## REVIEW



# Tumor metastasis: Mechanistic insights and therapeutic interventions

Mengmeng Liu<sup>1,2</sup> | Jing Yang<sup>1,2</sup> | Bushu Xu<sup>1,2</sup> | Xing Zhang<sup>1</sup>

<sup>1</sup> Melanoma and Sarcoma Medical Oncology Unit, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou, China

<sup>2</sup> State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou, China

#### Correspondence

Xing Zhang, Sun Yat-sen University Cancer Center, 651 Dongfeng Road East, Guangzhou 510060, China. Email: zhangxing@sysucc.org.cn

Mengmeng Liu and Jing Yang contributed equally to this work.

#### **Funding information**

National Natural Science Foundation of China, Grant/Award Numbers: 81772863, 82072958; China Postdoctoral Science Foundation, Grant/Award Number: 2020M683119

#### Abstract

Cancer metastasis is responsible for the vast majority of cancer-related deaths worldwide. In contrast to numerous discoveries that reveal the detailed mechanisms leading to the formation of the primary tumor, the biological underpinnings of the metastatic disease remain poorly understood. Cancer metastasis is a complex process in which cancer cells escape from the primary tumor, settle, and grow at other parts of the body. Epithelial-mesenchymal transition and anoikis resistance of tumor cells are the main forces to promote metastasis, and multiple components in the tumor microenvironment and their complicated crosstalk with cancer cells are closely involved in distant metastasis. In addition to the three cornerstones of tumor treatment, surgery, chemotherapy, and radiotherapy, novel treatment approaches including targeted therapy and immunotherapy have been established in patients with metastatic cancer. Although the cancer survival rate has been greatly improved over the years, it is still far from satisfactory. In this review, we provided an overview of the metastasis process, summarized the cellular and molecular mechanisms involved in the dissemination and distant metastasis of cancer cells, and reviewed the important advances in interventions for cancer metastasis.

#### **KEYWORDS**

cancer, epithelial-mesenchymal transition, immunotherapy, metastasis, targeted therapy, tumor microenvironment

# **1** | INTRODUCTION

Cancer is one of the main diseases threatening human health. According to the Global Cancer Statistics, 2020, there were 18.1 million new cancer cases and 9.6 million cancer-related deaths worldwide.<sup>1</sup> Despite the continuous development of medical technology, distant metastasis has already appeared at the time of diagnosis for many

patients. Furthermore, a large number of cancer patients, both early- and late-stage, may eventually develop metastatic diseases.<sup>2</sup> Metastatic lesions in distant organs are difficult to be cured by current therapeutic approaches, and metastasis accounts for about 90% of death in cancer patients.<sup>3</sup> In other words, the overwhelming problem highlighted by the cancer-associated deaths is, for the most part, metastatic cancer. In order to develop the

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

<sup>© 2021</sup> The Authors. MedComm published by Sichuan International Medical Exchange & Promotion Association (SCIMEA) and John Wiley & Sons Australia, Ltd.

methods for preventing or treating metastasis, it is inevitable to understand the cellular and molecular mechanisms of metastasis.

In contrast to numerous discoveries that reveal the detailed mechanisms leading to the formation of the primary tumor, research on metastatic cancer is still lagging behind. Stephen Paget and James Ewing proposed the "seed and soils" and the "mechanical metastasis" hypothesis, respectively, which laid the foundation for the study of tumor metastasis.<sup>4–6</sup> Cancer metastasis is a complicated process, which is regulated by various signaling pathways and modulated by the surrounding extracellular matrix (ECM). Tumor cells spread from the primary tumor mass to distant organs through blood vessels, lymphatic vessels, and transcoelomic routes.<sup>7,8</sup> In order to successfully metastasize from the prior site, tumor cells need to go through the following stages: local invasion, intravasation, survival in circulation, extravasation, and colonization.<sup>9</sup> Furthermore, multiple components in the tumor microenvironment (TME), including immune cells, stromal cells, chemokines, and cytokines are involved in a complex crosstalk with tumor cells that affects tumor growth and metastasis.<sup>10–12</sup> Surgery, chemotherapy, and radiotherapy are the three cornerstones of cancer treatment. With the deepening understanding of mechanisms of tumorigenesis and metastasis in recent years, a plethora of treatment approaches, including targeted therapy and immunotherapy have been established.<sup>13–15</sup> However, the underlying mechanisms of cancer metastasis are not yet fully understood, and the strategies for preventing and inhibiting cancer metastasis are also limited.

Here, we provided an overview of the metastasis process, summarized the mechanisms underlying the dissemination and distant metastasis of cancer cells, and reviewed the important advances in interventions targeting cancer metastasis.

# 2 | COMPONENTS AND MECHANISMS INVOLVED IN METASTASIS

Tumor metastasis is one of the hallmarks of tumor malignancy and one of the causes of tumor-related death.<sup>16</sup> The malignant behavior of tumor metastasis is mainly related to the malignant degree of the primary tumor, but one of the common features is that all metastases need to go through a cascade called "invasion-metastasis cascade."<sup>17</sup> The metastasis process begins when the tumor cells gain invasiveness and lose their adhesion to the surrounding matrix including basement membrane (BM) and ECM; tumor cells migrate out of the primary tumor and invade surrounding tissues.<sup>18</sup> LIU ET AL.

Subsequently, disseminated tumor cells penetrate blood vessels or lymph vessels into the circulation and respond to various resistance conditions such as shear force, anoikis, and immune surveillance in the circulation.<sup>19</sup> Only a small percentage of tumor cells can survive in the harsh conditions of circulation. After successfully entering the secondary site, tumor cells adhere to the endothelium of the target organ, and exudate and migrate into the organ parenchyma, which is the "pre-metastatic niche."<sup>20–22</sup> They either enter a long-term dormant state in the form of a single cell, or enter micrometastasis in the form of multiple cells, and finally begin to grow continuously to form clinical metastases (Figure 1).<sup>23,24</sup>

At the beginning of the whole metastasis process, the most important thing is that there are a group of invasive and plastic tumor cells in the cancer nest, which are collectively referred to as metastasis initiating cells.<sup>25</sup> Cancer stem cells (CSCs) have been proved to have the above characteristics and play a crucial role in tumor metastasis.<sup>26,27</sup> In addition to the role of the tumor itself, metastasis is induced by various factors, including genetic alation, abnormal epigenetic modifications, immune escape, and changes in the growth environment.<sup>9</sup> Here, we will describe in detail the major factors and mechanisms involved in the metastasis cascade.

# 2.1 | Key driver of metastasis: Epithelial-mesenchymal transition (EMT)

The EMT is the transition of epithelial cells to mesenchymal cells under certain physiological and pathological conditions, which was proposed by Greenberg and Hay in 1982.<sup>28–30</sup> Studies in the last decades have shown that EMT is an important molecular event for epithelial tumor cells to gain invasiveness and plays an important role in the development and metastatic dissemination of malignant tumor cells.<sup>31</sup> Tumor cells can obtain a higher mesenchymal phenotype through EMT, which manifests as reduced contact with surrounding cells and matrix, and enhanced cell migration and motility at the beginning of tumor cell invasion and metastasis.<sup>30</sup> Therefore, interrupting or reversing EMT can inhibit the invasion of malignant tumor cells and reduce the rate of tumor metastasis.<sup>32</sup> The process of EMT is shown in Figure 2.

Cancer cells undergoing and molecular EMT exhibit morphological changes, such as decreased expression of epithelial markers (e.g., E-cadherin, zonula occludens-1, and occluding) and increased expression of mesenchymal markers (e.g., N-cadherin protein, fibroblast-specific protein 1, and fibronectin).<sup>33,34</sup> Cell adhesion molecules, such as E-cadherin, N- cadherin, and  $\beta$ -catenin, are closely

# Wetastatic tumor Tumor cell CAF NK cell MSC NK cell MSC Neutrophil Tumor Tumor Primary tumor Tumor Exosomes Premetastatic niche Proliferation Angiogenesis Dissemination and distant metastasis Tropic sites of metastasis Tropic sites of metastasis (Acquisition of invasive potential) (EMT, adhesion, invasion, and survival in circulation) (Lung, liver, bone and brain)

**FIGURE 1** Overview of the metastatic cascade. Carcinoma cells escaping from the primary tumor migrate and invade through the basement membrane and extracellular matrix, enter the blood or lymphatic vessels, intravasate into the circulation, penetrate the blood or lymphatic vessels (extravasation), and adhere and grow in secondary sites. A variety of stromal cells, immune cells, and other molecular components surrounding the tumor provide signals that enhance the metastatic potential of cancer cells. Platelets and neutrophils can protect tumor cells by providing physical protection against shear stress, secreting mediators (such as transforming growth factor-beta (TGF- $\beta$ )), neutralizing the cytotoxicity of NK cells and favoring immune escape. Abbreviations: BM, basement membrane; CAF, cancer-associated fibroblast; CTC, circulating tumor cell; ECM, extracellular matrix; EMT, epithelial-to-mesenchymal transition; MSC, mesenchymal stem cell; NK cell, natural killer cell; RBC, red blood cell; TAM, tumor-associated macrophage

related to EMT and are regulated by EMT-related transcription factors (EMT-TFs), such as zinc finger E-boxbinding homeobox 1/2 (ZEB1/2), snai1/2 and twist.<sup>35–37</sup> The down-regulation of E-cadherin is accompanied by the upregulation of N-cadherin, which reduces the adhesion of cancer cells to epithelial cells and increases their adhesion to stromal cells, leading to subsequent invasion of tumor cells into matrix.<sup>38</sup>  $\beta$ -catenin has also been found to promote tumor metastasis, mainly driven by the ectopic expression of  $\beta$ -catenin.<sup>39</sup> Vimentin has been found to promote cell invasion by regulating the E-cadherin/ $\beta$ -catenin complex.<sup>40</sup> EMT-TFs can be regulated by signaling pathways such as Wnt/ $\beta$ -catenin, transforming growth factorbeta (TGF- $\beta$ ), and Notch; these factors work together to give tumor cells the characteristics required for metastasis under different conditions, thereby promoting the occurrence of metastasis.41-45

EMT is also involved in the process of tumor invasion and colonization in metastasis, which mainly depends on circulating tumor cells (CTCs) and CSCs.<sup>41,46,47</sup> CTCs are divided into three groups, including epithelial CTCs, hybrid epithelial/mesenchymal phenotype CTCs, and mesenchymal CTCs.<sup>48</sup> With the development of modern medical technology, tumor metastasis can be monitored throughout the process. Compared with epithelial CTCs, mesenchymal CTCs may be easier to metastasize because they are resistant to anoikis and chemotherapy and have a stronger ability to migrate to distant organs.<sup>49,50</sup> Previous studies reported that up-regulating the expression of EMT-TFs in breast cancer cells increased the expression of CSC-specific cell markers, improved the ability to form spheroids, and accelerated tumor formation of breast cancer cells in mice.<sup>51</sup> In non-CSCs, the activation of EMT promotes their conversion to CSCs.<sup>51–54</sup> Tumor formation and metastasis rely on the tumor-forming ability of tumor cells, which suggests that CSCs with tumor-initiating ability are the crucial prerequisite for disseminated cancer cells to establish metastatic colonies.<sup>55</sup>

MedComm

Another mechanism of EMT promoting metastasis is to protect tumor cells from immune cell-mediated killing.<sup>56–58</sup> Recent researches on breast cancer found that, compared with epithelial cancer cell lines, mesenchymal breast cancer cell lines can recruit more immunosuppressive regulatory T cells (Tregs) and M2 macrophages.<sup>57</sup> Compared with epithelial cancer cells, mesenchymal cancer cells are less responsive to anti-cytotoxic lymphocyteassociated protein 4 (CTLA4) immunotherapy.<sup>59</sup> In addition, the expressions of programmed death ligand-1 (PD-L1), PD-L2, and B7 homolog 3 (B7-H3) are up-regulated in tumor cells that have undergone EMT, which can facilitate the immune escape of cancer cells.<sup>58,60</sup>

During dormancy and colonization, the process of mesenchymal cells to transform to epithelial cells (MET) is required to help tumor cells survive in the foreign microenvironment.<sup>31</sup> Indeed, the role of MET in promoting tumor cell colonization at a secondary site during metastatic growth has been confirmed in numerous studies.<sup>31,61</sup> It is worth noting that the tightly interconnected transfer network of EMT and MET has been proved to be subject to epigenetic regulation mediated by DNA



**FIGURE 2** Cancer cells undergo EMT and invade into circulation. (A) A single transformed epithelial cell remains quiescent for a period of time. (B, C). The transformed cells proliferate and generate a small intraepithelial colony, accompanied by the formation of cancer stem cells. Cancer cells destroy the basement membrane, undergo EMT, and migrate and invade through the basement membrane and extracellular matrix. Normal extracellular matrix undergoes cancer-associated remodel. Meanwhile, cells and molecular components in tumor microenvironment (TME; CAFs, TAMs, neutrophils, MSCs...) surrounding the primary tumor enhance cancer cell survival, proliferation and metastasis. (D) Cancer cells escaping from primary tumors can invade into the circulation as single CTCs or multicellular CTC clusters. Abbreviations: BM, basement membrane; CAF, cancer-associated fibroblast; CTC, circulating tumor cell; EC, endothelial cell; ECM, extracellular matrix; EMT, epithelial-to-mesenchymal transition; MSC, mesenchymal stem cell; TAM, tumor-associated macrophage

methylation and histone modification.<sup>62–64</sup> For example, in breast cancer, histone methyltransferase nuclear receptor-binding SET domain 3 (NSD3) cooperates with enhancer of zeste homolog 2 (EZH2) and RNA polymerase II to stimulate the Notch pathway-mediated E-cadherin transcriptional inhibition to promote EMT and trigger tumor invasion and metastasis.<sup>65</sup> During the initiation of EMT, histone demethylase lysine (K) -specific demethylase 6A (KDM6A) is inhibited, leading to the transcriptional inhibition of epithelial genes, whereas during the MET process, the expression of KDM6A is restored, thereby reactivating the epithelial gene by the trimethylation of histone 3 lysine 27 (H3K27me3) and reversing the phenotype.<sup>66,67</sup> Therefore, in general, a series of epigenetic regulation of tumor cells makes EMT dynamically

change, giving tumor cells the ability to survive in different microenvironments.

#### 2.2 | The most important soil factor: TME

In addition to the characteristics of tumor cells, numerous studies have shown that TME is the soil for tumor cells and provides the necessary energy for tumor growth (Figure 3).<sup>68</sup> In the past two decades, many achievements have been made regarding the impact of TME on cancer metastasis.<sup>10,11,69,70</sup> TME can promote the occurrence and development of tumor by affecting metabolism, secretion, immunity, structure, and function of tumor cells, thereby playing an essential role in the overall





**FIGURE 3** TME involved in the processes of invasion-metastasis cascade. The cellular components in TME can be classified into cancer cells, stromal cells, and immune cells. These cells interact with each other through ligand-receptor interactions, and the secretion of cytokines, chemokines, exosomes, and extracellular vesicles, forming an evolving microenvironment. Cancer cells that are good at recruiting and establishing a supportive metastatic niche may be able to survive and initiate the process of proliferation and metastasis. The formation of the metastatic niche may occur before the arrival of cancer cells, also known as pre-metastatic niches. Here, we summarized the role of important cellular components in TME in tumor metastasis

cascade of tumor metastasis.<sup>12,71,72</sup> The members in TME are generally classified into two categories: cellular components and extracellular components. Cellular components mainly include immune cells, cancer-associated fibroblasts (CAFs), mesenchymal stem cells (MSCs), and endothelial cells.<sup>73–75</sup> Extracellular components, such as tumor-secreted extracellular vesicles (EVs), growth factors, chemokines, and cytokines are also closely involved in tumor metastasis.<sup>76</sup> Cellular components and extracellular components restrict and influence each other, which form a mutual feedback network.

# 2.2.1 | Immune cells

Under normal circumstances, the immune system will recognize foreign antigens to initiate autoimmunity and passive immunity to eliminate harmful pathogens. For this reason, tumor immunity has become a hot spot in the field of tumor research. Studies have pointed out that some types of immune cells are closely related to tumor occurrence and development and play an essential role in tumor metastasis.<sup>72,77</sup> Among the immune cells in TME, tumorassociated macrophages (TAMs), Tregs, and neutrophils are noteworthy for their roles in tumor development and metastasis.

#### TAMs

According to the expression of specific markers, differentiation status, and functional role in the immune system, macrophages are conventionally classified into two major phenotypes, M1 and M2, which can be converted to each other.<sup>78</sup> M1 macrophages promote inflammation responses against tumor cells, whereas M2 macrophages tend to exert immune suppressive effects. M2 macrophages (M2-TAMs) express abundant arginase-1, mannose receptor, and scavenger receptors, and secrete a large number of anti-inflammatory cytokines such as interleukin-4 (IL-4), interleukin-10 (IL-10), and interleukin-13 (IL-13).<sup>79,80</sup> In general, researchers tend to consider TAMs as M2-like phenotype-acquired macrophages.<sup>81</sup> The cytokines and chemokines secreted by tumor cells or fibroblasts could recruit more M2-TAMs in TME.<sup>82-84</sup> TAMs are elevated in TME and are associated with poor clinical prognoses.<sup>85</sup>

MedComm

The role of TAMs in promoting metastasis involves multiple mechanisms. First, TAMs participate in the regulation of the EMT process by enhancing the expression of Ncadherin and snail, secreting various soluble factors such as IL-1 $\beta$ , IL-8, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), and TGF- $\beta$ , and secreting a number of proteolytic enzymes.<sup>86–88</sup> Second, TAMs promote vascularization of tumor cells through stimulating the formation of new tumor vessels and the remodeling of the established vascular system into a more tortuous and leaky form.<sup>89,90</sup> Third, TAMs promote intravasation of tumor cell, favor tumor cell survival in the circulation, and promote extravasation of tumor cells.<sup>91</sup> Last, TAMs are important determinants for the formation of pre-metastatic niches. Notably, TAMs contribute to the immune escape of tumor cells throughout the metastasis process.81

# Tregs

There are mounting studies that convincingly demonstrated that Tregs play a prominent role in promoting metastasis. Tregs have a strong immunosuppressive function in TME by inhibiting adaptive and innate immune responses.<sup>92–96</sup> Here, we summarized the main mechanisms.<sup>97</sup> First, Tregs secrete inhibitory cytokines, including IL-10, TGF- $\beta$ , and IL-35.<sup>98-100</sup> Tregs can inhibit the function of CD8+ T cells and dendritic cells (DCs) through membrane-bound TGF- $\beta$ . For example, Tregs impede CD8+T cell-mediated anti-tumor immune responses by inhibiting interleukin-2 (IL-2) production and activating the TGF- $\beta$  signaling pathway.<sup>92</sup> It has been reported that TGF- $\beta$ 1 secreted by Tregs in breast cancer can induce the up-regulation of IL-17Rb and promote tumor lymph node metastasis.<sup>101</sup> Second, Tregs can kill effector cells by granzymes and perforin and diminish the function of T cells and natural killer (NK) cells through inhibitory receptors, such as CTLA-4, lymphocyte activation gene 3 (LAG-3), and programmed cell death 1 (PD-1).<sup>102-104</sup> Tregs can also suppress CD8+ T cells and NK cells secretion of interferon- $\gamma$  (IFN- $\gamma$ ).<sup>105</sup> Olkhanud et al. found that chemokine receptor (CCR) 4+Tregs are mainly recruited at the lung metastasis site in breast cancer, and the Tregs secret  $\beta$ -galactoside-binding proteins to induce the apoptosis of NK cells, thereby promoting tumor metastasis.<sup>95</sup> Third, Tregs affect effector cell functions by interfering with cell metabolism mainly through the following three ways: depriving IL-2, promoting the production of adenosine, and transferring a large number of cyclic adenosine monophosphate to effector T cells.<sup>106–109</sup> Fourth, Treg induces DCs tolerance through the expression of inhibitory receptors CTLA-4 and LAG-3, with the latter further inhibiting T-cell capacity through indoleamine 2,3-dioxygenase.<sup>110,111</sup> Finally, factors produced by myeloid-derived suppressor cells (MDSCs) and Tregs form positive feedback loops to reinforce the suppressive microenvironment.<sup>112–114</sup>

In addition, it has been found that Tregs could secrete vascular endothelial growth factor A (VEGF-A) in metastatic ovarian cancer, thereby promoting angiogenesis and tumor cell dissemination.<sup>115</sup> TGF- $\beta$ 1 secreted by Tregs can promote tumor metastasis by enhancing EMT.<sup>116</sup> Some preclinical studies have shown that Tregs were closely related to lymph node metastasis and peritoneal metastasis and suggested that Treg may be involved in pre-metastasis niche reprogramming to promote metastasis.<sup>101,117</sup> Therefore, eliminating or alleviating the immunosuppressive state of Tregs may suppress the progression of tumor metastasis.

