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Arbuscular mycorrhizal fungi enhance soil carbon sequestration in the coalfields, northwest China

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Carbon storage is affected by photosynthesis (P_n) and soil respiration (R_s), which have been studied extensively in natural and agricultural systems. However, the effects of P_n and R_s on carbon storages in the presence of arbuscular mycorrhizal fungi (AMF) in coalfields remain unclear. A field experiment was established in 2014 in Shendong coal mining subsidence area. The treatments comprised two inoculation levels (inoculated with or without 100 g AMF inoculums per seedlings) and four plant species [wild cherry (*Prunus discadenia* Koebne L.), cerasus humilis (*Prunus dictyneura* Diels L.), shiny leaf Yellow horn (*Xanthoceras sorbifolium* Bunge L.) and apricot (*Armeniaca sibirica* L.)]. AMF increased P_n of four species ranging from 15.3% to 33.1% and carbon storage, averaged by 17.2% compared to controls. Soil organic carbon (OC), easily extractable glomalin-relation soil protein (EE-GRSP), and total glomalin-relation soil protein (T-GRSP) were significantly increased by AMF treatment. The effect of AMF on the sensitivity of R_s depended on soil temperature. The results highlighted the exponential models to explain the responses of R_s to soil temperature, and for the first time quantified AMF caused carbon sequestration and R_s . Thus, to our knowledge, AMF is beneficial to ecosystems through facilitating carbon conservation in coalfield soils.

Carbon storage depends on the balance between carbon sequestration by plant P_n and carbon release to atmosphere through R_s ^{1,2}. Therefore, if plant P_n or R_s is altered, carbon balance will be affected. R_s , especially seasonal variations, are significantly affected by soil temperature and moisture^{3,4}. However, the impact of soil microbes on R_s or carbon storage remains unclear. So far, quantified contribution of soil microbes, especially AMF has not been reported yet. AMF are obligate plant symbionts that associates with the roots of more than 80% vascular land plants, which significantly enhance long-term success of mine site reclamation⁵. Studies showed that AMF are helpful for building up a productive, healthy, and sustainable post-mine land ecosystem with vegetation cover. The positive impact of AMF on reclaimed soil fertility and plant communities succession has been well documented^{6–8}. However, few studies focused on the effects of AMF on R_s in post-mine land. A recent study conducted in Germany grassland communities reported that AMF stimulated R_s on pasture soil, leading to elevated CO_2 level and temperature, with most carbon sequestered in belowground parts, making R_s an important component of carbon balance^{9,10}. Factors such as soil moisture, soil temperature, CO_2 enrichment, and precipitation changes were all affected R_s . However, relevant studies remain scarce and the complexity of various interactions that affect R_s (such as soil temperature vs moisture, soil temperature vs microbes, moisture vs microbes interactions) are still poorly understood.

R_s includes intergated CO_2 flux of root respiration, mycorrhizal respiration (considered as part of autotrophic respiration), and heterotrophic respiration, which are controlled by carbon supply and temperature^{9,10}. Carbon supply is an important R_s impact factor, which primarily depends on plant productivity and generally responds positively to CO_2 enrichment with the increased P_n ¹¹. Numerous studies confirmed that carbon storage depended on plant P_n and R_s via AMF in greenhouse or field experiments^{7,9–11}.

AMF usually form symbiotic associations with trees in diverse forests. The fungi rely on host plant to obtain carbon supply and utilize 5–20% of the net photosynthate of the symbiotic system¹¹. Global forest soil releases 24 Pg Carbon per year into atmosphere via CO_2 efflux and generates CO_2 from a widely variety of belowground organisms, with AMF as the dominant carbon source¹². AMF as an important carbon and soil CO_2 efflux source

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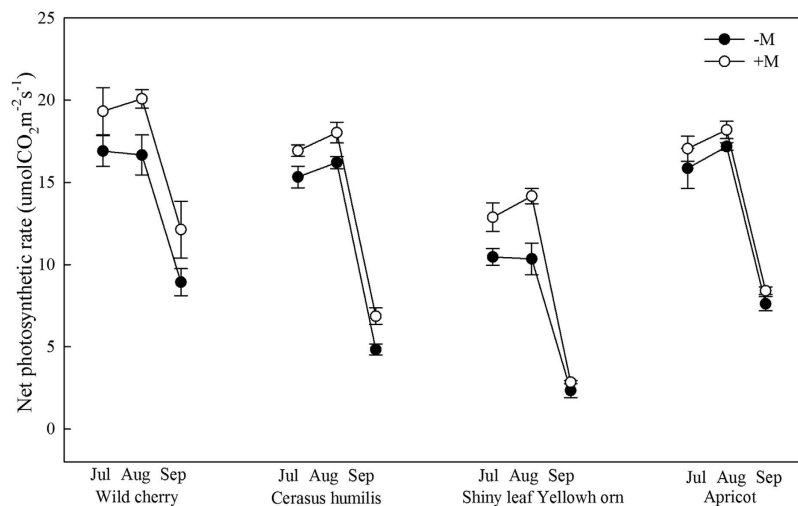


Figure 1. Net photosynthetic rate of four species in different growth stage. Average monthly variation in P_n ($n = 18$) in wild cherry, cerasus humilis, shiny leaf Yellowh orn, and apricot from July to September. Values represented means \pm SE. -M and +M denoted without and with AMF inoculation.

have been well studied under various temperature and moisture conditions¹². A pulse-labeling experiment showed that in temperate prairie, about 4–6% photoassimilates were facilitated via AMF¹⁰. Simultaneously, AMF enhanced 16% soil CO₂ efflux when *Lolium perenne* roots were colonized and 8% carbon supply was derived from AMF in a barley field¹³. To date, no studies have been carried out on the contribution of AMF on carbon storage in coal mining soil.

Respiratory quotient (Q_{10}) is defined as the temperature sensitivity of R_s which is derived from substrate availability, which is the respiration variation ratio when soil temperature rises 10 °C. Q_{10} is largely affected by a series of environmental factors, such as soil physicochemical properties and soil moisture, etc. However, few studies have quantified the effects of AMF on apparent Q_{10} from different tree species. Thus, knowledge about the function of AMF on R_s , P_n , and carbon storage is insufficient¹⁴.

Notably, CO₂ concentration indirectly influences R_s and carbon allocation from host species to fungus, suggesting that root participates in carbon fixation^{15–17}. Most colonized plants enhanced productivity or nutritional status of the plant symbioses system when exposed to higher CO₂ concentration¹⁸. In addition, mycorrhizal hyphae assisted carbon re-distribution from aboveground parts to roots via utilizing the photosynthate and provides multiple adherent agents such as extracellular polymer, and amino acid, etc than non-colonized plants¹⁹. Briefly, AMF enhance soil carbon proportion in higher CO₂ environment^{20–22}. However, lacking a better understanding of these responses in coal mining soil limited to prediction soil carbon storage when AMF existed in current scenarios.

Thus, the experiment planted wild cherry, cerasus humilis, shiny leaf Yellowh orn and apricot in coal mining soil and half seedlings were inoculated with AMF inoculums. Based on the experimental results, two hypotheses was proposed that: (i) plant species preferentially facilitate their photosynthetic ability and increase R_s rates, leading to different carbon storage amount; and (ii) soil temperature rather than soil moisture is sensitive to R_s due to the extreme drought. The future goal is to investigate the effect of AMF on carbon sequestration in coal mining fields.

