lipid using non-chlorinated solvents, *Analyst* 124 (11) (1999) 1711–1718.
- [44] FAO/WHO, Evaluation of Certain Food Additives and Contaminants (Technical Report No. 751). World Health Organisation-Food Agriculture Organisation, Cambridge University Press, 1987.
- [45] US EPA, Risk-based Concentration Table, United States Environmental Protection Agency, Washington, Philadelphia PA, 2009.
- [46] Q.T. Jiang, T.K.M. Lee, K. Chen, H.L. Wong, J.S. Zheng, J.P. Giesy, K.K.W. Lo, N. Yamashita, P.K.S. Lam, Human health risk assessment of organochlorines associated with fish consumption in a coastal city in China, *Environ. Pollut.* 136 (1) (2005) 155–165.
- [47] A. Malik, P. Ojha, K.P. Singh, Levels and distribution of persistent organochlorine pesticide residues in water and sediments of Gomti River (India)—a tributary of the Ganges River, *Environ. Monit. Assess.* 148 (1–4) (2009) 421–435.
- [48] INDIASTAT. www.Indiastat.com (2018).
- [49] Z. Rehana, A. Malik, M. Ahmad, Genotoxicity of the Ganges water at Narora (UP), India, *Mutat. Res. Toxicol.* 367 (4) (1996) 187–193.
- [50] N.P. Agnihotri, V.T. Gajbhiye, M. Kumar, S.P. Mohapatra, Organochlorine insecticide residues in Ganga river water near Farrukhabad, India, *Environ. Monit. Assess.* 30 (2) (1994) 105–112.
- [51] A. Agarwal, R. Prajapati, O.P. Singh, S.K. Raza, L.K. Thakur, Pesticide residue in water—a challenging task in India, *Environ. Monit. Assess.* 187 (2) (2015) 1–21.
- [52] EEC, Council directive 80/778/EEC, Off. J. Eur. Commun. L.229 (Aug. 30th) (1980) 11.
- [53] F. Eissa, K. Ghanem, M. Al-Sisi, Occurrence and human health risks of pesticides and antibiotics in Nile tilapia along the Rosetta Nile branch, Egypt, *Toxicol. Rep.* 7 (2020) 1640–1646.
- [54] M.W. Aktar, M. Paramasivam, D. Sengupta, S. Purkait, M. Ganguly, S. Banerjee, Impact assessment of pesticide residues in fish of Ganga river around Kolkata in West Bengal, *Environ. Monit. Assess.* 157 (1–4) (2009) 97–104.
- [55] S. Samanta, Organochlorine pesticide residues in fish of River Ganga in West Bengal, *Pesticide Res. J.* 18 (1) (2006) 104–108.
- [56] H.O. Esser, P. Moser, An appraisal of problems related to the measurement and evaluation of bioaccumulation, *Ecotoxicol. Environ. Saf.* (6) (1982) 1131–48.
- [57] V. Andreu, Y. Picó, Determination of currently used pesticides in biota, *Anal. Bioanal. Chem.* 404 (9) (2012) 2659–2681.
- [58] F. Ernst, B. Alonso, M. Colazzo, L. Pareja, V. Cesio, A. Pereira, A. Márquez, E. Errico, A.M. Segura, H. Heinzen, A. Pérez-Parada, Occurrence of pesticide residues in fish from south American rainfed agroecosystems, *Sci. Total Environ.* 631 (2018) 169–179.
- [59] W.X. Wang, P.S. Rainbow, Comparative approaches to understand metal bioaccumulation in aquatic animals, *Comp. Biochem. Physiol. Part C: Toxicol. Pharmacol.* 148 (4) (2008) 315–323.
- [60] F.A. Gobas, J.B. Wilcockson, R.W. Russell, G.D. Haffner, Mechanism of biomagnification in fish under laboratory and field conditions, *Environ. Sci. Technol.* 33 (1) (1999) 133–141.