#### Neutrophils

Neutrophils, as innate immune cells, protect the body from infection by foreign microorganisms and promote inflammatory responses under normal conditions.<sup>118</sup> However, it is worth noting that neutrophils have been found to promote tumor metastasis in a variety of ways, including inhibiting anti-tumor T cells and influencing tumor cell invasion.<sup>119</sup> In a mouse model of sarcoma, pleomorphic neutrophils were found to reduce the expression of intercellular adhesion molecule 1 in tumor cells, thereby increasing tumor motility and facilitating the initiation of metastasis.<sup>120</sup> In addition, pro-metastatic neutrophils activate various signaling pathways to promote EMT by secreting pro-EMT-related cytokines, including IL-8 and IL-17A.121,122 In the process of metastasis, neutrophils bind with CTCs in circulation to protect CTCs from immune killing, increase CTCs invasiveness, and promote metastasis.<sup>123</sup> A large number of neutrophils were found in the secondary sites.<sup>124,125</sup> In terms of mechanism, recruited neutrophils promote the formation of pre-metastatic niche and support angiogenesis mainly by forming an immunosuppressive microenvironment and secreting neutrophil elastase.<sup>126</sup>

#### 2.2.2 | Cancer-associated fibroblasts (CAFs)

CAFs are one of the most common fibroblasts in tumor tissues. CAFs are derived from a variety of different fibroblasts, including normal fibroblasts stimulated by exosomes, bone marrow MSCs induced by TGF- $\beta$ , and epithelial cells transformed by EMT.<sup>127–129</sup> Since CAFs originate from many kinds of cells and have apparent heterogeneity, there is no clear marker that can distinguish CAFs from normal fibroblasts. CAFs are the primary source of various factors and enzymes of TME and are one of the critical members involved in tumor metastasis.

In previous studies, it was believed that the role of CAFs in tumor metastasis is mainly manifested in the reconstruction of ECM structure.<sup>130</sup> However, current studies have found that CAFs promote EMT-mediated tumor metastasis in ovarian cancer, bladder cancer, and breast cancer cells in various ways, including the secretion of TGF- $\beta$  and exosomes.<sup>131</sup> The increased autocrine and paracrine TGF- $\beta$  signaling in the mesenchymal transcription factor forkhead box F2 (FOXF2) deficient base-like breast cancer cells induces EMT to mediate tumor metastasis.<sup>132</sup> In turn, TGF- $\beta$  silences FOXF2 expression by up-regulating the post-transcriptional regulator (miR-182-5p) of FOXF2 and promotes breast cancer metastasis.<sup>133</sup> In addition, chemokine (C-X-C motif) ligand 11 (CXCL11) secreted by CAFs promotes the metastasis of liver cancer by up-regulating the expression of circUBAP2.<sup>134</sup> In order to gain a deeper understanding of the role of CAFs in the tumor, Wang et al. analyzed CAFs by single-cell sequencing combined with RNA-sequence and divided CAFs into four types.<sup>136</sup> One of the subtypes has high glycolytic activity and is termed MeCAFs. It was found that pancreatic ductal adenocarcinoma patients with abundant MeCAFs had a higher risk of metastasis but had a significantly better response to immunotherapy.<sup>136</sup> These results indicate that abnormal glycolysis directly or indirectly affects tumor metastasis. CAFs up-regulate carnitine palmitoyltransferase 1A (CPT1A) to cause reduction of fatty acid oxidation, leading to peritoneal metastasis of colorectal cancer.<sup>137</sup>

Fibroblasts play an important role in both the primary tumor site and the secondary tumor site. Fibroblasts distributed in the metastatic site are called metastasisassociated fibroblasts (MAFs), which promote angiogenesis and the formation of immunosuppressive microenvironment in pre-metastatic niche.<sup>138</sup> The main difference between MAFs and primary tumor-related fibroblasts is that MAFs have a stronger ability to inhibit the antitumor effect of immune cells, which is reflected explicitly in the higher levels of CC motif chemokines ligand 2 (CCL2), CXCL12, and interferon-related genes secreted by MAFs.<sup>139</sup> Additionally, in metastatic liver cancer and gastric cancer, it was found that the decrease of CD3+ infiltrating lymphocytes and the increase of TAM cells were related to the high levels of MAFs, partly due to the chemokine CXC receptor 4 (CXCR4) signal transduction in  $\alpha$ -smooth muscle actin ( $\alpha$ -SMA)+ MAFs.<sup>140</sup> In another study conducted by Costa et al., fibroblasts that inhibit tumor immunity were discovered, named cancerassociated fibroblasts subset 1 (CAF-S1).<sup>141</sup> Furthermore, it was found that the presence of CAF-S1 in breast cancer promoted bone metastasis of cancer cells.<sup>142</sup>

# MedComm

# 2.2.3 | MSCs

More and more studies have recognized the importance of MSCs in regulating tumor metastasis at the initial tumor site and distant metastasis sites. At the beginning of tumor metastasis, MSCs support the generation of the cancer-promoting microenvironment by transforming into CAFs and increase the motility of cancer cells through secreting growth factors and chemokines in autocrine and paracrine manners.<sup>128</sup> Pendergast et al. co-cultured MSCs and lung cancer cells and found that MSCs can mediate EMT to promote lung cancer metastasis by activating the abelson- matrix metalloprotein 9 (ABL-MMP9) signaling pathway.<sup>143</sup> In the late stage of tumor metastasis, MSCs participate in the colonization and metastatic growth of tumor cell. For example, in ovarian cancer, MET of cancer-associated MSCs mediated by Wilms' tumor 1 (WT1) and EZH2 promote metastatic tumors growth in distant organs.<sup>135</sup>

MSCs can also promote tumor metastasis by regulating anti-tumor immunity. As an immune checkpoint, PD-L1 is one of the crucial members involved in tumor immune escape and treatment resistance.<sup>144</sup> Studies have found that PD-L1 is highly expressed on tumor-associated MSCs, and MSCs can up-regulate the expression of PD-L1 in a variety of tumor cells. Hamidreza et al. reported that MSCs promoted the up-regulation of PD-L1 expression in breast cancer cells by secreting CCL5.<sup>145</sup> Sun et al. found that IL-8 secreted by MSCs can increase the expression of PD-L1 in gastric cancer cells.<sup>146</sup> Further experiments suggested that MSCs can increase the binding of PD-L1 to the transcription factor CCCTC-binding factor, thereby affecting the stemness of gastric cancer cells and leading to tumorigenesis.<sup>147</sup>

# 2.2.4 | Tumor-associated endothelial cells (TECs)

The role of TECs in metastasis is complex. In the process of tumor metastasis, TECs play an important role in obtaining anti-anoikis properties of CTCs, which is one of the mechanisms by which TECs promote tumor cell metastasis to secondary organs.<sup>148</sup> In addition, studies have found that biglycan in TECs is significantly higher than that in normal endothelial cells.<sup>149</sup> TECs-biglycan activates nuclear factor- $\kappa$ B (NF-kB) and extracellular signalregulated kinase (ERK) signal transduction to stimulate the metastasis of tumor cells expressing toll-like receptors (TLR) and promote tumor angiogenesis of tumors by activating the TLR signaling pathway.<sup>150</sup> IL-6 secreted by tumor cells can activate the signal transducers and activators of transcription 3 (STAT3)-VEGF pathway of lymphatic endothelial cells and encourage lymphatic metastasis of the tumor.<sup>151</sup>

#### 2.2.5 | Extracellular matrix (ECM)

ECM is a network of various proteins represented by collagen and macromolecules, which maintain tissue structure outside tumor cells.<sup>18</sup> A hallmark of metastasis is ECM degradation.<sup>152</sup> In the process of metastasis, it is necessary to enhance the invasion ability of tumor cells and reconstruct the ECM structure.<sup>153</sup> ECM reconstruction is a relatively complex process, which requires the participation of multiple chemokines.<sup>154,155</sup> Matrix metalloproteinases (MMPs) are a group of enzymes that can directly degrade collagen and multiple connexins in the process of ECM degradation. Among matrix MMPs family, MMP-2 and MMP-9 are the two important enzymes involved in the EMT process, which specifically degrade gelatin, collagen, elastin, and fibronectin, thereby inducing invasion and metastasis of tumor cells.<sup>156</sup> Furthermore, Musashi-1 can promote the degradation of ECM by upregulating Timp3 to encourage the expression of MMP-9 in breast cancer, and thus regulating cell-ECM adhesion.<sup>157</sup>

### 2.2.6 | Cytokines and chemokines

The involvement of various cytokines/chemokines, along with their receptors and signaling axis in the promotion of tumor metastasis has been well-studied. The convoluted cross-talk between various cell types and these secreted factors helps drive the sequence of events that lead to tumor metastasis.<sup>158,159</sup> These cytokines/chemokines are involved in ECM remodeling, tumor invasion, EMT, angiogenesis, pre-metastatic niche reprogramming, extravasation, and modulating stromal cells and immune cells.<sup>115,154,160-162</sup> The cytokines/chemokines including TNF-α, IL-8, CCL20, CXCL5, CXCL12.<sup>163-166</sup> CXCR2, CXCR3, and CXCR4, up-regulate EMT-TFs (snail and ZEB1), promote MMP-2 expression, and accelerate EMT of cancer cells, thereby promoting tumor metastasis.<sup>160,165,166</sup> Cytokine IL-8 mainly down-regulates E-cadherin through activating the Wnt/ $\beta$ -catenin pathway in ovarian cancer, thereby promoting tumor invasion.<sup>163</sup> Some cytokines can promote the expression of chemokines and mesenchymal transformation. For example, TNF- $\alpha$  has been reported to promote the up-regulation of CXCL10 and activate the phosphoinositide3-kinase/protein kinase B (PI3K/AKT) pathway to inhibit the phosphorylation of GSK-3 $\beta$ , leading to the up-regulation of snail and promoting tumor metastasis.  $^{160}$ 

Cytokines/chemokines, such as VEGF, TGF- $\beta$ , IL-1, CXCL8, and CCL21 are involved in promoting angiogenesis and lymphatic formation.<sup>167–171</sup> Among the known proangiogenic factors. VEGF is the most effective cvtokine in promoting angiogenesis. By directly binding to receptors on endothelial cells, VEGF induces endothelial cell proliferation and promotes tumor metastasis.<sup>167</sup> VEGF-C and VEGF-D in the VEGF family can activate the generation of lymphatic vessels, recruit chemokines, such as CCL21, CCL27, and CCL28, to lymphatic vessels and promote the occurrence of lymph node metastasis.<sup>172,173</sup> Cytokines/chemokines also play an indispensable role in promoting the recruitment of immune cells in the tumor microenvironment. For example, the recruitment of Treg requires the participation of CCL20 and CCL22, while the recruitment of TAMs requires the participation of CCL2.174-177 Single-cell sequencing analysis revealed that chemokine CCL5 is an important mediator of CTC immune escape.<sup>162</sup> CCL5 can promote immune escape and metastasis of CTCs by recruiting Tregs, which further suggests that chemokines can promote immune escape.<sup>162</sup> In addition, MDSCs secrete TGF- $\beta$  in esophageal cancer, which can increase the expression of PD-1 on tumor-infiltrating CD8+ T cells, leading to immunotherapy resistance.<sup>178,179</sup>

Cytokines and chemokines are involved in the organotropism of tumor metastasis. Lung metastasis is associated with the high levels of CXCL1, CXCL9, and CXCL10 secreted by human pulmonary artery endothelial cells or lung fibroblasts.<sup>179,180</sup> Ricardo et. Al. reported that blocking CXCL5/CXCR2 signaling can inhibit the bone colonization of breast cancer cells.<sup>181</sup> Blocking CXCL12/CXCR4 signaling significantly impair the metastasis of breast cancer cells to regional lymph nodes and lung, which indicate that chemokines and their receptors play a key role in determining the destination of metastasis.<sup>182,183</sup> Overall, the potential of cytokines and chemokines to induce metastasis is mainly achieved by promoting EMT, angiogenesis, immunosuppression, and pre-metastatic niche reprogramming.

## 2.2.7 | Tumor-secreted extracellular vesicles

EVs secreted by tumors are the key mediators of cell-tocell contact in the local and remote microenvironments.<sup>184</sup> EVs are exceptional cargo for various nucleic acid, proteins, and lipids in TME, and play a prominent role in the metastasis of the tumor to distant organs.<sup>183–190</sup> Tumor-associated EVs in breast cancer can promote breast cancer colonization by changing the composition and structure of lung tissue fibroblasts.<sup>185</sup> It was found that EVs secreted by bladder cancer cells mediate their intercellular communication with human lymphatic endothelial cells through long non-coding RNA (lncRNA) ELNAT1 and promote lymph node metastasis in a SUMOylation-dependent manner.<sup>186</sup>

In nasopharyngeal carcinoma (NPC), EVs migrate from highly metastatic to poorly metastatic NPC cells, mediate intercellular communication, and enhance the metastatic potential of poorly metastatic NPC cells by inducing the up-regulation of epidermal growth factor receptor (EGFR) and down-regulation of reactive oxygen species (ROS).<sup>191</sup> Mechanistically, overexpression of EGFR mediated by EGFR-rich EVs down-regulates intracellular ROS levels through regulating PI3K/AKT pathway, thereby promoting the metastatic potential of NPC with poorly metastatic ability.<sup>191</sup>

The role of EVs in tumor metastasis depends on their components; the function of EVs may be different for different tumor metastasis sites. For example, in brain metastasis, EVs mainly mediate pre-metastasis regulation by destroying the blood-brain barrier or by distinct increasing the expression of cell migration-inducing and hyaluronan-binding protein in brain-tropic EVs.<sup>187,188</sup> In hepatocellular carcinoma, nidogen 1 (NID1) in EVs promote tumor cell colonization and extrahepatic metastasis by enhancing angiogenesis and lung endothelial permeability and promoting the formation of pre-metastasis niches in the lung.<sup>189</sup> EVs-NID1 also activates fibroblasts that secrete TNF receptor 1 (TNFR1), promote lung colonization of tumor cells, and augment the growth and motility of cancer cells.<sup>189</sup> EVs in bone metastasis mainly mediate cancer-induced osteolysis to produce a suitable microenvironment for tumor cells.<sup>190</sup> In addition, EVs can also promote the formation of pre-metastasis niches by stimulating CAFs, inducing MSC differentiation, and regulating tumor immunity.<sup>183</sup>

# 2.2.8 | Neutrophil extracellular traps (NETs)

NETs are net-like structures composed of DNA histone complexes, and proteins released by neutrophils in response to infection or inflammatory cytokines were discovered by Brinkmann et al. in 2004.<sup>192</sup> NETs are initially discovered to be one of the host defense mechanisms of neutrophils against pathogens, and they are also involved in the progression of sterile inflammation-related diseases, such as autoimmune diseases, diabetes, and carcinoma.<sup>193</sup> In the absence of infection, NETs can also be stimulated by cancer cells and CAFs, and hijack anti-tumor immune system to promote tumor cell proliferation and metastasis.<sup>193</sup>

**MedComm** 

The levels of plasma NETs in patients with lung cancer, cervical cancer, and pancreatic cancer are significantly higher than those in healthy controls and are related to tumor recurrence and metastasis.<sup>194</sup> In tumor metastasis, NETs are mainly involved in the spread of cancer cells and awakening dormant tumor cells. NETs can induce EMT of primary tumor cells in the early stage of metastasis and lead to the activation of inflammatory signaling pathways such as NF-kB and STAT3, thereby enhancing the mobility and invasiveness of cancer cells and promoting metastatic growth.<sup>195,196</sup>

In the circulation, NETs wrap CTCs to avoid contact between CTCs and immune cells and can also form a physical barrier for immune escape.<sup>197,198</sup> The molecular mechanisms by which NETs can trap CTCs have been investigated, and it has been found that expression of  $\beta$ 1-integrin on both cancer cells and NETs is important for the adhesion of CTCs to NETs.<sup>197,198</sup> After spreading to other sites, cancer cells from primary tumors often do not start growing immediately but enter a dormant state. Disseminated cancer cells can remain dormant for years or even decades before they relapse or "wake up" as metastatic cancer.<sup>199</sup> Studies have found that the formation of NETs induced by sustained inflammation is required for awakening dormant cancer.<sup>199</sup> Two NET-associated proteases (neutrophil elastase and MMP9) sequentially cleave laminin, and then NET-remodeled laminin facilitates the proliferation of dormant cancer cells by activating integrin  $\alpha 3\beta 1$ signaling and focaladhesion kinase (FAK)/ERK/myosin light chain kinase (MLCK)/Yes associated protein (YAP) signaling.<sup>199</sup>

# 2.3 | A double-edged knife in tumor metastasis: Autophagy

Autophagy is generally defined as a lysosome-dependent mechanism of intracellular degradation and recycling process, which is highly conserved in all eukaryotes.<sup>200–202</sup> Several forms of autophagy have been discovered. In all types of autophagy, autophagosomes are the hallmark morphological characteristics, and the formation of autophagosomes is the core process of this dynamic process.<sup>200,203</sup> Autophagy is a dynamic physiological process that maintains cell homeostasis when the body encounters stress, including nutrient deprivation, hypoxia, and infection.<sup>204,205</sup> It has been reported that autophagy plays a key role in the prevention of cancer and other diseases by promoting cellular senescence and antigen presentation and preventing genomic instability and necrosis.<sup>206</sup> However, autophagy also plays a vital role in the occurrence and progression of tumor.<sup>207,208</sup> In many types of tumors, including melanoma, breast cancer, and liver cancer, the expressions of autophagy-related

proteins light chain 3 and beclin-1 in tumor tissues of patients with distant metastasis are higher than in tumor tissues of patients with non-metastatic tumor, suggesting that autophagy is closely related to tumor metastasis.<sup>209–211</sup> Partly due to the heterogeneity of tumor cells, autophagy is a double-edged sword in the process of tumor metastasis.<sup>212</sup>

In the early stage of metastasis, autophagy mainly suppresses the occurrence of metastasis by regulating the antitumor immune response.<sup>213–215</sup> Autophagy-related protein 5(ATG5) is considered to be closely related to autophagy. In pancreatic ductal adenocarcinoma, mice with ATG5 knockout developed more tumors and metastases than control mice. Moreover, compared with the control mice, the expression of cytokines that regulate macrophage chemoattraction and differentiation into M2 macrophages was up-regulated in the tumors of mice with ATG5 knockout. Meanwhile, the number and activity of M2-TAMs were substantially higher in ATG5 knockout mice, indicating that autophagy can inhibit TAMs infiltration and limit tumor metastasis.<sup>214</sup> In addition, studies have found that autophagy can stimulate tumor-related spontaneous inflammation by releasing high mobility group box 1 and enhance the antitumor immune response of DCs to limit metastasis.<sup>215</sup> In the later stage of tumor metastasis, autophagy reduces the adhesion between tumor cells and ECM by regulating the activity of the Rho family so as to promote tumor migration and invasion.<sup>216</sup> In addition, autophagy can enhance tumor invasion and promote the progress of tumor metastasis by inhibiting anoikis of CTCs, maintaining the stemness of CSCs, and reawakening dormant tumor cells.<sup>210,217,218</sup>

Another significant aspect of autophagy in metastasis is involved in the regulation of EMT. P53, an oncogene, also plays an important role in regulating the relationship between EMT and autophagy.<sup>219,220</sup> Under normal circumstances, P53 exists in the cytoplasm and inhibits autophagy. However, under cellular stress, P53 translocates from the cytoplasm to the nucleus and induces autophagy.<sup>221,222</sup> In addition to autophagy, recent studies have found that P53 inhibits EMT and metastasis by affecting the TFs involved in the EMT process.<sup>223</sup>

It is well known that AKT/ mammalian target of rapamycin (mTOR) signaling pathway participates in modulating the EMT process.<sup>224</sup> A large number of studies have found that oncogenes activate mTOR-related pathways to promote tumor metastasis by inhibiting autophagy.<sup>225</sup> Suppressor of cytokine signaling-5 (SOCS5), a member of the SOCS protein family, has also been found to promote tumor cell invasion and metastasis in hepatocellular tumors by up-regulating PI3K/Akt/mTOR-mediated autophagy pathway.<sup>226</sup> Interestingly, opposite findings have been reported in NPC. Annexin A1 (ANXA1)

inhibits autophagy and promotes EMT by activating PI3K/Akt/mTOR pathway.<sup>227</sup> ANXA1 inhibits autophagy by activating PI3K/Akt/mTOR pathway, leading to the upregulation of tumor cell migration and invasion capabilities, EMT-like changes, and metastasis in vivo.<sup>227</sup> During this process, autophagy inhibited by ANXA1 induces EMT-like alterations, possibly by inhibiting autophagy-mediated Snail degradation.<sup>227</sup>

Autophagy can also be induced by activating the adenosine monophosphate activated protein kinase (AMPK) pathway that promotes tumor metastasis.<sup>228</sup> For example, tight junction protein 1 (CLDN1) promotes EMT and metastasis of esophageal cancer cells by triggering autophagy in vivo and in vitro.<sup>229</sup> Mechanically, CLDN1 induces autophagy by up-regulating the expression of Unc-51-like autophagy activating kinase 1 (ULK1) through AMPK/signal transducer and activator of transcription 1 (STAT1)/ULK1 signaling pathway.<sup>229,230</sup> In general, autophagy is a double-edged sword in the process of tumor metastasis, and the mechanisms involved are very intricate and require further research.

