# Research Article

# Schizophrenia: A Pathogenetic Autoimmune Disease Caused by Viruses and Pathogens and Dependent on Genes

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Many genes have been implicated in schizophrenia as have viral prenatal or adult infections and toxoplasmosis or Lyme disease. Several autoantigens also target key pathology-related proteins. These factors are interrelated. Susceptibility genes encode for proteins homologous to those of the pathogens while the autoantigens are homologous to pathogens' proteins, suggesting that the risk-promoting effects of genes and risk factors are conditional upon each other, and dependent upon protein matching between pathogen and susceptibility gene products. Pathogens' proteins may act as dummy ligands, decoy receptors, or via interactome interference. Many such proteins are immunogenic suggesting that antibody mediated knockdown of multiple schizophrenia gene products could contribute to the disease, explaining the immune activation in the brain and lymphocytes in schizophrenia, and the preponderance of immune-related gene variants in the schizophrenia genome. Schizophrenia may thus be a "pathogenetic" autoimmune disorder, caused by pathogens, genes, and the immune system acting together, and perhaps preventable by pathogen elimination, or curable by the removal of culpable antibodies and antigens.

#### 1. Introduction

Over 600 genes have been implicated in schizophrenia in association studies, supporting the contention that multiple genes of small effect contribute to this condition [1, 2] (see http://www.polygenicpathways.co.uk/schizgenesandfunc.htm for association references). These genes cluster together in clearly defined signalling networks related to the diverse subpathologies of schizophrenia [3–7]. Epistasis between genes within these same signalling networks markedly affects the degree of risk-promotion [8–10], in part, explaining the inconsistency in genetic association studies.

Schizophrenia has also been associated with prenatal complications including maternal rubella (German measles) [11], influenza [12, 13], Varicella zoster (chicken pox) [14], Herpes (HSV-2) [15], common cold infection with fever [16], or poliovirus infection [17] while in childhood or adulthood, coxsackie virus infection (in neonates [18]) or Lyme disease (vectored by the Ixodes tick and Borrelia Burgdorferri) or Toxoplasmosis have been reported as risk

factors [19, 20] (see Table 1). The human endogenous retrovirus, HERV-W, has also been implicated in schizophrenia [21]. A number of schizophrenia-related genes are implicated in the life cycles of these pathogens, suggesting an interplay between genes and risk factors [22].

Many schizophrenia genes relate to the immune network [5, 6, 22, 23]. Immune activation is also observed in the schizophrenic brain [24, 25] or in lymphocytes [26–29]. A number of autoantigens/autoantibodies to key schizophrenia-related proteins have also been reported. These include dopamine, serotonin, acetylcholine, and NMDA receptors; inter alia (Table 2). Maternal immune activation in animal models has also been shown to generate phenotypes relevant to schizophrenia in the offspring [30].

As shown below, genes, risk factors, and immunity can be linked together forming a unifying pathway whose elements are interdependent. Dysfunction of this network which is conditional upon interactions between its three branches may be responsible for schizophrenia.

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500



#### Rubella 304 hits



HSV-2 242 hits



No filter



#### Rhinovirus 284 hits



Chromosome 10 versus viral prot 119857 hits Distribution of 10867 blast hits on query sequence



FIGURE 1: Continued.

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FIGURE 1: Screenshots of the pictorial representation of the viral BLAST results against the human proteome. The streaks dotted throughout the human genome/proteome represent the areas of homology, some with contiguous sequences of 5 or more amino acids. The number of hits is shown for each virus or pathogen. The figure also shows the total coverage of human chromosome 10 by viral gene homologues. The top set of figures were from unfiltered blasts while the bottom set of 6 figures represent filtered blasts using the query "schizophrenia".

The larger font illustrates highly antigenic regions of DISC1 (and of the viral homologues). The boxes represent the alignment position and the blue letters 100% identity.

>gi|61742823|ref|NP\_061132.2| disrupted in schizophrenia 1 protein isoform L [Homo sapiens]



>gi|61742823|ref|NP\_061132.2| disrupted in schizophrenia 1 protein isoform L [Homo sapiens]



FIGURE 2: Continued.



FIGURE 2: Continued.



FIGURE 2: Continued.



(c) The homology of viral risk factors to the highly antigenic regions of DISC1 (and of the viral homologues)

FIGURE 2: (a) Varicella protein alignments within DISC1: the boxed regions show the region of alignment, and the blue letters denote 100% identity. This is not an alignment of the whole Varicella proteome but represents fragments of the same or different Varicella proteins that align with DISC1 fragments (vatches). The larger font delineates highly antigenic regions of DISC1 with an antigenicity index of >0.8 (Figure 4). (b) Other viral vatches within the DISC1 protein. The vatches are colour or format coded in relation to the different viruses. (c) Viral vatches for the risk factors implicated in schizophrenia in relation to the highly immunogenic regions of DISC1.

#### 2. Methods

The human herpesvirus 2 genome (NC\_001798) as well as those of the rhinovirus (NC\_001490), rubella (NC\_001545.1) and Varicella zoster (NC\_001348.1) and HERV-W (NP\_055405.3: env polyprotein) viruses, Borrelia Burgdorferri (NC\_011728) and T. Gondii (NC\_001799: Partial genome) were screened against the human proteome using the NCBI BLAST server and the Entrez query filter "schizophrenia". The HERV-W, influenza, HSV-2 and rubella viruses were also screened unfiltered (Translated pathogen genome versus human proteins: BlastX) [31]. The BLAST algorithm detects overall homology between entire gene or protein sequences, and it is necessary to set parameters to low significance levels in order to detect short intraprotein consensus homology. The parameters used were: Expect 20,000, E value = 100,000; matrix PAM30. The original BLAST results are stocked at http://www.polygenicpathways.co.uk/blasts.htm. Information for all abbreviations is available at this site, provided by the NextBio highlighting service.

BLAST files were scanned by an online tag cloud generator producing tags sized according to gene word occurrences http://www.tagcloud-generator.com/ generator.php#anker. Word occurrences were counted using a "Highlightall add-in" for Firefox https://addons .mozilla.org/en-US/firefox/addon/4240/.

Antigenicity (B-cell epitope prediction) was estimated using the BepiPred server http://www.cbs.dtu.dk/services/ BepiPred/[32] (Table 4).

Kegg pathway analysis [33] of 632 schizophrenia susceptibility gene candidates was performed using Kegg mapper http://www.genome.jp/kegg/tool/color\_pathway.html. The results of this analysis are available at http://www .polygenicpathways.co.uk/keggszgenes.htm. Venn diagrams were constructed online at http://www.bioinformatics.org/ gvenn/index.htm[34].

Genes and risk factors with at least one positive association are included in this study. Although certain genes and risk factors are clearly more important than others, and problems of replication in both gene and risk factor studies abound, gene, gene, and gene/environment interactions may explain some of the heterogeneity. For example many schizophrenia-related genes are involved in the life cycle of T. Gondii, but may be irrelevant if this pathogen is not encountered. Similarly T. Gondii infection may have little effect is such gene variants are not present. Pathway analyses of genome wide association data, and previous studies, are showing that the risk-promoting effects of many genes in similar pathways are better predictors of risk, than when treating each gene in isolation (see Section 1). Although some of these factors may be false positives, many genes and risk factors may have a role to play in certain conditions, but the greater import of genes such as DISC1 or neuregulin is recognised.

#### 3. Results

Pictograms of selected BLAST results are shown in Figure 1. The initial sweep of unfiltered BLASTs returned 125–304 hits, but this number was markedly increased when using the filter "schizophrenia" (14,088 Hits for HSV-2). For unfiltered sweeps, the viral homologues are longer, while the filtered sweeps return shorter contiguous sequences nevertheless including multiple matches of pentapeptides or more.

Viral-human matches are characterised by short contiguous amino acid matches of 5 or more amino acids, that are identical in viral and human proteins, defined as vatches (viral matches). These are exemplified, for DISC1 in Figure 2. Hexapeptide matches have also been described for the influenza H5N1 virus and this study also highlighted homologies with DISC1, reelin and neurexin, inter alia [35]. The entire length of a human protein can be composed of



FIGURE 3: (a) Venn diagrams of the number of Schizophrenia gene products (N = 632) with homology to the rubella, HERV-W and influenza viruses. The singleton in SZ-genes was different on each occasion: Thus, all genes are covered. (b) The viral matching spectra of DISC1, neuregulin, the dopamine D2 receptor and transcription factor 4. The *Y*-axis depicts the number of word occurrences on the original BLAST results page. Note the logarithmic axis. (c) The number of pathogens expressing proteins with homology to the protein products of schizophrenia susceptibility genes. Those marked by an asterisk are within the 30 top-ranked genes in SZ-gene http://www.szgene.org/.



FIGURE 4: The antigenicity (B-cell epitope prediction) of DISC1: the amino acid sequences with an index of >0.35 are considered as epitopes. A value of 0.8 was chosen to define highly antigenic regions as seen in Figure 2. The amino acid sequences of these highly antigenic regions are shown.

many overlapping, intercalated vatches, related to multiple viral species. However, the viral spectrum is distinct for each protein as shown in Figure 3 for DISC1, neuregulin, the D2 dopamine receptor and transcription factor 4. Each is homologous to proteins from a large spectrum of viruses, but this spectrum is distinct for each protein. Interestingly, all are homologous to proteins from the hepatitis C virus. Several studies have noted that Hepatitis C infection is associated with schizophrenia, but this has generally been interpreted in terms of a schizophrenia life style that favours infection, rather than viewing Hepatitis C as a risk-promoting factor [36–39]. These data may challenge this assumption.

All of the pathogens implicated in schizophrenia express proteins with homology to multiple schizophrenia susceptibility gene products (Table 3). The profile of each individual pathogen is again specific for different types of gene product, but all target key members of the schizophrenia network including dopamine, serotonin and glutamate receptors as well as neuregulin and growth-related or DISC1 related pathways. This is the case even when no filter is used. Interestingly, both the rubella and the influenza viruses target members of the translation initiation complex, which has been implicated in myelination and oligodendrocyte survival [4, 40]. Oligodendrocyte cell loss and myelination defects are prominent in the schizophrenic brain [41–44].

The degree of overlap between the rubella, HERV and influenza viruses and schizophrenia gene products is shown

by the Venn diagrams in Figure 3. All but one schizophrenia gene product was covered by various permutations and similar data were recovered for other pathogens. All schizophrenia gene products (N = 632) were homologous to proteins expressed by one or more of these pathogens. However, only 16 proteins were common to all 8 pathogens (Figure 3). These included neuregulin (NRG1) and DISC1, dopamine (DRD5), glutamate (GRIA4, GRID1, GRM3, GRM7) GABA (GABBR1) and serotonin (HTR7) receptors, a presynaptic protein regulating glutamate release (synapsin SYN3) and HOMER2, a member of the postsynaptic scaffold, all of which are key elements relating to the pathology of schizophrenia.

Other proteins within this class included neurocan (CSPG5), a chondroitin sulphate proteoglycan expressed in oligodendrocytes that inhibits neurite outgrowth and regulates axonal growth [45–47]. It is also involved thalamocortical projection development [48]. ARHGEF10 is a rho Guanine-nucleotide exchange factor that controls myelination [49]. NDUFV2 is a subunit of the mitochondrial respiratory chain and its protein expression levels are reduced in the frontal cortex and striatum in schizophrenia [50]. PPP3CC Calcineurin gamma (PPP3CC) plays a role in dopamine receptor signalling [51, 52]. Calcineurin knockout mice show defects in prepulse inhibition and other phenotypes related to schizophrenia [53]. Calcineurin is highly expressed in the immune system and regulates the expression of numerous



Red lines = direct interaction with DISC1 Blue lines = separation from DISC1 by one binding partner

FIGURE 5: The DISC1 interactome see http://www.polygenicpathways.co.uk/discforum.htm. Proteins in red are homologous to Rubella proteins.

cytokines [54]. MAP6 is a microtubule protein that controls synaptic organisation, in particular of glutamatergic synapses where it controls the expression of the glutamate transporter and presynaptic genes, synaptophysin and GAP-43, spinophilin and MAP2. [55, 56] KCNH2 is a potassium channel that plays a role in the development of neural crest cells [57] and in lymphocyte proliferation [58]. PRSS16 is a serine protease involved in autoimmunity and the presentation of self-antigens within the thymus [59].

So, by a random bioinformatics process, trawling the entire human proteome, asking simply which proteins are homologous to those of the pathogens implicated in schiz-ophrenia, we arrive at a small set of proteins related to synaptic and dendritic function, myelination, neuregulin and DISC1 pathways, glutamate, dopamine, GABA and serotonin transmission, and immune regulation that are the cornerstones of schizophrenia pathology [3, 60–62].

3.1. Autoantigens in Schizophrenia. Many autoantibodies have been reported in schizophrenia. The pathogens implicated in schizophrenia also express proteins that are homologous to these autoantigens. Again the profile of each autoantigen or pathogen is distinct as shown in Table 2.

3.2. DISC1. DISC1 is a key "hub gene" in schizophrenia linked, via its interactome, to many other schizophrenia susceptibility gene products [3, 63–66]. Its viral homology is illustrated in Figure 2. The Varicella virus is homologous to DISC1 in several regions, over its entire length, many matches in regions of high immunogenicity. These figures illustrate the types of matches seen in other proteins and shows that the vatches are often part of larger gapped consensus sequences. Interestingly, Varicella infection also results in the production of antibodies to pericentrin, a DISC1 binding partner [67].

DISC1 is a highly immunogenic protein, as predicted by B-cell epitope prediction (Figure 4). Autoantibodies to DISC1 have not been reported in schizophrenia. However, the viral risk factors implicated in schizophrenia express proteins that are homologous to the highly antigenic regions of the DISC1 protein, as shown in Figure 2. These viral proteins are equally antigenic and antiviral antibodies might also thus be expected to target multiple regions of the DISC1 protein.

3.3. Viral Proteins Are Part of the DISC1 Interactome. DISC1 and many of its binding partners, or other members of



HSV-2 filter "dopamine receptor"

FIGURE 6: A screen shot of the HSV-2 BLAST results using the filter "dopamine receptor". The repeated patterns correspond to dopamine receptors on different chromosomes as shown in Table 1. Homology with glutamate, serotonin, GABA, acetylcholine and other receptors is also noted.

its interactome, contain vatches that are homologous to proteins expressed by the Rubella virus (Figure 5). (Other viruses also display this property, although the interactome members targeted are distinct, and specific for each virus (see http://www) .polygenicpathways.co.uk/vatches.htm). Upon infection, viruses might therefore be considered as extraneous spurs to these types of protein/protein networks, and are likely to markedly affect their integrity. Indeed, several viruses, including herpes simplex, hepatitis C, Epstein-Barr, the cytomegalovirus, adenovirus and Coxsackie virus are known to bind to DISC1 interaction partners (Table 4).

3.4. Viral DNA within the Human Genome. The insertion of viral DNA into the human genome had until recently been thought to be the preserve of retroviruses. However the incorporation of DNA into mammalian genomes has recently been demonstrated on a large scale for both RNA and DNA viruses. Viral integration may be mediated by nonhomologous recombination with chromosomal DNA or, in the case of RNA viruses, by interactions with host chromosomal retrotransposons [68, 69]. It has also been shown the herpes virus HHV-6 can be transmitted from parent to child via chromosomal integration [70]. The BLAST analyses of the viruses detailed in this paper, and of others at http://www.polygenicpathways.co.uk/blasts.htm clearly show that viral DNA from many species is present within the human genome. This viral homology may well cover the entire human genome. For example, a Blast of human chromosome 10 against all viral genomes (almost 3,000 viral forms) yielded 119,857 hits with entire coverage of 135.5 million bases. Viral DNA is thus both inter and intragenic (Figure 1). It has been proposed that retroviral integration, into paternal and maternal gene lines, inserting several genes at once and effectively creating a new being, is responsible for evolutionary saccades [71]. The fact that RNA and DNA nonretroviruses can also be so incorporated has important implications in this area.

The HSV-2 virus is homologous to several dopamine receptors and the BLAST pictogram shows how the same virus provokes repeating patterns in the human proteome (Figure 6). The same is true of the Herpes simplex virus (HSV-1) which is homologous to multiple lipoprotein receptors as well as to multiple kinases or of the cytomegalovirus which expresses proteins homologous to many chemokine receptors (see http://www.polygenicpathways.co.uk/blasts.htm). One interpretation of this, given the ability of chromosomal integration, is that repeated viral visits to the human genome over millions of years are responsible for the creation of gene families.