Results

P_n and R_s temporal dynamics. P_n was significantly greater with AMF inoculation in all the four species from July to September (Fig. 1). Specifically, AMF treatment increased P_n in wild cherry, cerasus humilis, shiny leaf Yellowh orn, and apricot by 21.2%, 15.0%, 53.1%, and 7.4%, respectively. Meanwhile, P_n in all four tree species increased from July to August and then strikingly decreased from August to September (Fig. 1).

In all tree species, R_s was significantly decreased from July to September, with slight decrease from July to August and reduced sudden reduction of 21.3–26.9% in September (Fig. 2).

Cumulative carbon measures. Annual cumulative carbon in AMF treated wild cherry, cerasus humilis, shiny leaf Yellowh orn, and apricot were about 1706, 1784, 2049 and 2065 g C.m⁻².yr⁻¹ (Fig. 3a–d). Compared to the corresponding controls, the annual cumulative carbon levels in the four tree species were increased by 28.3%, 44.6%, 31.8% and 33.5%, respectively (Fig. 3a–d). In growing season, AMF increased the carbon storage by 29.4%, 45.7%, 32.4% and 34.4% ($P < 0.0001$) (Fig. 3a–d). AMF also significantly increased the annual cumulative R_s carbon in the four tree species by 32.4%, 36.6%, 25.2% and 29.9%, when compared to the corresponding controls ($P < 0.0001$) (Fig. 4a–d).

Soil temperature sensitivity to R_s . R_s was calculated using exponential function (Eq. 1), temperature dynamic showed over 70% R_s variation in AMF treated groups and the correlation of R_s showed only 40% variation in control groups (Table 1). Apparent Q_{10} and the coefficient b were decreased significantly in control

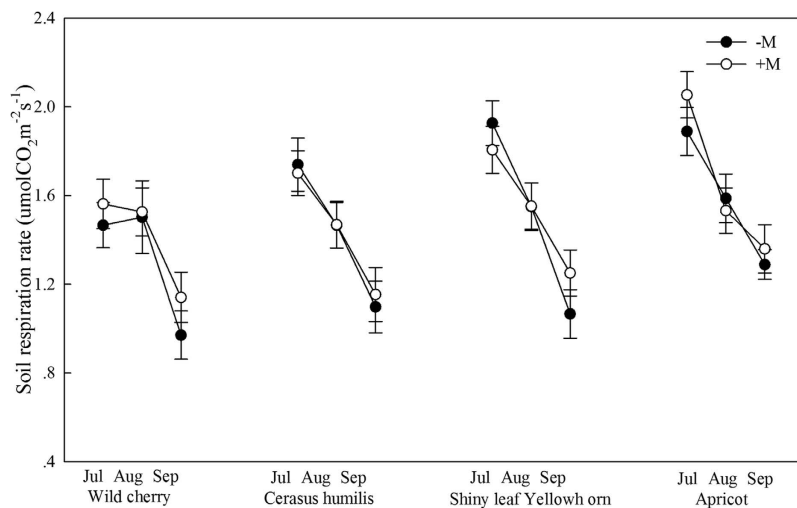


Figure 2. Soil respiration rate of four species in different growth stage. Average monthly variation in R_s ($n = 18$) in wild cherry, cerasus humilis, shiny leaf Yellowhorn, and apricot from July to September. Values represented means \pm SE. -M and +M denoted without and with AMF inoculation.

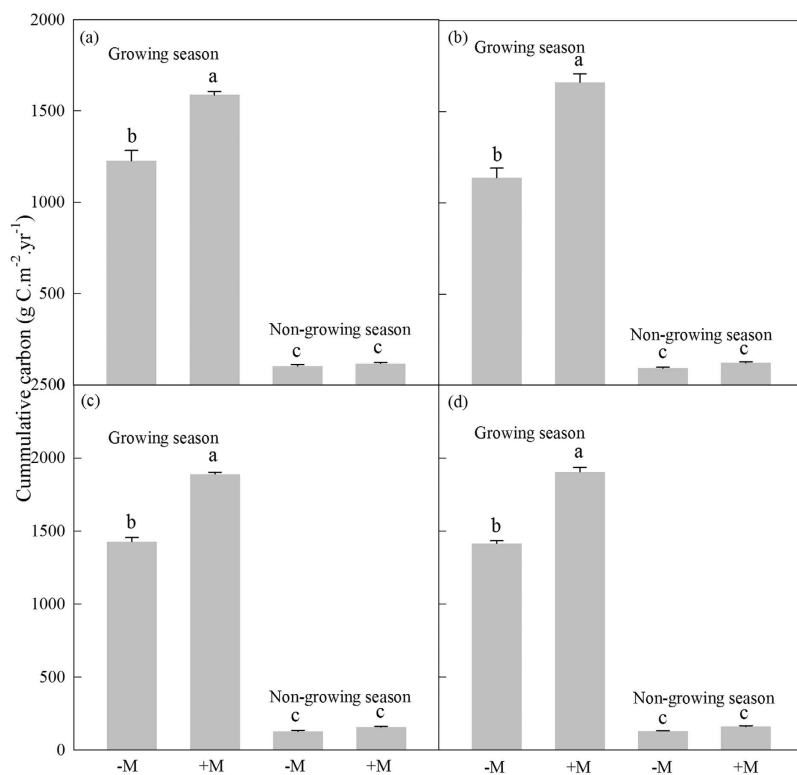


Figure 3. Carbon storage in growing and non-growing season. (a–d) Indicated wild cherry, cerasus humilis, shiny leaf Yellowhorn, and apricot, respectively. Bar represented means \pm SE. -M and +M denoted without and with AMF inoculation. Different lowercase letters indicated that -M and +M were significantly different at 5% level by LSD.

apricot and wild cherry ($P < 0.05$) (Table 1; Fig. 5a,d). However, AMF decreased previous two parameters in cerasus humilis and shiny leaf Yellowhorn (Fig. 5b,c). Apparent Q_{10} responded differently to temperature (the precipitation in 2014 was higher than 10 year's average precipitation) in August from that in July or September (Fig. 6a–d).

Effects of AMF inoculation on carbon-linked parameters. In control of wild cherry, cerasus humilis, shiny leaf yellowhorn, and apricot, the production of OC, EE-GRSP and T-GRSP almost did not change with

