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Exploration and determination of algal role as Bioindicator to evaluate water quality – Probing fresh water algae

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ABSTRACT

Objectives: To explore the algal floral diversity and its role to determine water quality.**Methods:** The regular monthly collection of algal and water samples was made during 2018. Unicellular algae were preserved in 2 to 3% formalin while macroalgae in 4% formalin. Microphotographs of algae were taken at the biotechnological Lab of PCSIR Lahore, Pakistan. Palmer pollution index was used to determine water quality.**Results:** The study identified 201 algal species distributed among 57 genera, 42 families, 25 orders, 10 classes and 7 divisions. The total score of Algal Genus Pollution Index of Banjosa Lake, Ali Sojal Dam, Dothan Dam, Drake Dam and Rawalakot Nullah (city) were 14, 9, 10, 18 and 25 respectively. It was revealed that Banjosa Lake has probable organic pollution, Ali Sojal Dam and Dothan Dam showed lack of organic pollution, Drake Dam indicated moderate pollution while Rawalakot Nullah (City) indicated confirm high organic pollution.**Conclusion:** We strongly recommend the conservation and managed status of algal species for sustainable resource of algal- derived products in future. It was revealed that the water quality of Banjosa Lake, Drak Dam and Rawalakot Nullah was affected from anthropogenic activities and needs to be managed.© 2021 Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

Algae are diverse and large group of simple, photosynthetic, unicellular and multicellular organisms. They have very simple body called thallus which cannot be differentiated into true roots,

stems and leaves and lack vascular tissues. They are very diverse organisms in size, form, structure, color, habit and habitat. Algae occupy variety of habitats due to their vast ecological amplitude and distributed across the oceans, rivers, lakes, ponds and streams. They are present from the depth of ocean to the highest peaks of the world. Linnaeus for the first time coined the term 'Algae (Linnaeus, 1753). Fig. 1.

Algae are major source of food for aquatic organism and play an important role in aquatic food chain or food web (Galloway et al., 2012). They are good source of natural human diet (Gupta and Pandey, 2007). Microalgae are good source of biofuel and higher productivities than traditional crops (Khattoon and Pal, 2015). Many crucial antibiotics and medicines are obtained from these

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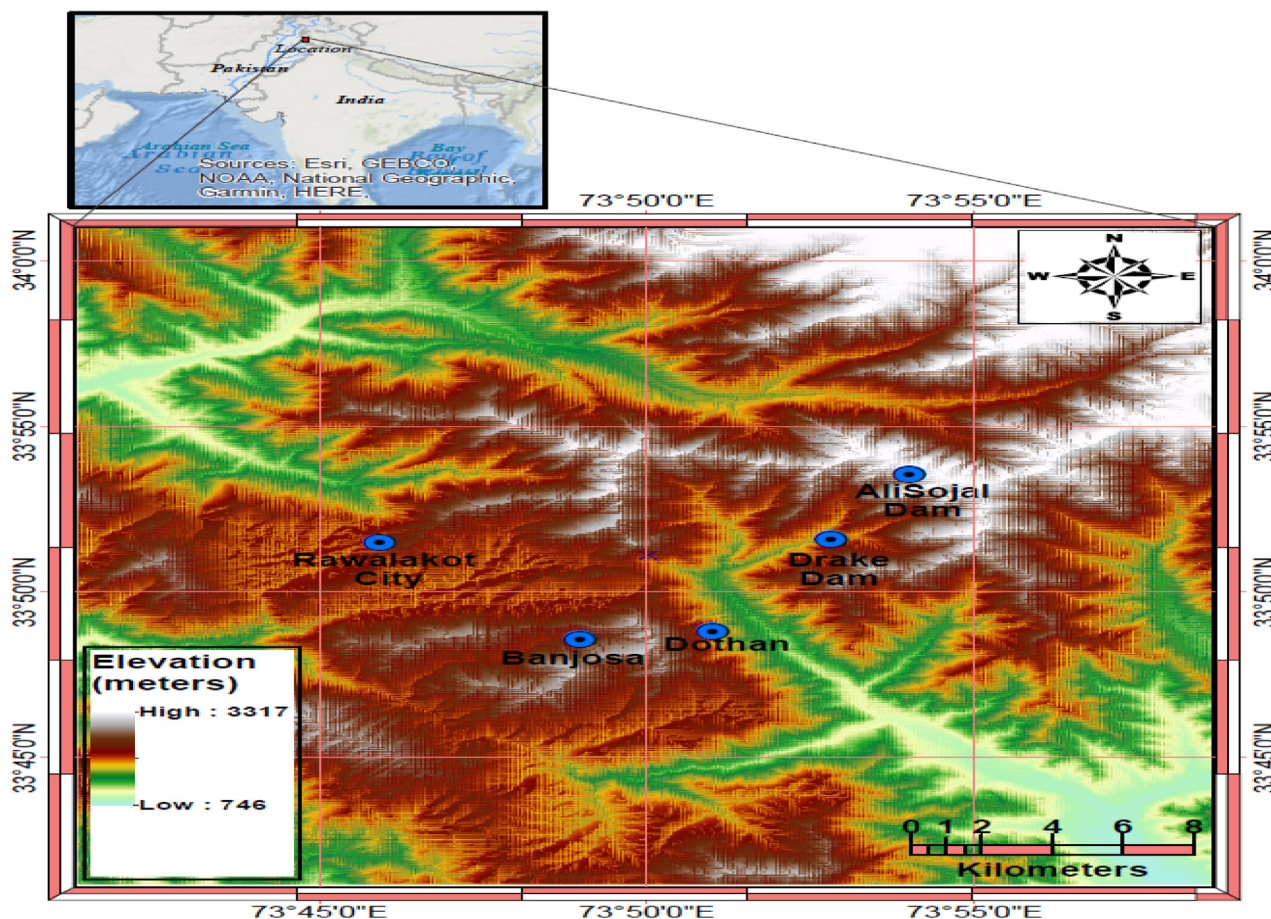