It is also possible that viral/human homology reflects convergent viral evolution, although this is difficult to reconcile with the presence of viral DNA in intergenic regions, for which there would be little evolutionary drive or selective pressure. It is also plausible that a bidirectional transfer of human and viral DNA could be at work.

For whatever reason, the result is that human proteins resemble those expressed by a multitude of today's viruses and other pathogens. Upon infection, these pathogens are thus able to interfere with the function of their human counterparts in a number of ways (see below).

3.5. Copy Number Variations and the Effects of Parental Age on Risk. Repeated viral insertion could well explain copy number variations, which are associated with a number of diseases, including schizophrenia [72, 73]. As their number increases, so will the number of matches to the same viral proteins, thus increasing the risk of viral interference and autoimmunity. As viral infection can be passed from parent to child via chromosomal integration, perhaps this is also why both paternal and maternal older age have been reported as risk factors in schizophrenia and other disorders [74, 75].

3.6. KEGG Pathway Analysis of Schizophrenia Susceptibility Genes. The color-coded pathways for this analysis are posted at http://www.polygenicpathways.co.uk/keggszgenes.htm. It confirmed the involvement of a number of polygenic pathways, including long-term potentiation and oxidative stress [3] growth factor/neuregulin pathways [121], neuroactive ligand pathways (dopamine/serotonin/glutamate and others) as well as dopamine metabolism pathways [9]. In the context of this review, a large number of immune-related pathways are traced out by these genes, together with many pathogenrelated pathways, including toxoplasmosis, which heads the list (Table 5). The involvement of schizophrenia related genes in the life cycles of pathogens has been the subject of a previous review [22] and this relationship is supported by this analysis. Other pathogen related pathways relating to amoebiasis, Staphylococcus aureus and Helicobacter pylori infection, might indicate the involvement of other pathogens in schizophrenia, although such pathways could also be considered as generic pathways related to many pathogens.

There is no specific viral life cycle pathway within the KEGG dataset. However, viruses use adhesion molecules as receptors, endocytosis for cellular entry and the intracellular actin and tubulin networks for migration to and from the nucleus, mediated via dynein and kinesin motors. They also subjugate intracellular vesicular trafficking pathways, and are able to subvert both lysosomal and phagosomal pathways. Their exit may depend upon exocytosis, or by apoptotic or other means of killing their host cell [122]. These pathways are heavily represented within the schizophrenia gene analysis.

*3.7. Mechanisms of Action.* Individual proteins are homologous to multiple viral proteins, which nevertheless are specific for a spectrum of viruses, while individual viruses are homologous to a large but specific subset of human proteins.

Our proteomes therefore contain proteins with sequences exactly matching those in the current virome, and in the proteomes of bacteria and other pathogens, which are also subject to phage or viral infection. Pathogens' proteins are therefore homologous to receptors, transporters, peptide messengers, growth factors, and other protein products of diverse gene families. Upon infection, surrogate dopamine, NMDA serotonin and other receptors, as well as transporters and enzymes are made available, which in effect may steal the ligands of their human counterparts. It is already known that the dopaminergic ligand, amantadine, binds to the influenza virus [123], which expresses proteins homologous to dopamine receptors (Table 3). When homologous to peptide ligands, viral proteins may occupy and block or perhaps stimulate their cognate receptors, or use them for entry, as is the case with the AIDS virus and the CCR5 and CXCR4 chemokine receptors [124].

This is illustrated by the Norovirus (Norwalk) which causes vomiting sickness. The virus expresses proteins homologous to monoamine and other amine oxidases as well as to a number of dopamine and monoamine transporters (Table 6). Dopamine subversion by the viral homologues would be expected to increase dopamine levels resulting in emesis, thus explaining the recurrent vomiting produced by infection.

The potential interference by viruses within protein/ protein networks is well illustrated by the homology of rubella proteins to DISC1 and other members of its interactome, and by the fact that many viruses have indeed been found to bind to these components (Table 4).

The homologous human proteins of the viral risk factors implicated in schizophrenia correspond to the genomic

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Pre- and perinatal maternal infection	Juvenile (in offspring)	Adult
Rubella (first trimester) [76]: Influenza (first trimester) [13] Influenza or common cold with fever (second trimester) [16]	Mumps or cytomegalovirus infection (0–12 years old) [77]	HSV-1 seropositivity related to grey matter volume [78]
Poliovirus (second trimester) [17]	Coxsackie B5 infection perinatally [18]	HSV-1 (in Afro-Americans) or HHV-6 seropositivity: Inverse correlation with HSV-2 and cytomegalovirus [79]
Measles, Varicella zoster or polio (seropositivity at birth) [14]	Childhood meningitis (0–4 years old) [80]	Borna disease virus seropositivty [81]
HSV-2 (antibodies assayed at the end of pregnancy) [82]		Coronavirus seropositivity [83]
Influenza B (seropositivity at birth) [84]		Elevated retrovirus HERV-W transcripts [85]
Toxoplasmosis (antibodies during pregnancy) [86]		Measles virus seropositivity [87]
		Hepatitis C [38]
		Toxoplasmosis [88]
		Correlation with the incidence of Lyme disease (Borrelia) [20]

TABLE 1: Some of the pathogens implicated in Schizophrenia, either in relation to maternal infection, or to infection in later life.

TABLE 2: Pathogens expressing proteins with homology to the autoantigens reported in schizophrenia. The size of the tags is proportional the number of pathogen's proteins that are homologous to the autoantigen. Note that the profile is different for each pathogen. The original BLAST files can be found at http://www.polygenicpathways.co.uk/blasts.htm.

Autoantigen reference	Pathogens
CHRNA7 Nicotinic cholinergic receptor [89]	Rhinovirus T. Gondii Varicella Borrelia Herpesvirus 2 _
CHRM1 Muscarinic cholinergic receptor [90]	<u>Herpesvirus 2 varicella</u> Borrelia Influenza T.Gondii Rhinovirus Rubella
DRD2 Dopamine receptor [82]	Varicella Herpesvirus 2 Influenza - Rubella Rhinovirus T. Gondii
GRIN1 NMDA receptor subunit [91]	<u>T. Gondii Herpesvirus 2 Influenza Varicella</u> Borrelia Rubella Rhinovirus
ELANE Leukocyte elastase [92]	<sub>ressat</sub> <u>Rhinovirus</u> <u>Borrelia</u> <u>Influenza</u> <u>Rubella</u> <u>Varicella</u>
OPRM1 Opioid receptor [93]	Varicella Rubella Borrelia Herpesvirus Influenza T. Gondii Rhinovirus
NGF Nerve growth factor [92]	Rhinovirus Borrelia Herpesvirus 2 Varicella Rubella T. Gondii Manaza
HTR1A Serotonin receptor [93]	<u>Rubella</u> Rhinovirus Varicella I.Gondii Influenza Herpesvirus 2 Borrelia
HSP60 Heat shock protein 60 [94]	Influenza _ Herpesvirus 2 T.Gondii Rubella Borrelia Varicella
HSPA12A Heat shock protein 70 [95]	Influenza T. Gondii Varicella Rubella Herpesvirus 2 Borrelia
HSP90 Heat shock protein 90 [95]	Rhinovirus Influenza Herpesvirus 2 Materia Varicella T. Gondii
PAM/MYC [94]	Herpesvirus 2 Rubella _ and I.Gondii Influenza Rhinovirus
S100B [96]	<u> </u>
STRN Striatin [96]	<u>Rubella</u> mara <u>T. Gondii</u> mara <u>Borrelia</u> <u>Herpesvirus 2</u> varicela

locations of 632 schizophrenia susceptibility genes (see Venn diagrams). Both negative and positive genetic association results have been reported for these many genes and it now seems plausible that, in some cases, this may be due to the presence or absence of active infection with these and other pathogens, and that DNA assays have been detecting pathogen as well as human DNA in the blood samples used for assay. There is evidently no way of discriminating viral or bacterial double-stranded DNA from human DNA.

This is not specific to schizophrenia, as the viruses implicated in Alzheimer's disease (HSV-1, HIV-1, HHV-6 and the cytomegalovirus) [125–127] are also homologous to proteins encoded by Alzheimer's disease susceptibility genes see http://www.polygenicpathways.co.uk/blasts.htm [128].

It seems that a viable interpretation, given the same phenomenon in these diseases, is that these genes are susceptibility genes precisely because they encode for proteins with homology to the viral risk factors. Infection and genetics TABLE 3: Human proteins with homology to proteins expressed by pathogens. The size of the tags reflects the number of pathogen's proteins that are homologous to the human protein: the filters used are described. The number of schizophrenia susceptibility genes within each of these datasets is shown in the left-hand column. Certain genes are classified according to family and are highlighted in red. Gene definitions and the original BLAST files can be found at http://www.polygenicpathways.co.uk/blasts.htm. Note that the homologues are often clustered in families (e.g., HTR1A, HTR2A, HTR3A, HTR3B, HTR3E, HTR5A, and HTR7).

Pathogen	Human protein Homologues
HERV-W	Dopamine related ALDH1A2 COMT DDC DRD2 DRD5
Filter	SLC18A2TH Serotonin related HTR1A HTR2A HTR3A HTR3B
"schizophrenia"	HTR3E HTR5A HTR7 Glutamate related DAO GRIA2 GRIA3
Number of SZ	GRIA4 GRID1 GRIN2A GRIN2D GRIN3A GRIK3 GRIK5 GRM2
genes = 103 ABCA13	GRM3 GRM7 Synaptic CABIN1 CPLX2 DTNBP1 DRP2 GRIP1
ACSM1 AHI1 ALDH1A2 ANK3 ARHGAP18 ARHGEF10 ARVCF	RPGRIP1L HOMER1 HOMER2 HOMER3 HIPK3 SPTAN1
ASTNI BRDI CACNG2 CHRFAM7A CIT CLINTI COMT CPLX2 CSF2RB CSPG5 CTNND2 DAO DGKI DISCI	SYN2 SYN3 RGS9 SNX6 GABA related GABRB1 GABRG1
DRD2 DRD3 DRD5 DTNBP1 EGR4 FEZ1 GABBR1 GFRA1 GFRA3 GNPAT GPC1 GPR85 GRIA3 GRIA4	GABBR1 Cholinergic CHRFAM7A DISC1 related ATF7IP
GRID1 GRIK3 GRIN2A GRIN2D GRM3 GRM4 GRM7 HDAC4 HIPK3 HIST1H2AG HIST1H2AH HLA-DRB1	DISC1 FEZ1 MLC1 PCM1 PCNT Myelin related MBP MOG
HOMER1 HOMER2 HTR1A HTR2A HTR5A HTR7 IMPA2 JARID2 KCNH2 LIF MAP6 MAPT MCHR1	MPZL1 NOTCH4 Translation initiation EIF3D EIF4A2
MCTP2 MDGAI MEGF10 MLC1 MOG MPZLI MUTED NALCN NDUFV2 NEUROG1 NOS1 NOTCH4	Neuregulin and growth EGR4 GFRA1 CSF2RB _ GFRA3 NRXN1
NRG1 NRG3 NRXN1 NTNG2 PCM1 PCNT PDE4B PDE7B PI4KA PIK3C2G PLXNA2 PPP3CC PRSS16	NLGN4X RET UTRN Oxidative stress NDUFV2 ATP2A2 CBR1
QK1 RAPGEF6 RELN RPGRIP1L RSRC1 SLC18A2 SP4 SPRY4	NDUFS1 Channels Calcium CACNA1B CACNG2 Sodium
ST8SIA2 SULT4A1 SYN2 SYN3 TAAR6 TH YWHAH ZNF184 ZNF804A	NALCN Potassium KCNH2 KCNH7 KCNH6 KCNN2 Immune/cytokine LIF
	TPI1 Structural MAP1A MAPT MYO1D MAP1B TBCB TUBB2A Signalling
	DGKI GNB1 IMMT PDE10A PIK3C2G PI4KA PAK3
	PPP3CC GNAI2 RAPGEF6 RASF7 STK24 DiGeorge DGCR14
	ABCA7 ADAM28 AHI1 ANK2 ANK3 ANKHD1 APOL4 APP

	AP3B2 ASTN1 BACE1 BRPF3 BRD1 CARTPT CEP63 CLDN10
	CLINT1 CTNND2 CYFIP1 CYP26B1 DCDC2 DKK1
	DLD $DNAJC6$ DOCK9 DZIP1 FNIP1 GDA $GNPAT$ GPC1
	GPR125 HDAC4 HDAC10 HNTPNU HIST1H2AH HSPD1
	JARID2 KIF13A LAMB2 LANCL2 LSAMP
	MCHR1MDGA1 MCTP2 MTIF3 MUTED NC5C NBEAL2
	PAFAH1B1 PAICS PCDHA1 PCDH11Y PLXNA2 PNP0 PRSS16
	PTGFRN RAI1 RSRC1 SEL1L3SEMA3A _ SIGMAR1 SIRPB1
	SLC25A3(PO4) SLCO3A1(anion) SP4 STAB1 ST6GAL2 STRN SULT4A1
	SPRY4 MOON ST8SIAZ _ YWHAH ZBT20 ZNF184 ZNF804A OMG VGF MEGF10 ERVWE1 MOON
	SPRY4 NOON STRSIA2 WWHAH ZBT20 ZNF184 ZNF804A OMG VGF _ MEGF10 ERVWE1 POER HIST1H2AG TMEM200C OF NERA ATAY LERUT _ OLFM1 CELSEN _ ARACAN _ ARCF _ COOCHI EPNE _ PROOR ADMITSH _ ARHGAP18 NRCAM NOVA1 CON DIEG GPR85 _ G GRIK1 KONG _ CNTN4 _ GRE2 OPINI RELN HNRNPA2B1 INA ALDH1A3 _ VAR2 _ G
HSV-2	SPRY4 NOVE ST8SIA2 _ YWHAH ZBT20 ZNF184 ZNF804A OMG VGF MEGF10 ERVWE1 NEED HIST1H2AG TMEM200C OF NEW _ ATCAY LIBERT _ OLFM1 CELSRI _ MOLAN _ ANGE _ COCCHI EPNE _ FROM AMMENT _ ARHGAP18 NRCAM NOVA1 ONL DED GPR85 _ GRIK1 NOOS _ CNTN4 _ DRE _ OPMEN RELN HNRNPA2B1 INA ALDH1A3 _ VAR2
HSV-2 Filter	SPRY4 NOOR STRSM2 WHAT ZBT20 ZNF184 ZNF804A OMG VGF _ MEGF10 ERVWE1 NOOD HIST1H2AG TMEM200C OF NOOD ALLOW LOWFF OLFM1 CENT _ MEGA1 & MOVE _ COCCUL BYNG _ MOVE _ MEGF10 ERVWE1 NOOD NCAM NOVA1 ONE OF GPR85 _ GRIK1 NOOS CNTN4 _ MOVE ON MIR RELN HNRNPA2B1 INA ALDH1A3 _ WAR2 _ T KIF21Awa LWCZ _ CHT65 Dopamine related COMT DDC DRD1 DRD3 DRD4 DRD5 TH PEMT Serotonin related ITT2 HTT66 HTT77 HTT54 Glutamate related DAO DAOA GRIA4
HSV-2 Filter "schizophrenia"	SPRY4 1000 ST851A2 WHAH ZBT20 ZNF184 ZNF804A ONG VGF _ MEGF10 ERVWE1 1000 HIST1H2AG TMEM200C OF 1000 A COLOR OF OLFM1 COLOR OF A COL
HSV-2 Filter "schizophrenia" Number of SZ	SPRY4 HOW STREAD _ YWHAH ZBT20 ZNF184 ZNF804A OMG VGF _ MEGF10 ERVWE1 HORS HIST1H2AG TMEM200C OF HOR , ATCH HORT _ OLFM1 CHERT _ OLFM1 CHERT _ OKO OF ON RELA HINRNPA2B _ MARGE _ ARHGAP18 NCAM NOVA1 ON ORD GPR85 _ GRIK1 NOOS _ CNTN4 _ OKO OF ON RELA HINRNPA2B I INA ALDH1A3 _ VAR2 _ C HIST1H2AM UMC _ ORD Dopamine related comt DDC DRD1 DRD3 DRD4 DRD5 TH PEMT Serotonin related HIRRA HTR6 HTR7 HIRSA Glutamate related DAO DAOA GRIA4 SUCIAHGUUT GRID1 GRIN2D GRIN3A GRM3 GRM4 PROH
HSV-2 Filter "schizophrenia" Number of SZ genes = 166	SPRY4 noor STBSIA2 YWHAH ZBT20   Intra ZNF804A ong vor megrio ervwei _ ore and ore and ore and ore and and ore and and ore and and and ore and
HSV-2 Filter "schizophrenia" Number of SZ genes = 166 ABCA13 ACSL6 ADAM12 ADCYAP1 ADSS AH11	SPRY4 wow STBSIA2 _ WWHAH ZBT20 _ ZNF184 ZNF804A _ OMG VGF MEGF10 ERWWE1 wow   HIST1HZAG TMEM200C _ OF WAQ _ ACCAR _ UNUTI _ OLEFM1 _ OLEM MOLT _ MOLT _ OCCAR _ MOLT _ MOLT _ MOLT _ ARHGAP18 MICAM NOVA1 _ ONE OF GPR85 GRIK1 _ ONE _ CNTN4 _ ONE _ ONE RELN HINRNPARB1 _ INA ALDHIA3 _ UNA2