## 2.4 | Lipid metabolism

Under hypoxic conditions, tumor cells mainly rely on glycolysis to obtain sufficient energy.<sup>231</sup> This metabolic mode different from normal cells is also called tumorrelated metabolic rearrangement.<sup>232</sup> Glycolysis is essential to maintain the malignant proliferation behavior of the tumor, but there are relatively few studies on the effect of lipid metabolism on tumor progression. Obesity has been recognized as a risk factor for cancer, so investigating lipid metabolism in the tumor is necessary.<sup>233,234</sup> The results of researches in the past 10 years have shown that abnormal lipid metabolism in the TME is one of the key steps involved in tumor metastasis.<sup>235</sup> Therefore, correcting abnormal lipid metabolism may prevent the occur-

It has been found that a variety of enzymes involved in lipid anabolism and catabolism are related to tumor metastasis, including adenosine triphosphate (ATP) citrate lyase (ACLY), fatty-acid synthase (FASN), stearoyl-CoA desaturases (SCD), and monoacylglycerol lipases.<sup>236</sup> ACLY and SCD-1 are associated with facilitated colon cancer metastasis, and FASN can promote retroperitoneal metastasis of ovarian cancer.<sup>237,238</sup> Identifying tumor metastasis initiating cells is considered to be one of the most challenging problems in the field of tumor metastasis research. In recent years, fatty acid receptor CD36 has been shown to play an essential role in the metastasis of oral cancer because CD36 is positively correlated with lymph node metastasis and can be used as a marker of metastasis initiating cells.<sup>239</sup> Palmitic acid or a high-fat diet can enhance the metastatic ability of CD36+ tumor cells. Subsequent studies have found that CD36+ cells are associated with a poor prognosis of ovarian cancer, and blocking CD36 can attenuate tumor metastasis.<sup>240</sup> These findings suggest that the initiating cells of tumor metastasis may depend on lipids, and blocking CD36 can reverse tumor metastasis.

With the development of proteomics and metabolomics, new discoveries have been made about cancer metabolic disorder and its correlation with metastasis. In breast cancer, it was found that the levels of phospholipid are different between mammary epithelial cells and breast cancer cells, as well as between breast cancer cells with different levels of aggressiveness.<sup>241</sup> In short, lipid metabolic reprogramming is associated with breast cancer carcinogenesis and metastasis.<sup>241</sup> In pancreatic cancer, fatty acid synthesis was found to maintain the stemness of pancreatic cancer cells, indicating that abnormal lipid metabolism can make cancer cells more invasive and promote the spread of tumor cells.<sup>242</sup> Promoting cholesterol biosynthesis may promote tumor metastasis.

More and more molecules regulating lipid metabolism have been proved to promote tumor metastasis by regulating EMT, including apolipoprotein C, sterol regulatory element-binding transcription protein 1, stromal-interaction molecule 1, human hydroxysteroid dehydrogenase-like 2, and cytosolic phospholipase  $A2\alpha$ .<sup>243–246</sup> In triple-negative breast cancer, nicotinamide adenine dinucleotide phosphate (NADP) steroid dehydrogenase-like (NSDHL), a cholesterol metabolic enzyme, has been reported to be a potential metastatic driver.<sup>247</sup> The functions of NSDHL rely on its enzyme activity in the biosynthesis of cholesterol and is mediated by the NSDHL-TGF $\beta$ R2 signaling pathway.<sup>247</sup>

TME has a unique lipid structure, which is characterized by being rich in sphingolipids and cholesterol and is called lipid rafts.<sup>248</sup> The role of lipid rafts in tumor metastasis is different from other lipid metabolism-related regulatory molecules. Rina et al. reported that palmitoylated CD44 is wrapped in lipid rafts, and its binding to promigration binding partners (such as Ezlin) is restricted, thereby inhibiting cancer metastasis and spread.<sup>249</sup> Moreover, another study reported that squalene synthase could promote lung cancer metastasis by activating TNFR1, NF-Kb, and matrix metallopeptidase 1, and destroying the lipid raft structure.<sup>250</sup> Therefore, lipid rafts mainly play an inhibitory role in tumorigenesis and metastasis. For cells in TME, abnormal lipid metabolism may be a potential mechanism to promote tumor metastasis. For example, lipid accumulation in TME can lead to CD8+T cell dysfunction and promote TAMs differentiation.<sup>251,252</sup> In addition, the abnormal elevation of fatty acid synthase in CAFs may enhance the aggressiveness of tumor cells.<sup>253</sup> Therefore, lipid metabolism reprogramming plays an important role in regulating the formation of the pro-metastatic TME.

MedComm

# 2.5 | Long non-coding RNA

LncRNA is a standard non-coding RNA with a length of more than 200.<sup>254</sup> Researches in recent years have shown that lncRNA plays an important role in tumorigenesis.<sup>254</sup> In fact, lncRNA plays a double-edged role in tumor metastasis. LncRNA may be involved in all the processes of metastasis, including EMT, tumor invasion and migration, and tumor cell colonization in secondary sites. We briefly summarized the representative lncRNA that have been shown to be relevant with metastasis in vivo and in vitro (Table 1).<sup>255–280</sup>

As mentioned above, EMT is an essential step in the initiation of tumor metastasis. It has been reported that LncRNA H19 can mediate the transformation of tumor cells into mesenchyme and is significantly elevated in bladder cancer, breast cancer, and colorectal cancer.<sup>255–257</sup> H19 directly binds to EZH2 in bladder cancer, resulting in a reduction in EMT epithelial marker E-cadherin.<sup>255</sup> In colorectal cancer, H19 acts as a competing endogenous RNA, leading to the up-regulation of ZEB1/2 protein expression.<sup>257</sup> However, in prostate cancer, the opposite phenomenon has been observed, that is, H19 inhibits tumor metastasis, mainly by encoding miR-675 to mediate the down-regulation of TGFBI.<sup>258</sup> Similar to H19, lncRNA-PNUTS serves as a competitive sponge for miR-205 and miR-200, leading to the up-regulation of EMT in breast cancer.<sup>259</sup> LncRNA LINC00460 is increased in colon cancer, and it induces EMT and promoteS tumor growth and metastasis by enhancing the expression of high-mobility group AT-hook 1 and decreasing the expression of Ecadherin.<sup>260</sup>

The influence of lncRNA on tumor invasion and migration is also one of the mechanisms by which it affects tumor metastasis. For example, colon cancer-associated transcript 2 (CCAT2) and RNA associated with metastasis-11 (RAMS11) have been reported to participate in the process of tumor invasion and migration. CCAT2 is related to the stability of microsatellites; knockdown of CCAT2 inhibits tumor invasion and migration in mouse models. CCAT2 may interact with TCF7L2 to activate Myc transcription and Wnt signaling pathways and thus promote metastasis.<sup>281,282</sup> RAMS11 is overexpressed in colon cancer with liver metastasis. Knockout of RAMS11 gene using CRISPR-Cas9 technology can reduce the invasion and migration of colon cancer cells in vitro and reduce liver metastasis in mouse model.<sup>283</sup> Lnc01232 was found to promote metastasis by inhibiting ubiquitination, upregulating HNRNPA2B1, and activating the mitogen-activated

# TABLE 1 Important metastasis-related long non-coding RNAs

Metastasis- related long non-coding				<b>D</b> (
RNA	Cancer type	Function	Effect	Reference
H19	Bladder cancer	Bind to enhancer of zeste homolog 2 (EZH2) to downregulate E-cadherin	Inhibit	255
	Prostate cancer	Encode miR-675 to mediate the down-regulation of TGF-β1	Promote	258
	Colorectal cancer	Upregulate zinc finger E-box-binding homeobox 1/2 protein	Promote	257
PNUTS	Breast cancer	Competitive sponge for miR-205 and miR-200 and enhancing epithelial-mesenchymal transition (EMT)	Promote	259
LINC00460	Colon cancer	Enhance the expression of high-mobility group AT-hook 1	Promote	260
Lnc01232	Pancreatic cancer	Upregulate HNRNPA2B1, and activate the MAPK/ERK signaling	Promote	261
MALAT1	Colorectal cancer	Regulate the miR-106b-5p via SLAIN2	Promote	262
TPA	Breast Cancer	Activate TGF- $\beta$ signaling pathway	Promote	263
PVT1	Colon Cancer	Downregulate tumor suppressor miR-152-3p	Promote	264
LIMT	Breast cancer	Suppress tumor cells motility	Inhibit	265
SPRY4-IT1	Bladder cancer	Bind to miR-101-3p to upregulate the expression of EZH2	Promote	266
TRERNA1	Gastric cancer	Regulating CDH1 to upregulate SNAI1	Promote	267
NEF	Hepatocellular carcinoma	Suppress Wnt/β-catenin signaling to activate expression of FOXA2	Inhibit	268
HOXD-AS1	Hepatocellular carcinoma	Competitive bind to miR-130a-3p to upregulate the expression of EZH2 and MMP2	Promote	269
CYTOR	Colon cancer	Activate Wnt/ $\beta$ -catenin signaling to enhance EMT	Promote	270
JPX	Lung cancer	Activate Wnt/β-catenin signaling pathway to upregulate Twist1 expression	Promote	271
LINC00662	Colon cancer	Activate extracellular signal-regulated kinase (ERK) signaling pathway	Promote	272
ID2-AS1	Hepatocellular carcinoma	Activate HDAC8/ID2 signaling pathway to decrease Twist expression	Inhibit	273
RPPH1	Colorectal cancer	Interact with TUBB3 to reduce E-cadherin levels and induce macrophages M2 polarization	Promote	274
SATB2-AS1	Colorectal cancer	Regulate SATB2 to decrease MMP9 and vimentin	Inhibit	275
NORAD	Lung cancer Breast cancer	Bind and sequester S100P to suppress S100P pro-metastatic signaling pathway	Inhibit	276
URRCC	Renal cancer	Enhance EGFL7 expression to suppress P-AKT/FOXO3 signaling pathway	Promote	277
FEZF1-AS1	Colorectal cancer	Activate PKM2/signal transducers and activators of transcription 3 signaling pathway	Promote	278
ADAMTS9-AS2	Salivary adenoid cystic carcinoma	Activate PI3K/Akt and MEK/ERK signaling pathway	Promote	279
GAS5	Pancreatic cancer.	Regulate miR-221/SOCS3 to suppress EMT and cancer stem cells self-renewal	Inhibit	280

protein kinase/extracellular signal-regulated kinase (MAPK/ERK) signaling pathway in pancreatic cancer.<sup>261</sup>

As the first lncRNA found to be associated with metastasis, metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) has been shown to be associated with poor prognosis of lung cancer and breast cancer, and its expression is significantly increased in patients with metastatic cancer.<sup>284,285</sup> In the breast cancer mice model lacking the promoter of MALAT1 or MALAT1, tumor differentiation and E-cadherin are increased, whereas lung metastasis is significantly reduced, indicating that MALAT1 may serve as essential factors for tumor cell colonization at distant metastasis sites.<sup>285</sup>

With the progress in understanding the functions of lncRNAs, increasing evidence has indicated that lncRNAs play a role in the physiological and pathological processes of malignancies. Although many findings need further verification, it is certain that lncRNAs can carry out diverse functions in carcinogenesis and metastasis. However, the specific role and mechanisms of different lncRNAs in cancer metastasis need to be further elucidated.

# 3 | INTERVENTIONS FOR TUMOR METASTASIS

Metastatic disease is the major contributor to cancerrelated mortality. Therefore, there is an urgent need for effective cancer treatments that can eliminate large solid tumors and disseminated and metastatic nodules, while simultaneously preventing tumor recurrence. With the deepening understanding of the molecular mechanism of tumorigenesis and metastasis in recent years, a plethora of treatment approaches, including targeted therapy and immunotherapy have been established for antitumor treatments.<sup>13,286,287</sup> Although the emergence of new therapies, such as those based on immune checkpoint inhibitors (ICIs), is rapidly changing the treatment modality of metastatic patients in some cases, the current standard of cancer treatment for localized disease is still usually based on surgery, chemotherapy, and radiotherapy.<sup>288,289</sup> In addition, the combination therapies are receiving more attention and are being actively evaluated.<sup>290</sup> We summarize the interventions for metastatic diseases based on preclinical and clinical evidence.

# 3.1 | Surgery, chemotherapy, and radiotherapy

Surgery, chemotherapy, and radiotherapy are the three cornerstones of tumor treatment. According to the purpose of treatment, surgical treatment can be divided into radMedComm

ical surgery, local surgery, and palliative surgery.<sup>291</sup> For patients with early-stage tumor, radical surgery can prevent and reduce tumor recurrence and metastasis. However, for most patients, due to the insidious tumor-related symptoms, distant metastasis has already occurred at the time of diagnosis. In the past, it was considered that metastatic tumors cannot be treated surgically, but a number of recent studies and clinical practices have confirmed that local surgery can be performed on some patients with localized metastasis, which can reduce the tumor burden and may achieve the goal of radical cure.<sup>292–295</sup> For example, palliative surgery for patients with intraperitoneal metastases such as liver cancer and ovarian cancer can alleviate their symptoms and improve their quality of life.<sup>296,297</sup>

For intervention and treatment of metastatic tumors, surgery alone is not enough. Dormant CTCs is the main cause of early postoperative metastasis. Therefore, many clinical studies have been carried out to reduce the probability of tumor metastasis. Neoadjuvant chemotherapy and adjuvant chemotherapy are currently widely used in the treatment of cancer because these interventions can kill CTCs and reduce the possibility of metastasis while reducing the size of the preoperative tumor and preserving some functional organs.<sup>298-301</sup> Except for a small number of patients, chemotherapy, such as 5-FU, Adriamycin, and platinum, is the first-line choice for most patients with advanced cancer.<sup>301-304</sup> It has been observed that diverse malignancy patients with metastatic lesions benefit from chemotherapy, especially patients with lymphoma and leukemia, and some patients can even obtain a durable response.305,306

However, conventional maximum-dose chemotherapy cannot be tolerated by many patients with advanced cancer due to severe side effects and ultimately leads to treatment failure. In addition, it has been found that chemotherapy may induce the production of pro-metastatic cytokines and chemokines, thereby inducing tumor metastasis.<sup>307</sup> Therefore, many studies have been carried out to optimize chemotherapy regimens, and some progress has been made. Recent studies have shown that metronomic chemotherapy can inhibit angiogenesis and reduce the expression of pro-metastatic cytokines and chemotaxis, thereby reducing tumor recurrence and the distant spread of tumor cells.<sup>308,309</sup> These results indicate that low-dose metronomic chemotherapy may be effective in the prevention and treatment of tumor metastasis.

Radiotherapy has been recognized as a radical treatment for some early-stage cancer, such as NPC, and progress has also been made in the treatment of patients with metastatic cancer.<sup>295,310</sup> Compared with radiotherapy alone, the combination of radiotherapy and other therapies has better therapeutic effects on metastatic MedComm

cancer, especially when combined with chemotherapy. For locally advanced NPC, concurrent chemoradiotherapy has been approved as a standard regimen by the National Comprehensive Cancer Network Guidelines.<sup>310</sup> Recently, two Phase III clinical trials from China proposed that induction chemotherapy before concurrent chemoradiotherapy can significantly improve the prognosis of patients with NPC.<sup>311,312</sup> It can be reflected that multidisciplinary combination therapy will be the main means to improve the prognosis of patients with metastatic cancer.

# 3.2 | Targeted therapy for metastasis

For most patients with advanced-stage cancer, surgery and radiotherapy are difficult to achieve satisfactory therapeutic effects, which is also the reason for the high tumorrelated mortality. In the process of tumor metastasis from initiation to colonization, the occurrence of each step is the result of the joint action of some specific genes and signaling pathways. Blocking one of these steps may block the formation of metastases. Therefore, the development of drugs targeting these targets may provide an alternative for patients with advanced tumors.

# 3.2.1 | Targeting EMT and cell motility

Blocking EMT is one of the key strategies to prevent tumor cells from spreading from the primary tumor site. As a marker of mesenchymal cells, N-cadherin is elevated in metastatic tumors and is associated with a poor prognosis.<sup>33,38</sup> ADH-1 is the first humanized antibody selectively targeting N-cadherin, which has been proved to improve the prognosis of patients with tumor-expressing N-cadherin.<sup>313</sup> Recently, a study showed that blocking Ncadherin by ADH-1 can also inhibit the expression of PD-L1 and the recruitment of Tregs in TME and augment the cytotoxicity of tumor-infiltrating lymphocytes (TILs) against tumor cells.<sup>314</sup> The steroid receptor coactivator (SRC) tyrosine kinase family is one of the central members that mediate EMT.<sup>45</sup> Drugs targeting SRC, including dasatinib, bosutinib, and saracatinib, have been proved to inhibit tumor growth and prolong patient survival.315 When combined with other treatment modalities, the antitumor effect is stronger. In a Phase III prospective clinical trial (NCT01584648), the 3-year progression-free survival rate of patients receiving dabrafenib plus trametinib was 22% and 12% for patients receiving dabrafenib monotherapy, and the 3-year overall survival rate was 44% and 32%, respectively. Furthermore, in the subgroup with normal lactate dehydrogenase and less than three metastatic sites,

the 3-year overall survival rate of patients receiving combination therapy reached 62%.<sup>316</sup>

The motility of tumor cells is one of the critical factors that determine whether the metastasis can proceed smoothly, and the integrin family can affect cell motility by regulating cell adhesion.<sup>37</sup> Therefore, targeting the integrin family can weaken the motility of tumor cells. Currently, drugs targeting the integrin family mainly include cilengitide, intetumumab, 264RAD, and MK-0429, which have been proven to prolong the survival of cancer patients in preclinical studies and prospective clinical studies.<sup>317–320</sup> The allosterisms of actin and myosin in ECM promote structural remodeling and tumor cell polarization during tumor cell migration; Ras homolog gene family, member A (RhoA), MMP, and non-muscle Myosin-II are involved in the allosterisms of these two proteins.<sup>130</sup> Small molecule inhibitors of molecules regulating actin and myosin have been shown to attenuate tumor cell motility and inhibit tumor metastasis in vivo and in vitro, but their effectiveness in humans needs to be confirmed by further clinical studies.<sup>156,321</sup>

After separation from ECM, only a small percentage of cells can survive and achieve subsequent metastasis. The main reason is that this small part of tumor cells has acquired the properties of anti-anoikis.<sup>19,322</sup> Therefore, tumor metastasis can be further inhibited by promoting tumor cell anoikis and eliminating anoikis resistance. T0070907 inhibits PPAR- $\gamma$  and leads to cell death by reducing adhesion and inducing anoikis.<sup>323</sup> In addition, galectin-3 inhibitors have been shown to be effective in the treatment of thyroid cancer. The mechanism is mainly attributed to the inhibition of anoikis resistance, which is expected to provide a new strategy for inhibiting tumor metastasis.<sup>324</sup>

## 3.2.2 | Targeting CAFs

CAFs have also been proven to be important members involved in promoting tumor metastasis, and targeting CAFs have gradually attracted more attention in the treatment of metastatic tumor.<sup>130</sup> The activity of CAFs promoting tumor metastasis is mediated by a variety of signaling pathways, especially TGF- $\beta$  and EGFR signaling pathways, which may provide targets for anti-metastasis.<sup>325–327</sup> In a variety of malignancies, including bladder cancer, breast cancer, colorectal cancer, and pancreatic cancer, TGF- $\beta$ 1 can induce the transformation of resident normal fibroblasts into CAFs and lead to the differential expression of  $\alpha$ -SMA and fibroblast activation protein (FAP) genes (specific markers of CAFs) through the typical TGF- $\beta$  signaling pathway.<sup>161,328–330</sup> These data suggest that TGF- $\beta$ 1 plays a role in promoting CAFs production.