Fig. 1. Map of the study area.

organisms. They are used by pharmaceutical companies to prepare drugs which are used to treat various diseases like cancer, Acquired immune deficiency syndrome (AIDS), Arthritis, respiratory diseases and infections due to viruses, bacteria and fungi (V, Vadlapudi, 2012). Algae are involved in water purification from nutrients and pollutants (Biggs and Kilroy 2000). Algae are considered as very good bioindicator of water quality due to their rapid response to pollutants. Algae are important biological organism for purification of waters bodies because they absorb organic and inorganic pollutants, heavy metals and radioactive substances (Alp et al., 2012). Microalgae are bioindicators of eutrophication and effectively used to assess the quality of water (Kumar and Amit, 2012). The dominance of green algae and diatoms indicate the oligotrophic conditions while abundance of blue green algae indicate the eutrophic conditions of water bodies (Musharaf et al., 2011).

(Abdul Aziz et al., 2003) undertook algal study on the Arabian Gulf of the coastal waters of Saudi Arabia. They reported 35 genera of unicellular algae belonging to Bacillariophyta, Dinophyta and Cyanophyta and found that algae showed luxurious growth in summer due to nutrients status and temperature conditions. Similar algal studies were conducted by (Khomayis and Al-Harbi, 2003) on marine fouling area of Sharm Obhour, Saudi Arabia and recorded 22 species of Bacillariophyta and 2 species of Cyanophyta and Dynophyta each. An important study regarding algae of different groups at Batkhela, Malakand, Pakistan was carried out and total of 63 algal species belonging to Cyanophyta, Chlorophyta and Bacillariophyta were recorded (Barkatullah et al., 2013). An important study to explore fresh water algal flora of Jauharabad district, Pakistan was conducted and species were taxonomically described (Zarina et al., 2013). Some studies on algal flora were

carried out in Azad Jammu & Kashmir (Khuhawari et al., 2009, Naz et al., 2009, Haq et al., 2012, and Ali et al., 2006).

Due to immense importance and contribution of algae and attempt was made to document the presence of valuable algal species. The objectives of this particular study were to conduct an extensive research for collection, identification and classification of algal flora to determine Phycological diversity and its role to determine water quality from the study area for the first time. This research work is significant because this is an up to date novel work. The paper should be of interest to readers in the areas of Botany, Ecology, Conservation biology, Agriculture and Environmental sciences etc.

2. Materials and methods

2.1. Study site

Rawalakot is headquarter of district Poonch Azad Jammu & Kashmir, Pakistan. It located in temperate region at an elevation of 5374 feet and the Latitude 33°51'32.18"N, Longitude 73°45'34.93"E. The climate of Rawala Kot can be divided into four seasons. Rawala Kot has mild and warm temperature during the spring and summer, while snow falls in the winter. Maximum and minimum temperatures can be 38 C and –1 C in summer and winter respectively.

2.2. Sampling sites

For present study, algal and water samples were collected from five randomly selected sampling Sites; Banjosa Lake, Ali Sojal Dam,

Dothan Dam, Drak Dam and Rawalakot City (Nallah) were selected as sampling stations.

2.3. Sampling of algae

The regular monthly collection of algae was made from January 2018 to December 2018. Epiphytic algae and desmid flora were collected with pipette. Filamentous algal species were collected with the help of forceps. Tooth brush was used to collect diatoms while macro-algae were picked up with hands. The collected algal samples were transferred into bottles and were labelled.

2.4. Preservation of algae

Unicellular algae were preserved in 2 to 3% formalin while macro algae were kept in 4% formalin (Mason, 1967).

2.5. Laboratory studies and identification of algae

The algal samples were taken to molecular & biotechnological lab, food and biotechnological research Centre PCSIR, Lahore, Pakistan for microphotography. Microphotographs of algal species were taken with the camera attached with the microscope (MT5300H-Japan). The specimens of algal species were identified with the help of authentic literature upto species level or even up to variety level (Hustedt 1930, Majeed 1935, Smith 1950, Presscot, 1962, Tilden, 1910).

2.6. Analysis of algae

The collected material was assessed on the basis of morphological, cytological and reproductive characters during microscopic examination. After detail analysis, algae were documented and tabulated for evaluation of algal biodiversity from the study area.

2.7. Palmer pollution index

Palmer pollution index was used to evaluate water quality.

3. Results

The present study identified 201 algal species consisting of 57 genera, 42 families, 25 orders, 10 classes and 7 divisions from the study area during 2018. The maximum number of families (18) were contributed by division Bacillariophyta, followed by Chlorophyta (14) families and Cyanophyta (5) families. Euglenophyta contributed 2 while Glucophyta and Dinophyta contributed 1 family each. (Table.1). The Bacillariophyta showed dominancy with (47%) frequency followed by Chlorophyta and Cyanophyta 38% and 19% frequency respectively. The minor divisions Euglenophyta, Charophyta and Dinophyta showed poor representation with (2%), (1.5%) and (1%) frequency respectively (Fig. 2).