BTN3A1 CACNA1C CCE CCKAR CDKN2A CHN2 CHRM5 CHRNA7 CIT C CSF2RA CSF2RB CSPG5 CYTB DAO DAOA DBH DGCR2 DGCR6 DISC1 D DLG4 DLX1 DRD2 DRD2 DRD4 DRD5 DTNBP1 EC ENO2 FABP7 FEZ1 FGF FXR1 FZD3 GABBR1 GA GABRP GCLM GFRA1 G GNB1L GNL3 GRIA3 GF GRID1 GRIN2A GRIN2D GRM3 GRM4 GRM5 GR HIST1H2AG HIST1H2AF HOMER1 HOMER2 HSP HTR1A HTR2A HTR2C H HTR6 HTR7 IL1A IMPA JARID2 KCNH2 KCNN3 MAG MAP4 MAP6 MAP MC5R MCTP2 MDGA1 MEGF10 MICB MLC1 M MUTED MYH9 MYT1L NDE1 NDEL1 NDUFV2 NEUROG1 NOS1AP NO NPAS3 NPTN NRG1 NRG NRXN1 NTF3 NUMBL C PADI2 PCM1 PDE4B PD PDLIM5 PEMT PLA2G40 PLAA PLXNA2 PNPO POM121L2 PPP1R1B PPI PRODH PRSS16 PTBP2 RAPGEF6 RELN RGS4 RPGRIP1L RTN4R SHIS. SIRT5 Influenza Filter "schizophrenia Number of SZ

DC60	GFRA1 NRG1 NRG3 NRXN1 NTF3 NPTN FGF14csf2rb NTNG
OMT	Myelin related MAG MPZLI NOTCH4 RTN4R Oxidative stress GCLM ATP6
0KK4	ND2 Channels Calcium CACNA1C Potassium KCNN3 KCNH2
3 GR2	Immune/cytokine IL1A MICB TRAFSIPI Structural MAP4 MAP6 MYH9
14 ABRA1 FNAL	MYT1L Signalling ARHGAP18 ARHFGEF10 GNAL
UA4	MAPK14 IMPA2 PDE4D PLAA PLA2G4C PPP1R1B RAPGEF6
M7 H A12A	UHMK1 DiGeorge DGCR2 DGCR6
HTR5A 2 KMO	ABCA13 ADAM12 ADSS AHI1 ARVCF BRD1 BT2NA2 CHN2
K14 PZL1	CSPG5 DLX1 ENO2 FBCL21FCYD6 GRP78 JARID2
ND2	
ГСН4	KPNA3NPAS3 NUMBL OPRM1 PCDH8 PLXNA2 PNPO POM121L2
G3 DPRM1 E4D	PRSS16 (INCOMPARATION - 1994) TAAR6 VRK2 - TXNDC5 1 GNL3 HTR2C NDUFV2 FXR1 - MC5R dBH (DK62A
0	NEUROGI PADI2 CCKAR MLC1 FABET ASTRE CHRNA7 PPEKE TUBA8 PTBP2 GRM7 TSPAN8 1798 HSPA12A SYN3 DKK4 C CYB
P3CC	CCDC60 754.05 HIST1H2AH SLC06A1(ANION) MEGF10 8768716
4.5	ZNF74 smarcci btn3a1 ASTN1 TSPO slc17A3 deci singer (spece scenerary) ufd1l $ZNF804A$ chrms
A3	YWHAZ MUTED SIRT5 HRH2 - SPARCL1 HTTEA CAUNT HIST1H2BJ
	Dopamine related COMT DRD1 DRD3 DRD5 Serotonin related HTR1A
	HTR2C HTR5A HTR6 HTR7 TPH1 Glutamate related DAO DAOA GRIA3
ı"	GRIA4 GRINZA GRIN2D GRIN3A GRM5 KMO SRR SLC6A5(GLY) SLC17A7(PO4)
	SLC17A1 <sub>SLC17A3</sub> GABA –related GABRA1 <sub>GABBR1GABRP</sub> Synaptic

genes = 167 ABCA13 DTNBP1 DLG4 HOMER1 RPGRIP1L SYN2 SYN3

ACSL6 ADAM12 ADCYAP1 ADSS AHI1 ALDH1A2 ALDH3B1 ALK ANK3 ARHGAP18 ARVCF ASTN1 ASTN2 ATP6 BRD1 BTN3A1 CCDC60 CCKAR CDKN2A CHN2 CHRM5 CHRNA7 CIT COMT CSF2RA CSF2RB CSPG5 CYTB DAO DAOA DBH DGCR2 DGCR6 DISC1 DKK4 DLG4 DLX1 DRD2 DRD3 DRD4 DRD5 DTNBP1 EGR2 ENO2 FABP7 FEZ1 FGF14 FXR1 FZD3 GABBR1 GABRA1 GABRP GCLM GFRA1 GNAL GNB1L GNL3 GRIA3 GRIA4 GRID1 GRIN2A GRIN2D GRM3 GRM4 GRM5 GRM7 HIST1H2AG HIST1H2AH HIST1H2BJ HOMER1 HOMER2 HSPA12A HTR1A HTR2A HTR2C HTR5A HTR6 HTR7 IL1A IMPA2 JARID2 KCNH2 KCNN3 KMO MAG MAP4 MAP6 MAPK14 MC5R MCTP2 MDGA1 MEGF10 MICB MLC1 MPZL1 MUTED MYH9 MYT1L ND2 NDE1 NDEL1 NDUFV2 NEUROG1 NOS1AP NOTCH4 NPAS3 NPTN NRG1 NRG3 NRXN1 NTF3 NTNG1 NUMBL OPRM1 PADI2 PCM1 PDE4B PDE4D PDLIM5 PEMT PLA2G4C PLAA PLXNA2 PNPO POM121L2 PPP1R1B PPP3CC PRODH PRSS16 PTBP2 RAPGEF6 RELN RGS4 RPGRIP1L RTN4R SHISA5 SIRT5 SLC17A1 SLC17A3 SLC17A7 SLC1A3 SLC1A4 SLC6A5 SMARCC1 SNAP29 SP4 SPARCL1 SRR SYN2 SYN3 SYNGR1 SYT11 TH TPH1 TRAF3IP1 TSNAX TSPAN8 TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A

SYNGAP1syngr1 SYT11 DISC1 related DISC1 CIT NDE1 PCM1 PDE4B MLC1 TSNAX Neuregulin and growth ALK NRG1 CSF2RB GFRA1 NEUROG1 CSF2RA EGR2 NTNG1 NTF3 Myelin related MAG NOTCH4 MPZL1 RTN4R Cholinergic CHRM5 CHRM5 Oxidative stress CYTB GCLM Channels Calcium CACNAIC Potassium KCNN3 KCNH2 Immune/Cytokine IL1A TRAF3IP1 Structural MAP6 MYTH Signalling ARHFGEF10 GNL3 MAPK14PLA2G4C PPP3CC UHMK1 DiGeorge DGCR2 DKK4 \_ YWHAE ZDHHC8 SPARCL1 MC5R FORMER CABINI FROM ASTN1 FEED = IN FISALEA ALDH3B1 ARVCF \_\_ UFDIL \_ PPPIRIB \_\_ SIRT5 DEC SLC06A1(ANION) \_\_ MCTP2 ATE ATE MITTED FBCL21 JARID2 NARY2 PCDH8 \_ MEGFIO \_ SHISAS NDELI PLXNA2 TSV CSPG5 \_ TXNDC5 ALEMIA CHN2 \_\_ NPAS3 (TOTA \_ DLX1 (MAN FRA \_ FXR1 SP4 (GRM5 (GRM5 ) ( NUMBL \_ \_ \_ SMARCC1 ZNF74 \_ BRD1 \_ KPNA3 BTN3A1 GRV HIST1H2AG TOTA HIST1H2AH FABP7 \_\_\_\_ MOTO POLO HRH2 ACSL6 ADAM12 HIST1H2BJ MICB ADSS - ARHGAP18 - ABCA13 YWHAZ TAAR6 ADCYAP1 GRP78 PADI2

Rhinovirus	Dopamine related DBH COMT DRD1 DRD2 DRD3 DRD3 TH SLC18A2
Filter	Serotonin related HTR1A HTR2C HTR6 HTR7 TDO2 Glutamate related
"schizophrenia"	DAO DAOA GRIN1 GRIN2A GRIK3 GRM4 GRM5 GRM7 SLC1A3(GLUT)
Number of SZ	SLC1A4(GLUT) SLC17A3 SLC17A7(PO4/glut) GABA –
genes = 176	related GABRA1GABRB2 GABBR1 Synaptic HOMER1 HOMER2
ABCA13 ACSL6 ADAM12 ADCYAP1 ADSS AHI1	RPGRIP1L NOS1 NOS1AP SYNGRI SYT11RGS4 DISC1 related _ DISC2 CIT
ALDH3B1 ANK3 ANKK1 APOL2 ARHGAP18 ADUCEE10 ADVCE ASTN1	FEZ1 NDEL1 PDE4B PCM1 MLC1 Neuregulin and growth
ASTN2 BRD1 BTN3A1 C10orf120 CACNA1C CCKAR	NRG1NRG3 NPTN CSF2RA EGR4 FGF14 GFRA3 NRXN1
CHI3L1 CHL1 CHRFAM7A CHRM5 CHRNA7 CIT CLDN5	NTF3 Myelin related MAG NOTCH4 Cholinergic CHRNA7 CHRFAM7A
CLINTI CLOCK CNP COMT CSF2RA CSF2RB CTLA4 CTNND2 DAO DAOA DBH DGCR2 DGCR5 DGCR6 DGKI	Oxidative stress $ALDH3B1$ ND2 ND4 NDUFV2 Channels Calcium CACNA1B
DISC1 DKK4 DLX1 DPYSL2 DRD2 DRD3 DRD4 DRD5	CACNAIC Potassium KCNH2 Immune /Cytokine LIF Structural MAP4
EGR4 ENO2 ESR1 FABP7 FEZ1 FGF14 FXR1 FZD3	MAP6 signalling ARHGEF10 GNL3 ARHGAP18 IMPA2 PDE7B PK3C2G
GABBRI GABRAI GABRB2 GABRP GFRA3 GNB1L GNL3 GNPAT GRIA1 GRIA3 GRIA4	PPP1R1B RAPGEF6 UHMK1 PLAA PNPLA8 PLA2G4C
GRIK3 GRIN1 GRIN2A GRIN2D GRM3 GRM4 GRM5	VRK2 DiGeorge DGCR2 DGCR5 DGCR6 and a ABCA13 ITIH3 JARID2
GRM7 HIST1H2BJ HLA-DRB1 HOMER1 HOMER2 HSPA12A	FXR1 === PADI2 GNB1L ORC3L SIL1 == ZNF804A RELN = DKK4 adcyapi POM121L2
HTRTA HTREA HTREE HTR6 HTR7 IL1A IMPA2 INTS6 ITIH3 JARID2 KCNH2 KPNA3	$ADSS_{-} HSPA12A_{-} PPP3cc_{-} MYH9 INTS6_{SP4} ESR1_{PB1} TMEM108_{-} PFN4$
LIF MAG MAP4 MAP6 MAPK14 MC5R MCHR1	CHL1 AK1 and and CLOCK PNPO SPARCL1 and OPCML ZDHC8 NUMBL and SYN3
MCHR2 MCTP2 MDGA1 MICB MLC1 MUTED MYH9 MYT1L	PTBP2 - TSPAN8 - ST8S1A2 ASTN1 HIST1H2BJ NPAS3 - MCTP2
ND2 ND4 NDEL1 NDUFV2 NOS1 NOS1 NOS1AP NOTCH4	APOL2 TXNDC5 SGCR6 CTIND2 C100rf120 ANK3 HRH2 CLINTI PRSS16
NPAS3 NP1N NRG1 NRG3 NRXN1 NTF3 NUMBL OPCML OPRM1 PADI2 PCM1 PCQAP	HTR2A MRCL3 MRLC2 DLX1 _ GLRA2 MICB PCQAP == == CLDN5
PDE4B PDE7B PDLIM5 PFN4 PLA2G4C PLAA PNPLA8	CHI3L1

PNPO POM121L2 PPP1R1B PPP3CC PRODH PRSS16 PTBP2 RAPGEF6 RELN RGS4 RPGRIP1L SHISA5 SIL1 SIRT5 SLC17A3 SLC17A7 SLC18A2 SLC1A3 SLC1A4 SLC1A6 SLC6A5 SMARCC1 SP4 SPARCL1 SULT4A1 SYN2 SYN3 SYNGR1 SYT11 TAAR6 TD02 TH TMEM108 TSPAN8 TXNDC5 UHMK1 VRK2 ZBED4 ZNF184 ZNF804A	BTN3A1 ANKKI MACF1 SHISA5 MDGA1 produt ZBED4 CTLA4 znf184
Rubella	Dopamine related COMT DBH DRD3 DRD4 DRD5 PEMT
Filter "schizophrenia"	Serotonin related HTR1A HTR2A HTR2C HTR5A HTR7 TPH1 Glutamate related DAO DAOA GRIA3 GRIA4 GRID1 GRIN1 GRIN2A
Number of SZ	GRM3 GRM4 GRM5 GRM7 KMO SLC1A4(GLUT) SLC17A3 GABA –
genes $= 179$	
ADAM12 ADCYAP1 ADSS Ahii Aldh1A2 Aldh3B1	related GABRAI GABRAI GABRB2 GABRP Cholinergic CHRM5 Synaptic
ANK3 ARHGAP18 ARHGEF10 ARVCF ASTN1 ASTN2 ATP6	CABIN1 DTNBP1 NOSIAP RPGRIP1L SNAP29 SYN2 SYN3 RGS4
BDNF BRD1 BTN2A2 CACNA1C CCDC60 CCKAR	SYNGAP1syngri sytti DISC1 related DISC1 PCM1 PDE4B NDEL1
CHRNA7 CLOCK COMT CSF2RA CSF2RB CSPG5	FEZ1 FZD3 Neuregulin and growth NRG1 ERBB1 ERBB3
CYTB DAO DAOA DBH DGCR2 DGCR6 DISC1 DKK4	ERBB4 GFRA1 NTF3 NTNG1 FGF14 Myelin related NOTCH4 RTN4R
DRD2 DRD3 DRD4 DRD5 DTNBP1 EGR2 ENO2 ERBB2	QK1 Oxidative stress CYTB GCLM ND2 NDUFV2 Channels Calcium
ERBB3 ERBB4 FABP7 FBXL21 FEZ1 FGF14 FXR1 FXYD6 FZD3 GABBR1 GABRA1	Potassium KCNN3 KPNA3Immune/cytokine ILIA TRAF3IP1 Structural
GABRB2 GABRP GCLM GFRA1 GNAL GNB1L GNL3	MAP6 Mytil Signalling ADCYAP1AK1 ARHGEF10 CDKN2A GNAL
GPC1 GRIA3 GRIA4 GRID1 GRIK3 GRIN1 GRIN2A GRIN2D GRM3 GRM4 GRM5	GNB1L GNL3 mapk14 IMPA2 plaa pla3G4C repgef6 PDE4D PPP1R1B
GRM7 HIST1H2AG HIST1H2AH HIST1H2BJ HLA-	DiGeorge DGCR2 DGCR6 IBBEE MEGF10 NDEI IBSTIELAARDA TXNDC5 ASCL6 SHISAS ZNF804A
DRB1 HOMER1 HOMER2 HTR1A HTR2A HTR2C HTR5A	JARID2 FXYD6 BTN2A2