As the first oral small-molecule selective inhibitor of TGF- $\beta$ 1, galunisertib has been proved to have anti-tumor effects in clinical trials of liver cancer, pancreatic cancer, and glioma.<sup>331–333</sup> In addition, the therapeutic effect of combination gemcitabine with galunisertib is significantly better than gemcitabine monotherapy in advanced pancreatic cancer (combination group 8.9 months vs. monotherapy group 7.1 months).<sup>332</sup> In a Phase II trial, the median overall survival of galunisertib combined with the multitarget tyrosine kinase inhibitor (TKI) sorafenib in the treatment of metastatic liver cancer was 18.8 months.<sup>331</sup> Researchers have also developed a fusion protein, M7824, which blocks both PD-L1 and TGF- $\beta$  signaling pathways; M7824 has shown potent anti-tumor activity in preclinical studies.<sup>334</sup> Subsequently, a Phase I clinical study was conducted in advanced lung cancer patients, and M7824 showed tolerable toxicities and preliminary anti-tumor effects.335

In addition, a variety of cytokines secreted by CAFs act as ligands on the janus kinase-signal transducer and activator of transcription (JAK/STAT) pathway, participate in the activation of this pathway, and promote tumor metastasis.<sup>336–338</sup> For example, CAFs-derived IL-6 can activate the STAT3 pathway to promote EMT of tumor cells, thereby facilitating metastasis.<sup>62,339</sup> Therefore, antagonizing those cytokines or blocking JAK/STAT signaling can inhibit the occurrence of tumor metastasis. The JAK inhibitor ruxolitinib has been shown to inhibit the invasion and migration of breast cancer, lung cancer, and colorectal cancer cells in vivo.<sup>336,340,341</sup> Two Phase II clinical trials have shown that ruxolitinib combined with gemcitabine improved overall survival in patients with advanced HER2negative cancer or advanced pancreatic cancer.<sup>342</sup> The IL-6 receptor inhibitor tocilizumab and IL-6 antagonist siltuximab have been shown to augment the anti-tumor activity of chemotherapeutic agents in preclinical trials, and further clinical studies are expected.<sup>343,344</sup>

Blocking the activity of CAFs directly can also play an anti-tumor metastasis role. FAP is one of the markers of activated CAFs, and targeting FAP could transfer activated CAFs into a quiescent state. The monoclonal antibody targeting FAP (sibrutuzumab) and the small molecule inhibitors of FAP (F19 and PT100) were well-tolerated, but no obvious clinical anti-tumor effect was observed in Phase I clinical trials, and subsequent structural adjustments may be needed.<sup>345–347</sup>

# 3.2.3 | Targeting angiogenesis

Angiogenesis is considered to be one of the components involved in tumor metastasis. A variety of smallmolecule tyrosine kinase inhibitors and monoclonal antiMedComm

body against angiogenesis have been approved as firstline or second-line therapy in a variety of advanced malignancies, including pazopanib targeting vascular endothelial growth factor receptor (VEGFR), imatinib targeting platelet-derived growth factor receptor (PDGFR), bevacizumab targeting VEGF, and multi-targeted receptor TKI anlotinib.<sup>348–351</sup> In addition, the combined application of existing drugs and the development of new drugs inhibiting angiogenesis are being explored in pre-clinical and clinical trials.<sup>352</sup> Several clinical trials of advanced lung cancer have shown that the combination of targeted drugs such as gefitinib, erlotinib, and anlotinib with radiotherapy can enhance the efficacy of anti-brain metastasis and are expected to be applied in clinical practice.<sup>354-356</sup> There has been a large amount of literature reviewing the therapies targeting angiogenesis, so we will not describe them in detail.

# 3.3 | Immunotherapy

# 3.3.1 | Targeting TAMs and Tregs

Regulating tumor immune microenvironment is another crucial therapeutic strategy to interfere with tumor metastasis. There is increasing evidence that TAMs and Tregs in the microenvironment promote tumor development and metastasis.<sup>93,353</sup> Targeting TAM and Tregs is a promising strategy to modify the immunosuppressive TME and prevent metastasis.

A large number of studies have shown that CC chemokine receptors are essential mediators involved in tumor metastasis.<sup>357</sup> Among them, CCR1, CCR2, CCR3, and CCR5 promote the recruitment of TAMs in the TME, and CCR4 mainly mediates the recruitment of Tregs. indicating that CC chemokines can be used as potential anti-tumor pharmacological targets.<sup>82,175</sup> In recent years, CCR inhibitors have shown strong anti-tumor effects in preclinical and clinical studies.<sup>358</sup> For example, in a Phase I clinical study, CCR5 antagonists have demonstrated their ability to inhibit liver metastasis in colon cancer.<sup>359</sup> A preclinical experiment has shown that mogamulizumab, which antagonizes CCR4, can reduce tumor lung metastasis, and a Phase III clinical trial has demonstrated that antibodies targeting CCR4 are effective in treating patients with cutaneous T lymphoma.<sup>360</sup> Furthermore, mogamulizuma combined with the immune checkpoint inhibitor nivolumab may provide a new strategy for the treatment of advanced or metastatic solid tumors.<sup>361</sup> Another monoclonal antibody targeting CCR, AMG 820, also showed preliminary anti-tumor ability.<sup>362</sup>

Colony-stimulating factor 1 (CSF1) can enhance macrophage recruitment and activation into a pro-tumoral

TAM phenotype and plays a role in promoting metastasis.<sup>363,364</sup> The pleiotropic signaling of CSF1 receptor (CSF1R) supports multiple functions of macrophage, including proliferation, differentiation, and migration.<sup>365</sup> The CSF1/CSF1R axis has received much attention, and approaches targeting the ligands or receptors are currently being clinically developed. Pexidartinib, which target KIT, CSF1R, and Fms-like tyrosine kinase 3 (FLT3), has been approved by the US Food and Drug Administration (FDA) for the treatment of symptomatic tenosynovial giant cell tumor in adult patients.<sup>366</sup> Emactuzumab, a humanized monoclonal antibody targeting CSF1/CSF1R, inhibit tumor cell proliferation and metastasis by blocking the activity of CSF1R-dependent TAMs, suppressing the recruitment of TAMs to the microenvironment, and enhancing the T-cell infiltration.<sup>367</sup> However, the antitumor ability of emactuzumab is not evident in a Phase I clinical trial, and further studies are needed.367

The CD4+ Treg cell population in the TME is the main cell group leading to the inactivation of effector T cells and is also one of the factors that promote tumor progression and metastasis. Therefore, targeting CD4+ Treg cells directly is a promising strategy to effectively prevent tumor metastasis and progression. IT1208, a defucosylated humanized anti-CD4 monoclonal antibody, demonstrated tolerable toxicities and encouraging preliminary efficacy in a Phase I clinical trial involving 11 patients with advanced solid tumors.<sup>368</sup> In addition, a microsatellite-stable colon cancer patient receiving IT1208 treatment showed increased infiltration of both CD4+ and CD8+ T cells in the tumor and achieved a durable partial response.<sup>368</sup> These results suggest that Tregs-targeted drugs, such as IT1208, are expected to provide a new idea for immunotherapy, but further researches are needed.

## 3.3.2 | ICIs

A major hallmark of T-cell exhaustion is the increased expression of multiple immune checkpoints, such as PD-1, CTLA-4, LAG-3, T-cell immunoglobulin-3 (TIM-3), and T-cell immunoglobulin and ITIM domain (TIGIT).<sup>369–371</sup> Immune checkpoints have distinct ligands and inhibit anti-tumor activity of T cell through multiple mechanisms; T cell function decreases with the increase of immune checkpoint expression.<sup>373</sup> Recent advances in cancer immunotherapy have shown that ICIs, including inhibitors of PD-1, PD-L1, CTLA-4, LAG-3, TIM-3, and TIGIT, can improve the clinical response and survival of patients with a broad spectrum of metastatic cancers, such as melanoma, non-small lung cancer, and renal cell cancer (Table 2).<sup>13,369,371,374–376</sup> ICIs limit the inhibitory pathway of T cells and stimulate the activation of effector T

cells to enhance the anti-tumor immune response. So far, at least eight different ICIs have been approved by the US FDA for the treatment of more than a dozen different cancers, including melanoma, kidney cancer, lymphoma, colorectal cancer, lung cancer, head and neck carcinoma, liver cancer, and sarcoma.<sup>377</sup>

Among them, immune checkpoint therapy-targeting PD-1/PD-L1 pathway has achieved remarkable success in various types of tumors. For example, the most dramatic effects have been observed in metastatic melanoma, a malignancy that only slightly responds to conventional chemotherapy, and the historical average survival time of metastatic melanoma patients is less than 1 year.<sup>378</sup> Surprisingly, when combined with anti-CTLA4 and anti-PD-1 therapy, nearly 60% of the patients can achieve radiographic responses, with a median survival time of more than 3 years.<sup>379,380</sup> In a Phase II trial, 42 patients with brain metastases from non-small-cell lung cancer (NSCLC) were recruited to receive PD-1 targeting inhibitor pembrolizumab, 11 of whom responded, and the median followup time was 8.3 months.<sup>381</sup> In addition, clinical studies have shown that the PD-L1 inhibitor atezolizumab can significantly improve the prognosis of patients with advanced or metastatic solid tumors.<sup>382</sup> Pembrolizumab and atezolizumab have been approved as first-line or second-line therapies for some patients with advanced or metastatic solid tumors, such as NSCLC, kidney cancer, breast cancer, and melanoma.<sup>381–383</sup>

However, only a limited number of patients can benefit from immunotherapy, and some patients who initially respond to immunotherapy may eventually relapse and progress.<sup>384</sup> Therefore, a large number of pre-clinical and clinical studies have investigated the combination of immunotherapy with other therapies, including chemotherapy, radiotherapy, or targeted therapy, as well as the combination of ICIs targeting different immune checkpoints to overcome the dilemma. To date, many therapies combined with immunotherapy and chemotherapy or anti-angiogenic drugs have been recommended as first-line treatments for multiple solid tumors.<sup>385,386</sup> For example, in patients with squamous NSCLC, the addition of pembrolizumab to chemotherapy (carboplatin plus paclitaxel or nab-paclitaxel) result in significantly better overall survival and progression-free survival than chemotherapy alone.<sup>387</sup> As a result, the combination of pembrolizumab with chemotherapy is recommended as the first-line treatment for patients with advanced squamous NSCLC.<sup>388</sup> In advanced renal cell carcinoma, the FDA approved pembrolizumab combined with lenvatinib or axitinib as the first-line treatment.<sup>389,391</sup> Among them, pembrolizumab combined with axitinib is the first anti-PD-1 antibody plus targeted drug combination therapy approved by the FDA.<sup>390,391</sup> Moreover, it has been reported

LIU ET AL.

TABLE 2 Important immune checkpoint inhibitors under clinical trial and on the market

**MedComm** 

T 4			DI III	0 11 11
larget	Phase I	Phase II	Phase III	On the market
PD-1	CS1003	MGA012	Cemiplimab	Nivolumab
	ZKAB001	GLS-010	Camrelizumab	Pembrolizumab
	MK-3475	Balstilimab	HLX10	Sintilimab
	PF-06801591	SG001	Penpulimab	Tislelizumab
	AGEN1777	BGB A317	REGN2810	Dostarlimab
	609A	Retifanlimab	Spartalizumab	Toripalimab
	AMP-224	Zimberelimab	JS001	
			PF-06801591	
			INCMGA00012	
			BCD-100	
			IBI308	
			JNJ-63723283	
PD-L1	LY3300054	STI-3031	ZKAB001	Atezolizumab
	KN035	CS1001	SHR-1316	Avelumab
	BMS-936559	BGB-A333		Durvalumab
	HLX20	LP002		Camrelizumab
	MSB2311	Bintrafusp alfa		
	BCD-135			
CTLA4	CS1002	Quavonlimab	Tremelimumab	Ipilimumab
	BCD-145	AGEN1884	MDX-010	
	ADU-1604	BCD-217		
	ONC-392	BMS-986218		
	ADG126	CP 675,206		
	ADG116	IBI310		
TIGIT	JS006	EOS-448	BGB-A1217	
	ASP8374	Ociperlimab	Tiragolumab	
	COM902	BMS-986207		
	AZD2936	Etigilimab		
	EOS-448			
	IBI939			
LAG-3	REGN3767	IMP321		
	TSR-033	Relatlimab		
	Sym022	LAG525		
		INCAGN02385		
TIM-3	Sym023	TSR-022		
	INCAGN2390	MBG453		
	LY3321367	BMS-986258		
	SHR-1702	INCAGN02390		
		Cobolimab		
VISTA	JNJ-61610588			
В7-Н3	MGD009		Enoblituzumab	
Dual PD-1/PD-L1		IBI318		
Dual PD-1/TIGIT		AZD2936		
Dual PD-1/TIM-3	RO7121661	AZD7789		

(Continues)

#### TABLE 2 (Continued)

Target	Phase I	Phase II	Phase III	On the market
Dual PD-1/LAG-3	RO7247669		MGD013	
	MGD013			
Dual PD-1/VEGF		AK112		
Dual PD-1/CTLA4		AK104		
		BCD-217		
Dual PD-L1/LAG-3	FS118	RO7247669		
	IBI323			
Dual PD-L1/TIM-3	LY3415244	RO7121661		
Dual PD-L1/4-1BB		ABL503		
Dual PD-L1/VISTA	CA-170			
Dual PD-L1/TGF- $\beta$	Y101D	SHR1701	M7824	

Note: All the data source information is from ClinicalTrials.gov.

Abbreviations: CTLA4, cytotoxic lymphocyte-associated protein 4; LAG-3, lymphocyte activation gene 3; PD-1, programmed cell death 1; PD-L1, programmed death ligand-1; TIGIT, T cell immunoglobulin and ITIM domain; TIM-3, T-cell immunoglobulin-3.

that radiotherapy can improve the response of patients with metastatic NSCLC to immunotherapy.<sup>392</sup> Overall, a broad range of immunotherapy-based combination therapies have been approved for the treatment of cancer, and a large number of preclinical studies and clinical trials are ongoing to explore more possibilities for combination therapies in the treatment of metastatic cancer.

# 3.4 | Others

In recent years, due to the abundance of natural herbal compounds and the diversity of their chemical compositions, they have received more and more attention as antitumor drugs. Triptolide, anthocyanidins, and gigantol have been confirmed to inhibit the proliferation and invasion of NSCLC cells, glioblastoma cells, and bladder cancer cells in vivo and in vitro.<sup>393–395</sup> Studies have found that the antitumor effects of certain compounds are mainly achieved by inhibiting EMT.<sup>394,395</sup> In addition, the chemical compound DZ-50 and the natural product curcumol can inhibit tumor metastasis by weakening the resistance of tumor cells to anoikis.<sup>396,397</sup> However, their anti-tumor effects need to be confirmed by further clinical studies.

The failure of metastatic cancer treatment may be partly due to the inability of drugs to persist in the blood or lymph and the inability of drugs to cross certain natural barriers. For example, for patients with metastatic brain cancer, the difficulty in treatment is that most anti-tumor drugs cannot cross the blood-brain barrier. The emergence of nanomedicine has brought revolutionary updates to tumor treatment. Due to the small molecular weight of nanometers and the ability to carry specific pharmacophores to distant metastatic regions, nano-related chemotherapy and nano-related immunotherapy have demonstrated outstanding potential in tackling metastatic cancer.<sup>398,399</sup>

The combination of different nano-polymer materials (such as hydrogels, micelles, and nanoparticles) and chemotherapeutic drugs, small molecule inhibitors, or immunotherapy drugs, has successfully improved the control rate of metastatic tumor.400-403 For example, losartan-loaded injectable peptide hydrogel can inhibit the formation of CAFs collagen, thereby increasing the concentration of adriamycin at the metastatic sites and killing tumor cells.<sup>400</sup> Succinobucol is a small molecule inhibitor targeting vascular cell adhesion molecule-1, which is believed to be able to reverse lung metastasis, but the results in in vivo experiments are not satisfactory. Then, He et al. successfully designed succinobucol on ph-responsive wormlike micelles as a novel nanomedicine. Both in vivo and in vitro experiments demonstrated that succinobucol could be successfully delivered to the site of metastasis through ph-responsive wormlike micelles, thus improving treatment efficiency.<sup>401</sup> In addition, nanotechnology can enhance the anti-tumor effect of immunotherapy by rebuilding the tumor immune microenvironment, enhancing the anti-tumor effect of immune cells, enhancing the effectiveness of anti-tumor cytokines, and awakening tumor cells to release antigens.<sup>398,399</sup> Moreover, nanotechnology may reduce the side effects of immunotherapy.

# 4 | CONCLUSIONS AND FUTURE PERSPECTIVES

Metastasis accounts for the great majority of cancerrelated deaths, which is the result of the interaction between tumor cells and TME components. In contrast to numerous discoveries that have revealed the detailed mechanisms leading to the formation of primary tumor, the biological underpinnings of metastatic disease are still poorly understood. However, some progress has been made in elucidating the cellular and molecular mechanisms that drive cancer metastasis over the past decades.

Despite the diversity of tumor types, the mechanisms of metastasis have some commonalities. The invasionmetastasis cascade involves angiogenesis, detachment of metastatic cells from the primary tumor, invasion through the BM and ECM rounding tumor, intravasation of the metastatic cells into the blood and lymphatic vessels and survival during hematogenous transit, adhesion of these cells to the endothelium of capillaries, extravasation through vascular walls into the parenchyma of distant tissues, and colonization at the target organ site.<sup>17,23,404</sup> The EMT and anoikis resistance of tumor cells are the main forces to promote the metastasis cascade, and multiple components in TME are inducers to ensure the success of metastasis by reprogramming the pre-metastasis niche and promoting metastatic cell growth.21,35,325 EMT modulates cell-to-cell or cell-to-matrix adhesion and allows cells to increase their migration and invasion capabilities by forming invasive protrusions.<sup>405</sup> Metastatic cells develop an ability to resist anoikis, enabling them to adapt and survive when detached from the ECM or in the absence of the ECM.<sup>19</sup> The interactions between tumor and ECM components are essential for EMT and acquisition of tumor invasive abilities, and TME is also closely involved with all the processes of metastasis.<sup>69</sup> As presented earlier, immune cells (e.g., TAMs, Treg, and neutrophil), stromal cells (e.g., CAFs, MSCs, and TECs), chemokines, cytokines, and growth factors in TME, are involved in a complex crosstalk with tumor cells that affects tumor growth and metastasis.<sup>69,79,138,406</sup> Furthermore, recent studies have emphasized the role of autophagy, lipid metabolism, and lncRNA in tumor metastasis.<sup>212,240,255</sup>

Those discoveries have brought new insight into our understanding of cancer metastasis. For most patients, due to the insidious tumor-related symptoms, distant metastasis has already appeared at the time of diagnosis.<sup>1</sup> In addition to the three cornerstones of tumor treatment, surgery, chemotherapy, and radiotherapy, novel treatment approaches, including targeted therapy and immunotherapy, have been established in patients with metastatic cancer. According to the chronological order and biological characteristics of metastasis, intervention strategies can be roughly classified into four types: early prevention, blocking metastasis-specific pathways, reducing angiogenesis, and enhancing anti-tumor immunity. For low-grade early malignancies, preoperative neoadjuvant chemotherapy and postoperative adjuvant chemotherapy can prevent and reduce the probability of recurrence **MedComm** 

and metastasis to a certain extent.<sup>298,301</sup> For advanced or metastatic tumors, targeted therapies, especially drugs targeting metastasis-specific pathways and angiogenesis have been approved for the treatment of most types of tumors.<sup>15</sup> What is more exciting is that immunotherapy, especially ICIs, is revolutionizing the treatment of malignancies and has demonstrated their potential as "cancer terminators." The advent of immunotherapy and combination therapies based on immunotherapy has provided more options for patients with metastatic cancer. Despite many challenges, we hold an optimistic view of the future prospect of immunotherapy.

In summary, this review provided our current understanding of the mechanisms that underlie the dissemination and metastatic outgrowth of cancer cells. As presented above, although many critical obstacles are still lying ahead, tumor metastasis-related pathways and TME represent novel and attractive directions, which may change the landscape of cancer therapy in the future. However, further researches and more endeavors in this area are needed.

#### ACKNOWLEDGMENTS

This work was supported by the National Natural Science Foundation of China (No. 81772863 and 82072958) and the China Postdoctoral Science Foundation-funded project (No. 2020M683119). This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

#### CONFLICT OF INTEREST

We declare that we do not have any commercial or associative interest that represents a conflict of interest in connection with the work submitted.

#### ETHICS APPROVAL

Not applicable.

#### **DATA AVAILABILITY STATEMENT** Not applicable.

#### AUTHOR CONTRIBUTIONS

ML and YJ: data curation, formal analysis, and writin goriginal draft. BX: data curation and formal analysis. XZ: conceptualization, supervision, validation, and writing review editing. All authors read and approved the final manuscript.

#### REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin. 2020;70(1):7-30.
- Jiang WG, Sanders AJ, Katoh M, et al. Tissue invasion and metastasis: Molecular, biological and clinical perspectives. *Semin Cancer Biol.* 2015;35:S244-S275.