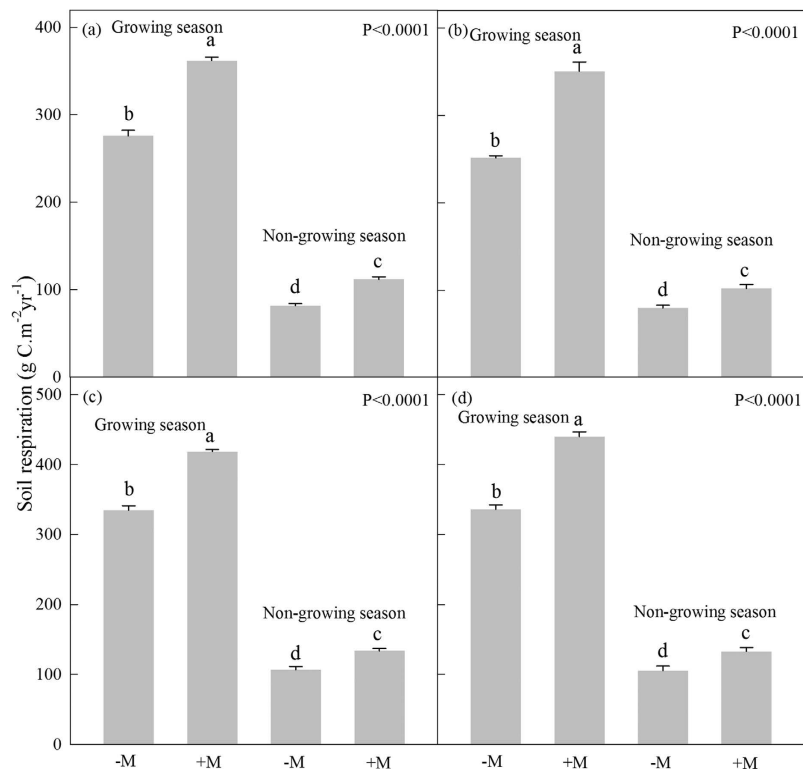


Figure 4. Soil carbon releases via R_s in growing and non-growing seasons. (a–d) Showed that wild cherry, cerasus humilis, shiny leaf Yellowh orn, and apricot, respectively. Bar represented means \pm SE. –M and +M denoted without and with AMF inoculation, respectively. Different lowercase letters indicated that –M and +M were significantly different at 5% level by LSD.

Species	–M			+M		
	SR	r^2	Q_{10}	SR	r^2	Q_{10}
Wild cherry	$1.18e^{0.05T}$	0.42	2.13	$1.09e^{0.09T}$	0.73	2.67
Cerasus humilis	$1.21e^{0.04T}$	0.49	2.01	$1.28e^{0.05T}$	0.69	2.51
Shiny leaf Yellowh orn	$1.27e^{0.05T}$	0.39	1.87	$1.31e^{0.08T}$	0.75	2.58
Apricot	$1.35e^{0.07T}$	0.44	1.63	$1.42e^{0.07T}$	0.71	2.61

Table 1. The exponential model of Q_{10} value. Exponential relationship between the R_s and soil temperature (T °C; July to September) and apparent Q_{10} values.

time (Table 2). In contrast, the production of soil OC, EE-GRSP and T-GRSP in AMF treated of four species increased significantly over time (Table 2). After three months of treatment, the increment of OC, EE-GRSP and T-GRSP in AMF treated groups were all significant (27–37%, 34–45% and 19–22%, respectively (Table 2). After five months of AMF treatments, compared to the control, the levels of OC, EE-GRSP and T-GRSP in AMF treated groups were remarkable enhanced by 52–61%, 55–70% and 36–44%, respectively (Table 2). Collectively, AMF had a positively impacted on soil carbon-linked parameters over time in all of the four tree species (Table 2).

Discussion

The present study firstly examined photosynthetic ability, R_s and carbon storage in respond to AMF inoculation in the coalfields. Under AMF treatment, carbon storage was derived from belowground parts of the tree species. Meanwhile, the temporal variability strongly responded to R_s and annual cumulative carbon with exponential models.

P_n and R_s responses to AMF inoculation. AMF inoculation incurred a series changes in P_n , R_s and carbon storage via close symbiosis with host plants, leading to enhanced carbon production and improved nutrition status against the detrimental effects^{14,23–27}. Meanwhile, AMF accelerated the organic matter (OM) decomposition rates²⁸ and provided some raw materials, which elevated CO_2 concentration in micro circumstance^{29,30} and facilitated soil carbon turnover in the agro-or grassland via CO_2 stimulation effects³¹. In the moist forest, abundant and diverse AMF (e.g. tropical forests) species created a transient carbon sink via N transferring to reduce carbon lost. This implied that AMF promotes carbon production and storage via increasing P_n , which ameliorated the

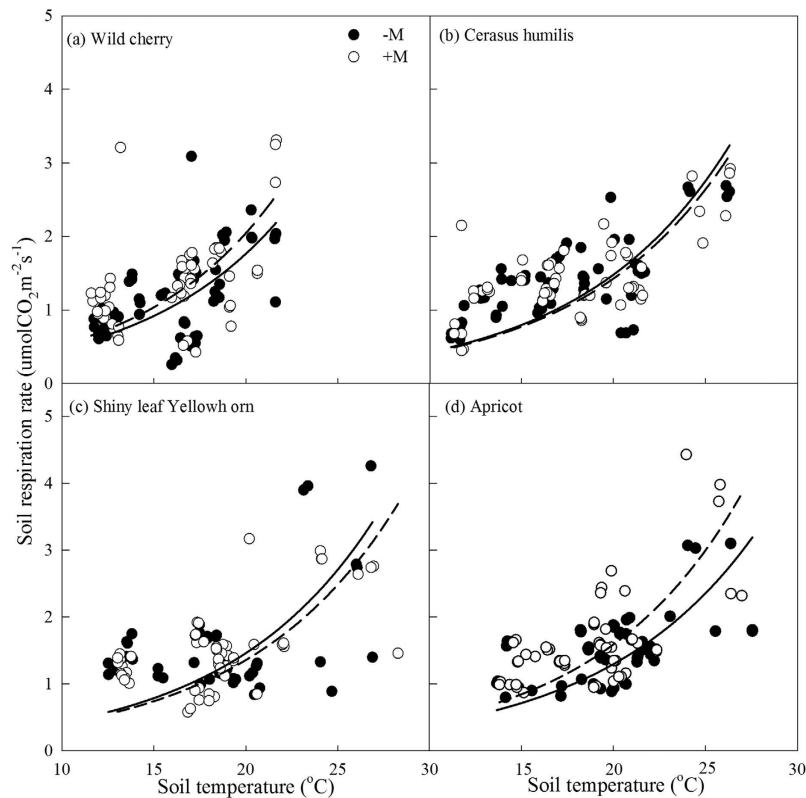


Figure 5. Exponential models of R_s based on soil temperature. Soil temperature was plotted against R_s for plant species with or without AMF inoculation ($n = 36$). (a–d) Indicated that wild cherry, cerasus humilis, shiny leaf Yellowhorn, and apricot, respectively. Solid and hollow circles represented data from without and with AMF inoculation. Solid and dash lines represented the fitted curve of R_s to soil temperature without and with AMF inoculation, respectively.

negative effect caused by unfavorable environment³². Recently, strong evidences showed that the colonized plants have higher P_n and ribulose-1,5-bisphosphate carboxylase/oxygenase activity than non-colonized plants, which compensated greater photosynthate^{33,34}. Carbon was produced directly from P_n , and AMF perhaps supported the higher metabolism of plants and greater productivity of carbon in the harsh environment^{35–39}. Furthermore, soil temperature decreasing inhibited substrate movement, lowering microbial activity, which decreased plant P_n and belowground carbon allocation. Temperature also reduced P_n and the belowground coupling, which potentially affected the substrate availability and carbon cycling⁴⁰. The results demonstrated that AMF can increase P_n of the plant, thus increasing carbon allocation.