JARID2 KCNH2 KCNN3 KMO	TSPO CCDC60 YWHAE KCNH2 DWD MLC1 CSPG5 PLXNA2 – MCTP2 GPC1 VRK2
KPNA3 KREMEN1 MAG MAP4 MAP6 MAPK14 MC5R MCTP2 MDGA1 MEGF10	CCKAR ASTN2 - Sciance of Price FXR1 ASTN1 (1990) MCB (1997) SHE NECT (1992) SMARCCI - BOY
MICB MLC1 MPZL1 MUTED MYH9 MYT1L ND2 NDE1	TH HRH2 ADAM12 pcdH8 ww ZNF74 YWHAZ - atta - CHN2 and TSPAN8
NDEL1 NDUFV2 NEUROG1 NOS1 NOS1AP NOTCH4 NPAS3 NPTN NRG1 NRG3	PER1 TUBA8 ARHGAP18 NEW MUTE ENO2
NRGN NRXNI NTF3 NTNG1 NTNG2 NUMBL OPRM1	$TAAR6 \_\ HIST1H2BJ KREMEN1 SLCO6A1 HIST1H2AH CLOCK PSSIS PADI2$
PADI2 PCM1 PDE4B PDE4D PDLIM5 PEMT PLAA PLXNA2 POM121L2 PPP1R1B PPP3CC	$PDLIM5 \ MDGA1 \ \ \text{cance ADSS } \text{ZdhHcs} \ \ - \ AHI1 \ \ - \ \text{OPRM1 } \text{ ufdil } \text{NPAS3} \ \ - \ \text{NPAS3} \ \ - \ \text{OPRM1} \ \ \text{NPAS3} \ \ - \ \text{NPAS3} \ \ \ - \ \text{NPAS3} \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \$
PROTHER STATES S	MC5R POMI2IL2 DKK4
TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A	
TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella	Dopamine related COMT DDC DRD2 DRD5 Serotonin related
TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C HTR7 HTR3A HTR3D TPH2 TPH1 Glutamate related GRIN1
TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter "schizophrenia"	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C HTR7 HTR3A HTR3D TPH2 TPH1 Glutamate related GRIN1 GRNA GRIN2A GRIK1 GRIK5 GRM2 GRM7 SLC6A5(Gly) SLC17A6
TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter "schizophrenia" Number of SZ	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C <sub>HTR7</sub> HTR3A HTR3D TPH2 <sub>TPH1</sub> Glutamate related GRIN1 GRINA GRIN2A GRIK1 GRIK5 GRM2 GRM7 SLC6A5(Gly) SLC17A6 GABA –related GABRA5 GABBR1 Synaptic <sub>DLGAP2</sub>
TUBA8 TXNDC5 UFDIL UHMKI VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter "schizophrenia" Number of SZ genes = 75	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C htr7 HTR3A HTR3D TPH2 tph1 Glutamate related GRIN1 GRINA GRIN2A GRIK1 GRIK5 GRM2 GRM7 SLC6A5(Gly) SLC17A6 GABA -related GABRA5 GABBR1 Synaptic dlgap2 RPGRIP1LNOS1AP RGS9 SPTBN4 HIPK3 stx7 syngr1 DISC1
TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter "schizophrenia" Number of SZ genes = 75 ABCA13 ACSL6 ALDH1A2 ANK3 ANKK1 ARHGAP18	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C http://http://http://gitage.com//gitage.co
TUBA8 TXNDC5 UFDIL UHMKI VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter "schizophrenia" Number of SZ genes = 75 ABCA13 ACSL6 ALDH1A2 ANK3 ANKKI ARHGAP18 ASTNI BRDI BTN3A2 CACNAIC CHRFAM7A	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C htr7 htr3A htr3D tPh2 tphi Glutamate related GRIN1 GRIN2A GRIK1 GRIK5 GRM2 GRM7 SLC6A5(Gly) SLC17A6 GABArelated GABRA5 GABBR1 Synaptic dlGAP2 RPGRIP1LNOS1AP RGS9 SPTBN4 HIPK3 stx7 syngr1 DISC1 related DISC1 CIT FEZ1 PCNT PCM1 PDE4B Neuregulin and growth NRG1 NRG3 NTF3 NTNG2 CSF2RA NRXN1 EGRP2 NRXN3 NLGN4X
TUBA8 TXNDC5 UFDIL UHMKI VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A Varicella Filter "schizophrenia" Number of SZ genes = 75 ABCA13 ACSL6 ALDH1A2 ANK3 ANKKI ARHGAP18 ASTNI BRDI BTN3A2 CACNAIC CHRFAM7A CHRM5 CHRNA7 CNTNAP2 COMT CSPG5 CTNND2 CTXN3 DAO DBH DGCR2	Dopamine related COMT DDC DRD2 DRD5 Serotonin related HTR2C htr7 HTR3A HTR3D TPH2 tph1 Glutamate related GRIN1 GRIN2A GRIN2A GRIK1 grik5 grm2 grm7 SLC6A5(Gly) SLC17A6 GABA - related GABRA5 gabbr1 Synaptic dlgap2 RPGRIP1LNOS1AP RGS9 SPTBN4 HIPK3 stx7 syngr1 DISC1 related DISC1 CIT FEZ1 PCNT PCM1 PDE4B Neuregulin and growth NRG1 NRG3 NTF3 NTNG2 CSF2RA NRXXN1 EGR2 NRXN3 NLGN4X UTRN VGF Myelin related NOTCH1 OMG Cholinergic CHRM5

HTR6 HTR7 IL1A IMPA2

Calcium CACNA1C Potassium KCNH7 KCNQ5 KCNH2

GNPAT GRIK3 GRIN1

GRIN2A GRIN2D GRM5 GRM7 HDAC3 HDAC4 HIPK3 HLA-DRB1 HTR2C HTR7 KCNH2 KREMEN1 MAP6 MAPK14 MCHR1 MCTP2 MICB MYT1L NALCN NEUROG1 NOS1 NOS1AP NPAS3 PDE7B PI4KA PLAA PLXNA2 PTBP2 RAPGEF6 RPGRIP1L SEMA3D SLC17A6 SLC6A5 SLCO6A1 SP4 SPARCL1 SULT4A1 TAAR6 TPH1 TRMT2A UHMK1 VRK2 ZNF804A

T.Gondii

Filter

Immune/cytokine IL1RAPL1 LIFR Structural NEFM MAP1A MAP1B MAP6 MYHP MYTIL Signalling CDC42 GNBI DAG1 DGKI UHMK1 GUCY1A2 PI4KA PLAA PRKCG PDE10A RAPGEF6 GSK3A RASSF7 VRK2 DiGeorge DGCR2 DGCR14 MACF1 NR3C1 ... ESR1 CTNND2 APP ... SE2 \_ GPM6A INA STAB1 PCDHB11 FIGN ... CPNE2 AP3B2 SULT4A1 CELF4 MICB ..... MCHR1 CNTNAP2 AP3D1 MCF2 YWAH ANKHD1 PCDH11Y PSMD10 GABRD .... SLC6A2(NE) w ORC3L PTBP2 w ZNF804A CSPG5 NUDT6 - HDAC4 - NCOA7 CHRNA7 PCDH8 AP3M1 w TRMT2A TRIM3 ZBER4 RNH1 ANKK1 MBLN2 SEMA3B SLC26A6 CLDN10 ... NPAS1 BRD1 CTXN3 HRH2 ... BACE1 ASTN1 ARHGAP18 (GLY) DOCK9 APOL3 SPARCL1 KIF13A MECP2 SIRPB1 MAGEL2 TAAR6 EPHA6 ATP6V1A NRCAM ... USP4 LMNB2 CAMKV MCTP2 PCDHA1 UGGT2 PADI2 FNIP1 KIF21A EPB41LI ARVILZ - O ACLY NS - AVK3 - KIAA0513 ABCA13 SORBS1 .... PTGFRN SEMA3D RET RAIL SLCO6A1 SLC25A3 SP4 LANCL2 MARCH SRSF6 CACNAIB SETD2 HDAC3 GABRAI ATP5AI Dopamine related COMT DBH DRDS DRD3 DRD4 PEMT SLC18A2 TH Serotonin related HTR1A HTR2A HTR2C HTR5A TPH1 Glutamate "schizophrenia"

related DAOA GRIA4 GRIN1 GRIN2A GRIN2C GRIN2D GRIK3

SZ genes = 182ADAM12 ADCYAP1 ADSS AHI1 ALDH1A2 ALDH3B1 ANK3 ARHGAP18 ARHGEF10 ARVCF ASTN1 ASTN2 ATP6 BRD1 BTN2A2 BTN3A1 CACNA1C CCDC60 CCKAR CDKN2A CHN2 CHRM5 CHRNA7 CLOCK COMT CSF2RA CSF2RB CSPG5 CYTB DAO DAOA DBH DGCR2 DGCR6 DISC1 DKK4 DLX1 DRD2 DRD3 DRD4 DRD5 DTNBP1 EGR2 ENO2 ERBB2 ERBB3 ERBB4 FABP7 FBXL21 FEZ1 FGF14 FXR1 FXYD6 FZD3 GABBR1 GABRA1 GABRB2 GABRP GCLM GFRA1 GNAL GNB1L GNL3 GRIA3 GRIA4 GRID1 GRIK3 GRIN1 GRIN2A GRIN2B GRIN2D GRM3 GRM4 GRM5 GRM7 HIST1H2AG HIST1H2AH HIST1H2BJ HLA-DRB1 HOMER1 HOMER2 HSPA12A HTR1A HTR2A HTR2C HTR5A HTR6 HTR7 IL1A IMPA2 JARID2 KCNH2 KCNN3 KMO KPNA3 KREMEN1 MAG MAP4 MAP6 MAPK14 MC5R MCTP2 MDGA1 MEGF10 MICB MLC1 MPZL1 MUTED MYH9 MYT1L ND2 NDE1 NDEL1 NDUFV2 NEUROG1 NOS1 NOS1 NOS1AP NOTCH4 NPAS3 NPTN NRG1 NRG3 NRXN1 NTF3 NTNG1 NTNG2 NUMBL OPRM1 PADI2 PCM1 PDE4B PDE4D PDLIM5 PEMT PLA2G4C PLAA PLXNA2 PNPO POM121L2 PPP1R1B PPP3CC PRODH PRSS16 PTBP2 QK1 RAPGEF6 RELN RGS4 SHISA5 SIRT5 SLC17A1 SLC17A3 SLC17A7 SLC18A2 SLC1A3 SLC6A5 SLC06A1

GRM3 GRM4 PRODH KMO SLC6A5(GLY) SLC17AI GABA -related GABRP Synaptic DTNBPI CABINI HOMER1 NOS1NOS1AP SYNGAP1 SYN2 SYN3 SNAP29 SYNGR1 SYT11RGS4 DISC1 related DISC1 FEZ1 FZD3 NDE1PDE4B TSNAX MLC1 Neuregulin and growth ERBB2 ERBB3 EGR2 FGF14 GFRAI NEUROG1 NTF3 NRXN1 NTNG2Myelin related MAG MPZL1 QK1 Cholinergic CHRNA7 Oxidative stress ATP6 GCLM ND2 NDUFV2 Immune/cytokine IL1A MICB TRAF3IP1 Signalling ADCYAP1 ARHGEF10 CDKN2A GNAL GNL3 ARHGAP18 MAPK14 GNB1L PLA2G4C PLAA PPP3CC PPP1R1B RAPGEF6 UHMK1 VRK2 Structural MAP4 MYTILMYH9 TUBA8 DiGeorge DGCR2 DGCR6 Circadian CLOCK PER1 AHI1 AK1 ALDH3B1 ANK3 ARVCF ASTN2 BRD1 BTN2A2 BTN3A1 CCDC60 CCKAR CHN2 CSPG5 DKK4 ENO2 FABP7 FXYD6 GRP78 HIST1H2AH HIST1H2AG HIST1H2BJ KPNA3 KREMEN1 PRAME SICIASIGLUTD MDGA1 MC5R MCTP2 DRD2 CHRM5 ..... NPAS3 PNPO PTBP2 RELN SHISA5 SIRT5 TAAR6 TSPAN8 TXNDC5 YWHAE YWHAZ ZDHHC8 ZNF74 CSF2RB DEDI SRR GEDI GARAN SA - - SMACLI - ZNF64A - - - TSPO JARID2 - HTTM - - -

HOMER2 --- ALDH1A2 -- PDE4D MEGFIG SLCOGA1 MOTOR DLX1 ASCL6 PRSS16 UFD1L GRAN OPRM1 --- HRH2 KONG CYTR CR2RA --- NRGI PLXNA NREI FXR1 --- IMPA GRAR SMARCC1 NPTN NRG3 GRARE NINGI PCDHR SLC17A3 GRAF SLC17A7(PO4) --- --- DAG SMARCC1 SNAP29 SP4 3 SPARCL1 SRR SYN2 SYN H SYNGRI SYT11 TAAR6 T TPH1 TRAF3IP1 TSNAX TSPAN8 TUBA8 TXNDC5 UFD1L UHMK1 VRK2 YWHAE ZDHHC8 ZNF74 ZNF804A

#### Borrelia

#### Burgdorferri

#### Number of SZ

#### genes = 167

ABCA13 ACSL6 ADCYAP1 ADSS AGBL1 AHI1 AKT1 ALDH1A2 ALDH3B1 ANK3 ANKK1 APOL2 ARHGAP18 ARHGEF10 ASTN1 ASTN2 BRD1 CACNG2 CCDC60 CCKAR CHL1 CHN2 CHRM5 CHRNA7 CIT CLINT1 CLU CNP CNTNAP2 CPLX2 CSF2RA CSPG5 CTNND2 DAOA DBH DGCR2 DGKI DISC1 DLG1 DPYSL2 DRD2 DRD3 DRD4 DRD5 DYM EFNB2 EGR2 EGR4 ENO2 ERBB4 FBXL21 FEZ1 FGF14 FOXP2 FXR1 FZD3 GABBR1 GAD1 GCLC GFRA2 GNAL GNPAT GPR85 GRIA1 GRIA3 GRIA4 GRID1 GRIK3 GRIN1 GRIN2D GRM3 GRM4 GRM5 GRM7 GULP1 HLA-DRB1 HOMER2 HSPA12A HTR1A HTR2A HTR6 HTR7 IFNG IMPA2 IPO5 JARID2 KCNH2 KCNN3 KREMEN1 LRRTM1 MAOB MAP4 MAP6 MCHR2 MCTP2 MEGF10 MLC1 MYL12B MYT1L NALCN ND2 NDE1 NDEL1 NDUFV2 NOS1 NPTN NRG1 NRG3 NRXN1 NTNG2 NUMBL PADI2 PCM1

Serotonin HTR1A HTR3C HTR6 HTR3D HTR3E SLC6A5 TDO2 Glutamate DAOA GLUL GRIA3 GRIA4 GRIN1 GRIK1 GRIK3 GRIK5 GRIN2A GRM2 GRM3 GRM4 PRODH SLC1A3 SLC1A5 SLC1A4