The highest number of species were represented by Naviculaceae and Fragiellariaceae while minimum by Glucocystaceae (Table 1). Maximum number of species identified were 95 which were contributed by Bacillariophyta followed by Chlorophyta 76 species. Minimum number of species were represented by Glaucophyta and Dinophyta with one species each. Maximum number of genera (25) were contributed by Bacillariophyta followed by Chlorophyta and Cyanophyta by 21 and 6 genera respectively. Euglenophyta was represented by 3 genera while rest of the divisions were recorded with 1 genus each. The division with maximum number of orders was Chlorophyta which contributed 11 orders followed by Bacillariophyta 6 and Cyanophyta 4 orders while remaining divisions gave up 1 order each (Fig. 3). The largest

genus was Navicula which was represented by 14 species followed by spirogyra and Cymbella with 11 species each. Minimum number of species were contributed by Troschia and Lyngbya (Fig. 4 and Table 1). The division Bacillariophyta showed dominancy with (47%) frequency followed by Chlorophyta and Cyanophyta 38% and 19% respectively. The minor divisions Euglenophyta, Charophyta and Dinophyta showed poor representation with (2%), (1.5%) and (1%) frequency respectively. Fig. 5.

3.1. Evaluation of organic pollution by Palmer's algal genus

Pollution of surface water has become major environmental problem. Organic pollution in water bodies leads to eutrophication which in turn accelerate the growth of certain type of algal species in water body. Algae are taken as natural indicator to environmental conditions because they form blooms in water bodies and show vigorous growth. There are many studies by various authors who showed strong relationship among algal species and polluted and unpolluted water. They are influenced by factors like mixing of water masses, light, temperature, salinity and nutrients. Bioindicator organisms can be used to identify the effects of pollutants in aquatic ecosystems.

Different environmental conditions affect the occurrence and distribution of algae. Cyanobacteria mass is influenced by the hydrogen ion concentration (pH) followed by temperature, light, soil type and available nutrients. Their mass can be recorded in the form of the frequency as well as the intensity of biotic community due to eutrophication. Various studies on algal species confirmed their role for the assessment of pollution in water bodies. (Kolkwitz and Marsson, 1950) defined five zones on the degree of pollution in water bodies and proposed the use of aquatic organisms as bioindicators to evaluate the water quality.

(Werner, 1977) proposed nine different zones on the basis of organic pollution in water bodies. Werner proposed zones were Coprozoic, Polysaprobic, Mesosaprobic, Oligosaprobic, and Katharobic. Each zones was found to be different on the basis of physico-chemico and biological characteristics. He listed the indicator species of these zones which showed changes in their growth pattern in response to pollutants in water bodies. Polysaprobic zone was characterized by the complete absence of algae except for blue green algae *Spirulina* and green algae *Euglena viridis*. Blue green algae showed dominancy in alpha-Mesozoic while diatoms and green algae were dominant in Beta-Mesozoic zones. Members of Dinophyta and Charophyta were found only in oligosaprobic zones.

Palmer (1969) proposed pollution index on the basis of algal genus and species present in water bodies. This pollution index which is effectively used to analyze the water quality for high or low organic pollution (Table 2). The pollution tolerant genera of algae were recorded from all the sites of the study area and a pollution index score was assigned to each algal genus. Pollution index score of 20 or more confirms high organic pollution within the water body. A score of 15–19 is an evidence of probable organic pollution. Pollution index scores from 10 to 14 indicates moderate organic pollution, while index score 0–10 indicates lack of organic pollution in waters.

During our present study algal genera were used to determine the water quality of freshwater bodies in the study area in which the total score of algal genus pollution index of Banjosa Lake, Ali Sojal Dam, Dothan Dam, Drake Dam and Rawalakot City (Nullah) were found to be 14, 9, 10, 18 and 25 respectively. It was concluded that Banjosa Lake has probable organic pollution while Ali Sojal Dam and Dothan Dam, showed lack of organic pollution, Drake Dam indicated probable organic pollution and Rawalakot Nullah indicated confirm high organic pollution (Table 3).

Table 1
Algal flora explored from freshwater bodies of the study area.