Carbon partitioning responses to AMF inoculation. Due to its ability to enhance carbon allocation, AMF has been used in agricultural field and grasslands, but information about carbon storage is omitted in mining soil. Carbon accumulation tended to be higher with AMF inoculation in this study in the growing season. AMF sequestered greater carbon via enhanced P_n and photosynthate production, which was transferred from aboveground to roots or microbes. AMF counteracted carbon loss of respiration, due to increase productivity and nutrient acquisition, especially carbon sequestration^{35,41}. In the non-growing seasons, AMF preferentially allocated photosynthate from tree branches to belowground parts, increasing the EE-GRSP and T-GRSP of carbon stocks^{14,38}. Estimate, AMF utilized a large proportion (about 20%) of photosynthates^{19,42–45}, leading to more carbon retention via symbiosis^{46–50}. Consequently, AMF inoculation resulted in remarkable greater carbon sequestration.

Nonlinear response to R_s . AMF affected carbon storage and partition, which has been discussed in the previous study⁵¹. However, no data have shown whether interactions between species and temperature affect AMF facilitated carbon sequestration⁵². As literatures indicated, carbon productivity of the plants was greater with AMF, due to higher availability of nutrients from the soil environments⁵³. The beneficial effect of AMF in carbon sequestration was considered a result of improved availability of nutrients and altered carbon allocation caused by AMF^{54–55}. The exponential models successfully illustrated the changes of R_s in response to soil temperature variation and captured monthly dynamics of all tree species used in the experiment.

Hysteresis between R_s and soil temperature exists in various ecosystems and vegetations^{56–57}. In general, the decoupling of R_s in response to soil temperature is attributed to confound effect, such as precipitation or

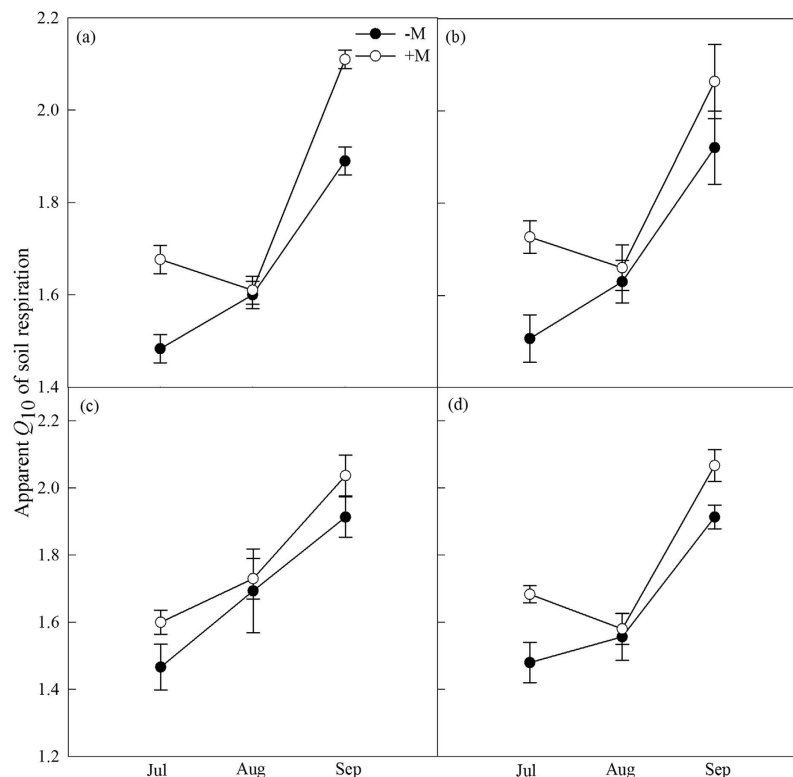


Figure 6. Apparent Q_{10} values represents the sensitivity of soil respiration and soil temperature when rises 10°C . Monthly apparent Q_{10} values variation of four tree species without or with AMF inoculation from July to September. Values represented means \pm SE. (a–d) Indicated wild cherry, cerasus humilis, shiny leaf Yellowh orn, and apricot of tree species, respectively.

Indicator	Duration (Month)	Wild cherry		Cerasus humilis		Shiny leaf Yellowh orn		Apricot	
		–M	+M	–M	+M	–M	+M	–M	+M
OC (g kg^{-1})	0	2.36aA	2.35cA	2.42aA	2.39cA	2.53aA	2.48cA	2.44aA	2.46cA
	3	2.42aB	3.18bA	2.51aB	3.27bA	2.65aB	3.35bA	2.51aB	3.13bA
	5	2.55aB	3.67aA	2.63aB	3.84aA	2.67aB	3.76aA	2.56aB	3.83aA
EE-GRSP (mg kg^{-1})	0	313aA	319cA	298aA	305cA	313aA	321cA	318aA	325cA
	3	329aB	428bA	332aB	443bA	321aB	439bA	339aB	453bA
	5	333aB	496aA	339aB	518aA	327aB	508aA	341aB	512aA
T-GRSP (mg kg^{-1})	0	897aA	903cA	916aA	904cA	889aA	901cA	898aA	883cA
	3	915aB	1087bA	923aB	1078bA	908aB	1093bA	923aB	1077bA
	5	936aB	1231aA	945aB	1249aA	926aB	1287aA	930aB	1272aA

Table 2. Different soil carbon-linked fractions variations. Note: values are means of three replicates. Values followed by the small lowercase letters are not significantly different among different inoculation durations (month) of one tree species with or without AMF (+M or –M) at the 5% level by LSD (vertical comparisons); values followed by the same capital letters are not significantly different between –M and +M at the same durations within the same tree species at the 5% level by LSD (horizontal comparisons) in the identical durations (months). –M, +M represented without and with added mycorrhizal fungi inoculums at 100 g per tree species. Effect of added AMF inoculums on soil OC, EE-GRSP, T-GRSP in four tree species with three inoculation durations.

physiological drought caused hysteresis, which leads to decreased AMF activity and subsequently lower CO_2 production⁵⁷. Over all, the R_s -temperature relationship was affected by P_n , litterfall, and soil microbe activities^{58–61}. However, the detailed mechanisms remain unclear. In this study, due to the existence of hysteresis, R_s in control groups was higher than AMF inoculation groups under a given temperature, which was in agreement with the previous hypothesis⁶². Most likely, AMF plants accumulated more fresh litter in a given temperature than non-colonized plants. Q_{10} was often used to represent the temperature measurement depth in greenhouse or field experimental models^{14,63}. In this work, apparent Q_{10} increased in AMF treated groups, indicating that AMF

increases the sensitivity of R_s to soil temperature. Apparent Q_{10} value was lower in summer than in cooler season, which might be because of higher substrate availability and severe soil moisture limitation in summer. In consistent with the previous publication¹⁴, the experiment data showed that R_s strongly depends on soil temperature in the study.

Previous studies have used exponential model but ignored the function of AMF⁶⁴. The results suggested that AMF plays a key role in R_s and carbon stocks. AMF and soil temperature in combination modulate apparent Q_{10} in coalfields.

Conclusions

This study provided new insights into the impacts of AMF on R_s and carbon storage in coalfields. AMF enhances carbon sequestration and the sensitivity of R_s to temperature. Higher AMF infection rate significantly enhances R_s and P_n , the positive AMF function is through promoting plant growth, especially increasing leaf area, chlorophyll content, and the Q_{10} value. These results clearly demonstrated that AMF infection increased organic matter and glomalin which may be associated with the enhancement of carbon storage in soil. The model of R_s to soil temperature should be reassessed to take into account of the interaction between the soil volumetric moisture and mycorrhizal fungus. Thus, further study will focus on long-term AMF effect on carbon stock in a tree species specific manner.