GABA GABBR1 GABRB2 GAD1 GAD2 Cholinergic

Dopamine ALDH1A2 DRD2 DRD3 DRD4 DRD5 MAOB

CHRM5 synaptic BLOC1S1 CABIN1 CNTN4 CPLX2 DLG1 GPRASP2GRIP1 HOMER2 HOMER3 RGS4 RGS9 SHANK3 SNAP29 SPTAN1 SYNGAP1 STXBP1 SYNGR1 SYN2 SYN3 SYT5 DISC1 related DISC1 FEZ1 IMMT MLC1 NDE1 NDEL1 PCM1 PCNT PDE4B TSNAX Neuregulin/ growth EGR4 EGR2ERBB4 FGF14 NRG1 NRG3 Immune IL1RAPL1 LIFR TPI1 Signalling CSNK1D GSK3A IMPA2 PIP4K2A PLA2G4C PLA2G4D SH3GL2 STK24 Channels CACNA1B CACNG2 KCNH5 KCNH6 . KCNN2 PCNT PDE4B PDE7B PGBD1 PICK1 PIK3C2G PIP4K2A PLA2G4C PLAA PLXNA2 PNPLA8 POM121L2 PPP1R1B PPP3CC PRKAG2 PRODH PRSS16 QK1 RAPGEF6 RELN RGS4 RPGRIP1L RSRC1 RTN4R SEMA3D SHANK3 SLC17A1 SLC17A6 SLC17A7 SLC1A2 SLC1A3 SLC1A6 SLC6A5 SLC06A1 SMARCA2 SMARCC1 SNAP29 SPARCL1 SYN2 SYN3 SYNGR1 TAAR6 TCF4 TDO2 TRAF3IP1 TRMT2A TSNAX UFD1L USP46 VRK2 ZBED4 ZDHHC8 ZNF804A

# GFAP MAP1B MAP2 MAP4 MYT1L NEFL TUBA1A TUBA1B Oxidative stress ATP5A1 ATP6V1A COX1 CRYM GCLC ND5 NDUFS3 NDUFV2 PRDX1 - AKT1

AADAT ABCA7 ADAM22 ADAMTS4 ADIPOQ AGBL1 AKR1D1 AHI1 ALOX12 AP3D1 AP3M1 APOL2 APOL3 APOL4 APOL6 ARHGDIA ARID4B ASTN1 ATCAY ATP2B2 BAP1 BIVM BRD1 BTN3A1 CAD CAP1 CCDC60 CCDC141 CCKAR CDC42EP3 CEP63 CHL1 CHN2 CLU CNTNAP2 CPS1 CTNND2 DBP DDAH1 DZIP1 DNAJC6 DOCK9 DPYSL2 DRP2 EFNB2 EIF4A2 ENO2 ENTPD4 EPHA6 ERLIN1 ERMN FABP3 FARSA FBXL21 FIGN FNIP1 FOLH1 FOLH1B FOXP2 FSTL1FTOFZD3 GAPDH GLRA2 GNPAT GMPS GPR18 GPR50 GPR125 HNRNPA2B1 HRH2 HS3ST2 HS6ST3 HSD11B1 HSPA8 HSPD1 ITIH4 KIAA0513 KREMEN1 LRRTM1 MMS22L MYL12B NCOA7 NLGN4X NOVA1 NR4A3 NRCAM NTNG2 NUBPL OLFM1 OPHN1 PADI2 PAK2 PAK3 PCDHA3 PDE7B PDE10A PGAM1 PGBD1 PGK1 PGPPNPLA8 PLXNA2 POMC POM121L2 PRKAG1 PRKAG2 PRKAG3 PYGB RAI1 RAPGEF6 RELN RIMS2 RIT2 RORB RSRC1 SEL1L3 SEMA3ASEMA3D SEPT4 SIM1SLC32A1 SLC17A1 SLC17A7 SLC24A5 SLC25A14 SLITRK2 SMARCA2 SMARCC1

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	SMARCE1SPARCL1 SRD5A2 STRNTAAR6 TH1L
	TMTC4 TRIM3 UBAC2 UNC5C UGGT2
	UQCRC1USP46 UTRN YWHAZ ZBED4 ZBTB20 ZDHHC8
	RECENNEZ KONH7 ALDH1A3 MTNR1A BRPF1 EMBLL ZNF804A COMMEN NR4A2 HEDTER BLOKK AKT _ HSPD1 _ OFCC1 BRIGAL MCHR2 MEGF10 ANKK1 PCDH11Y COX2 SEMA3B DGCR2 SLCO6A1 PICK1 AND HEMADA SLC25A14 TRMT2A ATHORIE VRK2 PCDH41 DHD1 ND2 ATP2A2 GSPT1 _ WACN _ MCTP2 HTR2A = CHRNA7 CHEMA GRIAT PACHE RCAN1 SLC26A6 KOME2 _ ACSL6 _ NOS1 MAD _ FAMBA1 ST7 DD0 ACTRE ERVWE1 _ OMA GNAL SEPT7 NBEAL2 _ DARRET OMAE KIF21A NS3 ACSS _ JANEZ PSMD10 _ KCHRE _ NUMBL RIC3 . BRPF3 MBLN2 SPA17 KCNN3 SLC1A6 LSAMP REMARA CHEMA AND HEMADA PIKAC2G GRM7 ACHE POS JAGI NRXN3 ATF7IP UFD1L MAGE2 PCDH12 MPTDA NEFH DGKI _ CHEM ALDH3B1 RLBP1 MCT2 _ SLC1TA6 MAP6 NDUFS1 TCF4 SNX6 PPP3CC SGCE _ ME3 ORC3 CICINES TRAFET WA BPRF3 ATP6VD1 DCDC2 PPT1 KOM9 _ WDR1 _ HSPA8 CRYAB _ SLC1A2 APP TMEM200A _ CLINT1 HDAC9 _ PAICS MTH3 NPTN PPP1R1B ORES NR3C1 OK1 PLAA PURA _ DEFD _ SMG6 ASTN2 PRKAR2A MPDZ SC5DL
	Human gene products
HEKV-W no	Glutamate related GLUL Synaptic SPTNB5 GNB2 SHANK3
filter	PDZRN4 DISC1 related DISC1 Neuregulin and growth FLT4 NEURL4
Number of SZ	NLGN4X NRG3 Myelin related MYL6B Oxidative stress
genes $= 13$	ATP11A COX11 Channels Calcium CATSPERG
DTNBPI HLA-DRBI MADILI MEDI2 MYOI8B NOSI NRG3	CACNG2 ITPR3 Potassium KCNN1 KCNJ16 Oxidative
SHANK3 SP4 XRCC1	stress PYROXD2 Immune /cytokine cirra DEFA7P
	TNFAIP3 CSF3R Structural COL6A3 DNM2 MYO18B
	MYBPC1 Signalling PDE4C PLA2R1 PLCB3 PLHDA2
	PLA2G4A PRICKLE1(wnt) ABCA2 ABCA5 ABLIM3
	ADAM29 ADAMYTSL2 AIM1L AKR1B15 ALG9 ANKRD11 ANXA1
	ARID2 ARSE BET1L CCDC51 CENPE CEP152
	CDH11 CHD23 CYFIP1 CYP1A2 DHTKD1 DPYSL5 DST EDC4

ERV3 ERVWE2 EXOC5 FAM65A FBN1FBN3 FEM1AGPSM1 HELB HERV-V1HERV-V2 HIRIP3 HSD3B7 HSPD1 INTS2 IPO4 ITGB6 кіаа1731 KRTAP12-1 LAMB2 LOC100288413 LPHN2 LRP2 MAD11 1 MAU2 MBOAT1 MED12 MED12 MINPP1 MMD2MUC12 NAP1L5 NER NCAPD3 NDC80 NXPH1 OAS1 PCDHB5 PCDHB6 PCYT1B PDMD3 PHE3 PPYR1 PRDM10 PRMT2 PRSS22 PSMC4 PTPRU RABAC1 RGL1 RRM1 RG9MTD3 RIMBP3 RIMBP3B RNF213 RUNDC3B RXFP3 SARM1 SCAMP3 SHH SP4 SKIV2L2 SLC28A3 SIK SMC2 SPXW5 SRGAP1 STIM2 TCHH TEX11 THEM5 TIMM44TRIM39 TKTL1 TMEM11 TMF1 TP63 TRAK1 TRIM39R TRIM47 TSHR TTC18 USH2A USP31 VMAC VPD13D WDR20 WDR62 WNK2 XIRP1 XRCC1 ZBTB6 ZDHHC11 ZKSCAN2 ZNF34 ZNF99 ZNF355P TTN GNA12 \_\_\_\_\_ OPSF2 FBXW5 \_\_\_\_ DISC1 AROLUI FAMAGE LSM14B CKAP5 UTRN ERVERDE1 SHITC1 HELT ANKFY1 \_\_\_ Influenza: no **Dopamine related** DRD1 DRD2 D(1A) dopamine receptor DRD2 DRD4 DRD5 - SLC5A3 Serotonin HTR1B HTR1D HTR5A SLC5A4 Glutamate receptors and Number of SZ release GLURI GRIA1 GRIK1 GRIN3B Synaptic genes = 24

filter

ACE CACNA1C DISC1 DRD2 DRD4 DRD5 ERBB3 GNAO1 GRIA1 HLA-DRB1 HTR1B HTR5A IL4 IPO5 KREMEN1 LRRTM1 ND2 NOS1 NRG1 NRG2 PCNT PNPLA8 PSEN2 SLC40A1 PDZRN3 syntaxin 16 Neuregulin and growth ERBB3 NRG1 NRG2 UTRN DISC1 related DISC1 PCNT Translation initiation EIF3A EIF3K EIF4G2 Oxidative stress ACOX2 cytochrome c oxidase subunit II NADH dehydrogenase subunit 1 NADH dehydrogenase subunit 2 Glycine receptors GLRAI GLRA2 GLRA3 GLRA4 Calcium voltage-dependent L-type calcium channel subunit Immune/cytokine NFATC4 TNFRSF1B TLR4 TNFSF10 immunoglobulin heavy chain IFT122 Signalling NFKBIL2 PIK3CA SOS1 Structural ACTRT2 COL6A5 ABCE6ACE ACTRT1 ADNP AKAP6 ANXA1 ANXA4 APOBEC1 APIG2 AQP7 ARMC7 ASB1 BAT2L1 CCDC135 CCS CCT6B CDC14A CDKL3 CENPA CEP192 CNGA4 CNOT1 CNTNAP5 CP CREG2 CRIM1CRIPAK CSEIL DCLRE1A DDX5 DENND3 DOCK1 EDNRA EFHC1ECHDC2 ESYT1 EXOC6 FAM123A FCHSD1 FUBP1GCNT6 GLT8D2 GLTSCRI GNAO1 GPR153 GSDMD HTT IPO5 ISM2 KIFC2 KREMEN1 KRT76 LIPN LRRTM1 LRRC61 LRRC14Blrrigi \_ MAN1B1map7di MBOAT7 mrap MTMR2 MGATSB NOP58 NR2F6 NWD1 PLEC PHC3 PHLPP2 POM121L12 PPFIA4 PRL prx \_ RAB15 RBPJ RPM12B RSPH9 RHOBTB2 SCAPER SEC22A

	SENP7 SIPA1L2 SLC22A15 SLC26A9 SLC40A1 SMTLN1 SNAI1 SNTG1 STAC3
	$TABC1_{{\tt TAS2R20}} {\tt TCHH} TMEM63A TMEM143_{{\tt TMEM163}} {\tt TMIGD2} TM4SF4 \_ {\tt rsph9}$
	ANLEZ TIN RAMA TAS2R31 TRIM33 TROAP TYK2 SPC25 THADA UPF2
	UBR3 umodl1 (Defined AP1G2 (Press) XPO5 ZFP2 ZNF219 ZFPM1 ZNF391 LIPF
	COLAM NFI PSEN2 PARS DEECT PIAS2 GTPBPI oxiom-dumed, unhug-degradue, Linguidad Cadoual 1983PI SNEK GASG IL4 TMTC4 GNA11 HOMER SPTY2DI DNAH8 AMI
Rubella: no filter	Cholinergic CHRNA3 ACHE Dopamine receptors dopamine receptor isoform short
	DRD4 Serotonin receptors HTR3C HTR3D Glutamate receptors GRM2
Number of SZ	gluR7 GRIK2 GRIK3 GRIK4 GRIK5 NMDA receptor subunit epsilon
genes = 21  disci drd2 Drd4 egr4 enah erbb2	2 NMDA receptor subunit epsilon-3 NMDA receptor subunit
FLNB GNAS GRIK2 GRIK3 GRIK4 GRIN2B GRIN2D HLA- A HI A-B HI A-DRBI NOSI	epsilon-4 Real GABA GABA transporter 1 Synaptic
NRG2 NRXN1 PRODH SHANK3	GRASP HIPK4 SYNPO PDZD7 SHANK3 SYNCRIP DISC 1 related DISCI
	Neuregulin and growth erbB-2 NRG2 GDF1 IGFBP3 Translation initiation
	EIF3A E2F1 Channels Calcium CACNAB1 CACNA1E Signalling
	GRB10 RASSF7 INPPLI PTPRK PPP1R14C MOREN Immune cytokine
	HLA-A HLA-B MDR/TAP IL17D Structural ACTN4 COLIAI
	COL7A1MYH14
	ABTB2 ADAMTSL5 adraid $\operatorname{AKAP5}$ AKNA AIPL1 arhgap30 baalc BST1
	CECR6 CHAC2 CKAP4 CTDP1 DCAF8L2 ENAH EPN3 FARS2
	FLNB  HCG4P6 HMGN1 INA  IQSEC2 ITGA8 ITGB4  KlHl4
	KLHDC4 LARP1 LEMD2 MED23 MFSD6 MON1B
	MRVIL myoib NCRNA00265 $NSAP1$ NTN1 NXF5 OPMCL $OS9$ otop1

# PARP9 PGAM5 PIGZ POM121 PTGER3 PYCRL

RNF128 SALL3 SASH1 SEC16A SEMA6C SEMAZ SRP68

# SLC4A3 SLC20A2 SOX4 SPAST SPHKAP SRD5A1 SRCAP

#### STARD9 STRA13 TFAP2B THEG TFAP2C

## TFAP2E THSD7A TJP3 TMC1 TMEM31 TMC8 TNKS2 TNRC18

## UNC5A VDRIP WAPAL WIPF3 WDR86

Z3CH3 Z3CH4 ZC3HAV1 ZFP36L2 ZP1 ZYX

001	PICHI	Shineseen		ID5 WDR20
PARVA	nor HMGAI	party.	80	008
REM2	ELAVLI	MAN2A2	10078 iku.	SSPO
hnRNP Q2	PRJ5 NRBP2	with web and and the Trans		TRAP2A RADPA
CCDC80	erer: PCSK7	un.r	RASAL3	KIPC3
QNAS havenue to	we MELTIO	PHC2 NAME	NLRP9	
ТЛРО2 ноок2	500 MAY 200	CDR2L	AGXT2	MEGF6 thyroid
receptor interactor	M attaches car	integrin, alpha 5		0 .co. au

therefore appear to be interdependent. The pathogens may promote disease if the human genes encode for homologous products, and the genes promote disease if the homologous pathogen is encountered. Such interdependence likely explains the heterogeneous data in both gene and risk factor association studies.

Other pathogens, including Borrelia Burgdorferri and T. Gondii have also been implicated in schizophrenia. These too express many homologous proteins to both viral and human proteomes. These parasites tend to be associated with schizophrenia in adulthood, while viral infections are predominantly prenatal risk factors. These may have primed the antibody network to respond to homologous antigens expressed by Borrelia or T. Gondii, suggesting that detection and elimination of these pathogens may be of therapeutic benefit in adult life.

Schizophrenia is a neurodevelopmental disorder [129, 130] and, as the risk-promoting effects of viruses are related

to maternal infection, it is possible that knockdown or interference of foetal proteins by viral-induced antibodies targeting their human counterparts may contribute to the neurodevelopmental disturbances observed in schizophrenia. Indeed DISC1, neuregulin, ERBB4, FEZ1 or COMT knockout mice display many of the pathological and behavioural symptoms associated with schizophrenia [131– 135]. Viral interference with these same proteins might be expected to promote the same effects, but on a massive scale, targeting many relevant proteins at once. It is also possible that such autoantibodies play a role in the comorbid conditions associated with schizophrenia, for example autoimmune disease such as Thyrotoxicosis, celiac disease, acquired haemolytic anaemia, interstitial cystitis, or Sjogren's syndrome [136].