Divisions	Class	Order, Family, Genus and Species name
Cyanophyta	Cyanophyceae	Order: Chroococcales
		Family: Chroococcaceae
		1. <i>Chroococcus turgidus</i> (Kutzing) Naegeli
		2. <i>Chroococcus limneticus</i> Lemmermann
		Family: Microcystaceae
		3. <i>Gleocapsa punctata</i> Nageli
		4. <i>Gleocapsa bituminosa</i> Kutzing
		Order: Nostocales
		Family: Nostocaceae
		5. <i>Nostoc muscorum</i> C.A. Agardh
		Order: Oscillatoriales
		Family: Oscillatoriaceae
		6. <i>Lyngbya birgei</i> G.M. Smith
		7. <i>Lyngbya martensiana</i> Meneghini ex Gomont
		8. <i>Lyngbya majuscula</i> Harvey ex Gomont
		9. <i>Oscillatoria tenuis</i> C.A. Agardh
		10. <i>Oscillatoria limosa</i> C.A. Agardh
		11. <i>Oscillatoria princeps</i> Vaucher
		12. <i>Oscillatoria sancta</i> (Kutzing) Gomont
13. <i>Oscillatoria fracta</i> G.W.F. Carlson		
14. <i>Oscillatoria acuta</i> Bruhl		
15. <i>Oscillatoria chilensis</i> Biswas		
16. <i>Oscillatoria obscura</i> Bruhl & Biswas		
17. <i>Oscillatoria curviceps</i> C.A. Agardh		
18. <i>Oscillatoria anguina</i> Bory ex Gomont		
Order: Synechococcales		
Family: Merismopediaceae		
19. <i>Merismopedia glauca</i> (Ehrenberg) Kutzing		
Chlorophyta	Chlorophyceae	Order: Chlorococcales
		Family: Oocystaceae
		20. <i>Ankistrodesmus falcatus</i> var. <i>radiates</i> (Chod) Lemmermann
		Order: Chlamydomonadales
		Family: Volvocaceae
		21. <i>Volvox aureus</i> Ehrenberg
		22. <i>Pandorina morum</i> Bory
		Family: Chlorococcaceae
		23. <i>Schroederia setigera</i> (Schroeder) Lemmermann
		Order: Sphaeropleales
		Family: Scenedesmaceae
		24. <i>Scenedesmus quadricauda</i> (Turpin) Brebisson
		25. <i>Scenedesmus opoliensis</i> Richter
		26. <i>Scenedesmus protuberans</i> Fritsch and Rich
		27. <i>Scenedesmus longus</i> Meyen
		28. <i>Scenedesmus dimorphus</i> (Turpin) Kutzing
		29. <i>Scenedesmus carinatus</i> (Lemmermann) Chodat
		30. <i>Scenedesmus communis</i> E. Hegewald
		31. <i>Scenedesmus abundans</i> var. <i>longicada</i> G.M Smith
		32. <i>Scenedesmus quadricauda</i> var. <i>maxima</i> West & GS West
		33. <i>Scenedesmus abundans</i> (Kirchner) Chodat
		34. <i>Tetraedron regulare</i> Kutzing
		35. <i>Tetraedron caudatum</i> (Corda) Hansgirg
		36. <i>Tetraedron trigonum</i> var. <i>minus</i> Reinch
		37. <i>Troschia aspera</i> Reinsch
		38. <i>Coelastrum microporum</i> Nacgeli
		39. <i>Coelosphaerium kuetzinginum</i>
		Family: Hydrodictyaceae
		40. <i>Pediastrum duplex</i> Meyen
		41. <i>Pediastrum duplex</i> var. <i>gracillimum</i> West & GS West
		42. <i>Pediastrum simplex</i> var. <i>echnulatum</i> Wittrock
		43. <i>Pediastrum biwae</i> var. <i>ovatum</i> (Ehrenberg) Tiffany
		44. <i>Pediastrum simplex</i> var. <i>duodenarium</i> (Bailey) Rabenhorst
		45. <i>Pediastrum boryanum</i> var. <i>longicorne</i> Raciborski
		Order: Oedogoniales
		Family: Oedogoniaceae
		46. <i>Oedogonium macrandrium</i> Wittrock
		47. <i>Oedogonium cardiacum</i> (Hassall) Wittrock
		48. <i>Oedogonium majus</i> (Hansgirg) Tiffany
Order: Tetrasporales		
Family: Sphaerocystidaceae		
49. <i>Sphaerocystis schroeteri</i> R. Chodat		
Order: Microsporaes		

(continued on next page)

Table 1 (continued)

Divisions	Class	Order, Family, Genus and Species name
		Family: Microsporaceae 50. <i>Microspora quadrata</i> Hazen 51. <i>Microspora willeana</i> Lagerheim 52. <i>Microspora tumidula</i> Hazen
		Order: Chetophorales Family: Chaetophoraceae 53. <i>Draparnaldia plumose</i> (Vaucher) C.A. Agardh 54. <i>Chaetophora lobata</i> F.Schrank 55. <i>Chaetophora elegans</i> (Roth) Agardh
	Ulvophyceae	Order: Ulvotrichales Family: Ulotrichaceae 56. <i>Ulothrix gemilata</i> Kützing 57. <i>Ulothrix zonata</i> (Weber & Mohr) Kützing 58. <i>Ulothrix aqualis</i> Kützing Order: Cladophorales Family: Cladophoraceae 59. <i>Cladophora glomerata</i> (Linnaeus) Kützing 60. <i>Cladophora oligoclona</i> Kützing
	Zygnematophyceae	Order: Zygnematales Family: Zygnemataceae 61. <i>Spirogyra communis</i> (Hassall) Kützing 62. <i>Spirogyra bififormis</i> C.C.Jao 63. <i>Spirogyra maxima</i> Link in C.G.Nees 64. <i>Spirogyra neglecta</i> (Hassall) Kützing 65. <i>Spirogyra subsalsa</i> Link in C.G.Nees 66. <i>Spirogyra tetrapla</i> Transeau 67. <i>Spirogyra elongata</i> (Vaucher) Kützing 68. <i>Spirogyra fluviatilis</i> Hilse 69. <i>Spirogyra punctiformis</i> Trasca 70. <i>Spirogyra rectangularis</i> Transeau 71. <i>Spirogyra catenaeformis</i> (Hassall) Kützing 72. <i>Zygnema tenue</i> Kützing 73. <i>Zygnema sterile</i> Transeau 74. <i>Zygnema insigni</i> (Hassall) 75. <i>Zygnema aplanosporum</i> Stancheva, J.D.Hall & Sheath 76. <i>Mougetia micropora</i> Taft 77. <i>Mougetia viridis</i> (Kützing) Wittrock Order: Desmidiaceae Family: Desmidiaceae 78. <i>Cosmarium formosulum</i> Hoffman 79. <i>Cosmarium botrytis</i> Meneghini 80. <i>Cosmarium speciosum</i> Lundell 81. <i>Cosmarium granatum</i> Brebisson 82. <i>Cosmarium nitidulum</i> Denotaris 83. <i>Cosmarium subtumidum</i> Nordstedt 84. <i>Staurastrum rzoskae</i> Meyen ex Ralfs Family: Closteriaceae 85. <i>Closterium acutum</i> (Lyngbye) Brebisson 86. <i>Closterium parvulum</i> Naegeli 87. <i>Closterium leibleinii</i> Kützing 88. <i>Closterium lanceolatum</i> Kützing 89. <i>Closterium intermedium</i> Ralfs 90. <i>Closterium littorale</i> Gay 91. <i>Closterium lunula</i> (Mueller) Nitzsch 92. <i>Closterium pseudolunula</i> Borge 93. <i>Closterium acerosum</i> var. <i>elongatum</i> Brebisson 94. <i>Closterium striolatum</i> Ehrenberg
	Trebouxiophyceae	Order: Chlorellales Family: Chlorallaceae 95. <i>Chlorella conductrix</i> Brandt
Charophyta	Charophyceae	Order: Charales Family: Characeae 96. <i>Chara coralina</i> Klein ex C.L.Willdenow 97. <i>Chara globular</i> Thuiller 98. <i>Chara vulgaris</i> Linnaeus