# MedComm

- Gupta GP, Massague J. Cancer metastasis: building a framework. *Cell*. 2006;127(4):679-695.
- 4. Paget S. The distribution of secondary growths in cancer of the breast. 1889. *Cancer Metastasis Rev.* 1989;8(2):98-101.
- 5. Fidler IJ, Poste G. The "seed and soil" hypothesis revisited. *Lancet Oncol.* 2008;9(8):808.
- Scanlon EF. James Ewing lecture. The process of metastasis. Cancer. 1985;55(6):1163-1166.
- Brown M, Assen FP, Leithner A, et al. Lymph node blood vessels provide exit routes for metastatic tumor cell dissemination in mice. *Science*. 2018;359(6382):1408-1411.
- Turajlic S, Swanton C. Metastasis as an evolutionary process. Science. 2016;352(6282):169-175.
- 9. Lambert AW, Pattabiraman DR, Weinberg RA. Emerging biological principles of metastasis. *Cell*. 2017;168(4):670-691.
- Lunt SJ, Chaudary N, Hill RP. The tumor microenvironment and metastatic disease. *Clin Exp Metastasis*. 2009;26(1):19-34.
- Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med.* 2013;19(11):1423-1437.
- 12. Zhou Z, Lu ZR. Molecular imaging of the tumor microenvironment. *Adv Drug Deliv Rev.* 2017;113:24-48.
- Edwards SC, Hoevenaar WHM, Coffelt SB. Emerging immunotherapies for metastasis. *Br J Cancer*. 2021;124(1):37-48.
- Chyuan IT, Chu CL, Hsu PN. Targeting the tumor microenvironment for improving therapeutic effectiveness in cancer immunotherapy: focusing on immune checkpoint inhibitors and combination therapies. *Cancers (Basel)*. 2021;13(6):1188.
- Yang H, Kuo YH, Smith ZI, Spangler J. Targeting cancer metastasis with antibody therapeutics. *Wiley Interdiscip Rev Nanomed Nanobiotechnol.* 2021;13(4):e1698.
- 16. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674.
- Nguyen DX, Bos PD, Massague J. Metastasis: from dissemination to organ-specific colonization. *Nat Rev Cancer*. 2009;9(4):274-284.
- Theocharis AD, Skandalis SS, Gialeli C, Karamanos NK. Extracellular matrix structure. Adv Drug Deliv Rev. 2016;97:4-27.
- 19. Simpson CD, Anyiwe K, Schimmer AD. Anoikis resistance and tumor metastasis. *Cancer Lett.* 2008;272(2):177-185.
- Liu Y, Cao X. Characteristics and significance of the premetastatic niche. *Cancer Cell*. 2016;30(5):668-681.
- 21. Peinado H, Zhang H, Matei IR, et al. Pre-metastatic niches: organ-specific homes for metastases. *Nat Rev Cancer*. 2017;17(5):302-317.
- 22. Wang H, Pan J, Barsky L, et al. Characteristics of pre-metastatic niche: the landscape of molecular and cellular pathways. *Mol Biomed*. 2021;2(1):3.
- Chambers AF, Groom AC, MacDonald IC. Dissemination and growth of cancer cells in metastatic sites. *Nat Rev Cancer*. 2002;2(8):563-572.
- Klein CA. Framework models of tumor dormancy from patient-derived observations. *Curr Opin Genet Dev.* 2011;21(1):42-49.
- Celia-Terrassa T, Kang Y. Distinctive properties of metastasisinitiating cells. *Genes Dev.* 2016;30(8):892-908.
- Visvader JE, Lindeman GJ. Cancer stem cells in solid tumours: accumulating evidence and unresolved questions. *Nat Rev Cancer*. 2008;8(10):755-768.

- Malanchi I, Santamaria-Martinez A, Susanto E, et al. Interactions between cancer stem cells and their niche govern metastatic colonization. *Nature*. 2011;481(7379):85-89.
- Yang J, Antin P, Berx G, et al. Guidelines and definitions for research on epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol.* 2020;21(6):341-352.
- Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelialmesenchymal transitions in development and disease. *Cell*. 2009;139(5):871-890.
- 30. Greenburg G, Hay ED. Epithelia suspended in collagen gels can lose polarity and express characteristics of migrating mesenchymal cells. *J Cell Biol*. 1982;95(1):333-339.
- Bakir B, Chiarella AM, Pitarresi JR, Rustgi AK. EMT, MET, plasticity, and tumor metastasis. *Trends Cell Biol.* 2020;30(10):764-776.
- Davis FM, Stewart TA, Thompson EW, Monteith GR. Targeting EMT in cancer: opportunities for pharmacological intervention. *Trends Pharmacol Sci.* 2014;35(9):479-488.
- Huber MA, Kraut N, Beug H. Molecular requirements for epithelial-mesenchymal transition during tumor progression. *Curr Opin Cell Biol*. 2005;17(5):548-558.
- Cano A, Perez-Moreno MA, Rodrigo I, et al. The transcription factor snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression. *Nat Cell Biol.* 2000;2(2):76-83.
- Hotz B, Arndt M, Dullat S, Bhargava S, Buhr HJ, Hotz HG. Epithelial to mesenchymal transition: expression of the regulators snail, slug, and twist in pancreatic cancer. *Clin Cancer Res.* 2007;13(16):4769-4776.
- van Roy F. Beyond E-cadherin: roles of other cadherin superfamily members in cancer. *Nat Rev Cancer*. 2014;14(2):121-134.
- Li ZH, Zhou Y, Ding YX, Guo QL, Zhao L. Roles of integrin in tumor development and the target inhibitors. *Chin J Nat Med*. 2019;17(4):241-251.
- Nieman MT, Prudoff RS, Johnson KR, Wheelock MJ. N-cadherin promotes motility in human breast cancer cells regardless of their E-cadherin expression. *J Cell Biol.* 1999;147(3):631-644.
- Morgan RG, Pearn L, Liddiard K, et al. Gamma-catenin is overexpressed in acute myeloid leukemia and promotes the stabilization and nuclear localization of beta-catenin. *Leukemia*. 2013;27(2):336-343.
- 40. Liu LK, Jiang XY, Zhou XX, Wang DM, Song XL, Jiang HB. Upregulation of vimentin and aberrant expression of Ecadherin/beta-catenin complex in oral squamous cell carcinomas: correlation with the clinicopathological features and patient outcome. *Mod Pathol.* 2010;23(2):213-224.
- Jin W. Role of JAK/STAT3 signaling in the regulation of metastasis, the transition of cancer stem cells, and chemoresistance of cancer by epithelial-mesenchymal transition. *Cells*. 2020;9(1):217.
- Yuan X, Wu H, Han N, et al. Notch signaling and EMT in nonsmall cell lung cancer: biological significance and therapeutic application. *J Hematol Oncol.* 2014;7:87.
- 43. Zhang N, Ji J, Zhou D, et al. The interaction of the senescent and adjacent breast cancer cells promotes the metastasis of heterogeneous breast cancer cells through Notch signaling. *Int J Mol Sci.* 2021;22(2):849.

- Cai J, Fang L, Huang Y, et al. Simultaneous overactivation of Wnt/beta-catenin and TGFbeta signalling by miR-128-3p confers chemoresistance-associated metastasis in NSCLC. *Nat Commun.* 2017;8:15870.
- Patel A, Sabbineni H, Clarke A, Somanath PR. Novel roles of Src in cancer cell epithelial-to-mesenchymal transition, vascular permeability, microinvasion and metastasis. *Life Sci.* 2016;157:52-61.
- 46. Qi LN, Xiang BD, Wu FX, et al. Circulating tumor cells undergoing EMT provide a metric for diagnosis and prognosis of patients with hepatocellular carcinoma. *Cancer Res.* 2018;78(16):4731-4744.
- Fan F, Samuel S, Evans KW, et al. Overexpression of snail induces epithelial-mesenchymal transition and a cancer stem cell-like phenotype in human colorectal cancer cells. *Cancer Med.* 2012;1(1):5-16.
- Yu M, Bardia A, Wittner BS, et al. Circulating breast tumor cells exhibit dynamic changes in epithelial and mesenchymal composition. *Science*. 2013;339(6119):580-584.
- 49. Guan X, Ma F, Li C, et al. The prognostic and therapeutic implications of circulating tumor cell phenotype detection based on epithelial-mesenchymal transition markers in the first-line chemotherapy of HER2-negative metastatic breast cancer. *Cancer Commun (Lond)*. 2019;39(1):1.
- Zhang Q, Rong Y, Yi K, Huang L, Chen M, Wang F. Circulating tumor cells in hepatocellular carcinoma: single-cell based analysis, preclinical models, and clinical applications. *Theranostics*. 2020;10(26):12060-12071.
- Mani SA, Guo W, Liao MJ, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell*. 2008;133(4):704-715.
- Chaffer CL, Brueckmann I, Scheel C, et al. Normal and neoplastic nonstem cells can spontaneously convert to a stem-like state. *Proc Natl Acad Sci U S A*. 2011;108(19):7950-7955.
- 53. Guo W, Keckesova Z, Donaher JL, et al. Slug and Sox9 cooperatively determine the mammary stem cell state. *Cell*. 2012;148(5):1015-1028.
- 54. Okuda H, Kobayashi A, Xia B, et al. Hyaluronan synthase HAS2 promotes tumor progression in bone by stimulating the interaction of breast cancer stem-like cells with macrophages and stromal cells. *Cancer Res.* 2012;72(2):537-547.
- Ye X, Tam WL, Shibue T, et al. Distinct EMT programs control normal mammary stem cells and tumour-initiating cells. *Nature*. 2015;525(7568):256-260.
- Jiang Y, Zhan H. Communication between EMT and PD-L1 signaling: New insights into tumor immune evasion. *Cancer Lett.* 2020;468:72-81.
- 57. Dongre A, Rashidian M, Reinhardt F, et al. Epithelial-tomesenchymal transition contributes to Immunosuppression in breast carcinomas. *Cancer Res.* 2017;77(15):3982-3989.
- Noman MZ, Janji B, Abdou A, et al. The immune checkpoint ligand PD-L1 is upregulated in EMT-activated human breast cancer cells by a mechanism involving ZEB-1 and miR-200. *Oncoimmunology*. 2017;6(1):e1263412.
- Kursunel MA, Taskiran EZ, Tavukcuoglu E, et al. Small cell lung cancer stem cells display mesenchymal properties and exploit immune checkpoint pathways in activated cytotoxic T lymphocytes. [published online ahead of print, 2021 Jul 6].

Cancer Immunol Immunother. 2021. https://doi.org/10.1007/ s00262-021-02998-1

- 60. Lou Y, Diao L, Cuentas ER, et al. Epithelial-mesenchymal transition is associated with a distinct tumor microenvironment including elevation of inflammatory signals and multiple immune checkpoints in lung adenocarcinoma. *Clin Cancer Res.* 2016;22(14):3630-3642.
- Jolly MK, Ware KE, Gilja S, Somarelli JA, Levine H. EMT and MET: necessary or permissive for metastasis? *Mol Oncol.* 2017;11(7):755-769.
- 62. Lin WH, Chang YW, Hong MX, et al. STAT3 phosphorylation at Ser727 and Tyr705 differentially regulates the EMT-MET switch and cancer metastasis. *Oncogene*. 2021;40(4):791-805.
- Lee J, You JH, Kim MS, Roh JL. Epigenetic reprogramming of epithelial-mesenchymal transition promotes ferroptosis of head and neck cancer. *Redox Biol.* 2020;37:101697.
- Skrypek N, Goossens S, De Smedt E, Vandamme N, Berx G. Epithelial-to-mesenchymal transition: epigenetic reprogramming driving cellular plasticity. *Trends Genet*. 2017;33(12):943-959.
- Jeong GY, Park MK, Choi HJ, et al. NSD3-induced methylation of H3K36 activates notch signaling to drive breast tumor initiation and metastatic progression. *Cancer Res.* 2021;81(1):77-90.
- 66. Terashima M, Ishimura A, Wanna-Udom S, Suzuki T. Epigenetic regulation of epithelial-mesenchymal transition by KDM6A histone demethylase in lung cancer cells. *Biochem Biophys Res Commun.* 2017;490(4):1407-1413.
- 67. Taube JH, Sphyris N, Johnson KS, et al. The H3K27me3demethylase KDM6A is suppressed in breast cancer stem-like cells, and enables the resolution of bivalency during the mesenchymal-epithelial transition. *Oncotarget*. 2017;8(39):65548-65565.
- 68. Giraldo NA, Sanchez-Salas R, Peske JD, et al. The clinical role of the TME in solid cancer. *Br J Cancer*. 2019;120(1):45-53.
- Neophytou CM, Panagi M, Stylianopoulos T, Papageorgis P. The role of tumor microenvironment in cancer metastasis: molecular mechanisms and therapeutic opportunities. *Cancers* (*Basel*). 2021;13(9):2053.
- Ren B, Cui M, Yang G, et al. Tumor microenvironment participates in metastasis of pancreatic cancer. *Mol Cancer*. 2018;17(1):108.
- Ye LY, Chen W, Bai XL, et al. Hypoxia-induced epithelial-tomesenchymal transition in hepatocellular carcinoma induces an immunosuppressive tumor microenvironment to promote metastasis. *Cancer Res.* 2016;76(4):818-830.
- 72. Hibino S, Kawazoe T, Kasahara H, et al. Inflammation-induced tumorigenesis and metastasis. *Int J Mol Sci.* 2021;22(11):5421.
- Maia A, Wiemann S. Cancer-associated fibroblasts: implications for cancer therapy. *Cancers (Basel)*. 2021;13(14):3526.
- Karnoub AE, Dash AB, Vo AP, et al. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature*. 2007;449(7162):557-563.
- Maishi N, Hida K. Tumor endothelial cells accelerate tumor metastasis. *Cancer Sci.* 2017;108(10):1921-1926.
- Maacha S, Bhat AA, Jimenez L, et al. Extracellular vesiclesmediated intercellular communication: roles in the tumor microenvironment and anti-cancer drug resistance. *Mol Cancer*. 2019;18(1):55.

- Yang L, Lin PC. Mechanisms that drive inflammatory tumor microenvironment, tumor heterogeneity, and metastatic progression. *Semin Cancer Biol*. 2017;47:185-195.
- Wang N, Liang H, Zen K. Molecular mechanisms that influence the macrophage m1-m2 polarization balance. *Front Immunol.* 2014;5:614.
- 79. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell*. 2010;141(1):39-51.
- Movahedi K, Laoui D, Gysemans C, et al. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. *Cancer Res.* 2010;70(14):5728-5739.
- Lin Y, Xu J, Lan H. Tumor-associated macrophages in tumor metastasis: biological roles and clinical therapeutic applications. *J Hematol Oncol.* 2019;12(1):76.
- Korbecki J, Grochans S, Gutowska I, Barczak K, Baranowska-Bosiacka I. CC chemokines in a tumor: a review of pro-cancer and anti-cancer properties of receptors CCR5, CCR6, CCR7, CCR8, CCR9, and CCR10 ligands. *Int J Mol Sci.* 2020;21(20):7619.
- Mantovani A, Allavena P, Sozzani S, Vecchi A, Locati M, Sica A. Chemokines in the recruitment and shaping of the leukocyte infiltrate of tumors. *Semin Cancer Biol*. 2004;14(3):155-160.
- Marcovecchio PM, Thomas G, Salek-Ardakani S. CXCL9expressing tumor-associated macrophages: new players in the fight against cancer. *J Immunother Cancer*. 2021;9(2):e002045.
- 85. Sica A, Schioppa T, Mantovani A, Allavena P. Tumourassociated macrophages are a distinct M2 polarised population promoting tumour progression: potential targets of anti-cancer therapy. *Eur J Cancer*. 2006;42(6):717-727.
- Fu XT, Dai Z, Song K, et al. Macrophage-secreted IL-8 induces epithelial-mesenchymal transition in hepatocellular carcinoma cells by activating the JAK2/STAT3/Snail pathway. *Int J Oncol.* 2015;46(2):587-596.
- Helm O, Held-Feindt J, Grage-Griebenow E, et al. Tumorassociated macrophages exhibit pro- and anti-inflammatory properties by which they impact on pancreatic tumorigenesis. *Int J Cancer.* 2014;135(4):843-861.
- Vasiljeva O, Papazoglou A, Kruger A, et al. Tumor cellderived and macrophage-derived cathepsin B promotes progression and lung metastasis of mammary cancer. *Cancer Res.* 2006;66(10):5242-5250.
- Lin EY, Li JF, Gnatovskiy L, et al. Macrophages regulate the angiogenic switch in a mouse model of breast cancer. *Cancer Res.* 2006;66(23):11238-11246.
- Lin EY, Pollard JW. Tumor-associated macrophages press the angiogenic switch in breast cancer. *Cancer Res.* 2007;67(11):5064-5066.
- Lewis CE, De Palma M, Naldini L. Tie2-expressing monocytes and tumor angiogenesis: regulation by hypoxia and angiopoietin-2. *Cancer Res.* 2007;67(18):8429-8432.
- 92. Chen ML, Pittet MJ, Gorelik L, et al. Regulatory T cells suppress tumor-specific CD8 T cell cytotoxicity through TGF-beta signals in vivo. *Proc Natl Acad Sci U S A*. 2005;102(2):419-424.
- McNally A, Hill GR, Sparwasser T, Thomas R, Steptoe RJ. CD4+CD25+ regulatory T cells control CD8+ T-cell effector differentiation by modulating IL-2 homeostasis. *Proc Natl Acad Sci U S A*. 2011;108(18):7529-7534.

- 94. Du Y, Chen X, Lin XQ, Wu W, Huang ZM. Tumor-derived CD4+CD25+ Tregs inhibit the maturation and antigenpresenting function of dendritic cells. *Asian Pac J Cancer Prev.* 2015;16(7):2665-2669.
- Olkhanud PB, Baatar D, Bodogai M, et al. Breast cancer lung metastasis requires expression of chemokine receptor CCR4 and regulatory T cells. *Cancer Res.* 2009;69(14):5996-6004.
- 96. Huang L, Guo Y, Liu S, et al. Targeting regulatory T cells for immunotherapy in melanoma. *Mol Biomed*. 2021;2(1):11.
- Li C, Jiang P, Wei S, Xu X, Wang J. Regulatory T cells in tumor microenvironment: new mechanisms, potential therapeutic strategies and future prospects. *Mol Cancer*. 2020;19(1):116.
- Toomer KH, Malek TR. Cytokine signaling in the development and homeostasis of regulatory T cells. *Cold Spring Harb Perspect Biol.* 2018;10(3):a028597.
- Budhu S, Schaer DA, Li Y, et al. Blockade of surface-bound TGF-beta on regulatory T cells abrogates suppression of effector T cell function in the tumor microenvironment. *Sci Signal*. 2017;10(494):eaak9702.
- 100. Wei X, Zhang J, Gu Q, et al. Reciprocal expression of IL-35 and IL-10 defines two distinct effector Treg subsets that are required for maintenance of immune tolerance. *Cell Rep.* 2017;21(7):1853-1869.
- 101. Huang SC, Wei PC, Hwang-Verslues WW, et al. TGF-betal secreted by Tregs in lymph nodes promotes breast cancer malignancy via up-regulation of IL-17RB. *EMBO Mol Med.* 2017;9(12):1660-1680.
- 102. Bos PD, Plitas G, Rudra D, Lee SY, Rudensky AY. Transient regulatory T cell ablation deters oncogene-driven breast cancer and enhances radiotherapy. J Exp Med. 2013;210(11):2435-2466.
- 103. Camisaschi C, Casati C, Rini F, et al. LAG-3 expression defines a subset of CD4(+) CD25(high)Foxp3(+) regulatory T cells that are expanded at tumor sites. *J Immunol*. 2010;184(11):6545-6551.
- Delgoffe GM, Woo SR, Turnis ME, et al. Stability and function of regulatory T cells is maintained by a neuropilin-1semaphorin-4a axis. *Nature*. 2013;501(7466):252-256.
- 105. Liu C, Chikina M, Deshpande R, et al. Treg cells promote the SREBP1-dependent metabolic fitness of tumorpromoting macrophages via repression of CD8(+) T cellderived interferon-gamma. *Immunity*. 2019;51(2):381-397.e6.
- 106. Ohta A, Kini R, Ohta A, Subramanian M, Madasu M, Sitkovsky M. The development and immunosuppressive functions of CD4(+) CD25(+) FoxP3(+) regulatory T cells are under influence of the adenosine-A2A adenosine receptor pathway. *Front Immunol.* 2012;3:190.
- 107. Carmenate T, Ortiz Y, Enamorado M, et al. Blocking IL-2 signal in vivo with an IL-2 antagonist reduces tumor growth through the control of regulatory T cells. *J Immunol.* 2018;200(10):3475-3484.
- Spolski R, Li P, Leonard WJ. Biology and regulation of IL-2: from molecular mechanisms to human therapy. *Nat Rev Immunol.* 2018;18(10):648-659.
- 109. Park YJ, Ryu H, Choi G, et al. IL-27 confers a protumorigenic activity of regulatory T cells via CD39. *Proc Natl Acad Sci U S A*. 2019;116(8):3106-3111.
- 110. Liu J, Zhang H, Jia L, Sun H. Effects of Treg cells and IDO on human epithelial ovarian cancer cells under hypoxic conditions. *Mol Med Rep.* 2015;11(3):1708-1714.