Autoantibodies to several proteins have been reported in schizophrenia (muscarinic, nicotinic, dopaminergic and NMDA receptors, *inter alia*, (Table 2) and all are homologous

DISC1 partner gene symbol	Protein name	Viral binder
ACTG1	Actin, cytoplasmic 2	HIV-1 [97] HSV1 [98]
ACTN2	Actinin, alpha 2	Hepatitis C [99]
АКАР9	A-kinase anchor protein 9	Epstein-Barr [97]
ATF4	Cyclic AMP-dependent transcription factor ATF-4	HSV1 [98]
ATF5	Cyclic AMP-dependent transcription factor ATF-5	HTLV1 [100]
BICD1	Protein bicaudal D homolog 1	Cytomegalovirus [101]
C14orf135	Uncharacterized protein C14orf135 precursor	Hepatitis C [102]
DCTN1	Dynactin-1	HSV1 [98]
DCTN2	Dynactin subunit 2	Dynactins are involved in the transport of the adenoviruses, HSV-1, the hantaan virus, HTLV-1 and the poliovirus [103–108]
DNAJC7	DnaJ homolog subfamily C member 7	Part of a complex forming the coxsackie virus receptor [109]
DYNC1H1	Dynein heavy chain, cytosolic	Adenovirus (in a complex with dynactin and NDEL1) [110]
EEF2	Elongation factor 2	Epstein Barr [111]
EIF3S3	Eukaryotic translation initiation factor 3 subunit 3	Hepatitis C [112]
FEZ1	Fasciculation and elongation protein zeta 1 (zygin I)	JC Polyomavirus [113]
HERC2	HECT domain and RCC1-like domain-containing protein 2	Papillomavirus 16 [114]
KIF3C	Kinesin-like protein KIF3C	HIV-1 [115]
MATR3	Matrin-3	HSV1 [98]
NDEL1	Nuclear distribution protein nudE-like 1	Part of a complex involved in Adenovirus transport (with dynactin and cytoplasmic dynein) [110]
PAFAH1B1	Platelet-activating factor acetylhydrolase IB subunit alpha	Binds to Poliovirus P3 protein and HIV-1 Tat [116, 117]
PCNT	Pericentrin	Involved in the microtubular transport of the adenovirus [118]
PGK1	Phosphoglycerate kinase 1	Epstein-Barr [119]
SMARCE1	SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subfamily E member 1	HSV-1 [97]
STX18	Syntaxin-18	Papillomavirus [119]
TNKS	Tankyrase-1	Epstein-Barr [120]
TUBB	Tubulin beta chain	Epstein-Barr [119]
YWHAE	14-3-3 protein epsilon	Hepatitis C [97]: L:Epstein-Barr [119]
YWHAQ	14-3-3 protein theta	HIV [97] HSV1 [98]
YWHAZ	14-3-3 protein zeta/delta	HSV1 [98]: Epstein-Barr [119]

TABLE 4: Viruses reported to bind to DISC1 interactome partners.

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T = T 1	C 1 · 1 ·	$1 \cdot V = V = O O$	1 1 1 1	1 1 1 1 1 1 1 1
TABLE 5: The number of	of schizophrenia gene	products in KEGG r	bathways related to immunity	, and viral or pathogen life cycles.

Dathogon pathwaya		Viral pathwaya		Immuna	
Pathogen pathways		virai patnways	Viral pathways		
Toxoplasmosis	16	Focal adhesion	20	Cytokine-cytokine receptor interaction	26
Chagas disease	15	Cell adhesion molecules (CAMs)	19	Jak-STAT signaling pathway	16
Amoebiasis	13	Regulation of actin cytoskeleton	17	Systemic lupus erythematosus	13
Leishmaniasis	12	Protein processing in endoplasmic reticulum	13	T cell receptor signaling pathway	13
Viral myocarditis	8	Endocytosis	12	Phagosome	12
Staphylococcus aureus infection	7	Phagosome	12	Allograft rejection	11
Epithelial cell signaling in Helicobacter pylori infection	6	Gap junction	11	Hematopoietic cell lineage	11
Malaria	6	Tight junction	11	Antigen processing and presentation	10
Tryptophan metabolism	6	Adherens junction	6	Fc epsilon RI signaling pathway	10
NOD-like receptor signaling pathway	4	ECM-receptor interaction	6	Apoptosis	10
Vibrio cholerae infection	4	Oocyte meiosis	5	Graft-versus-host disease	9
Bacterial invasion of epithelial cells	3	SNARE interactions in vesicular transport	4	Autoimmune thyroid disease	8
E.coli infection	3			Chemokine signaling pathway	8
RIG-I-like receptor signaling pathway	3	Basal transcription factors	3	Leukocyte transendothelial migration	8
Cytosolic DNA-sensing pathway	2	Spliceosome	2	Natural killer cell mediated cytotoxicity	8
Shigellosis	2	Aminoacyl-tRNA biosynthesis	1	Adipocytokine signaling pathway	7
		Base excision repair	1	Asthma	7
		RNA degradation	1	Intestinal IgA production	5
				Toll-like receptor signaling pathway	5
				Complement and coagulation cascades	4
				B cell receptor signaling pathway	3
				TGF-beta signaling pathway	3
				Lysosome	2
				Regulation of autophagy	2
				Fc gamma R-mediated phagocytosis	1
				Primary immunodeficiency	1

to proteins expressed by the risk factors in schizophrenia. The effects of antibody knockdown have not been analysed for any schizophrenia related proteins, but have been reported for the microtubule-related protein *tau*, in relation to Alzheimer's disease. In mice, *tau* immunisation produces *tau* hyperphosphorylation, neurofibrillary tangles and axonal damage as seen in the human condition [137]. *Tau* (MAPT) is homologous to Herpes simplex (HSV-1) and a number of other pathogens. Such effects are relevant to the autoantigens observed in schizophrenia.

Schizophrenia is also a degenerative disease in adolescence or adulthood, characterised by oligodendrocyte cell loss, impaired synaptic connectivity and pyramidal cell dendrite shrinkage [41, 138–140], In the light of the above homologies it seems likely that such degenerative changes may relate to autoimmune-related attack of these diverse compartments. Indeed there is evidence for microglial activation in the schizophrenic brain [141] and several studies have reported changes in the cytokine profile in the brain, CSF or peripheral immune compartments [24, 142–146]. TABLE 6: Human homologues of Norwalk virus proteins.

Dopamine metabolisers	Amine transporters	Others
AOC2 amine oxidases AOC3"" KDM1A amine oxidase demethylase KDM1B" MAOA monoamine oxidase MAOB "" RNLS renalase amine oxidase SMOX spermine oxidase SPR sepiapterin reductase Monoamine synthesis cofactor SULT1A1 sulphotransferases SULT1A3 monoamine metabolite sulphation SULT1A4	SLC6A2 (Noradrenaline) SLC6A3 (Dopamine) SLC18A1vesicular monoamine SLC18A2"" SLC22A2 organic cation SLC22A3 extraneuronal monoamine SLC29A4 (Na <sup>+</sup> /H <sup>+</sup> )	CADPS2 amine release activator CDCA7 cell division cycle associated 7 CDCA7L IL411 cytokine PICK1 postsynaptic scaffold

3.8. Clinical Implications in Schizophrenia and Other Conditions. These data suggest that susceptibility gene products are the vehicles enabling the risk-promoting effects of pathogenic risk factors, via the interactions described above, and that the two are indispensable for the genesis of schizophrenia. Pathogen detection and elimination or vaccination, particularly prior to pregnancy might be expected to reduce the incidence of schizophrenia and also to be of clinical benefit in adulthood. Interestingly, vitamin D is able to stunt the growth of T. Gondii [147] and low levels of this vitamin, both prenatally and in adulthood, have been associated with schizophrenia risk, although abnormally high levels are also a risk factor [148]. Pharmaceutical effort in this direction may also vastly improve the armoury and safety of drugs against parasites such as T. Gondii and Borrelia.

Autoimmunity, involving several key schizophreniarelated proteins may well be a consequence of pathogen infection, and related to viral/human protein homology. Antigen and antibody removal by immunoadsorption techniques might therefore also be if clinical benefit.

This scenario suggests a novel and probably common class of "pathogenetic" autoimmune disease caused by pathogens but dependent on our genes. Indeed, the same phenomenon has been observed in Alzheimer's disease where the risk factor herpes simplex expresses proteins containing peptide matches to the products of multiple susceptibility genes [128]. Work from Kanduc's laboratory has also shown that 30 viral proteomes, including many nonretroviruses, contain multiple pentapeptide matches to many human proteins [149]. This is corroborated by data posted at http://www.polygenicpathways.co.uk/blasts.htm which shows, inter alia, that Bornavirus proteins, a virus implicated in Bipolar disorder [150], display this type of homology in relation to Bipolar disorder susceptibility gene products, that the coronavirus implicated in Parkinson's disease [151] expresses proteins homologous to the PARK7 gene product and to dopaminergic and oxidative stressrelated proteins, and that multiple sclerosis autoantigens are homologous to the products of the Epstein-Barr virus which has been implicated in this disorder [152]. Our genomes and polymorphisms determine which vatches we possess, which

pathogens match these sequences and which pathogenrelated disorder we might develop. Environmental variables, and vaccination, determine which pathogens we encounter and our immune system (HLA-antigens and immune background determined soon after birth) may determine how we deal with these pathogens. With the power of current day bioinformatics, it should be possible to rapidly identify all vatches in the human proteome and to pair them with the various pathogenic species and human diseases. This would greatly aid our understanding of the implication of pathogens in disease and may lead to radically new therapies and prevention strategies in many disorders.

#### References

- T. J. Crow, "How and why genetic linkage has not solved the problem of psychosis: review and hypothesis," *American Journal of Psychiatry*, vol. 164, no. 1, pp. 13–21, 2007.
- [2] M. J. Owen, N. Craddock, and M. C. O'Donovan, "Schizophrenia: genes at last?" *Trends in Genetics*, vol. 21, no. 9, pp. 518–525, 2005.
- [3] C. J. Carter, "Schizophrenia susceptibility genes converge on interlinked pathways related to glutamatergic transmission and long-term potentiation, oxidative stress and oligodendrocyte viability," *Schizophrenia Research*, vol. 86, no. 1–3, pp. 1–14, 2006.
- [4] C. J. Carter, "eIF2B and oligodendrocyte survival: where nature and nurture meet in bipolar disorder and schizophrenia?" *Schizophrenia Bulletin*, vol. 33, no. 6, pp. 1343–1353, 2007.
- [5] A. K. Khler, S. Djurovic, L. M. Rimol et al., "Candidate gene analysis of the human natural killer-1 carbohydrate pathway and perineuronal nets in schizophrenia: B3GAT2 Is associated with disease risk and cortical surface area," *Biological Psychiatry*, vol. 69, no. 1, pp. 90–96, 2011.
- [6] P. Jia, L. Wang, H. Y. Meltzer, and Z. Zhao, "Common variants conferring risk of schizophrenia: a pathway analysis of GWAS data," *Schizophrenia Research*, vol. 122, no. 1–3, pp. 38–42, 2010.
- [7] C. O'Dushlaine, E. Kenny, E. Heron et al., "Molecular pathways involved in neuronal cell adhesion and membrane scaffolding contribute to schizophrenia and bipolar disorder

susceptibility," *Molecular Psychiatry*, vol. 16, no. 3, pp. 286–292, 2010.

- [8] K. K. Nicodemus, A. J. Law, E. Radulescu et al., "Biological validation of increased schizophrenia risk with NRG1, ERBB4, and AKT1 epistasis via functional neuroimaging in healthy controls," *Archives of General Psychiatry*, vol. 67, no. 10, pp. 991–1001, 2010.
- [9] M. E. Talkowski, G. Kirov, M. Bamne et al., "A network of dopaminergic gene variations implicated as risk factors for schizophrenia," *Human Molecular Genetics*, vol. 17, no. 5, pp. 747–758, 2008.
- [10] K. K. Nicodemus, J. H. Callicott, R. G. Higier et al., "Evidence of statistical epistasis between DISC1, CIT and NDEL1 impacting risk for schizophrenia: biological validation with functional neuroimaging," *Human Genetics*, vol. 127, no. 4, pp. 441–452, 2010.
- [11] A. S. Brown, P. Cohen, S. Greenwald, and E. Susser, "Nonaffective psychosis after prenatal exposure to rubella," *American Journal of Psychiatry*, vol. 157, no. 3, pp. 438–443, 2000.
- [12] W. Adams, R. E. Kendell, E. H. Hare, and P. Munk-Jorgensen, "Epidemiological evidence that maternal influenza contributes to the aetiology of schizophrenia. An analysis of Scottish, English, and Danish data," *British Journal of Psychiatry*, vol. 163, pp. 522–534, 1993.
- [13] A. S. Brown, M. D. Begg, S. Gravenstein et al., "Serologic evidence of prenatal influenza in the etiology of schizophrenia," *Archives of General Psychiatry*, vol. 61, no. 8, pp. 774–780, 2004.
- [14] E. F. Torrey, R. Rawlings, and I. N. Waldman, "Schizophrenic births and viral diseases in two states," *Schizophrenia Research*, vol. 1, no. 1, pp. 73–77, 1988.
- [15] P. B. Mortensen, C. B. Pedersen, D. M. Hougaard et al., "A Danish National Birth Cohort study of maternal HSV-2 antibodies as a risk factor for schizophrenia in their offspring," *Schizophrenia Research*, no. 1–3, pp. 257–263, 2010.
- [16] G. Stober, E. Franzek, and H. Beckmann, "Pregnancy infections in mothers of chronic schizophrenic patients. The significance of differential nosology [Schwangerschaftsinfektionen bei Muttern von chronisch Schizophrenen. Die Bedeutung einer differenzierten Nosologie]," *Nervenarzt*, vol. 65, no. 3, pp. 175–182, 1994.
- [17] J. Suvisaari, J. Haukka, A. Tanskanen, T. Hovi, and J. Lönnqvist, "Association between prenatal exposure to poliovirus infection and adult schizophrenia," *American Journal of Psychiatry*, vol. 156, no. 7, pp. 1100–1102, 1999.
- [18] P. Rantakallio, P. Jones, J. Moring, and L. von Wendt, "Association between central nervous system infections during childhood and adult onset schizophrenia and other psychoses: a 28-year follow-up," *International Journal of Epidemiology*, vol. 26, no. 4, pp. 837–843, 1997.
- [19] R. H. Yolken and E. F. Torrey, "Are some cases of psychosis caused by microbial agents? A review of the evidence," *Molecular Psychiatry*, vol. 13, no. 5, pp. 470–479, 2008.
- [20] M. Fritzsche, "Seasonal correlation of sporadic schizophrenia to Ixodes ticks and Lyme borreliosis," *International Journal of Health Geographics*, vol. 1, article 2, 2002.
- [21] H. Karlsson, S. Bachmann, J. Schröder, J. McArthur, E. F. Torrey, and R. H. Yolken, "Retroviral RNA identified in the cerebrospinal fluids and brains of individuals with schizophrenia," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 98, no. 8, pp. 4634– 4639, 2001.
- [22] C. J. Carter, "Schizophrenia susceptibility genes directly implicated in the life cycles of pathogens: cytomegalovirus,

influenza, herpes simplex, rubella, and Toxoplasma gondii," *Schizophrenia Bulletin*, vol. 35, no. 6, pp. 1163–1182, 2009.