Table 1 (continued)

Divisions	Class	Order, Family, Genus and Species name
Bacillariophyta	Bacillariophyceae	<p>Order: Bacillariales</p> <p>Family: Anomoeneidaceae</p> <p>99. <i>Anomoeneis vitrea</i> Pfitzer</p> <p>100. <i>Anomocoenis exilis</i> (Kutzing) Cleve</p> <p>101. <i>Anomocoenis serianus</i> (Brebisson) Cleve</p> <p>Family: Catenulaceae</p> <p>102. <i>Amphora holsatica</i> Hustedt</p> <p>Family: Achnanthidiaceae</p> <p>103. <i>Achanthes microcephala</i> (Kutzing) Cleve</p> <p>Family: Cocconeidaceae</p> <p>104. <i>Cocconeis planctula</i> Ehrenberg</p> <p>Family: Bacillariaceae</p> <p>105. <i>Denticula tennis</i> Kutzing</p> <p>106. <i>Nitzschia palea</i> (Kutzing) Wm. Smith</p> <p>107. <i>Nitzschia sublinearis</i> Hustedt</p> <p>108. <i>Nitzschia acicularis</i> (Kutzing) Wm. Smith</p> <p>109. <i>Nitzschia hungarica</i> Grunow</p> <p>110. <i>Nitzschia denticula</i> Grunow</p> <p>111. <i>Nitzschia palea</i> var. <i>tenuirostris</i> Grunow</p> <p>Order: Fragilariales</p> <p>Family: Fragilariaceae</p> <p>112. <i>Diatom vulgare</i> Bory</p> <p>113. <i>Diatoma anceps</i> (Ehrenberg) Kirchner</p> <p>114. <i>Fragillaria virescens</i> Ralfs</p> <p>115. <i>Fragillaria pinnota</i> Ehrenberg</p> <p>116. <i>Fragillaria capucina</i> Desmazieres</p> <p>117. <i>Fragillaria vaucheriae</i> (Kutzing) J.P. Peterson</p> <p>118. <i>Fragillaria intermedia</i> (Grunow) Grunow</p> <p>119. <i>Fragillaria crotonensis</i> var. <i>prolongata</i> Grunow</p> <p>120. <i>Stauroneis phoenicentron</i> (Nitzsch) Ehrenberg</p> <p>121. <i>Stauroneis acuta</i> Wm. Smith</p> <p>122. <i>Stauroneis phoenicentron</i> var. <i>amphilepta</i> (Ehrenberg) Cleve</p> <p>123. <i>Stauroneis anceps</i> var. <i>birostris</i> (Ehrenberg) Cleve</p> <p>124. <i>Synedra dorsiventralis</i> Mueller</p> <p>125. <i>Synedra ulna</i> (Nitzsch) Ehrenberg</p> <p>126. <i>Synedra acus</i> Kutzing</p> <p>127. <i>Synedra parasitica</i> (Wm. Smith) Hustedt</p> <p>128. <i>Synedra amphicephala</i> Kutzing</p> <p>129. <i>Synedra affinis</i> Kuetz</p> <p>130. <i>Synedra crystalina</i> (C.A. Agardh) Kutzing</p> <p>Order: Naviculales</p> <p>Family: Naviculaceae</p> <p>131. <i>Caloneis bacillum</i> (Grunow) Mereschkowsky</p> <p>132. <i>Gyrosigma acuminatum</i> (Kutzing) Cleve</p> <p>133. <i>Gyrosigma wormleyi</i> (Sullivant) Boyer</p> <p>134. <i>Gyrosigma Kuetzingii</i> (Grunow) Cleve</p> <p>135. <i>Gyrosigma scalproides</i> Rabenhorst</p> <p>136. <i>Navicula tuscula</i> (Ehrenberg) Grunow</p> <p>137. <i>Navicula gracilis</i> Ehrenberg</p> <p>138. <i>Navicula protracta</i> (Grunow) Cleve</p> <p>139. <i>Navicula exigua</i> (Gregory) Muller</p> <p>140. <i>Navicula veneta</i> Kutzing</p> <p>141. <i>Navicula salinarum</i> Grunow</p> <p>142. <i>Navicula rhyncocephala</i> Kutzing</p> <p>143. <i>Navicula radiosa</i> Kutzing</p> <p>144. <i>Navicula viridula</i> Kutzing</p> <p>145. <i>Navicula cuspidata</i> Kutzing</p> <p>146. <i>Navicula grimmei</i> Krasske</p> <p>147. <i>Navicula dicephala</i> (Ehrenberg) Wm. Smith</p> <p>148. <i>Navicula reinhardtii</i> (Grunow) Van Heurek</p> <p>149. <i>Navicula gregaria</i> Donkin</p> <p>Family: Rhopalodiaceae</p> <p>150. <i>Rhopalodia gibba</i> (Kutzing) Mueller</p> <p>151. <i>Rhopalodia gibba</i> var. <i>ventricosa</i> (Kutzing) H. Peragallo & M. Peragallo</p> <p>152. <i>Epithemia adnata</i> (Kutzing) Brebisson</p> <p>153. <i>Epithemia argus</i> (Ehrenberg) Kutzing</p> <p>Family: Pinnulariaceae</p> <p>154. <i>Pinnularia nobilis</i> Ehrenberg</p> <p>155. <i>Pinnularia braunii</i> (Grunow) Cleve</p> <p>156. <i>Pleurosigma angulatum</i> (Quekett) Wm. Smith</p> <p>157. <i>Pleurosigma salinarum</i> (Grunow) Grunow</p> <p>Family: Neidiaceae</p> <p>158. <i>Nedium iridis</i> (Ehrenberg) Pfitzer</p>