- 111. Ihara F, Sakurai D, Takami M, et al. Regulatory T cells induce CD4(-) NKT cell anergy and suppress NKT cell cytotoxic function. *Cancer Immunol Immunother*. 2019;68(12):1935-1947.
- 112. Hoechst B, Ormandy LA, Ballmaier M, et al. A new population of myeloid-derived suppressor cells in hepatocellular carcinoma patients induces CD4(+) CD25(+)Foxp3(+) T cells. *Gastroenterology*. 2008;135(1):234-243.
- Morello S, Pinto A, Blandizzi C, Antonioli L. Myeloid cells in the tumor microenvironment: role of adenosine. *Oncoimmunology*. 2016;5(3):e1108515.
- 114. Ghiringhelli F, Puig PE, Roux S, et al. Tumor cells convert immature myeloid dendritic cells into TGF-beta-secreting cells inducing CD4+CD25+ regulatory T cell proliferation. *J Exp Med.* 2005;202(7):919-929.
- 115. Facciabene A, Peng X, Hagemann IS, et al. Tumour hypoxia promotes tolerance and angiogenesis via CCL28 and T(reg) cells. *Nature*. 2011;475(7355):226-230.
- 116. Shi C, Chen Y, Chen Y, Yang Y, Bing W, Qi J. CD4(+) CD25(+) regulatory T cells promote hepatocellular carcinoma invasion via TGF-beta1-induced epithelial-mesenchymal transition. Onco Targets Ther. 2019;12:279-289.
- 117. Nunez NG, Tosello Boari J, Ramos RN, et al. Tumor invasion in draining lymph nodes is associated with Treg accumulation in breast cancer patients. *Nat Commun.* 2020;11(1):3272.
- 118. Liew PX, Kubes P. The neutrophil's role during health and disease. *Physiol Rev.* 2019;99(2):1223-1248.
- 119. Coffelt SB, Kersten K, Doornebal CW, et al. IL-17-producing gamma delta T cells and neutrophils conspire to promote breast cancer metastasis. *Nature*. 2015;522(7556):345-348.
- 120. Donadio AC, Remedi MM, Frede S, Bonacci GR, Chiabrando GA, Pistoresi-Palencia MC. Decreased expression of intercellular adhesion molecule-1 (ICAM-1) and urokinase-type plasminogen activator receptor (uPAR) is associated with tumor cell spreading in vivo. *Clin Exp Metastasis*. 2002;19(5):437-444.
- 121. Li XJ, Peng LX, Shao JY, et al. As an independent unfavorable prognostic factor, IL-8 promotes metastasis of nasopharyngeal carcinoma through induction of epithelial-mesenchymal transition and activation of AKT signaling. *Carcinogenesis*. 2012;33(7):1302-1309.
- 122. Li S, Cong X, Gao H, et al. Tumor-associated neutrophils induce EMT by IL-17a to promote migration and invasion in gastric cancer cells. *J Exp Clin Cancer Res.* 2019;38(1):6.
- Szczerba BM, Castro-Giner F, Vetter M, et al. Neutrophils escort circulating tumour cells to enable cell cycle progression. *Nature*. 2019;566(7745):553-557.
- Wculek SK, Malanchi I. Neutrophils support lung colonization of metastasis-initiating breast cancer cells. *Nature*. 2015;528(7582):413-417.
- 125. Tyagi A, Sharma S, Wu K, et al. Nicotine promotes breast cancer metastasis by stimulating N2 neutrophils and generating pre-metastatic niche in lung. *Nat Commun.* 2021;12(1):474.
- 126. Gong L, Cumpian AM, Caetano MS, et al. Promoting effect of neutrophils on lung tumorigenesis is mediated by CXCR2 and neutrophil elastase. *Mol Cancer*. 2013;12(1):154.
- 127. Ringuette Goulet C, Bernard G, Tremblay S, Chabaud S, Bolduc S, Pouliot F. Exosomes induce fibroblast differentiation into cancer-associated fibroblasts through TGF-beta signaling. *Mol Cancer Res.* 2018;16(7):1196-1204.

- 128. Yeon JH, Jeong HE, Seo H, et al. Cancer-derived exosomes trigger endothelial to mesenchymal transition followed by the induction of cancer-associated fibroblasts. *Acta Biomater*. 2018;76:146-153.
- 129. Tan HX, Cao ZB, He TT, Huang T, Xiang CL, Liu Y. TGFbetal is essential for MSCs-CAFs differentiation and promotes HCT116 cells migration and invasion via JAK/STAT3 signaling. Onco Targets Ther. 2019;12:5323-5334.
- Erdogan B, Webb DJ. Cancer-associated fibroblasts modulate growth factor signaling and extracellular matrix remodeling to regulate tumor metastasis. *Biochem Soc Trans.* 2017;45(1):229-236.
- 131. Fiori ME, Di Franco S, Villanova L, Bianca P, Stassi G, De Maria R. Cancer-associated fibroblasts as abettors of tumor progression at the crossroads of EMT and therapy resistance. *Mol Cancer*. 2019;18(1):70.
- 132. Wang QS, Kong PZ, Li XQ, Yang F, Feng YM. FOXF2 deficiency promotes epithelial-mesenchymal transition and metastasis of basal-like breast cancer. *Breast Cancer Res.* 2015;17:30.
- 133. Lu JT, Tan CC, Wu XR, et al. FOXF2 deficiency accelerates the visceral metastasis of basal-like breast cancer by unrestrictedly increasing TGF-beta and miR-182-5p. *Cell Death Differ*. 2020;27(10):2973-2987.
- 134. Liu G, Sun J, Yang ZF, et al. Cancer-associated fibroblastderived CXCL11 modulates hepatocellular carcinoma cell migration and tumor metastasis through the circUBAP2/miR-4756/IFIT1/3 axis. Cell Death Dis. 2021;12(3):260.
- 135. Fan H, Atiya HI, Wang Y, et al. Epigenomic reprogramming toward mesenchymal-epithelial transition in ovarian-cancerassociated mesenchymal stem cells drives metastasis. *Cell Rep.* 2020;33(10):108473.
- 136. Wang Y, Liang Y, Xu H, et al. Single-cell analysis of pancreatic ductal adenocarcinoma identifies a novel fibroblast subtype associated with poor prognosis but better immunotherapy response. *Cell Discov.* 2021;7(1):36.
- 137. Peng S, Chen D, Cai J, et al. Enhancing cancer-associated fibroblast fatty acid catabolism within a metabolically challenging tumor microenvironment drives colon cancer peritoneal metastasis. *Mol Oncol.* 2021;15(5):1391-1411.
- Wang Z, Liu J, Huang H, et al. Metastasis-associated fibroblasts: an emerging target for metastatic cancer. *Biomark Res.* 2021;9(1):47.
- 139. Gui Y, Aguilar-Mahecha A, Krzemien U, et al. Metastatic breast carcinoma-associated fibroblasts have enhanced protumorigenic properties related to increased IGF2 expression. *Clin Cancer Res.* 2019;25(23):7229-7242.
- 140. Chen IX, Chauhan VP, Posada J, et al. Blocking CXCR4 alleviates desmoplasia, increases T-lymphocyte infiltration, and improves immunotherapy in metastatic breast cancer. *Proc Natl Acad Sci U S A*. 2019;116(10):4558-4566.
- 141. Costa A, Kieffer Y, Scholer-Dahirel A, et al. Fibroblast heterogeneity and immunosuppressive environment in human breast cancer. *Cancer Cell*. 2018;33(3):463-479.e10.
- 142. Bonneau C, Elies A, Kieffer Y, et al. A subset of activated fibroblasts is associated with distant relapse in early luminal breast cancer. *Breast Cancer Res.* 2020;22(1):76.
- 143. Gu JJ, Hoj J, Rouse C, Pendergast AM. Mesenchymal stem cells promote metastasis through activation of an

ABL-MMP9 signaling axis in lung cancer cells. PLoS One. 2020;15(10):e0241423.

- 144. Kalbasi A, Ribas A. Tumour-intrinsic resistance to immune checkpoint blockade. Nat Rev Immunol. 2020;20(1):25-39.
- 145. Aboulkheyr Es H, Bigdeli B, Zhand S, Aref AR, Thiery JP, Warkiani ME. Mesenchymal stem cells induce PD-L1 expression through the secretion of CCL5 in breast cancer cells. J Cell Physiol. 2021;236(5):3918-3928.
- 146. Sun L, Wang Q, Chen B, et al. Gastric cancer mesenchymal stem cells derived IL-8 induces PD-L1 expression in gastric cancer cells via STAT3/mTOR-c-Mvc signal axis. Cell Death Dis. 2018;9(9):928.
- 147. Sun L, Huang C, Zhu M, et al. Gastric cancer mesenchymal stem cells regulate PD-L1-CTCF enhancing cancer stem cell-like properties and tumorigenesis. Theranostics. 2020;10(26):11950-11962.
- 148. Endemann DH, Schiffrin EL. Endothelial dysfunction. J Am Soc Nephrol. 2004;15(8):1983-1992.
- 149. Maishi N, Ohba Y, Akiyama K, et al. Tumour endothelial cells in high metastatic tumours promote metastasis via epigenetic dysregulation of biglycan. Sci Rep. 2016;6:28039.
- 150. Hu L, Zang MD, Wang HX, et al. Biglycan stimulates VEGF expression in endothelial cells by activating the TLR signaling pathway. Mol Oncol. 2016;10(9):1473-1484.
- 151. Lee E, Fertig EJ, Jin K, Sukumar S, Pandey NB, Popel AS. Breast cancer cells condition lymphatic endothelial cells within pre-metastatic niches to promote metastasis. Nat Commun. 2014;5:4715.
- 152. Kai F, Laklai H, Weaver VM. Force matters: biomechanical regulation of cell invasion and migration in disease. Trends Cell Biol. 2016;26(7):486-497.
- 153. Yuzhalin AE, Lim SY, Kutikhin AG, Gordon-Weeks AN. Dynamic matrisome: ECM remodeling factors licensing cancer progression and metastasis. Biochim Biophys Acta Rev Cancer. 2018;1870(2):207-228.
- 154. Savino W, Mendes-Da-Cruz DA, Smaniotto S, Silva-Monteiro E, Villa-Verde DM. Molecular mechanisms governing thymocyte migration: combined role of chemokines and extracellular matrix. J Leukoc Biol. 2004;75(6):951-961.
- 155. Lloyd AR, Oppenheim JJ, Kelvin DJ, Taub DD. Chemokines regulate T cell adherence to recombinant adhesion molecules and extracellular matrix proteins. J Immunol. 1996;156(3):932-938.
- 156. Jacob A, Prekeris R. The regulation of MMP targeting to invadopodia during cancer metastasis. Front Cell Dev Biol. 2015;3:4.
- 157. Bi X, Lou P, Song Y, et al. Msi1 promotes breast cancer metastasis by regulating invadopodia-mediated extracellular matrix degradation via the Timp3-Mmp9 pathway. Oncogene. 2021;40(29):4832-4845.
- 158. Yao M, Brummer G, Acevedo D, Cheng N. Cytokine Regulation of metastasis and tumorigenicity. Adv Cancer Res. 2016;132:265-367.
- 159. Singh S, Sadanandam A, Singh RK. Chemokines in tumor angiogenesis and metastasis. Cancer Metastasis Rev. 2007;26(3-4):453-467.
- 160. Wang Z, Ao X, Shen Z, et al. TNF-alpha augments CXCL10/CXCR3 axis activity to induce epithelial-

mesenchymal transition in colon cancer cell. Int J Biol Sci. 2021;17(11):2683-2702.

- 161. Tan HX, Xiao ZG, Huang T, Fang ZX, Liu Y, Huang ZC. CXCR4/TGF-beta1 mediated self-differentiation of human mesenchymal stem cells to carcinoma-associated fibroblasts and promoted colorectal carcinoma development. Cancer Biol Ther. 2020;21(3):248-257.
- 162. Sun YF, Wu L, Liu SP, et al. Dissecting spatial heterogeneity and the immune-evasion mechanism of CTCs by singlecell RNA-seq in hepatocellular carcinoma. Nat Commun. 2021:12(1):4091.
- 163. Wen J, Zhao Z, Huang L, Wang L, Miao Y, Wu J. IL-8 promotes cell migration through regulating EMT by activating the Wnt/beta-catenin pathway in ovarian cancer. J Cell Mol Med. 2020;24(2):1588-1598.
- 164. Ren T, Zhu L, Cheng M. CXCL10 accelerates EMT and metastasis by MMP-2 in hepatocellular carcinoma. Am J Transl Res. 2017:9(6):2824-2837.
- 165. Cheng Y, Song Y, Qu J, et al. The chemokine receptor CXCR4 and c-MET cooperatively promote epithelial-mesenchymal transition in gastric cancer cells. Transl Oncol. 2018;11(2):487-497.
- 166. Zhao J, Ou B, Han D, et al. Tumor-derived CXCL5 promotes human colorectal cancer metastasis through activation of the ERK/Elk-1/Snail and AKT/GSK3beta/beta-catenin pathways. Mol Cancer. 2017;16(1):70.
- 167. Dai J, Rabie AB. VEGF: an essential mediator of both angiogenesis and endochondral ossification. J Dent Res. 2007;86(10):937-950.
- 168. Voronov E, Shouval DS, Krelin Y, et al. IL-1 is required for tumor invasiveness and angiogenesis. Proc Natl Acad Sci U S A. 2003;100(5):2645-2650.
- 169. Bertolino P, Deckers M, Lebrin F, ten Dijke P. Transforming growth factor-beta signal transduction in angiogenesis and vascular disorders. Chest. 2005;128(6):585S-590S.
- 170. Bae WJ, Ahn JM, Byeon HE, Kim S, Lee D. PTPRDinactivation-induced CXCL8 promotes angiogenesis and metastasis in gastric cancer and is inhibited by metformin. J Exp Clin Cancer Res. 2019;38(1):484.
- 171. Xiong Y, Huang F, Li X, et al. CCL21/CCR7 interaction promotes cellular migration and invasion via modulation of the MEK/ERK1/2 signaling pathway and correlates with lymphatic metastatic spread and poor prognosis in urinary bladder cancer. Int J Oncol. 2017;51(1):75-90.
- 172. Karnezis T, Farnsworth RH, Harris NC, et al. CCL27/CCL28-CCR10 chemokine signaling mediates migration of lymphatic endothelial cells. Cancer Res. 2019;79(7):1558-1572.
- 173. Issa A, Le TX, Shoushtari AN, Shields JD, Swartz MA. Vascular endothelial growth factor-C and C-C chemokine receptor 7 in tumor cell-lymphatic cross-talk promote invasive phenotype. Cancer Res. 2009;69(1):349-357.
- 174. Kitamura T, Qian BZ, Soong D, et al. CCL2-induced chemokine cascade promotes breast cancer metastasis by enhancing retention of metastasis-associated macrophages. J Exp Med. 2015;212(7):1043-1059.
- 175. Marshall LA, Marubayashi S, Jorapur A, et al. Tumors establish resistance to immunotherapy by regulating Treg recruitment via CCR4. J Immunother Cancer. 2020;8(2):e000764.



- LIU ET AL.
- 176. Gobert M, Treilleux I, Bendriss-Vermare N, et al. Regulatory T cells recruited through CCL22/CCR4 are selectively activated in lymphoid infiltrates surrounding primary breast tumors and lead to an adverse clinical outcome. *Cancer Res.* 2009;69(5):2000-2009.
- 177. Zhang CY, Qi Y, Li XN, et al. The role of CCL20/CCR6 axis in recruiting Treg cells to tumor sites of NSCLC patients. *Biomed Pharmacother*. 2015;69:242-248.
- 178. Chen X, Wang L, Li P, et al. Dual TGF-beta and PD-1 blockade synergistically enhances MAGE-A3-specific CD8(+) T cell response in esophageal squamous cell carcinoma. *Int J Cancer*. 2018;143(10):2561-2574.
- 179. Pein M, Insua-Rodriguez J, Hongu T, et al. Metastasisinitiating cells induce and exploit a fibroblast niche to fuel malignant colonization of the lungs. *Nat Commun.* 2020;11(1):1494.
- 180. Chao CC, Lee CW, Chang TM, Chen PC, Liu JF. CXCL1/CXCR2 paracrine axis contributes to lung metastasis in osteosarcoma. *Cancers (Basel)*. 2020;12(2):459.
- 181. Romero-Moreno R, Curtis KJ, Coughlin TR, et al. The CXCL5/CXCR2 axis is sufficient to promote breast cancer colonization during bone metastasis. *Nat Commun.* 2019;10(1):4404.
- Muller A, Homey B, Soto H, et al. Involvement of chemokine receptors in breast cancer metastasis. *Nature*. 2001;410(6824):50-56.
- 183. Tao SC, Guo SC. Role of extracellular vesicles in tumour microenvironment. *Cell Commun Signal*. 2020;18(1):163.
- 184. Urabe F, Patil K, Ramm GA, Ochiya T, Soekmadji C. Extracellular vesicles in the development of organ-specific metastasis. *J Extracell Vesicles*. 2021;10(9):e12125.
- 185. Barenholz-Cohen T, Merkher Y, Haj J, et al. Lung mechanics modifications facilitating metastasis are mediated in part by breast cancer-derived extracellular vesicles. *Int J Cancer*. 2020;147(10):2924-2933.
- 186. Chen C, Zheng H, Luo Y, et al. SUMOylation promotes extracellular vesicle-mediated transmission of lncRNA ELNAT1 and lymph node metastasis in bladder cancer. *J Clin Invest.* 2021;131(8).
- 187. Rodrigues G, Hoshino A, Kenific CM, et al. Tumour exosomal CEMIP protein promotes cancer cell colonization in brain metastasis. *Nat Cell Biol*. 2019;21(11):1403-1412.
- Morad G, Carman CV, Hagedorn EJ, et al. Tumor-derived extracellular vesicles breach the intact blood-brain barrier via transcytosis. ACS Nano. 2019;13(12):13853-13865.
- 189. Mao X, Tey SK, Yeung CLS, et al. Nidogen 1-enriched extracellular vesicles facilitate extrahepatic metastasis of liver cancer by activating pulmonary fibroblasts to secrete tumor necrosis factor receptor 1. Adv Sci (Weinh). 2020;7(21):2002157.
- 190. Ma Q, Liang M, Wu Y, et al. Small extracellular vesicles deliver osteolytic effectors and mediate cancer-induced osteolysis in bone metastatic niche. *J Extracell Vesicles*. 2021;10(4): e12068.
- 191. Li F, Zhao X, Sun R, et al. EGFR-rich extracellular vesicles derived from highly metastatic nasopharyngeal carcinoma cells accelerate tumour metastasis through PI3K/AKT pathway-suppressed ROS. *J Extracell Vesicles*. 2020;10(1): e12003.

- Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science*. 2004;303(5663):1532-1535.
- Yang D, Liu J. Neutrophil extracellular traps: a new player in cancer metastasis and therapeutic target. *J Exp Clin Cancer Res.* 2021;40(1):233.
- 194. Qi JL, He JR, Liu CB, et al. Pulmonary staphylococcus aureus infection regulates breast cancer cell metastasis via neutrophil extracellular traps (NETs) formation. *MedComm*. 2020;1(2):188-201.
- 195. Zhu T, Zou X, Yang C, et al. Neutrophil extracellular traps promote gastric cancer metastasis by inducing epithelialmesenchymal transition. *Int J Mol Med.* 2021;48(1):127.
- 196. Martins-Cardoso K, Almeida VH, Bagri KM, et al. Neutrophil extracellular traps (NETs) promote pro-metastatic phenotype in human breast cancer cells through epithelial-mesenchymal transition. *Cancers (Basel)*. 2020;12(6):1542.
- Najmeh S, Cools-Lartigue J, Rayes RF, et al. Neutrophil extracellular traps sequester circulating tumor cells via betalintegrin mediated interactions. *Int J Cancer*. 2017;140(10):2321-2330.
- Cools-Lartigue J, Spicer J, McDonald B, et al. Neutrophil extracellular traps sequester circulating tumor cells and promote metastasis. *J Clin Invest*. 2013;123(8):3446-3458.
- 199. Albrengues J, Shields MA, Ng D, et al. Neutrophil extracellular traps produced during inflammation awaken dormant cancer cells in mice. *Science*. 2018;361(6409): eaao4227.
- 200. Xie Z, Klionsky DJ. Autophagosome formation: core machinery and adaptations. *Nat Cell Biol.* 2007;9(10):1102-1109.
- 201. Mizushima N, Komatsu M. Autophagy: renovation of cells and tissues. *Cell*. 2011;147(4):728-741.
- Kim KH, Lee MS. Autophagy–a key player in cellular and body metabolism. *Nat Rev Endocrinol.* 2014;10(6):322-337.
- Nakatogawa H. Mechanisms governing autophagosome biogenesis. Nat Rev Mol Cell Biol. 2020;21(8):439-458.
- 204. Kroemer G, Marino G, Levine B. Autophagy and the integrated stress response. *Mol Cell*. 2010;40(2):280-293.
- Yadav AK, Yadav PK, Chaudhary GR, et al. Autophagy in hypoxic ovary. *Cell Mol Life Sci.* 2019;76(17):3311-3322.
- 206. Glick D, Barth S, Macleod KF. Autophagy: cellular and molecular mechanisms. *J Pathol*. 2010;221(1):3-12.
- 207. Mizushima N, Levine B. Autophagy in human diseases. *N Engl J Med.* 2020;383(16):1564-1576.
- Ishaq M, Ojha R, Sharma AP, Singh SK. Autophagy in cancer: Recent advances and future directions. *Semin Cancer Biol.* 2020;66:171-181.
- 209. Lazova R, Camp RL, Klump V, Siddiqui SF, Amaravadi RK, Pawelek JM. Punctate LC3B expression is a common feature of solid tumors and associated with proliferation, metastasis, and poor outcome. *Clin Cancer Res.* 2012;18(2):370-379.
- 210. Peng YF, Shi YH, Ding ZB, et al. Autophagy inhibition suppresses pulmonary metastasis of HCC in mice via impairing anoikis resistance and colonization of HCC cells. *Autophagy*. 2013;9(12):2056-2068.
- 211. Zhao H, Yang M, Zhao J, Wang J, Zhang Y, Zhang Q. High expression of LC3B is associated with progression and poor outcome in triple-negative breast cancer. *Med Oncol.* 2013;30(1):475.

612 MedComm

- Kenific CM, Thorburn A, Debnath J. Autophagy and metastasis: another double-edged sword. *Curr Opin Cell Biol.* 2010;22(2):241-245.
- 213. Jiang GM, Tan Y, Wang H, et al. The relationship between autophagy and the immune system and its applications for tumor immunotherapy. *Mol Cancer*. 2019;18(1):17.
- 214. Gorgulu K, Diakopoulos KN, Ai J, et al. Levels of the autophagy-related 5 protein affect progression and metastasis of pancreatic tumors in mice. *Gastroenterology*. 2019;156(1):203-217.e20.
- 215. Teo Hansen Selno A, Schlichtner S, Yasinska IM, et al. High mobility group box 1 (HMGB1) induces toll-like receptor 4-mediated production of the immunosuppressive protein galectin-9 in human cancer cells. *Front Immunol.* 2021;12:675731.
- Kadandale P, Stender JD, Glass CK, Kiger AA. Conserved role for autophagy in Rho1-mediated cortical remodeling and blood cell recruitment. *Proc Natl Acad Sci U S A*. 2010;107(23):10502-10507.
- 217. Vera-Ramirez L, Vodnala SK, Nini R, Hunter KW, Green JE. Autophagy promotes the survival of dormant breast cancer cells and metastatic tumour recurrence. *Nat Commun.* 2018;9(1):1944.
- 218. Maycotte P, Jones KL, Goodall ML, Thorburn J, Thorburn A. Autophagy supports breast cancer stem cell maintenance by regulating IL6 secretion. *Mol Cancer Res.* 2015;13(4):651-658.
- 219. Tang Q, Su Z, Gu W, Rustgi AK. Mutant p53 on the path to metastasis. *Trends Cancer*. 2020;6(1):62-73.
- 220. Hu J, Kong S, Dong T, et al. Autophagy modulates mesenchymal-to-endothelial transition via p53. *Aging (Albany NY)*. 2020;12(21):22112-22121.
- Maiuri MC, Galluzzi L, Morselli E, Kepp O, Malik SA, Kroemer G. Autophagy regulation by p53. *Curr Opin Cell Biol.* 2010;22(2):181-185.
- 222. Yang Y, Karsli-Uzunbas G, Poillet-Perez L, et al. Autophagy promotes mammalian survival by suppressing oxidative stress and p53. *Genes Dev.* 2020;34(9-10):688-700.
- 223. Kim T, Veronese A, Pichiorri F, et al. p53 regulates epithelialmesenchymal transition through microRNAs targeting ZEB1 and ZEB2. *J Exp Med.* 2011;208(5):875-883.
- 224. Karimi Roshan M, Soltani A, Soleimani A, Rezaie Kahkhaie K, Afshari AR, Soukhtanloo M. Role of AKT and mTOR signaling pathways in the induction of epithelial-mesenchymal transition (EMT) process. *Biochimie*. 2019;165:229-234.
- 225. Kim YC, Guan KL. mTOR: a pharmacologic target for autophagy regulation. *J Clin Invest*. 2015;125(1):25-32.
- 226. Zhang M, Liu S, Chua MS, et al. SOCS5 inhibition induces autophagy to impair metastasis in hepatocellular carcinoma cells via the PI3K/Akt/mTOR pathway. *Cell Death Dis.* 2019;10(8):612.
- 227. Zhu JF, Huang W, Yi HM, et al. Annexin A1-suppressed autophagy promotes nasopharyngeal carcinoma cell invasion and metastasis by PI3K/AKT signaling activation. *Cell Death Dis.* 2018;9(12):1154.
- 228. Mihaylova MM, Shaw RJ. The AMPK signaling pathway coordinates cell growth, autophagy and metabolism. *Nat Cell Biol.* 2011;13(9):1016-23.
- 229. Wu J, Gao F, Xu T, et al. CLDN1 induces autophagy to promote proliferation and metastasis of esophageal squamous car-

cinoma through AMPK/STAT1/ULK1 signaling. *J Cell Physiol*. 2020;235(3):2245-2259.

- Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol.* 2011;13(2):132-141.
- Allen AE, Locasale JW. Glucose metabolism in cancer: The saga of pyruvate kinase continues. *Cancer Cell*. 2018;33(3):337-339.
- Biswas SK. Metabolic reprogramming of immune cells in Cancer Progression. *Immunity*. 2015;43(3):435-449.
- Annett S, Moore G, Robson T. Obesity and cancer metastasis: molecular and translational perspectives. *Cancers (Basel)*. 2020;12(12):3798.
- Fu Y, Zou T, Shen X, et al. Lipid metabolism in cancer progression and therapeutic strategies. *MedComm.* 2020;2(1):27-59.
- 235. Luo X, Cheng C, Tan Z, et al. Emerging roles of lipid metabolism in cancer metastasis. *Mol Cancer*. 2017;16(1):76.
- 236. He Y, Gao M, Cao Y, Tang H, Liu S, Tao Y. Nuclear localization of metabolic enzymes in immunity and metastasis. *Biochim Biophys Acta Rev Cancer*. 2017;1868(2):359-371.
- 237. Wen J, Min X, Shen M, et al. ACLY facilitates colon cancer cell metastasis by CTNNB1. *J Exp Clin Cancer Res*. 2019;38(1):401.
- Jiang L, Wang H, Li J, et al. Up-regulated FASN expression promotes transcoelomic metastasis of ovarian cancer cell through epithelial-mesenchymal transition. *Int J Mol Sci.* 2014;15(7):11539-11554.
- 239. Pascual G, Avgustinova A, Mejetta S, et al. Targeting metastasis-initiating cells through the fatty acid receptor CD36. *Nature*. 2017;541(7635):41-45.
- Ladanyi A, Mukherjee A, Kenny HA, et al. Adipocyte-induced CD36 expression drives ovarian cancer progression and metastasis. *Oncogene*. 2018;37(17):2285-2301.
- 241. Doria ML, Cotrim Z, Macedo B, et al. Lipidomic approach to identify patterns in phospholipid profiles and define class differences in mammary epithelial and breast cancer cells. *Breast Cancer Res Treat.* 2012;133(2):635-648.
- 242. Brandi J, Dando I, Pozza ED, et al. Proteomic analysis of pancreatic cancer stem cells: Functional role of fatty acid synthesis and mevalonate pathways. *J Proteomics*. 2017;150:310-322.
- 243. Fu H, He Y, Qi L, et al. cPLA2alpha activates PI3K/AKT and inhibits Smad2/3 during epithelial-mesenchymal transition of hepatocellular carcinoma cells. *Cancer Lett.* 2017;403:260-270.
- 244. Wang C, Yang Z, Xu E, et al. Apolipoprotein C-II induces EMT to promote gastric cancer peritoneal metastasis via PI3K/AKT/mTOR pathway. *Clin Transl Med.* 2021;11(8): e522.
- 245. Zhang N, Zhang H, Liu Y, et al. SREBP1, targeted by miR-18a-5p, modulates epithelial-mesenchymal transition in breast cancer via forming a co-repressor complex with Snail and HDAC1/2. *Cell Death Differ*. 2019;26(5):843-859.
- 246. Zhao H, Yan G, Zheng L, et al. STIM1 is a metabolic checkpoint regulating the invasion and metastasis of hepatocellular carcinoma. *Theranostics*. 2020;10(14):6483-6499.
- 247. Chen M, Zhao Y, Yang X, et al. NSDHL promotes triplenegative breast cancer metastasis through the TGFbeta signaling pathway and cholesterol biosynthesis. *Breast Cancer Res Treat*. 2021;187(2):349-362.
- 248. Greenlee JD, Subramanian T, Liu K, King MR. Rafting down the metastatic cascade: the role of lipid rafts in cancer

metastasis, cell Death, and clinical outcomes. *Cancer Res.* 2021;81(1):5-17.

- 249. Murai T, Maruyama Y, Mio K, Nishiyama H, Suga M, Sato C. Low cholesterol triggers membrane microdomain-dependent CD44 shedding and suppresses tumor cell migration. *J Biol Chem.* 2011;286(3):1999-2007.
- 250. Yang YF, Jan YH, Liu YP, et al. Squalene synthase induces tumor necrosis factor receptor 1 enrichment in lipid rafts to promote lung cancer metastasis. *Am J Respir Crit Care Med.* 2014;190(6):675-687.
- Manzo T, Prentice BM, Anderson KG, et al. Accumulation of long-chain fatty acids in the tumor microenvironment drives dysfunction in intrapancreatic CD8+ T cells. *J Exp Med.* 2020;217(8):e20191920.
- 252. Su P, Wang Q, Bi E, et al. Enhanced lipid accumulation and metabolism are required for the differentiation and activation of tumor-associated macrophages. *Cancer Res.* 2020;80(7):1438-1450.
- 253. Gong J, Lin Y, Zhang H, et al. Reprogramming of lipid metabolism in cancer-associated fibroblasts potentiates migration of colorectal cancer cells. *Cell Death Dis.* 2020;11(4):267.
- 254. Peng WX, Koirala P, Mo YY. LncRNA-mediated regulation of cell signaling in cancer. *Oncogene*. 2017;36(41):5661-5667.
- 255. Luo M, Li Z, Wang W, Zeng Y, Liu Z, Qiu J. Long non-coding RNA H19 increases bladder cancer metastasis by associating with EZH2 and inhibiting E-cadherin expression. *Cancer Lett.* 2013;333(2):213-221.
- 256. Zhou W, Ye XL, Xu J, et al. The lncRNA H19 mediates breast cancer cell plasticity during EMT and MET plasticity by differentially sponging miR-200b/c and let-7b. *Sci Signal*. 2017;10(483):eaak9557.
- 257. Liang WC, Fu WM, Wong CW, et al. The lncRNA H19 promotes epithelial to mesenchymal transition by functioning as miRNA sponges in colorectal cancer. *Oncotarget*. 2015;6(26):22513-22525.
- 258. Zhu M, Chen Q, Liu X, et al. lncRNA H19/miR-675 axis represses prostate cancer metastasis by targeting TGFBI. *FEBS J*. 2014;281(16):3766-3775.
- 259. Grelet S, Link LA, Howley B, et al. A regulated PNUTS mRNA to lncRNA splice switch mediates EMT and tumour progression. *Nat Cell Biol.* 2017;19(9):1105-1115.
- 260. Hou P, Meng S, Li M, et al. LINC00460/DHX9/IGF2BP2 complex promotes colorectal cancer proliferation and metastasis by mediating HMGA1 mRNA stability depending on m6A modification. J Exp Clin Cancer Res. 2021;40(1):52.
- 261. Meng LD, Shi GD, Ge WL, et al. Linc01232 promotes the metastasis of pancreatic cancer by suppressing the ubiquitinmediated degradation of HNRNPA2B1 and activating the A-Raf-induced MAPK/ERK signaling pathway. *Cancer Lett.* 2020;494:107-120.
- 262. Zhuang M, Zhao S, Jiang Z, et al. MALAT1 sponges miR-106b-5p to promote the invasion and metastasis of colorectal cancer via SLAIN2 enhanced microtubules mobility. *EBioMedicine*. 2019;41:286-298.
- 263. Li Q, Mo W, Ding Y, Ding X. Study of lncRNA TPA in promoting invasion and metastasis of breast cancer mediated by TGFbeta signaling pathway. *Front Cell Dev Biol*. 2021;9:688751.
- 264. Lai SW, Chen MY, Bamodu OA, et al. Exosomal lncRNA PVT1/VEGFA axis promotes colon cancer metastasis and stem-

ness by downregulation of tumor suppressor miR-152-3p. *Oxid Med Cell Longev.* 2021;2021:9959807.

- 265. Sas-Chen A, Aure MR, Leibovich L, et al. LIMT is a novel metastasis inhibiting lncRNA suppressed by EGF and downregulated in aggressive breast cancer. *EMBO Mol Med*. 2016;8(9):1052-1064.
- 266. Liu D, Li Y, Luo G, et al. LncRNA SPRY4-IT1 sponges miR-101-3p to promote proliferation and metastasis of bladder cancer cells through up-regulating EZH2. *Cancer Lett.* 2017;388:281-291.
- 267. Wu H, Hu Y, Liu X, et al. LncRNA TRERNA1 function as an enhancer of SNAI1 promotes gastric cancer metastasis by regulating epithelial-mesenchymal transition. *Mol Ther Nucleic Acids*. 2017;8:291-299.
- Liang WC, Ren JL, Wong CW, et al. LncRNA-NEF antagonized epithelial to mesenchymal transition and cancer metastasis via cis-regulating FOXA2 and inactivating Wnt/beta-catenin signaling. Oncogene. 2018;37(11):1445-1456.
- 269. Wang H, Huo X, Yang XR, et al. STAT3-mediated upregulation of lncRNA HOXD-AS1 as a ceRNA facilitates liver cancer metastasis by regulating SOX4. *Mol Cancer*. 2017;16(1): 136.
- 270. Yue B, Liu C, Sun H, et al. A positive feed-forward loop between LncRNA-CYTOR and Wnt/beta-catenin signaling promotes metastasis of colon Cancer. *Mol Ther*. 2018;26(5):1287-1298.
- 271. Pan J, Fang S, Tian H, et al. lncRNA JPX/miR-33a-5p/Twist1 axis regulates tumorigenesis and metastasis of lung cancer by activating Wnt/beta-catenin signaling. *Mol Cancer*. 2020;19(1):9.
- 272. Cheng B, Rong A, Zhou Q, Li W. LncRNA LINC00662 promotes colon cancer tumor growth and metastasis by competitively binding with miR-340-5p to regulate CLDN8/IL22 coexpression and activating ERK signaling pathway. J Exp Clin Cancer Res. 2020;39(1):5.
- 273. Zhou Y, Huan L, Wu Y, et al. LncRNA ID2-AS1 suppresses tumor metastasis by activating the HDAC8/ID2 pathway in hepatocellular carcinoma. *Cancer Lett.* 2020;469:399-409.
- 274. Liang ZX, Liu HS, Wang FW, et al. LncRNA RPPH1 promotes colorectal cancer metastasis by interacting with TUBB3 and by promoting exosomes-mediated macrophage M2 polarization. *Cell Death Dis.* 2019;10(11):829.
- 275. Xu M, Xu X, Pan B, et al. LncRNA SATB2-AS1 inhibits tumor metastasis and affects the tumor immune cell microenvironment in colorectal cancer by regulating SATB2. *Mol Cancer*. 2019;18(1):135.
- 276. Tan BS, Yang MC, Singh S, et al. LncRNA NORAD is repressed by the YAP pathway and suppresses lung and breast cancer metastasis by sequestering S100P. *Oncogene*. 2019;38(28):5612-5626.
- 277. Zhai W, Zhu R, Ma J, et al. A positive feed-forward loop between LncRNA-URRCC and EGFL7/P-AKT/FOXO3 signaling promotes proliferation and metastasis of clear cell renal cell carcinoma. *Mol Cancer*. 2019;18(1):81.
- Bian Z, Zhang J, Li M, et al. LncRNA-FEZF1-AS1 promotes tumor proliferation and metastasis in colorectal cancer by regulating PKM2 signaling. *Clin Cancer Res.* 2018;24(19):4808-4819.
- 279. Xie S, Yu X, Li Y, et al. Upregulation of lncRNA ADAMTS9-AS2 promotes salivary adenoid cystic carcinoma metastasis via

PI3K/Akt and MEK/Erk signaling. *Mol Ther*. 2018;26(12):2766-2778.

- 280. Liu B, Wu S, Ma J, et al. LncRNA GAS5 reverses EMT and tumor stem cell-mediated gemcitabine resistance and metastasis by targeting miR-221/SOCS3 in pancreatic cancer. *Mol Ther Nucleic Acids*. 2018;13:472-482.
- 281. Ling H, Spizzo R, Atlasi Y, et al. CCAT2, a novel noncoding RNA mapping to 8q24, underlies metastatic progression and chromosomal instability in colon cancer. *Genome Res.* 2013;23(9):1446-1461.
- 282. Redis RS, Vela LE, Lu W, et al. Allele-specific reprogramming of cancer metabolism by the long non-coding RNA CCAT2. *Mol Cell*. 2016;61(4):640.
- 283. Silva-Fisher JM, Dang HX, White NM, et al. Long non-coding RNA RAMS11 promotes metastatic colorectal cancer progression. *Nat Commun*. 2020;11(1):2156.
- 284. Ji P, Diederichs S, Wang W, et al. MALAT-1, a novel noncoding RNA, and thymosin beta4 predict metastasis and survival in early-stage non-small cell lung cancer. *Oncogene*. 2003;22(39):8031-8041.
- 285. Arun G, Diermeier S, Akerman M, et al. Differentiation of mammary tumors and reduction in metastasis upon Malat1 lncRNA loss. *Genes Dev.* 2016;30(1):34-51.
- 286. Wang X, Adjei AA. Lung cancer and metastasis: new opportunities and challenges. *Cancer Metastasis Rev.* 2015;34(2):169-171.
- 287. Ost P, Bossi A, Decaestecker K, et al. Metastasis-directed therapy of regional and distant recurrences after curative treatment of prostate cancer: a systematic review of the literature. *Eur Urol.* 2015;67(5):852-863.
- 288. Creutzberg CL, Lu KH, Fleming GF. Uterine Cancer: Adjuvant therapy and management of metastatic disease. *J Clin Oncol.* 2019;37(27):2490-2500.
- 289. Meko J, Rusch VW. Neoadjuvant therapy and surgical resection for locally advanced non-small cell lung cancer. *Semin Radiat Oncol.* 2000;10(4):324-332.
- 290. Pitroda SP, Chmura SJ, Weichselbaum RR. Integration of radiotherapy and immunotherapy for treatment of oligometastases. *Lancet Oncol.* 2019;20(8):e434-e442.
- 291. Wyld L, Audisio RA, Poston GJ. The evolution of cancer surgery and future perspectives. *Nat Rev Clin Oncol.* 2015;12(2):115-124.
- 292. Guner A, Son T, Cho I, et al. Liver-directed treatments for liver metastasis from gastric adenocarcinoma: comparison between liver resection and radiofrequency ablation. *Gastric Cancer*. 2016;19(3):951-960.
- 293. Evrard S, Torzilli G, Caballero C, Bonhomme B. Parenchymal sparing surgery brings treatment of colorectal liver metastases into the precision medicine era. *Eur J Cancer*. 2018;104:195-200.
- 294. Xu J, Fan J, Qin X, et al. Chinese guidelines for the diagnosis and comprehensive treatment of colorectal liver metastases (version 2018). *J Cancer Res Clin Oncol*. 2019;145(3):725-736.
- 295. Bhattacharya IS, Hoskin PJ. Stereotactic body radiotherapy for spinal and bone metastases. *Clin Oncol (R Coll Radiol)*. 2015;27(5):298-306.
- 296. Birgisson H, Enblad M, Artursson S, Ghanipour L, Cashin P, Graf W. Patients with colorectal peritoneal metastases and high peritoneal cancer index may benefit from cytoreductive surgery and hyperthermic intraperitoneal chemotherapy. *Eur J Surg Oncol.* 2020;46(12):2283-2291.