- [23] J. Sun, P. Jia, A. H. Fanous et al., "Schizophrenia gene networks and pathways and their applications for novel candidate gene selection," *PLoS One*, vol. 5, no. 6, Article ID e11351, 2010.
- [24] N. Müller, "The role of the cytokine network in the CNS and psychic disorders," *Nervenarzt*, vol. 68, no. 1, pp. 11–20, 1997.
- [25] M. Rothermundt, V. Arolt, and T. A. Bayer, "Review of immunological and immunopathological findings in schizophrenia," *Brain, Behavior, and Immunity*, vol. 15, no. 4, pp. 319–339, 2001.
- [26] S. Theodoropoulou, G. Spanakos, C. N. Baxevanis et al., "Cytokine serum levels, autologous mixed lymphocyte reaction and surface marker analysis in never medicated and chronically medicated schizophrenic patients," *Schizophrenia Research*, vol. 47, no. 1, pp. 13–25, 2001.
- [27] K. F. Tanaka, F. Shintani, Y. Fujii, G. Yagi, and M. Asai, "Serum interleukin-18 levels are elevated in schizophrenia," *Psychiatry Research*, vol. 96, no. 1, pp. 75–80, 2000.
- [28] M. C. O'Donnell, S. V. Catts, P. B. Ward et al., "Increased production of interleukin-2 (IL-2) but not soluble interleukin-2 receptors (sIL-2R) in unmedicated patients with schizophrenia and schizophreniform disorder," *Psychiatry Research*, vol. 65, no. 3, pp. 171–178, 1996.
- [29] R. Ganguli, B. S. Rabin, R. H. Kelly, M. Lyte, and U. Ragu, "Clinical and laboratory evidence of autoimmunity in acute schizophrenia," *Annals of the New York Academy of Sciences*, vol. 496, pp. 676–685, 1987.
- [30] M. Makinodan, K. Tatsumi, T. Manabe et al., "Maternal immune activation in mice delays myelination and axonal development in the hippocampus of the offspring," *Journal of Neuroscience Research*, vol. 86, no. 10, pp. 2190–2200, 2008.
- [31] S. F. Altschul, T. L. Madden, A. A. Schäffer et al., "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs," *Nucleic Acids Research*, vol. 25, no. 17, pp. 3389–3402, 1997.
- [32] J. E. Larsen, O. Lund, and M. Nielsen, "Improved method for predicting linear B-cell epitopes," *Immunome Research*, vol. 2, p. 2, 2006.
- [33] S. Goto, H. Bono, H. Ogata et al., "Organizing and computing metabolic pathway data in terms of binary relations," *Pacific Symposium on Biocomputing*, pp. 175–186, 1997.
- [34] M. Pirooznia, V. Nagarajan, and Y. Deng, "GeneVenn—a web application for comparing gene lists using Venn diagrams," *Bioinformation*, vol. 1, pp. 420–422, 2007.
- [35] D. Kanduc, "Describing the hexapeptide identity platform between the influenza A H5N1 and Homo sapiens proteomes," *Biologics*, vol. 4, pp. 245–261, 2010.
- [36] S. Himelhoch, J. F. McCarthy, D. Ganoczy et al., "Understanding associations between serious mental illness and hepatitis C virus among veterans: a national multivariate analysis," *Psychosomatics*, vol. 50, no. 1, pp. 30–37, 2009.
- [37] J. G. Baillargeon, D. P. Paar, H. Wu et al., "Psychiatric disorders, HIV infection and HIV/hepatitis co-infection in the correctional setting," *AIDS Care*, vol. 20, no. 1, pp. 124–129, 2008.
- [38] C. P. Carney, L. Jones, and R. F. Woolson, "Medical comorbidity in women and men with schizophrenia: a populationbased controlled study," *Journal of General Internal Medicine*, vol. 21, no. 11, pp. 1133–1137, 2006.
- [39] F. Cournos, K. McKinnon, and G. Sullivan, "Schizophrenia and comorbid human immunodeficiency virus or hepatitis C virus," *Journal of Clinical Psychiatry*, vol. 66, no. 6, pp. 27– 33, 2005.

- [40] M. S. van der Knaap, J. C. Pronk, and G. C. Scheper, "Vanishing white matter disease," *The Lancet Neurology*, vol. 5, no. 5, pp. 413–423, 2006.
- [41] N. Uranova, D. Orlovskaya, O. Vikhreva et al., "Electron microscopy of oligodendroglia in severe mental illness," *Brain Research Bulletin*, vol. 55, no. 5, pp. 597–610, 2001.
- [42] V. Vostrikov, D. Orlovskaya, and N. Uranova, "Deficit of pericapillary oligodendrocytes in the prefrontal cortex in schizophrenia," *World Journal of Biological Psychiatry*, vol. 9, no. 1, pp. 34–42, 2008.
- [43] V. M. Vostrikov, "Decreased numerical density of pericapillary oligodendrocytes in the cortex in schizophrenia," *Zhurnal Nevrologii i Psihiatrii imeni S.S. Korsakova*, vol. 107, no. 12, pp. 58–65, 2007.
- [44] Y. Hakak, J. R. Walker, C. Li et al., "Genome-wide expression analysis reveals dysregulation of myelination-related genes in chronic schizophrenia," *Proceedings of the National Academy* of Sciences of the United States of America, vol. 98, no. 8, pp. 4746–4751, 2001.
- [45] D. R. Friedlander, P. Milev, L. Karthikeyan, R. K. Margolis, R. U. Margolis, and M. Grumet, "The neuronal chondroitin sulfate proteoglycan neurocan binds to the neural cell adhesion molecules Ng-CAM/L1/NILE and N-CAM, and inhibits neuronal adhesion and neurite outgrowth," *Journal of Cell Biology*, vol. 125, no. 3, pp. 669–680, 1994.
- [46] A. Sandvig, M. Berry, L. B. Barrett, A. Butt, and A. Logan, "Myelin-, reactive glia-, and scar-derived CNS axon growth inhibitors: expression, receptor signaling, and correlation with axon regeneration," *Glia*, vol. 46, no. 3, pp. 225–251, 2004.
- [47] Z. J. Chen, Y. Ughrin, and J. M. Levine, "Inhibition of axon growth by oligodendrocyte precursor cells," *Molecular and Cellular Neuroscience*, vol. 20, no. 1, pp. 125–139, 2002.
- [48] H. P. Li, A. Oohira, M. Ogawa, K. Kawamura, and H. Kawano, "Aberrant trajectory of thalamocortical axons associated with abnormal localization of neurocan immunoreactivity in the cerebral neocortex of reeler mutant mice," *European Journal* of Neuroscience, vol. 22, no. 11, pp. 2689–2696, 2005.
- [49] K. Verhoeven, P. de Jonghe, T. van de Putte et al., "Slowed conduction and thin myelination of peripheral nerves associated with mutant rho Guanine-nucleotide exchange factor 10," *American Journal of Human Genetics*, vol. 73, no. 4, pp. 926–932, 2003.
- [50] D. Ben-Shachar and R. Karry, "Neuroanatomical pattern of mithochondrial complex I pathology varies between schizoprenia, bipolar disorder and major depression," *PLoS One*, vol. 3, no. 11, Article ID e3676, 2008.
- [51] E. Y. Yuen and Z. Yan, "Dopamine D<sub>4</sub> receptors regulate AMPA receptor trafficking and glutamatergic transmission in GABAergic interneurons of prefrontal cortex," *Journal of Neuroscience*, vol. 29, no. 2, pp. 550–562, 2009.
- [52] K. Fukunaga and Y. Takeuchi, "Novel intracellular signal transduction of dopamine D2 receptor in schizophrenia," *Tanpakushitsu Kakusan Koso*, vol. 51, no. 11, pp. 1602–1608, 2006.
- [53] T. Miyakawa, L. M. Leiter, D. J. Gerber et al., "Conditional calcineurin knockout mice exhibit multiple abnormal behaviors related to schizophrenia," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 15, pp. 8987–8992, 2003.
- [54] J. P. B. Viola and A. Rao, "Role of the cyclosporinsensitive transcription factor NFAT1 in the allergic response," *Memorias do Instituto Oswaldo Cruz*, vol. 92, supplement 2, pp. 147–155, 1997.

- [55] A. Andrieux, P. A. Salin, M. Vernet et al., "The suppression of brain cold-stable microtubules in mice induces synaptic defects associated with neuroleptic-sensitive behavioral disorders," *Genes and Development*, vol. 16, no. 18, pp. 2350– 2364, 2002.
- [56] A. Andrieux, P. A. Salin, and D. Job, "A role for the cytoskeleton in mental diseases? [Un rôle pour les microtubules dans les pathologies psychiatriques?]," *Pathologie Biologie*, vol. 52, no. 2, pp. 89–92, 2004.
- [57] A. Arcangeli, B. Rosati, A. Cherubini et al., "HERG- and IRKlike inward rectifier currents are sequentially expressed during neuronal development of neural crest cells and their derivatives," *European Journal of Neuroscience*, vol. 9, no. 12, pp. 2596–2604, 1997.
- [58] G. A. M. Smith, H. W. Tsui, E. W. Newell et al., "Functional up-regulation of HERG K channels in neoplastic hematopoietic cells," *Journal of Biological Chemistry*, vol. 277, no. 21, pp. 18528–18534, 2002.
- [59] J. Gommeaux, C. Grégoire, P. Nguessan et al., "Thymusspecific serine protease regulates positive selection of a subset of CD4<sup>+</sup> thymocytes," *European Journal of Immunology*, vol. 39, no. 4, pp. 956–964, 2009.
- [60] A. Buonanno, O. B. Kwon, L. Yan et al., "Neuregulins and neuronal plasticity: possible relevance in schizophrenia," *Novartis Foundation Symposium*, vol. 289, pp. 165–177, 2008.
- [61] P. Seeman, "Glutamate and dopamine components in schizophrenia," *Journal of Psychiatry and Neuroscience*, vol. 34, no. 2, pp. 143–149, 2009.
- [62] N. A. Uranova, V. M. Vostrikov, D. D. Orlovskaya, and V. I. Rachmanova, "Oligodendroglial density in the prefrontal cortex in schizophrenia and mood disorders: a study from the Stanley Neuropathology Consortium," *Schizophrenia Research*, vol. 67, no. 2-3, pp. 269–275, 2004.
- [63] C. Winter, T. J. Reutiman, T. D. Folsom et al., "Dopamine and serotonin levels following prenatal viral infection in mouse-Implications for psychiatric disorders such as schizophrenia and autism," *European Neuropsychopharmacology*, vol. 18, no. 10, pp. 712–716, 2008.
- [64] J. K. Millar, J. C. Wilson-Annan, S. Anderson et al., "Disruption of two novel genes by a translocation co-segregating with schizophrenia," *Human Molecular Genetics*, vol. 9, no. 9, pp. 1415–1423, 2000.
- [65] L. M. Camargo, Q. Wang, and N. J. Brandon, "What can we learn from the disrupted in schizophrenia 1 interactome: lessons for target identification and disease biology?" *Novartis Foundation Symposium*, vol. 289, pp. 208–216, 2008.
- [66] W. Hennah and D. Porteous, "The DISC1 pathway modulates expression of neurodevelopmental, synaptogenic and sensory perception genes," *PLoS One*, vol. 4, no. 3, Article ID e4906, 2009.
- [67] S. Rossner, U. Ueberham, R. Schlichs, J. R. Perez-Polo, and V. Bigl, "Neurotrophin binding to the p75 neurotrophin receptor is necessary but not sufficient to mediate NGFeffects on APP secretion in PC-12 cells," *Journal of Neural Transmission, Supplement*, no. 54, pp. 279–285, 1998.
- [68] A. Katzourakis and R. J. Gifford, "Endogenous viral elements in animal genomes," *PLoS Genetics*, vol. 6, no. 11, Article ID e1001191, 2010.
- [69] M. B. Geuking, J. Weber, M. Dewannieux et al., "Recombination of retrotransposon and exogenous RNA virus results in nonretroviral cDNA integration," *Science*, vol. 323, no. 5912, pp. 393–396, 2009.
- [70] K. Tanaka-Taya, J. Sashihara, H. Kurahashi et al., "Human herpesvirus 6 (HHV-6) is transmitted from parent to child

in an integrated form and characterization of cases with chromosomally integrated HHV-6 DNA," *Journal of Medical Virology*, vol. 73, no. 3, pp. 465–473, 2004.

- [71] K. Khodosevich, Y. Lebedev, and E. Sverdlov, "Endogenous retroviruses and human evolution," *Comparative and Functional Genomics*, vol. 3, no. 6, pp. 494–498, 2002.
- [72] A. S. Brown, "Prenatal infection as a risk factor for schizophrenia," *Schizophrenia Bulletin*, vol. 32, no. 2, pp. 200–202, 2006.
- [73] K. Ozawa, K. Hashimoto, T. Kishimoto, E. Shimizu, H. Ishikura, and M. Iyo, "Immune activation during pregnancy in mice leads to dopaminergic hyperfunction and cognitive impairment in the offspring: a neurodevelopmental animal model of schizophrenia," *Biological Psychiatry*, vol. 59, no. 6, pp. 546–554, 2006.
- [74] H. G. Kinnell, "Parental age in schizophrenia," *British Journal of Psychiatry*, vol. 142, p. 204, 1983.
- [75] J. Lopez-Castroman, D. D. Gómez, J. J. C. Belloso et al., "Differences in maternal and paternal age between Schizophrenia and other psychiatric disorders," *Schizophrenia Research*, vol. 116, no. 2-3, pp. 184–190, 2010.
- [76] A. S. Brown, P. Cohen, S. Greenwald, and E. Susser, "Nonaffective psychosis after prenatal exposure to rubella," *American Journal of Psychiatry*, vol. 157, no. 3, pp. 438–443, 2000.
- [77] C. Dalman, P. Allebeck, D. Gunnell et al., "Infections in the CNS during childhood and the risk of subsequent psychotic illness: a cohort study of more than one million Swedish subjects," *American Journal of Psychiatry*, vol. 165, no. 1, pp. 59–65, 2008.
- [78] K. M. Prasad, M. N. Bamne, B. H. Shirts et al., "Grey matter changes associated with host genetic variation and exposure to Herpes Simplex Virus 1 (HSV1) in first episode schizophrenia," *Schizophrenia Research*, vol. 118, no. 1–3, pp. 232–239, 2010.
- [79] D. W. Niebuhr, A. M. Millikan, R. Yolken, Y. Li, and N. S. Weber, "Results from a hypothesis generating case-control study: herpes family viruses and schizophrenia among military personnel," *Schizophrenia Bulletin*, vol. 34, no. 6, pp. 1182–1188, 2008.
- [80] A. L. Abrahao, R. Focaccia, and W. F. Gattaz, "Childhood meningitis increases the risk for adult schizophrenia," *World Journal of Biological Psychiatry*, vol. 6, no. 2, pp. 44–48, 2005.
- [81] H. Terayama, Y. Nishino, M. Kishi, K. Ikuta, M. Itoh, and K. Iwahashi, "Detection of anti-Borna Disease Virus (BDV) antibodies from patients with schizophrenia and mood disorders in Japan," *Psychiatry Research*, vol. 120, no. 2, pp. 201–206, 2003.
- [82] S. L. Buka, T. D. Cannon, E. F. Torrey, and R. H. Yolken, "Maternal exposure to herpes simplex virus and risk of psychosis among adult offspring," *Biological Psychiatry*, vol. 63, no. 8, pp. 809–815, 2008.
- [83] E. G. Severance, F. B. Dickerson, R. P. Viscidi et al., "Coronavirus immunoreactivity in individuals with a recent onset of psychotic symptoms," *Schizophrenia Bulletin*, vol. 37, no. 1, pp. 101–107, 2011.
- [84] L. M. Ellman, R. H. Yolken, S. L. Buka, E. F. Torrey, and T. D. Cannon, "Cognitive functioning prior to the onset of psychosis: the role of fetal exposure to serologically determined influenza infection," *Biological Psychiatry*, vol. 65, no. 12, pp. 1040–1047, 2009.
- [85] Y. Yao, J. Schröder, C. Nellåker et al., "Elevated levels of human endogenous retrovirus-W transcripts in blood cells from patients with first episode schizophrenia," *Genes, Brain* and Behavior, vol. 7, no. 1, pp. 103–112, 2008.