(continued on next page)

Table 1 (continued)

Divisions	Class	Order, Family, Genus and Species name
		159. <i>Nedium dubium</i> (Ehrenberg) Pfitzer
		Family: Sellaphoraceae
		160. <i>Sellaphora capitata</i> D.G.Mann & S.M. McDonald
		161. <i>Sellaphora pupilla</i> (Kutzing) Mereschkowsky
		Family: Amphipleuraceae
		161. <i>Frustulia rhomboids</i> (Ehrenberg) DeToni
		163. <i>Frustulia viridula</i> (Brebisson) DeToni
		Family: Stauroneidaceae
		164. <i>Craticula cuspidata</i> (Kutzing) D.G.Mann
		Order: Surirellales
		Family: Surirellaceae
		166. <i>Cymatopleura solea</i> (Brebisson) Wm.Smith
		167. <i>Cymatopleura elliptica f. spiralis</i> (Chase) Boyer
		168. <i>Surirella linearis</i> Wm.Smith
		169. <i>Surirella saronica</i> Auerswald
		170. <i>Surirella didyma</i> Kutzing
		171. <i>Surirella splendida</i> (Ehrenberg) Kutzing
		172. <i>Surirella linearis</i> var. <i>constricta</i> (Ehrenberg) Grunow
		173. <i>Surirella ovata</i> Kutzing
		174. <i>Surirella minuta</i> Kutzing
		175. <i>Surirella patella</i> Kutzing
		Order: Mastogloiales
		Family: Mastogloiaceae
		176. <i>Mastogloia smithii</i> Tshwait
		177. <i>Mastogloia smithii</i> var. <i>amphicephala</i> Grunow
		Order: Cymbellales
		Family: Cymbellaceae
		178. <i>Cymbella tumida</i> (Brebisson) Van Heurck
		179. <i>Cymbella leptoceros</i> (Ehrenberg) Grunow
		180. <i>Cymbella laevis</i> Naegeli
		181. <i>Cymbella lanceolata</i> (Brebisson) Van Heurck
		182. <i>Cymbella prostrata</i> (Berkeley) Cleve
		183. <i>Cymbella turgida</i> Gregory
		184. <i>Cymbella acqualis</i> Wm.Smith
		185. <i>Cymbella affinis</i> Kutzing
		186. <i>Cymbella parva</i> (Wm.Smith) Cleve
		187. <i>Cymbella aspera</i> (Ehrenberg) Cleve
		188. <i>Cymbella brehmii</i> C.A. Agardh
		Family: Gomphonemataceae
		189. <i>Gomphonema ventricosum</i> Gregory
		190. <i>Gomphonema constrictum</i> Ehrenberg
		191. <i>Gomphonema micropus</i> Kutzing
		192. <i>Gomphonema sphaerophorum</i> Ehrenberg
		193. <i>Gomphonema angustatum</i> (Kutzing) Grunow
		194. <i>Gomphonema parvulum</i> (Kutzing) Grunow
Euglenophyta	Euglenophyceae	Order: Euglenales
		Family: Euglenaceae
		195. <i>Euglena limnophila</i> Ehrenberg
		196. <i>Euglena sociabilis</i> P.A. Dangeard
		197. <i>Euglenaformis proxima</i> (Dangeard) M.S. Bennett
		Family: Phacaceae
		198. <i>Phacus longicauda</i> (Ehrenberg) Dujardin
		199. <i>Astasia cylindrical</i> Fromental
Glaucophyta	<u>Glaucophyceae</u>	Order: Glaucocystales
		Family: Glaucocystaceae
		200. <i>Gloescystis ampla</i> Kutzing
Dinophyta	Dinophyceae	Order: Phytodiniales
		Family: Phytodinaceae
		201. <i>Cystodinium steinii</i> Transeau

4. Discussion

The algae have been an interesting group of plants due to their primitive nature and worldwide distribution because of their capabilities to exist in variety of environmental conditions. Algae are the primary producers in all kind of aquatic ecosystems due to their photosynthetic ability. They are very much important organism from ecological, commercial and medical aspects. In Pakistan,

few algal studies have been conducted but little attention was given to explore algal flora of Azad Jammu & Kashmir. Our study in fresh water bodies showed rich diversity of algal species in which 3 species; *Phacus longicauda*, *Closterium leibeileinii*, and *Pediastrum duplex* var. *gracillimum* were new records for Azad Jammu & Kashmir. Our results are in agreement with the study on the diversity of algae carried out by Leghari et al., 2002 in which they identified 134 algal species from River Jhelum, Azad Jammu & Kashmir.