Materials and Methods

Study site and experimental set-up. This experiment was conducted at the ecological reclaimed region, which located in Daliuta town (39°18'N, 110°4'E), Shenmu County, Yulin City, Shaanxi Province, Northwest China at 1,200 m height above sea level. This site was a typical junction area of Shanxi, Shaanxi and Inner Mongolia of three Provinces and the south margin of Mu us desert in the loess plateau transition zone. About 70% precipitation falls from June to September, the 10-year (2005–2014) average total precipitation and the potential evaporation were about 150 mm and 2000 mm according to Shenmu Meteorological Station near the experiment site. The experimental location had a typical characteristic of continental climate, with annual average temperature about 8°C. The cumulative temperature above 0°C and 10°C were 3,550°C and 3,210°C, respectively; the frost-free period is 150 days and total solar radiation is 6,000 MJ m⁻² year⁻¹. The local soil was classified as Aeolian sandy (FAO/UNESCO, 1988) which contained 75% sand, 22% silt, and 3% clay. The topsoil physico-chemical properties were: soil OM 4.5 g kg⁻¹, total N 0.21 g kg⁻¹, Olsen P 5.3 mg kg⁻¹, exchangeable K 37.8 mg kg⁻¹ and pH value (1:2.5 soil: distilled water) 7.9.

All experiments were carried out in three replicates. The sub-plots of four tree species, wild cherry (*Prunus discadenia* Koebne L.), cerasus humilis (*Prunus dictyneura* Diels L.), shiny leaf Yellow horn (*Xanthoceras sorbifolium* Bunge L.), and apricot (*Armeniaca sibirica* L.) were used in the experiments. Each species was divided into two groups: AMF group and control group. AMF group received the inoculation of AMF at 100 g inoculums/seedling, while the control group did not contain any AMF inoculums.

The four seedlings were obtained from the forest bureau of Shenmu County, Yulin City, Shaanxi Province and Northwest China. Each seedling was transplanted into the plot at temperatures ranging from about 10–15°C in March 25, 2014 to April 5, 2014. The AMF colonization rate of wild cherry, cerasus humilis, shiny leaf Yellow horn, and apricot were 4.8%, 6.3%, 5.4%, and 7.2% before transplanting and 78.1%, 80.2%, 76.4%, and 81.7% at the end of monitoring stage. Two months after transplantation, over 90% of the seedlings were colonized with mycorrhizal fungus.

Each plot had an area of 20 × 12 m², which consisted of 6 rows, with 10 seedlings per row at 2 m row spacing. 100 g AMF was inoculated in root of AMF plants at seedling transplantation. *Funneliformis mosseae* BGXJ01 inoculums (Supplied by Beijing Academy of Agriculture and Forestry Sciences) were propagated on *Trifolium repens* (clover) for 12 weeks. 100 g inoculums included spores (10–20 spores/g), colonized root fragments (40 root fragments per gram inoculums, 85% root colonization), and external mycelium⁶⁵.

R_s determination. R_s was measured from July to September in individual plot with three replicates. Each measurement event was made in the middle of individual month (about 15th) using an Automated Soil Gas Flux System (Li-8100A, Li-cor Inc., Lincoln, Nebraska, USA) coupled with the small PVC chamber (20 cm × 10 cm). The instrument was permanently installed between two rows with identical distance. In each plot, measurements of R_s were taken inside three replicate PVC chambers with 20 cm in diameter and 10 cm in height, inserted to a depth of 2–3 cm in soil, capturing respirations. The CO₂ efflux from this collar mainly derived from soil organic matter decomposition, and root and microbial respiration. Meanwhile, soil temperature and volumetric moisture at the top of 5 cm was measured with the corresponding probes. Specifically, R_s was always measured between 8:00 and 12:00 in the morning in the sunny day without wind⁶⁶.

The cumulative R_s values were calculated by the method of Bremer *et al.*⁶⁷. Additionally, diel respiration (every 2 hours over 24-hour cycles) was periodically measured in all plots and individual collars. All of the measurements were made in four seasons from 2014 to 2015 due to time constraints. The diel measurements were used to calculate the annual growing season (July to September) cumulative R_s . Briefly, respiration measured during the daytime was assumed to be the daily maximum soil CO₂ efflux. Diel measurements were used to calculate the daily minimum efflux as a percentage of maximum efflux. The daily maximum and minimum efflux were used to calculate the average daily efflux. The cumulative flux as the product of average daily flux and the number of days was estimated between each measurement.

P_n determination. Plants assimilated CO₂ via P_n . To quantify photosynthetic performance of plant species, the measurement was with two AMF inoculation levels using a portable open system infrared gas analyzer for

net CO₂ assimilation rate of leaves in individual treatment (Li-6400, Li-cor Inc., Lincoln, Nebraska, USA). The experiment was performed from July to September in 2014, including three candidate trees within individual plot. Each measurement event was carried out at 08:00 to 18:00 in sunny day every 2 h, with three fully expanded healthy sun-exposed leaves of each species. Leaves were carefully positioned in the leaf chamber when they were exposed to the 1000 μmol m⁻² s⁻¹ saturating quantum flux level. Parameters including the instantaneous P_n, transpiration rate (T_r), stomatal conductance (G_s), intercellular CO₂ concentration (C_i), photosynthetically active radiation (PAR), atmosphere CO₂ concentration (C_a), atmosphere temperature (T_a), leaf temperature (T_{leaf}), and relative air humidity (RH) were tested.

Diurnal P_n amount was the area which surrounded by the curves of net photosynthetic rate > 0 and the time transverse in the diurnal curves variations of species P_n. According to the principle, the diurnal net photosynthetic amount was calculated as follows:

$$P = \sum_{i=1}^n [(p_{i+1} + p_i) \div 2 \times (t_{i+1} - t_i) \times 3600 \div 1000]$$

where P was the diurnal total net assimilation amount (mmol.m⁻².d⁻¹), p_i and p_{i+1} (μmol.m⁻².s⁻¹) was the instantaneous photosynthetic rate of the beginning and next measuring point, respectively; t_i and t_{i+1} was the time duration of the beginning and next measuring point; n was the numbers of determination times; one hour equaled to 3600 seconds; 1 mmol equaled to 1000 μmol.

Generally, the 20% diurnal assimilation photosynthates production was consumed by dark respiration, and converted the assimilation into the CO₂ diurnal sequestration as follows:

$$W_{CO_2} = P \times (1 - 0.2) \times 44/1000$$

where 44 was CO₂ mole mass; W_{CO₂} was CO₂ sequestration mass in the per unit leaf area (g.m⁻².d⁻¹).

According to the P_n reaction equation CO₂ + 4H₂O → CH₂O + 3H₂O + O₂, which the formula can be calculate the daily CO₂ absorption in the per unit land area of per plant as follow:

$$Q_{CO_2} = Y \times W_{CO_2}$$

Q_{CO₂} was the daily CO₂ sequestration in the per unit land area of per plant species (g.m⁻².d⁻¹).