- [86] A. S. Brown, C. A. Schaefer, C. P. Quesenberry, L. Liu, V. P. Babulas, and E. S. Susser, "Maternal exposure to toxoplasmosis and risk of schizophrenia in adult offspring," *American Journal of Psychiatry*, vol. 162, no. 4, pp. 767–773, 2005.
- [87] F. Dickerson, C. Stallings, A. Origoni, C. Copp, S. Khushalani, and R. Yolken, "Antibodies to measles in individuals with recent onset psychosis," *Schizophrenia Research*, vol. 119, no. 1–3, pp. 89–94, 2010.
- [88] G. P. Amminger, P. D. McGorry, G. E. Berger et al., "Antibodies to infectious agents in individuals at ultra-high risk for psychosis," *Biological Psychiatry*, vol. 61, no. 10, pp. 1215– 1217, 2007.
- [89] M. J. Chandley, M. N. Miller, C. N. Kwasigroch, T. D. Wilson, and B. E. Miller, "Increased antibodies for the α7 subunit of the nicotinic receptor in schizophrenia," *Schizophrenia Research*, vol. 109, no. 1–3, pp. 98–101, 2009.
- [90] T. Borda, R. Gomez, M. I. Berría, and L. Sterin-Borda, "Antibodies against astrocyte M1 and M2 muscarinic cholinoceptor from schizophrenic patients' sera," *Glia*, vol. 45, no. 2, pp. 144–154, 2004.
- [91] Y. Suzuki, T. Kurita, K. Sakurai, Y. Takeda, and T. Koyama, "Case report of anti-NMDA receptor encephalitis suspected of schizophrenia," *Seishin Shinkeigaku Zasshi*, vol. 111, no. 12, pp. 1479–1484, 2009.
- [92] I. V. Shcherbakova, T. M. Siryachenko, N. A. Mazaeva, V. G. Kaleda, S. A. Krasnolobova, and T. P. Klyushnik, "Leukocyte elastase and autoantibodies to nerve growth factor in the acute phase of schizophrenia and their relationship to symptomatology," *World Journal of Biological Psychiatry*, vol. 5, no. 3, pp. 143–148, 2004.
- [93] S. Tanaka, H. Matsunaga, M. Kimura et al., "Autoantibodies against four kinds of neurotransmitter receptors in psychiatric disorders," *Journal of Neuroimmunology*, vol. 141, no. 1-2, pp. 155–164, 2003.
- [94] X. F. Wang, D. Wang, W. Zhu, K. K. Delrahim, D. Dolnak, and M. H. Rapaport, "Studies characterizing 60 kDa autoantibodies in subjects with schizophrenia," *Biological Psychiatry*, vol. 53, no. 5, pp. 361–375, 2003.
- [95] J. J. Kim, S. J. Lee, K. Y. Toh, C. U. Lee, C. Lee, and I. H. Paik, "Identification of antibodies to heat shock proteins 90 kDa and 70 kDa in patients with schizophrenia," *Schizophrenia Research*, vol. 52, no. 1-2, pp. 127–135, 2001.
- [96] A. B. Poletaev, S. G. Morozov, B. B. Gnedenko, V. M. Zlunikin, and D. A. Korzhenevskey, "Serum anti-S100b, anti-GFAP and anti-NGF autoantibodies of IgG class in healthy persons and patients with mental and neurological disorders," *Autoimmunity*, vol. 32, no. 1, pp. 33–38, 2000.
- [97] A. Chatr-Aryamontri, A. Ceol, D. Peluso et al., "VirusMINT: a viral protein interaction database," *Nucleic Acids Research*, vol. 37, no. 1, pp. D669–D673, 2009.
- [98] C. J. Carter, "Herpes simplex: host viral protein interactions," WikiGenes, 2010.
- [99] S. Lan, H. Wang, H. Jiang et al., "Direct interaction between α-actinin and hepatitis C virus NS5B," *FEBS Letters*, vol. 554, no. 3, pp. 289–294, 2003.
- [100] E. Forgacs, S. K. Gupta, J. A. Kerry, and O. J. Semmes, "The bZIP transcription factor ATFx binds human T-cell leukemia virus type 1 (HTLV-1) tax and represses HTLV-1 long terminal repeat-mediated transcription," *Journal of Virology*, vol. 79, no. 11, pp. 6932–6939, 2005.
- [101] S. V. Indran, M. E. Ballestas, and W. J. Britt, "Bicaudal D1dependent trafficking of human cytomegalovirus tegument protein pp150 in virus-infected cells," *Journal of Virology*, vol. 84, no. 7, pp. 3162–3177, 2010.

- [102] Y. P. Huang, J. Cheng, S. L. Zhang et al., "Screening of hepatocyte proteins binding to F protein of hepatitis C virus by yeast two-hybrid system," *World Journal of Gastroenterology*, vol. 11, no. 36, pp. 5659–5665, 2005.
- [103] K. Radtke, D. Kieneke, A. Wolfstein et al., "Plus- and minusend directed microtubule motors bind simultaneously to herpes simplex virus capsids using different inner tegument structures," *PLoS Pathogens*, vol. 6, no. 7, Article ID e1000991, 2010.
- [104] K. H. Bremner, J. Scherer, J. Yi, M. Vershinin, S. P. Gross, and R. B. Vallee, "Adenovirus transport via direct interaction of cytoplasmic dynein with the viral capsid hexon subunit," *Cell Host and Microbe*, vol. 6, no. 6, pp. 523–535, 2010.
- [105] H. Si, S. C. Verma, M. A. Lampson, Q. Cai, and E. S. Robertson, "Kaposi's sarcoma-associated herpesvirusencoded LANA can interact with the nuclear mitotic apparatus protein to regulate genome maintenance and segregation," *Journal of Virology*, vol. 82, no. 13, pp. 6734–6746, 2008.
- [106] H. N. Ramanathan, D. H. Chung, S. J. Plane et al., "Dyneindependent transport of the Hantaan virus nucleocapsid protein to the endoplasmic reticulum-Golgi intermediate compartment," *Journal of Virology*, vol. 81, no. 16, pp. 8634– 8647, 2007.
- [107] B. Schramm, C. A. M. de Haan, J. Young et al., "Vacciniavirus-induced cellular contractility facilitates the subcellular localization of the viral replication sites," *Traffic*, vol. 7, no. 10, pp. 1352–1367, 2006.
- [108] I. J. Dorweiler, S. J. Ruone, H. Wang, R. W. Burry, and L. M. Mansky, "Role of the human T-cell leukemia virus type 1 PTAP motif in Gag targeting and particle release," *Journal* of Virology, vol. 80, no. 7, pp. 3634–3643, 2006.
- [109] K. Kobayashi, T. Sueyoshi, K. Inoue, R. Moore, and M. Negishi, "Cytoplasmic accumulation of the nuclear receptor CAR by a tetratricopeptide repeat protein in HepG2 cells," *Molecular Pharmacology*, vol. 64, no. 5, pp. 1069–1075, 2003.
- [110] K. H. Bremner, J. Scherer, J. Yi, M. Vershinin, S. P. Gross, and R. B. Vallee, "Adenovirus transport via direct interaction of cytoplasmic dynein with the viral capsid hexon subunit," *Cell Host and Microbe*, vol. 6, no. 6, pp. 523–535, 2010.
- [111] E. Johannsen, M. Luftig, M. R. Chase et al., "Proteins of purified Epstein-Barr virus," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 46, pp. 16286–16291, 2004.
- [112] A. J. Collier, J. Gallego, R. Klinck et al., "A conserved RNA structure within the HCV IRES eIF3-binding site," *Nature Structural Biology*, vol. 9, no. 5, pp. 375–380, 2002.
- [113] H. Sawa, T. Suzuki, Y. Orba, Y. Sunden, and K. Nagashima, "Recent research on the JC virus," *Brain and Nerve*, vol. 59, no. 2, pp. 101–108, 2007.
- [114] R. M. Vos, J. Altreuter, E. A. White, and P. M. Howley, "The ubiquitin-specific peptidase USP15 regulates human papillomavirus type 16 E6 protein stability," *Journal of Virology*, vol. 83, no. 17, pp. 8885–8892, 2009.
- [115] R. König, Y. Zhou, D. Elleder et al., "Global analysis of host-pathogen interactions that regulate early-stage HIV-1 replication," *Cell*, vol. 135, no. 1, pp. 49–60, 2008.
- [116] A. A. Kondratova, N. Neznanov, R. V. Kondratov, and A. V. Gudkov, "Poliovirus protein 3A binds and inactivates LIS1, causing block of membrane protein trafficking and deregulation of cell division," *Cell Cycle*, vol. 4, no. 10, pp. 1403–1410, 2005.
- [117] N. Epie, T. Ammosova, T. Sapir et al., "HIV-1 Tat interacts with LIS1 protein," *Retrovirology*, vol. 2, 2005.

- [118] Y. Verdier and B. Penke, "Binding sites of amyloid β-peptide in cell plasma membrane and implications for Alzheimer's disease," *Current Protein and Peptide Science*, vol. 5, no. 1, pp. 19–31, 2004.
- [119] I. Bossis, R. B. S. Roden, R. Gambhira et al., "Interaction of tSNARE syntaxin 18 with the papillomavirus minor capsid protein mediates infection," *Journal of Virology*, vol. 79, no. 11, pp. 6723–6731, 2005.
- [120] C. R. Brunetti, K. S. Dingwell, C. Wale, F. L. Graham, and D. C. Johnson, "Herpes simplex virus gD and virions accumulate in endosomes by mannose 6-phosphate-dependent and -independent mechanisms," *Journal of Virology*, vol. 72, no. 4, pp. 3330–3339, 1998.
- [121] D. T. Balu and J. T. Coyle, "Neuroplasticity signaling pathways linked to the pathophysiology of schizophrenia," *Neuro-science and Biobehavioral Reviews*, vol. 35, no. 3, pp. 848–870, 2011.
- [122] S. J. Flint, L. W. Enquist, V. R. Racaniello, and A. M. Skalka, *Principles of Virology*, ASM Press, Herndon, Va, USA, 2008.
- [123] M. R. Rosenberg and M. G. Casarotto, "Coexistence of two adamantane binding sites in the influenza A M2 ion channel," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 31, pp. 13866–13871, 2010.
- [124] J. Toma, J. M. Whitcomb, C. J. Petropoulos, and W. Huang, "Dual-tropic HIV type 1 isolates vary dramatically in their utilization of CCR5 and CXCR4 coreceptors," *AIDS*, vol. 24, no. 14, pp. 2181–2186, 2010.
- [125] C. J. Carter, "Interactions between the products of the Herpes simplex genome and Alzheimer's disease susceptibility genes: relevance to pathological-signalling cascades," *Neurochemistry International*, vol. 52, no. 6, pp. 920–934, 2008.
- [126] K. Honjo, R. van Reekum, and N. P. L. G. Verhoeff, "Alzheimer's disease and infection: do infectious agents contribute to progression of Alzheimer's disease?" *Alzheimer's* and Dementia, vol. 5, no. 4, pp. 348–360, 2009.
- [127] R. F. Itzhaki, M. A. Wozniak, D. M. Appelt, and B. J. Balin, "Infiltration of the brain by pathogens causes Alzheimer's disease," *Neurobiology of Aging*, vol. 25, no. 5, pp. 619–627, 2004.
- [128] C. J. Carter, "Alzheimer's disease: a pathogenetic autoimmune disorder caused by herpes simplex in a gene-dependent manner," *International Journal of Alzheimer's Disease*, vol. 2010, Article ID 140539, 17 pages, 2010.
- [129] S. A. Chance, M. M. Esiri, and T. J. Crow, "Ventricular enlargement in schizophrenia: a primary change in the temporal lobe?" *Schizophrenia Research*, vol. 62, no. 1-2, pp. 123–131, 2003.
- [130] W. Hennah and D. Porteous, "The DISC1 pathway modulates expression of neurodevelopmental, synaptogenic and sensory perception genes," *PLoS One*, vol. 4, no. 3, Article ID e4906, 2009.
- [131] L. Desbonnet, J. L. Waddington, and C. M. P. O'Tuathaigh, "Mutant models for genes associated with schizophrenia," *Biochemical Society Transactions*, vol. 37, no. 1, pp. 308–312, 2009.
- [132] H. Jaaro-Peled, "Gene models of schizophrenia: DISC1 mouse models," *Progress in Brain Research*, vol. 179, pp. 75– 86, 2009.
- [133] N. Sakae, N. Yamasaki, K. Kitaichi et al., "Mice lacking the schizophrenia-associated protein FEZ1 manifest hyperactivity and enhanced responsiveness to psychostimulants," *Human Molecular Genetics*, vol. 17, no. 20, pp. 3191–3203, 2008.

- [134] C. S. Barros, B. Calabrese, P. Chamero et al., "Impaired maturation of dendritic spines without disorganization of cortical cell layers in mice lacking NRG1/ErbB signaling in the central nervous system," *Proceedings of the National Academy* of Sciences of the United States of America, vol. 106, no. 11, pp. 4507–4512, 2009.
- [135] D. Babovic, C. M. O'Tuathaigh, A. M. O'Connor et al., "Phenotypic characterization of cognition and social behavior in mice with heterozygous versus homozygous deletion of catechol-O-methyltransferase," *Neuroscience*, vol. 155, no. 4, pp. 1021–1029, 2008.
- [136] W. W. Eaton, M. Byrne, H. Ewald et al., "Association of schizophrenia and autoimmune diseases: linkage of Danish national registers," *American Journal of Psychiatry*, vol. 163, no. 3, pp. 521–528, 2006.
- [137] H. Rosenmann, N. Grigoriadis, D. Karussis et al., "Tauopathy-like abnormalities and neurologic deficits in mice immunized with neuronal tau protein," *Archives of Neurology*, vol. 63, no. 10, pp. 1459–1467, 2006.
- [138] J. E. Black, I. M. Kodish, A. W. Grossman et al., "Pathology of layer V pyramidal neurons in the prefrontal cortex of patients with schizophrenia," *American Journal of Psychiatry*, vol. 161, no. 4, pp. 742–744, 2004.
- [139] N. S. Kolomeets and N. A. Uranova, "Synaptic contacts in schizophrenia: the study with immunocytochemical identification of dopaminergic neurons [Sinapticheskie kontakty pri shizofrenii: issledovaniie s immunotsitokhimicheskoi identifikatsiei dofaminergicheskikh neironov]," *Zhurnal Nevropatolgii i Psikhiatrii im. S S Korsakova*, vol. 97, no. 12, pp. 39–43, 1997.
- [140] P. M. Thompson, C. Vidal, J. N. Giedd et al., "Mapping adolescent brain change reveals dynamic wave of accelerated gray matter loss in very early-onset schizophrenia," *Proceedings* of the National Academy of Sciences of the United States of America, vol. 98, no. 20, pp. 11650–11655, 2001.
- [141] T. A. Bayer, R. Buslei, L. Havas, and P. Falkai, "Evidence for activation of microglia in patients with psychiatric illnesses," *Neuroscience Letters*, vol. 271, no. 2, pp. 126–128, 1999.
- [142] R. J. Holden and I. S. Pakula, "Immunological influences in attention-deficit disorder and schizophrenia: is there a link between these two conditions?" *Medical Hypotheses*, vol. 45, no. 6, pp. 575–587, 1995.
- [143] J. Söderlund, J. Schröder, C. Nordin et al., "Activation of brain interleukin-1β in schizophrenia," *Molecular Psychiatry*, vol. 14, no. 12, pp. 1069–1071, 2009.
- [144] Ł. Drzyzga, E. Obuchowicz, A. Marcinowska, and Z. S. Herman, "Cytokines in schizophrenia and the effects of antipsychotic drugs," *Brain, Behavior, and Immunity*, vol. 20, no. 6, pp. 532–545, 2006.
- [145] R. S. Smith and M. Maes, "The macrophage-T-lymphocyte theory of schizophrenia: additional evidence," *Medical Hypotheses*, vol. 45, no. 2, pp. 135–141, 1995.
- [146] M. P. Vawter, O. Dillon-Carter, F. Issa, R. J. Wyatt, and W. J. Freed, "Transforming growth factors β1 and β2 in the cerebrospinal fluid of chronic schizophrenic patients," *Neuropsychopharmacology*, vol. 16, no. 1, pp. 83–87, 1997.
- [147] R. Rajapakse, B. Uring-Lambert, K. L. Andarawewa et al., "1,25(OH)2D3 inhibits in vitro and in vivo intracellular growth of apicomplexan parasite *Toxoplasma gondii*," *Journal* of Steroid Biochemistry and Molecular Biology, vol. 103, no. 3-5, pp. 811–814, 2007.
- [148] J. J. McGrath, D. W. Eyles, C. B. Pedersen et al., "Neonatal vitamin D status and risk of schizophrenia: a populationbased case-control study," *Archives of General Psychiatry*, vol. 67, no. 9, pp. 889–894, 2010.

- [149] D. Kanduc, A. Stufano, G. Lucchese, and A. Kusalik, "Massive peptide sharing between viral and human proteomes," *Peptides*, vol. 29, no. 10, pp. 1755–1766, 2008.
- [150] D. E. Dietrich and L. Bode, "Human Borna disease virusinfection and its therapy in affective disorders," *APMIS*, vol. 116, no. 124, pp. 61–65, 2008.
- [151] E. Fazzini, J. Fleming, and S. Fahn, "Cerebrospinal fluid antibodies to coronavirus in patients with Parkinson's disease," *Movement Disorders*, vol. 7, no. 2, pp. 153–158, 1992.
- [152] B. A. Bagert, "Epstein-Barr virus in multiple sclerosis," *Current Neurology and Neuroscience Reports*, vol. 9, no. 5, pp. 405–410, 2009.