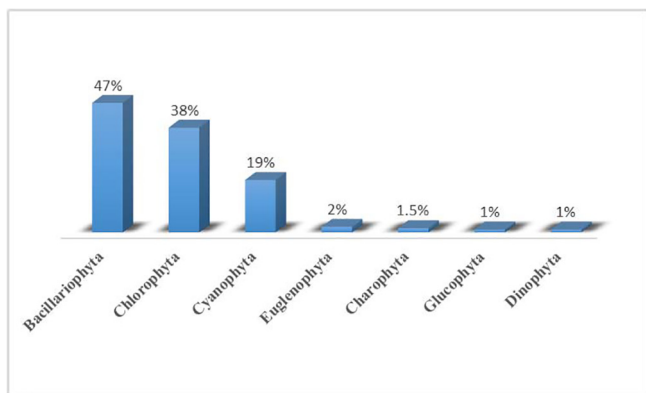


Fig. 2. Species percentage of identified taxa.

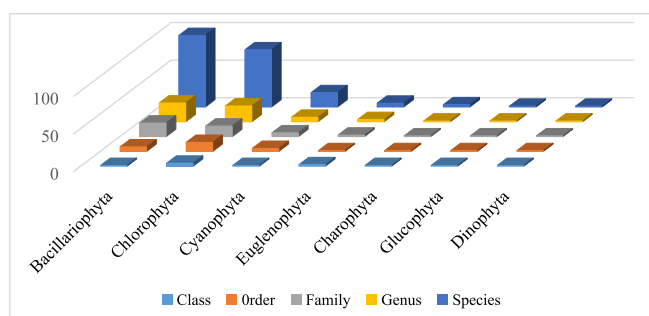


Fig. 3. Number of taxa within documented Divisions.

Similar algal research was undertaken and 195 algal species were documented from Punjab, Khyber Pakhtoonkhwa, Azad Kashmir, and north-eastern areas of Pakistan. The class Nostocophyceae was found to be more abundant which contributed 144 species as compared to the class Chlorophyceae which added 51 algal species. It was observed that these species appeared during spring and summer and disappeared in the autumn (Haq et al., 2012).

Previously some research work has been carried out on the algal diversity in Jauharabad District, Pakistan by Zarina et al., 2013. They identified four species of diatoms belonging to family Bacillariaceae, Gomphonemaceae and Nitzschiaceae. (Jang et al., 2014)

investigated the algal diversity in fresh water bodies from different localities of Swabi District, Pakistan and identified 22 genera with 35 species. Our studies are in accordance with (Aliya et al., 2009) during which they explored the fresh water habitats of Karachi city, Pakistan and reported 6 division, 33 families, 86 genera and 214 algal species. The divisions Cyanophycota and Volvophycota showed dominancy with (74.8%) frequency while the Euglenophycota and Chrysophyta showed poor representation of (2.8%) frequency. Variations in algal composition and diversity was analyzed in Sangju weir, Gyeongsangbuk do-province and it was found that it was varied at bottom, middle and surface layers of water due to formation of thermocline in water body. Micro algae were dominated by diatoms followed by green algae and blue green algae.

Bioindicator organisms indicate the effects of different pollutants in aquatic ecosystems. Phytoplanktons are reliable tool to evaluate the water quality of wetlands (Crosseti et al., 2008). Freshwater bodies and lakes are characterized on the basis of dominant algal group. We characterized water bodies of our study area on the basis of algal genus and concluded that Banjosa Lake has probable organic pollution, Ali Sojal Dam and Dothan Dam showed lack of organic pollution, Drake Dam indicated probable organic pollution while Rawalakot Nullah indicated confirm high organic pollution. Species of Chlorophyta like Chlymdomonas and Euglena, members of Bacillariophyta like Navicula, Synedra, Gomphonema, and blue green algae such as Oscillatoria and Phormidium grow in organic polluted waters (Palmer, 1969). (Anand, 2000) investigated the ecology of a diatom species and noticed their role as an indication of water quality parameters. The dominance of green algae and diatoms indicate the oligotrophic conditions while abundance of blue green algae indicate the eutrophic conditions of water bodies (Musharaf et al., 2011). Our findings agree with palmer (1969) and Musharaf et al. (2011).

5. Conclusion

The water of the fresh water bodies of Rawalakot is used for domestic, industrial and agricultural purposes. It was revealed through the current study that Rawalakot Nullah, Drak Dam and Banjosa Lake are declining due to pollution. Conservation and management of these water bodies is required in order to avail them for long term in a proper way. Sustainable water use and management is recommended for the subsistence of the water bodies in the study area. It is recommended for future studies to carry out

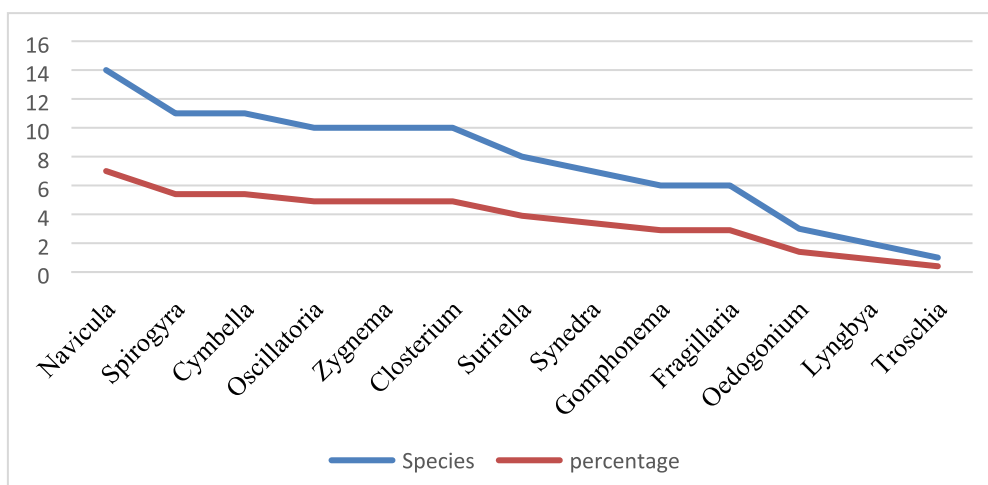


Fig. 4. Genera with their contribution for algal species.