The average CO₂ uptake per day of individual plants as follow:

$$Q_{CO_2} = Y \times p_{average} \times (1 - 0.2) \times 44/1000$$

$$P_{average} = (P_{Jul} + P_{Aug} + P_{Sep})/3$$

where Q_{CO₂} was the daily average CO₂ sequestration of each plant (g.d⁻¹); P_{average} was the daily assimilation in the per unit land area (mmol.m⁻².d⁻¹); Y was the total leaf area of each plant (m⁻²).

Soil OC, EE-GRSP and T-GRSP fractions. Soil samples were collected from 0–20 cm of the profile using an auger (35 mm diameter) in each corresponding monitoring stage in 2014. Three cores were collected from individual plot and combined to give one composite sample per plot of each tree species. The composite soil samples were air-dried and sieved through a 2 mm mesh and placed in plastic bags for chemical analysis.

A simple method for routine determination of soil OC by a modified Mebius procedure was described. It involved a digestion of the soil sample with an acidified dichromate (K₂Cr₂O₇-H₂SO₄) solution for 30 minutes in a Pyrex digestion tube in (a) 40 tube block digester preheated to 170 °C and (b) estimation of the un-reacted dichromate by titration of the cooled digest with an acidified solution of ferrous ammonium sulfate using the *N*-phenylanthranilic acid as an indicator.

0.25 g composite sample was extracted with 2 ml of extractant. EE-GRSP was extracted with 20 mM citrate solution; pH 7.0 at 121 °C for 30 min. T-GRSP was extracted with 50 mM citrate solution, pH 8.0 at 121 °C. 90 min was required for one soil sample and six additional sequential extractions. For the sequential extractions, the supernatant was removed by centrifugation at 10,000 × g for 5 min, 2 ml of 50 mM citrate, pH 8.0 was added to the residue, and samples were autoclaved for 60 min. Extraction of a sample continued until the supernatant showed no red-brown color typical of glomalins. Extracts from each replicate were pooled and then analyzed.

Citrate extractants were added to soil samples and disrupted by a brief (3 min) autoclave cycle. When necessary, the extractant was adjusted with HCl solution until the pH stabilized at 7.0 for 20 mM citrate or 8.0 for 50 mM citrate solution. Samples were then subjected to 121 °C for 90 min to extract T-GRSP or 30 min to extract EE-GRSP.

After extraction cycles completed, samples were centrifuged to remove soil particles (10,000 × g for 5 min), and protein in the supernatant was determined by the Bradford dye-binding assay with bovine serum albumin as standard. Concentration of glomalins was extrapolated to mg/g of soil particles by correcting the dry weight of coarse fragments >0.25 mm included in the weight of aggregates and the volume of extractant.

Data analysis. To assess the four plant species with and without AMF inoculation treatments to long-term exposure and potentially elucidate the role of AMF in previous response, the relationship of R_s and P_n, R_s and soil temperature, and carbon storage and AMF inoculation of the key issues have been a focus of this study.

The experiment was at two AMF inoculation levels with four plant species. To determine main effects of AMF inoculation on soil temperature and R_s variables, mixed model restricted maximum likelihood estimation with

repeated measures was used (PROC MIXED; version 8.0; SAS Institute Inc., Cary, NC, USA, 2003). Turkey's HSD multiple comparison was used to identify the differences between two AMF inoculation levels.

An exponential model was used to calculate soil temperature sensitivity on R_s ^{68,69}.

$$R_s = ae^{bT} \quad (1)$$

where R_s is soil CO₂ efflux ($\mu\text{mol m}^{-2} \text{s}^{-1}$), T is soil temperature ($^{\circ}\text{C}$) at 0–5 cm depth, a is the basal R_s , b is temperature sensitivity of CO₂ efflux. The respiratory quotient (Q_{10}) was calculated as $Q_{10} = e^{10b}$.

Regression coefficients, correlations, figures and curves were obtained by the Sigma-Plot software package (version 11.0, San Jose, California, USA). Analysis was performed to reveal the differences of two AMF inoculation levels with the relevant parameters; differences in means were revealed by LSD ($P < 0.05$) with SAS software package. To determine the effect of AMF on photosynthetic parameters, the data set was divided into two parts; the results were insensitive to variation in species groupings as the sample size was not unduly restricted. Differences in slopes were determined by a dummy variable representing the interaction between independent variable (AMF inoculation level) and plant species in the regression analyses.

Cumulative carbon calculations. To assess the effect of soil volumetric moisture on carbon accumulation, the cumulative carbon (Cumulative-C) was fitted using a combined exponential and quadratic function as follows:

$$\Delta_{\text{Cumulative-C}} = (ae^{bT})[2.12(\theta_v - \min \theta_v)(\max \theta_v - \theta_v)^c] \quad (2)$$

where θ_v is the volumetric moisture content (the minimum soil volumetric water content of our data set was 3.1% and the maximum was 37.6%, respectively) and c is the coefficient for soil moisture.