- [61] J. Hoekstra, A. Hart, H. Owen, M. Zeilmaker, B. Bokkers, B. Thorgilsson, H. Gunnlaugsdottir, Fish, contaminants and human health: quantifying and weighing benefits and risks, *Food Chem. Toxicol.* 54 (2013) 18–29.
- [62] S.S. Liu, C.L. Wang, J. Zhang, X.W. Zhu, W.Y. Li, Combined toxicity of pesticide mixtures on green algae and photobacteria, *Ecotoxicol. Environ. Saf.* 95 (2013) 98–103.
- [63] A.F. El Nahas, M.A. Abdel-Razek, N.M. Helmy, S. Mahmoud, H.A. Ghazy, Impaired antioxidant gene expression by pesticide residues and its relation with other cellular biomarkers in Nile Tilapia (*Oreochromis niloticus*) from Lake Burullus, *Ecotoxicol. Environ. Saf.* 137 (2017) 202–209.
- [64] A.M. Tsatsakis, D. Kouretas, M.N. Tzatzarakis, P.K. Stivaktakis, K.S. Tsarouhas, Golokhvast simulating real-life exposures to uncover possible risks to human health: a proposed consensus for a novel methodological approach, *Hum. Exp. Toxicol.* 36 (2017) 554–564.
- [65] A. Tsatsakis, M. Goumenou, J. Liesivuori, W. Dekant, A.F. Hernández, Toxicology for Real-life Risk Simulation - Editorial Preface to This Special Issue (2018), *Toxicology Letters*, 2018.
- [66] Anca Oana Docea, Eliza Gofita, Marina Goumenou, Daniela Calina, Otilia Rogoveanu, Marius Varut, Six months exposure to a real life mixture of 13 chemicals' below individual NOAELs induced non monotonic sex-dependent biochemical and redox status changes in rats, *Food Chem. Toxicol.* 115 (2018) 470–481.
- [67] N. Ronald Kostoff, Marina Goumenou, Aristidis Tsatsakis, The Role of Toxic Stimuli Combinations in Determining Safe Exposure Limits, 2018. *Toxicology Reports*.
- [68] E.A. Renieri, M. Goumenou, D.A. Kardonsky, V.V. Veselov, A.K. Alegakis, A. Buha, M.N. Tzatzarakis, A.E. Nosyrev, V.N. Rakitskii, M. Kentouri, A. Tsatsakis, Indicator PCBs in farmed and wild fish in Greece-Risk assessment for the Greek population, *Food Chem. Toxicol.* 127 (2019) 260–269.
- [69] M. Tzatzarakis, M. Kokkinakis, E. Renieri, M. Goumenou, M. Kavvalakis, E. Vakonaki, A. Chatzinikolaou, P. Stivaktakis, I. Tsakiris, A. Rizos, A. Tsatsakis, Multiresidue analysis of insecticides and fungicides in apples from the Greek market. Applying an alternative approach for risk assessment, *Food Chem. Toxicol.* 140 (2020) 111262.

- [70] Y. Wang, L. Lv, Y. Yu, G. Yang, Z. Xu, Q. Wang, L. Cai, Single and joint toxic effects of five selected pesticides on the early life stages of zebrafish (*Deniorerio*), *Chemosphere* 170 (2017) 61–67.
- [71] Y.B. Yohannes, Y. Ikenaka, A. Saengtienchai, K.P. Watanabe, S.M. Nakayama, M. Ishizuka, Concentrations and human health risk assessment of organochlorine pesticides in edible fish species from a Rift Valley lake—lake Ziway, Ethiopia, *Ecotoxicol. Environ. Saf.* 106 (2014) 95–101.
- [72] Z.U. SHAH, S. Parveen, Pesticides pollution and risk assessment of river Ganga: A review, *Heliyon* 7 (8) (2021) p.e07726, <https://doi.org/10.1016/j.heliyon.2021.e07726> (2121), In this issue.