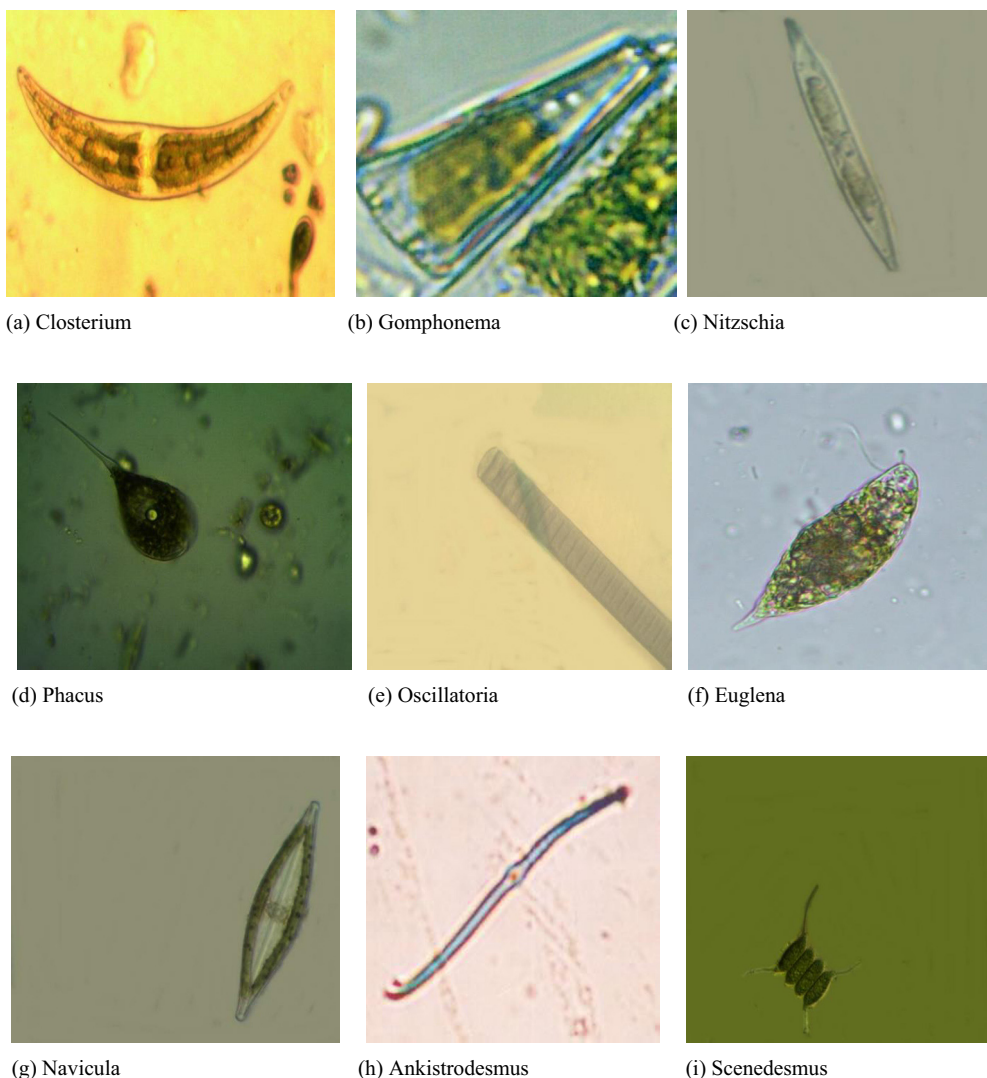


Fig. 5. (a-i) Pollution tolerating genera identified from the freshwater bodies of the study area.

Table 2
Palmer algal genus pollution index.

S. No.	Genus	Index	Genus	Index
1	Ankistrodesmus	4	Nitzschia	3
2	Closterium	1	Scenedesmus	4
3	Euglena	1	Syndra	2
4	Gomphonema	1	Pandorina	1
5	Chlorella	1	Oscillatoria	5
6	Navicula	3	Phacus	2
7	Cyclotella	5	Phormidium	1
8	Chlamydomonas	3	Stigeoclonium	2
9	Anacystis	2	Micractinium	1

molecular studies of algae along with phylogeny of the representative genera. The algal use for bio-fuel, bioremediation, medicinal, human algal diet and fish flora should be studied for commercial and industrial applications and its advantages to the mankind.

Table 3
Pollution indicating algal genera from water bodies of the study area.

Algal Genus	Banjosa Lake	Alisojal Dam	Dothan Dam	Drak Dam	Rawalakot City (Nullah)
Ankistrodesmus	4	–	–	4	4
Closterium	1	–	1	1	1
Euglena	1	–	–	1	1
Gomphonema	1	1	1	1	1
Chlorella	–	1	1	–	–
Navicula	3	–	–	3	3
Cyclotella	–	–	–	–	–
Chlamydomonas	–	–	–	–	–
Nitzschia	3	3	–	3	3
Scenedesmus	–	–	4	–	4
Syndra	2	2	2	–	2
Pandorina	–	–	1	–	1
Oscillatoria	–	–	–	5	5
Phacus	2	2	–	–	–
Phormidium	–	–	–	–	–
Stigeoclonium	–	–	–	–	–
Total	14	9	10	18	25

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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