References

- Hu, Z. Q., Wei, Z. Y. & Qin P. Concept and methods for soil reconstruction in mined land reclamation. *Soils*. **37**(1), 8–12 (In Chinese) (2005).
- Zhang, F. W. *et al.* The effect of coal-mining subsidence on water environment in the shenfu-dongsheng mining area. *Acta Geoscientia Sinica*. **28**(6), 521–527 (In Chinese)] (2007).
- McSweetney, K. & Jansen, I. J. Soil structure and associated rooting behavior in minesoils. *Soil Sci Soc of Am J*. **48**, 607–612, doi: 10.2136/sssaj03615995004800030028x (1984).
- Vitousek, P. M. *et al.* Human domination of Earth's ecosystems. *Science* **277**, 494–499, doi: 10.1126/science.277.5325.494 (1997).
- Levy, M. A. & Cumming J. R. Development of soils and communities of plants and arbuscular mycorrhizal fungi on West Virginia surface mines. *Environ Manage.* **54**(5), 1153–1162, doi: 10.1007/s00267-014-0365-0 (2014).
- Sinha, S. *et al.* Rhizosphere soil microbial index of tree species in a coal mining ecosystem. *Soil Biol Biochem.* **41**, 1824–1832, doi: 10.1016/j.soilbio.2008.11.022 (2009).
- Ussiri, D. A. N., Lal, R. & Jacinthe, P. A. Soil properties and carbon sequestration of afforested pastures in reclaimed minesoils of Ohio. *Soil Sci Soc of Am J*. **70**, 1797–1806, doi: 10.2136/2005.0352 (2006).
- Pedrol, N. *et al.* Soil fertility and spontaneous revegetation in lignite spoil banks under different amendments. *Soil Till Res.* **110**, 134–142, doi: 10.1016/2010.07.005 (2010).
- Friedlingstein, P. *et al.* Climate-carbon cycle feedback analysis: results from the C4 MIP model intercomparison. *J Climate*. **19**, 3337–3353, doi: http://dx.doi.org/10.1175/3800.1 (2006).
- Bahn, M. *et al.* Soil respiration in European grasslands in relation to climate and assimilate supply. *Ecosystems*. **11**, 1352–1367, doi: 10.1007/s10021-008-9198-0 (2008).
- Luo, Y. Q. & Zhou, X. H. *Soil respiration and the environment*. Academic Press, San Diego, California, USA (2010).
- Qi, Y., Xu, M. & Wu, J. G. Temperature sensitivity of soil respiration and its effects on ecosystem carbon budget: nonlinearity begets surprises. *Ecol Model.* **153**, 131–142, doi: 10.1016/S0304-3800(01)00506-3 (2002).
- Lee, X. H. *et al.* Rapid and transient response of soil respiration to rain. *Global Change Biol.* **10**, 1017–1026, doi: 10.1111/j.1529-8817.2003.00787.x (2004).
- Vicca, S. *et al.* Arbuscular mycorrhizal fungi may mitigate the influence of a joint rise of temperature and atmosphere CO₂ on soil respiration in grasslands. *International Journal of Ecology*. doi: 10.1155/2009/209768 (2009).
- Sanders, I. R. *et al.* Increased allocation to external hyphae of arbuscular mycorrhizal fungi under CO₂ enrichment. *Oecologia* **117**(4), 496–503, doi: 10.1007/s004420050685 (1998).
- Chapin, F. S., Schulze, E. D. & Mooney, H. A. The ecology and economics of storage in plants. *Annu Rev Ecol Syst.* **21**, 423–447, doi: http://www.jstor.org/stable/2097032 (1990).
- Treseder, K. K. A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and atmospheric CO₂ in field studies. *New Phytol.* **164**(2), 347–355, doi: 10.1111/j.1469-8137.2004.01159.x (2004).
- Diaz, S. Effects of elevated CO₂ at the community level mediated by root symbionts. *Plant Soil* **187**(2), 309–320, doi: 10.1007/BF00017096 (1996).
- Wilson, G. W. *et al.* Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: results from long-term field experiments. *Ecol Lett.* **12**(5), 452–461, doi: 10.1111/j.1461-0248.2009.01303.x (2009).
- Rillig, M. C. *et al.* Rise in carbon dioxide changes soil structure. *Nature* **400**(6745), 628, doi: 10.1038/23168 (1999).
- Heinemeyer, A. & Fitter, A. H. Impact of temperature on the arbuscular mycorrhizal (AM) symbiosis: growth responses of the host plant and its AM fungal partner. *J Exp Bot.* **55**(396), 525–534, doi: 10.1093/jxb/erh049 (2004).
- Hawkes, C. V. *et al.* Soil temperature affects carbon allocation within arbuscular mycorrhizal networks and carbon transport from plant to fungus. *Global Change Biol.* **14**(5), 1181–1190, doi: 10.1111/j.1365-2486.2007.01535.x (2008).
- Kaschuk, G. *et al.* Are the rates of photosynthesis stimulated by the carbon sink strength of rhizobial and arbuscular mycorrhizal symbiosis? *Soil Biol Biochem.* **41**, 1233–1244, doi: 10.1016/j.soilbio.2009.03.005 (2009).
- Tian, Y. H. *et al.* Synergistic effect of colonization with arbuscular mycorrhizal fungi improves growth and drought tolerance of *Plukenetia volubilis* seedlings. *Acta Physiol Plant.* **35**, 687–696, doi: 10.1007/s11738-012-1109-5 (2013).
- Cavagnaro, T. R. *et al.* Growth, nutrition, and soil respiration of a mycorrhiza-defective tomato mutant and its mycorrhizal wild-type progenitor. *Funct Plant Biol.* **35**, 228–235, http://dx.doi.org/10.1071/FP07281 (2008).
- Moyano, F. E., Kutsch, W. L. & Schulze, E. D. Response of mycorrhizal, rhizosphere and soil basal respiration to temperature and photosynthesis in a barley field. *Soil Biol Biochem.* **39**, 843–853, doi: 10.1016/j.soilbio.2006.10.001 (2007).
- Verbruggen, E. *et al.* Arbuscular mycorrhizal fungi-short-term liability but long-term benefits for soil carbon storage? *New Phytol.* **197**, 366–368, doi: 10.1111/nph.12079 (2013).

28. Zhang, J. *et al.* Glomalin-related soil protein responses to elevated CO₂ and nitrogen addition in a subtropical forest: Potential consequences for soil carbon accumulation. *Soil Biol Biochem.* **83**, 142–149, doi: 10.1016/j.soilbio.2015.01.023 (2015).
29. Wang, X. R. *et al.* Effect of co-inoculation with arbuscular mycorrhizal fungi and rhizobia on soybean growth as related to root architecture and availability of N and P. *Mycorrhiza* **21**, 173–181, doi: 10.1007/s00572-010-0319-1 (2010).
30. Cheng, L. *et al.* Arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO₂. *Science* **337**, 1084–1087, doi: 10.1126/science.1224304 (2012).
31. van Groenigen, K. J., Osenberg, C. W. & Hungate, B. A. Increased soil emissions of potent greenhouse gases under increased atmospheric CO₂. *Nature* **475(7355)**, 214–216, doi: 10.1038/nature10176 (2011).
32. Reich, P. B. *et al.* Nitrogen limitation constrains sustainability of ecosystem response to CO₂. *Nature* **440(7086)**, 922–925, doi: 10.1038/nature04486 (2006).
33. Nemeč, S. & Vu J. V. C. Effect of soil phosphorus and Glomus intraradices on growth, nonstructural carbohydrates, and photosynthetic activity of Citrus aurantium. *Plant Soil* **128**, 257–263, doi: 10.1007/BF00011117 (1990).
34. Smith, S. E. & Read D. J. *Mycorrhizal Symbiosis*. 2nd ed. London: Academic Press (1997).
35. Douds, D. D. Jr., Pfeffeer, P. E. & Schachar-Hill, Y. *Carbon partitioning, cost, and metabolism of arbuscular mycorrhizas*.-In Kapulnik, Y. & Douds, D. D., Jr. (ed.): *Arbuscular Mycorrhizas: Physiology and Function*. Kluwer Academic Publ., Dordrecht; pp. 107–130 (2000).
36. Davies, F. T., Potter, J. R. & Linuerman, R. G. Drought resistance of mycorrhizal pepper plants independent of leaf P concentration-response in gas exchange and water relations. *Physiol Plantarum* **87(1)**, 45–53, doi: 10.1111/j.1399-3054.1993.tb08789.x (1993).
37. Koide, R. T. Nutrient supply, nutrient demand and plant response to mycorrhizal infection. *New Phytol.* **117**, 365–386, doi: 10.1111/j.1469-8137.1991.tb00001.x (1991).
38. Wu, Q. S. *et al.* Arbuscular mycorrhiza mediates glomalin-related soil protein production and soil enzyme activities in the rhizosphere of trifoliolate orange grown under different P levels. *Mycorrhiza* **25**, 121–130, doi: 10.1007/s00572-014-0594-3 (2015).
39. Augé, R. M. Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. *Mycorrhiza* **11**, 361–365, doi: 10.1007/s005720100097 (2001).
40. Ruehr, N. K. *et al.* Drought effects on allocation of recent carbon: from beech leaves to soil CO₂ efflux. *New Phytol.* **184**, 950–961, doi: 10.1111/j.1469-8137.2009.03044.x (2009).
41. Sönmez, F. & Gülser, F. Effects of arbuscular mycorrhizal fungus and salicylic acid on nutrient uptake by maize (*Zea mays* L.) seedlings in cadmium contaminated media. *J. Int. Environmental Application & Science* **9(5)**, 608–613, doi: 280620985 (2014).
42. Kiers, E. T. *et al.* Reciprocal rewards stabilize cooperation in the mycorrhizal symbiosis. *Science* **333(6044)**, 880–882, doi: 10.1126/science.1208473 (2011).
43. Drigo, B. *et al.* Shifting carbon flow from roots into associated microbial communities in response to elevated atmospheric CO₂. *Proc Natl Acad Sci USA* **107(24)**, 10938–10942, doi: 10.1073/pnas.0912421107 (2010).
44. Tisdall, J. M., Smith, S. E. & Rengasamy, P. Aggregation of soil by fungal hyphae. *Aust J Soil Res.* **35(1)**, 54–60, doi: 10723.35400006225255.0050 (1997).
45. Sanders, I. R. *et al.* Increased allocation to external hyphae of arbuscular mycorrhizal fungi under CO₂ enrichment. *Oecologia* **117(4)**, 496–503, doi: 10.1007/s004420050685 (1998).
46. Treseder, K. K. & Allen, M. F. Mycorrhizal fungi have a potential role in soil carbon storage under elevated CO₂ and nitrogen deposition. *New Phytol.* **147(1)**, 189–200, doi: 10.1046/j.1469-8137.2000.00690.x (2000).
47. Alberton, O., Kuyper, T. W. & Gorissen, A. Taking mycorrhizal seriously: mycorrhizal fungal and plant responses to elevated CO₂. *New Phytol.* **167(3)**, 859–868, doi: 10.1046/j.1469-8137.2005.01458.x (2005).
48. Rillig, M. C. *et al.* Long-term CO₂ elevation affects soil structure of natural ecosystems. *Nature* **400**, 628, doi: 10.1038/23169 (1999).
49. Hu, S. *et al.* Nitrogen limitation of microbial decomposition in a grassland under elevated CO₂. *Nature* **409(6817)**, 188–191, doi: 10.1038/35051576 (2001).
50. Orwin, K. H. *et al.* Organic nutrient uptake by mycorrhizal fungi enhances ecosystem carbon storage: a model-based assessment. *Ecol Lett.* **14(5)**, 493–502, doi: 10.1111/j.1461-0248.2011.01611.x (2011).
51. Sanders, I. R. *et al.* Increased allocation to external hyphae of arbuscular mycorrhizal fungi under CO₂ enrichment. *Oecologia* **117(4)**, 496–503, doi: 10.1007/s004420050685 (1998).
52. Treseder, K. K. & Allen, M. F. Mycorrhizal fungi have a potential role in soil carbon storage under elevated CO₂ and nitrogen deposition. *New Phytol.* **147(1)**, 189–200, doi: 10.1046/j.1469-8137.2000.00690.x (2000).
53. Alberton, O., Kuyper, T. W. & Gorissen, A. Taking mycorrhizal seriously: mycorrhizal fungal and plant responses to elevated CO₂. *New Phytol.* **167(3)**, 859–868, doi: 10.1046/j.1469-8137.2005.01458.x (2005).
54. Rillig, M. C. *et al.* Long-term CO₂ elevation affects soil structure of natural ecosystems. *Nature* **400**, 628, doi: 10.1038/23169 (1999).
55. Hu, S. *et al.* Nitrogen limitation of microbial decomposition in a grassland under elevated CO₂. *Nature* **409(6817)**, 188–191, doi: 10.1038/35051576 (2001).
56. Orwin, K. H. *et al.* Organic nutrient uptake by mycorrhizal fungi enhances ecosystem carbon storage: a model-based assessment. *Ecol Lett.* **14(5)**, 493–502, doi: 10.1111/j.1461-0248.2011.01611.x (2011).
57. Shan, J., Morris, L. A. & Hendrick, R. L. The effects of management on soil and plant carbon sequestration in slash pine plantations. *J Appl Ecol.* **38(5)**, 932–941, doi: 10.1046/j.1365-2664.2001.00648.x (2001).
58. Rillig, M. C. & Mummey, D. L. Mycorrhizas and soil structure. *New Phytol.* **171(1)**, 41–53, doi: 10.1111/j.1469-8137.2006.01750.x (2006).
59. van der Heijden, M. G. *et al.* Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature* **396(6706)**, 69–72, doi: 10.1038/23932 (1998).
60. Langley, J. A. & Hungate, B. A. Mycorrhizal controls on belowground litter quality. *Ecology* **84(9)**, 2302–2312, doi: 10.1890/02-0282 (2003).
61. Iqbal, J. *et al.* Fungal endophyte infection increases carbon sequestration potential of southeastern USA tall fescue stands. *Soil Biol Biochem.* **44(1)**, 81–92, doi: 10.1007/s00442-014-5 (2012).
62. Tang, J., Baldocchi, D. D. & Xu L. Tree photosynthesis modulates soil respiration on a diurnal time scale. *Global Change Biol.* **11**, 1298–1304, doi: 10.1111/j.1365-2486.2005.00978.x (2005).
63. Baath, E. & Wallander, H. Soil and rhizosphere microorganisms have the same Q₁₀ for respiration in a model system. *Global Change Biol.* **9(12)**, 1788–1791, doi: 10.1046/j.1365-2486.2003.00692.x (2003).
64. Reichstein, M. & Janssens, I. A. Semi-empirical modeling of the response of soil respiration to environmental factors in laboratory and field conditions. In W. Kutsch, M. Bahn, A. Heinemeyer (Eds), *Soil Carbon Dynamics-an Integrated Methodology*. Cambridge University Press, Cambridge, UK (pp. 207–220), <http://hdl.handle.net/11858/00-001M-0000-000E-D8B7-6> (2009).
65. Plenck, C. & Morel, C. External phosphorus requirement of mycorrhizal and non-mycorrhizal barley and soybean plants. *Biol Fert Soils.* **21(4)**, 303–308, doi: 10.1007/BF00334907 (1996).
66. Heinemeyer, A. *et al.* Respiration of the mycelium in the arbuscular mycorrhizal symbiosis shows strong dependence on recent photosynthesis and acclimation to temperature. *New Phytol.* **171(1)**, 159–170, doi: 10.1111/j.1469-8137.2006.01730.x (2006).
67. Bremer D. J. *et al.* Responses of soil respiration to clipping and grazing in a tallgrass prairie. *J Environ Qual.* **27(6)**, 1539–1548, doi: 10.2134/jeq1998.00472425002700060034x (1998).
68. Zhou, X., Wan, S. Q. & Luo, Y. Q. Source components and interannual variability of soil CO₂ efflux under experimental warming and clipping in a grassland ecosystem. *Global Change Biol.* **13**, 761–775, doi: 10.1111/j.1365-2486.2007.01333.x (2007).
69. Wan, S. Q. & Luo, Y. Q. Substrate regulation of soil respiration in a tallgrass prairie: results of a clipping and shading experiment. *Global Biogeochem Cy.* **17(2)**, 1–23, doi: 10.1029/2002GB001971 (2003).

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Author Contributions

Z.-G.W. initialized the study; Conceived and designed the experiments: Z.-G.W and Y.-L.B. performed the experiments: Z.-G.W., B.J., Y.Z., W.-W.L., H.L., Z.-G.W. and Y.-L.B. performed the data analysis; Z.-G.W. and Y.-L.B. drafted the manuscript; contributed reagents/materials/analysis tools: Z.-G.W., Y.-L.B., B.J., Y.Z., S.-P.P., W.-W.L. and H.L.

Additional Information

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