- 297. Mercier F, Mohamed F, Cazauran JB, et al. An update of peritonectomy procedures used in cytoreductive surgery for peritoneal malignancy. *Int J Hyperthermia*. 2019;36(1):744-752.
- 298. Radovich M, Jiang G, Hancock BA, et al. Association of circulating tumor DNA and circulating tumor cells after neoadjuvant chemotherapy with disease recurrence in patients with triple-negative breast Cancer: preplanned secondary analysis of the BRE12-158 randomized clinical trial. *JAMA Oncol.* 2020;6(9):1410-1415.
- 299. Pierga JY, Bidard FC, Autret A, et al. Circulating tumour cells and pathological complete response: independent prognostic factors in inflammatory breast cancer in a pooled analysis of two multicentre phase II trials (BEVERLY-1 and -2) of neoadjuvant chemotherapy combined with bevacizumab. *Ann Oncol.* 2017;28(1):103-109.
- Pilewskie M, Morrow M. Axillary nodal management following neoadjuvant chemotherapy: a review. JAMA Oncol. 2017;3(4):549-555.
- Kosuge T, Sakamoto Y, Ueno H. Postoperative adjuvant therapy. J Hepatobiliary Pancreat Sci. 2011;18(6):792-796.
- Xiong HQ, Ajani JA. Treatment of colorectal cancer metastasis: the role of chemotherapy. *Cancer Metastasis Rev.* 2004;23(1-2):145-163.
- Meyer M, Seetharam M. First-line therapy for metastatic soft tissue sarcoma. *Curr Treat Options Oncol.* 2019;20(1):6.
- 304. Decatris MP, Sundar S, O'Byrne KJ. Platinum-based chemotherapy in metastatic breast cancer: current status. *Cancer Treat Rev.* 2004;30(1):53-81.
- 305. Hallek M. Chronic lymphocytic leukemia: 2020 update on diagnosis, risk stratification and treatment. Am J Hematol. 2019;94(11):1266-1287.
- Ansell SM. Hodgkin lymphoma: diagnosis and treatment. Mayo Clin Proc. 2015;90(11):1574-1583.
- 307. Karagiannis GS, Condeelis JS, Oktay MH. Chemotherapyinduced metastasis: molecular mechanisms, clinical manifestations, therapeutic interventions. *Cancer Res.* 2019;79(18): 4567-4576.
- 308. Shaked Y, Emmenegger U, Man S, et al. Optimal biologic dose of metronomic chemotherapy regimens is associated with maximum antiangiogenic activity. *Blood.* 2005;106(9):3058-3061.
- 309. Chan TS, Hsu CC, Pai VC, et al. Metronomic chemotherapy prevents therapy-induced stromal activation and induction of tumor-initiating cells. *J Exp Med.* 2016;213(13):2967-2988.
- Colevas AD, Yom SS, Pfister DG, et al. NCCN guidelines insights: head and neck cancers, version 1.2018. J Natl Compr Canc Netw. 2018;16(5):479-490.
- 311. Sun Y, Li WF, Chen NY, et al. Induction chemotherapy plus concurrent chemoradiotherapy versus concurrent chemoradiotherapy alone in locoregionally advanced nasopharyngeal carcinoma: a phase 3, multicentre, randomised controlled trial. *Lancet Oncol.* 2016;17(11):1509-1520.
- 312. Zhang Y, Chen L, Hu GQ, et al. Gemcitabine and cisplatin induction chemotherapy in nasopharyngeal carcinoma. *NEngl J Med.* 2019;381(12):1124-1135.
- 313. Perotti A, Sessa C, Mancuso A, et al. Clinical and pharmacological phase I evaluation of Exherin (ADH-1), a selective anti-Ncadherin peptide in patients with N-cadherin-expressing solid tumours. Ann Oncol. 2009;20(4):741-745.



- 314. Sun Y, Jing J, Xu H, et al. N-cadherin inhibitor creates a microenvironment that protect TILs from immune checkpoints and Treg cells. *J Immunother Cancer*. 2021;9(3).
- 315. Rivera-Torres J, San Jose E. Src tyrosine kinase inhibitors: new perspectives on their immune, antiviral, and senotherapeutic potential. *Front Pharmacol.* 2019;10:1011.
- 316. Long GV, Flaherty KT, Stroyakovskiy D, et al. Dabrafenib plus trametinib versus dabrafenib monotherapy in patients with metastatic BRAF V600E/K-mutant melanoma: long-term survival and safety analysis of a phase 3 study. *Ann Oncol.* 2017;28(7):1631-1639.
- 317. Reardon DA, Nabors LB, Stupp R, Mikkelsen T. Cilengitide: an integrin-targeting arginine-glycine-aspartic acid peptide with promising activity for glioblastoma multiforme. *Expert Opin Investig Drugs*. 2008;17(8):1225-1235.
- 318. Heidenreich A, Rawal SK, Szkarlat K, et al. A randomized, double-blind, multicenter, phase 2 study of a human monoclonal antibody to human alphanu integrins (intetumumab) in combination with docetaxel and prednisone for the firstline treatment of patients with metastatic castration-resistant prostate cancer. *Ann Oncol.* 2013;24(2):329-336.
- 319. Eberlein C, Kendrew J, McDaid K, et al. A human monoclonal antibody 264RAD targeting alphavbeta6 integrin reduces tumour growth and metastasis, and modulates key biomarkers in vivo. *Oncogene*. 2013;32(37):4406-4416.
- 320. Pickarski M, Gleason A, Bednar B, Duong LT. Orally active alphavbeta3 integrin inhibitor MK-0429 reduces melanoma metastasis. *Oncol Rep.* 2015;33(6):2737-2745.
- 321. Li X, Wang J. Mechanical tumor microenvironment and transduction: cytoskeleton mediates cancer cell invasion and metastasis. *Int J Biol Sci.* 2020;16(12):2014-2028.
- 322. Alanko J, Mai A, Jacquemet G, et al. Integrin endosomal signalling suppresses anoikis. *Nat Cell Biol.* Nov 2015;17(11):1412-1421.
- 323. Schaefer KL, Wada K, Takahashi H, et al. Peroxisome proliferator-activated receptor gamma inhibition prevents adhesion to the extracellular matrix and induces anoikis in hepatocellular carcinoma cells. *Cancer Res.* 2005;65(6):2251-2259.
- 324. Lee JJ, Hsu YC, Li YS, Cheng SP. Galectin-3 inhibitors suppress anoikis resistance and invasive capacity in thyroid cancer cells. *Int J Endocrinol.* 2021;2021:5583491.
- 325. Calon A, Tauriello DV, Batlle E. TGF-beta in CAF-mediated tumor growth and metastasis. *Semin Cancer Biol.* 2014;25:15-22.
- 326. Madeo A, Maggiolini M. Nuclear alternate estrogen receptor GPR30 mediates 17beta-estradiol-induced gene expression and migration in breast cancer-associated fibroblasts. *Cancer Res.* 2010;70(14):6036-6046.
- 327. Yu Y, Xiao CH, Tan LD, Wang QS, Li XQ, Feng YM. Cancerassociated fibroblasts induce epithelial-mesenchymal transition of breast cancer cells through paracrine TGF-beta signalling. *Br J Cancer*. 2014;110(3):724-732.
- 328. Lamprecht S, Sigal-Batikoff I, Shany S, et al. Teaming up for trouble: cancer cells, transforming growth factor-betal signaling and the epigenetic corruption of stromal naive fibroblasts. *Cancers (Basel)*. 2018;10(3):61.
- 329. Tan HX, Gong WZ, Zhou K, et al. CXCR4/TGF-beta1 mediated hepatic stellate cells differentiation into carcinoma-associated

fibroblasts and promoted liver metastasis of colon cancer. *Cancer Biol Ther.* 2020;21(3):258-268.

- 330. Huang M, Fu M, Wang J, et al. TGF-beta1-activated cancer-associated fibroblasts promote breast cancer invasion, metastasis and epithelial-mesenchymal transition by autophagy or overexpression of FAP-alpha. *Biochem Pharma*col. 2021;188:114527.
- 331. Kelley RK, Gane E, Assenat E, et al. A phase 2 study of galunisertib (TGF-betal Receptor Type I Inhibitor) and sorafenib in patients with advanced hepatocellular carcinoma. *Clin Transl Gastroenterol.* 2019;10(7):e00056.
- 332. Melisi D, Garcia-Carbonero R, Macarulla T, et al. Galunisertib plus gemcitabine vs. gemcitabine for first-line treatment of patients with unresectable pancreatic cancer. *Br J Cancer*. 2018;119(10):1208-1214.
- 333. Rodon J, Carducci MA, Sepulveda-Sanchez JM, et al. First-inhuman dose study of the novel transforming growth factorbeta receptor I kinase inhibitor LY2157299 monohydrate in patients with advanced cancer and glioma. *Clin Cancer Res.* 2015;21(3):553-560.
- 334. Lan Y, Zhang D, Xu C, et al. Enhanced preclinical antitumor activity of M7824, a bifunctional fusion protein simultaneously targeting PD-L1 and TGF-beta. *Sci Transl Med.* 2018;10(424):eaan5488.
- 335. Strauss J, Heery CR, Schlom J, et al. Phase I trial of M7824 (MSB0011359C), a Bifunctional fusion protein targeting PD-L1 and TGFbeta, in advanced solid tumors. *Clin Cancer Res.* 2018;24(6):1287-1295.
- 336. Subramaniam KS, Omar IS, Kwong SC, et al. Cancerassociated fibroblasts promote endometrial cancer growth via activation of interleukin-6/STAT-3/c-Myc pathway. *Am J Cancer Res.* 2016;6(2):200-213.
- 337. Biffi G, Oni TE, Spielman B, et al. IL1-Induced JAK/STAT Signaling is antagonized by TGFbeta to shape CAF heterogeneity in pancreatic ductal adenocarcinoma. *Cancer Discov.* 2019;9(2):282-301.
- 338. Heichler C, Scheibe K, Schmied A, et al. STAT3 activation through IL-6/IL-11 in cancer-associated fibroblasts promotes colorectal tumour development and correlates with poor prognosis. *Gut.* 2020;69(7):1269-1282.
- 339. Karakasheva TA, Lin EW, Tang Q, et al. IL-6 mediates crosstalk between tumor cells and activated fibroblasts in the tumor microenvironment. *Cancer Res.* 2018;78(17):4957-4970.
- 340. Patel MR, Dash A, Jacobson BA, et al. JAK/STAT inhibition with ruxolitinib enhances oncolytic virotherapy in non-small cell lung cancer models. *Cancer Gene Ther*. 2019;26(11-12):411-418.
- 341. Cataldi M, Shah NR, Felt SA, Grdzelishvili VZ. Breaking resistance of pancreatic cancer cells to an attenuated vesicular stomatitis virus through a novel activity of IKK inhibitor TPCA-1. *Virology*. 2015;485:340-354.
- 342. Hurwitz HI, Uppal N, Wagner SA, et al. Randomized, doubleblind, phase II study of ruxolitinib or placebo in combination with capecitabine in patients with metastatic pancreatic cancer for whom therapy with gemcitabine has failed. *J Clin Oncol.* 2015;33(34):4039-4047.
- 343. Alraouji NN, Al-Mohanna FH, Ghebeh H, et al. Tocilizumab potentiates cisplatin cytotoxicity and targets cancer stem cells

in triple-negative breast cancer. *Mol Carcinog*. 2020;59(9):1041-1051.

- 344. Guo Y, Nemeth J, O'Brien C, et al. Effects of siltuximab on the IL-6-induced signaling pathway in ovarian cancer. *Clin Cancer Res.* 2010;16(23):5759-5769.
- 345. Welt S, Divgi CR, Scott AM, et al. Antibody targeting in metastatic colon cancer: a phase I study of monoclonal antibody F19 against a cell-surface protein of reactive tumor stromal fibroblasts. *J Clin Oncol.* 1994;12(6):1193-1203.
- 346. Nemunaitis J, Vukelja SJ, Richards D, et al. Phase I trial of PT-100 (PT-100), a cytokine-inducing small molecule, following chemotherapy for solid tumor malignancy. *Cancer Invest.* 2006;24(6):553-561.
- 347. Scott AM, Wiseman G, Welt S, et al. A phase I dose-escalation study of sibrotuzumab in patients with advanced or metastatic fibroblast activation protein-positive cancer. *Clin Cancer Res.* 2003;9(5):1639-1647.
- 348. van der Graaf WT, Blay JY, Chawla SP, et al. Pazopanib for metastatic soft-tissue sarcoma (PALETTE): a randomised, double-blind, placebo-controlled phase 3 trial. *Lancet*. 2012;379(9829):1879-1886.
- Carlisle BG, Zheng T, Kimmelman J. Imatinib and the long tail of targeted drug development. *Nat Rev Clin Oncol.* 2020;17(1):1-3.
- 350. Garcia J, Hurwitz HI, Sandler AB, et al. Bevacizumab (Avastin(R)) in cancer treatment: a review of 15 years of clinical experience and future outlook. *Cancer Treat Rev.* 2020;86:102017.
- 351. Shen G, Zheng F, Ren D, et al. Anlotinib: a novel multitargeting tyrosine kinase inhibitor in clinical development. *J Hematol Oncol.* 2018;11(1):120.
- 352. Song Y, Fu Y, Xie Q, Zhu B, Wang J, Zhang B. Anti-angiogenic agents in combination with immune checkpoint inhibitors: a promising strategy for cancer treatment. *Front Immunol.* 2020;11:1956.
- 353. Cornel AM, Mimpen IL, Nierkens S. MHC class I downregulation in cancer: underlying mechanisms and potential targets for cancer immunotherapy. *Cancers (Basel)*. 2020;12(7):1760.
- 354. Akamatsu H, Harada H, Tokunaga S, et al. A phase II study of gefitinib with concurrent thoracic radiotherapy in Patients With unresectable, stage III non-small-cell lung cancer harboring EGFR mutations (WJOG6911L). *Clin Lung Cancer*. 2019;20(1):e25-e27.
- 355. Yang Z, Zhang Y, Li R, et al. Whole-brain radiotherapy with and without concurrent erlotinib in NSCLC with brain metastases: a multicenter, open-label, randomized, controlled phase III trial. *Neuro Oncol.* 2021;23(6):967-978
- 356. Kong C, Zhu X, Jiang M, et al. Anlotinib in combination with whole brain radiotherapy for advanced non-small-cell lung cancer with brain metastases progressive or developed after at least one lines of prior treatment. *Int J Radiat Oncol Biol Phys.* 2021;111(3S):e569.
- 357. Korbecki J, Kojder K, Siminska D, et al. CC chemokines in a tumor: A review of pro-cancer and anti-cancer properties of the ligands of receptors CCR1, CCR2, CCR3, and CCR4. *Int J Mol Sci.* 2020;21(21):8412.
- 358. Miao M, De Clercq E, Li G. Clinical significance of chemokine receptor antagonists. *Expert Opin Drug Metab Toxicol*. 2020;16(1):11-30.

- 359. Halama N, Zoernig I, Berthel A, et al. Tumoral immune cell exploitation in colorectal cancer metastases can be targeted effectively by Anti-CCR5 therapy in cancer patients. *Cancer Cell.* 2016;29(4):587-601.
- 360. Kim YH, Bagot M, Pinter-Brown L, et al. Mogamulizumab versus vorinostat in previously treated cutaneous T-cell lymphoma (MAVORIC): an international, open-label, randomised, controlled phase 3 trial. *Lancet Oncol.* 2018;19(9):1192-1204.
- 361. Doi T, Muro K, Ishii H, et al. A Phase I study of the anti-CC chemokine receptor 4 antibody, mogamulizumab, in combination with nivolumab in patients with advanced or metastatic solid tumors. *Clin Cancer Res.* 2019;25(22):6614-6622.
- 362. Papadopoulos KP, Gluck L, Martin LP, et al. First-in-human study of AMG 820, a monoclonal anti-colony-stimulating factor 1 receptor antibody, in patients with advanced solid tumors. *Clin Cancer Res.* 2017;23(19):5703-5710.
- 363. Zhu Y, Knolhoff BL, Meyer MA, et al. CSF1/CSF1R blockade reprograms tumor-infiltrating macrophages and improves response to T-cell checkpoint immunotherapy in pancreatic cancer models. *Cancer Res.* 2014;74(18):5057-5069.
- 364. Hung JY, Horn D, Woodruff K, et al. Colony-stimulating factor 1 potentiates lung cancer bone metastasis. *Lab Invest.* 2014;94(4):371-381.
- 365. Rojo R, Raper A, Ozdemir DD, et al. Deletion of a csflr enhancer selectively impacts CSF1R expression and development of tissue macrophage populations. *Nat Commun.* 2019;10(1):3215.
- Lamb YN. Pexidartinib: first approval. *Drugs*. 2019;79(16):1805-1812.
- 367. Gomez-Roca CA, Italiano A, Le Tourneau C, et al. Phase I study of emactuzumab single agent or in combination with paclitaxel in patients with advanced/metastatic solid tumors reveals depletion of immunosuppressive M2-like macrophages. *Ann Oncol.* 2019;30(8):1381-1392.
- 368. Shitara K, Ueha S, Shichino S, et al. First-in-human phase 1 study of IT1208, a defucosylated humanized anti-CD4 depleting antibody, in patients with advanced solid tumors. J Immunother Cancer. 2019;7(1):195.
- Maruhashi T, Sugiura D, Okazaki IM, Okazaki T. LAG-3: from molecular functions to clinical applications. *J Immunother Cancer*. 2020;8(2):e001014.
- Qin S, Xu L, Yi M, Yu S, Wu K, Luo S. Novel immune checkpoint targets: moving beyond PD-1 and CTLA-4. *Mol Cancer*. 2019;18(1):155.
- Harjunpaa H, Guillerey C. TIGIT as an emerging immune checkpoint. *Clin Exp Immunol*. 2020;200(2):108-119.
- Andrews LP, Yano H, Vignali DAA. Inhibitory receptors and ligands beyond PD-1, PD-L1 and CTLA-4: breakthroughs or backups. *Nat Immunol.* 2019;20(11):1425-1434.
- 373. Blackburn SD, Shin H, Haining WN, et al. Coregulation of CD8+ T cell exhaustion by multiple inhibitory receptors during chronic viral infection. *Nat Immunol.* 2009;10(1):29-37.
- 374. Kraman M, Faroudi M, Allen NL, et al. FS118, a bispecific antibody targeting LAG-3 and PD-L1, enhances T-Cell activation resulting in potent antitumor activity. *Clin Cancer Res.* 2020;26(13):3333-3344.
- 375. Dougall WC, Kurtulus S, Smyth MJ, Anderson AC. TIGIT and CD96: new checkpoint receptor targets for cancer immunotherapy. *Immunol Rev.* 2017;276(1):112-120.

- LIU ET AL.
- 376. Saleh R, Toor SM, Elkord E. Targeting TIM-3 in solid tumors: innovations in the preclinical and translational realm and therapeutic potential. *Expert Opin Ther Targets*. 2020;24(12):1251-1262.
- 377. Vaddepally RK, Kharel P, Pandey R, Garje R, Chandra AB. Review of indications of FDA-approved immune checkpoint inhibitors per NCCN guidelines with the level of evidence. *Cancers (Basel)*. 2020;12(3):738.
- 378. Robert C, Long GV, Brady B, et al. Nivolumab in previously untreated melanoma without BRAF mutation. *N Engl J Med.* 2015;372(4):320-330.
- 379. Larkin J, Chiarion-Sileni V, Gonzalez R, et al. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N Engl J Med.* 2015;373(1):23-34.
- 380. Wolchok JD, Chiarion-Sileni V, Gonzalez R, et al. Overall survival with combined nivolumab and Ipilimumab in advanced Melanoma. N Engl J Med. 2017;377(14):1345-1356.
- 381. Goldberg SB, Schalper KA, Gettinger SN, et al. Pembrolizumab for management of patients with NSCLC and brain metastases: long-term results and biomarker analysis from a non-randomised, open-label, phase 2 trial. *Lancet Oncol.* 2020;21(5):655-663.
- 382. Socinski MA, Jotte RM, Cappuzzo F, et al. Atezolizumab for first-line treatment of metastatic nonsquamous NSCLC. N Engl J Med. 2018;378(24):2288-2301.
- 383. Esteva FJ, Hubbard-Lucey VM, Tang J, Pusztai L. Immunotherapy and targeted therapy combinations in metastatic breast cancer. *Lancet Oncol.* 2019;20(3):e175-e186.
- 384. Li SJ, Chen JX, Sun ZJ. Improving antitumor immunity using antiangiogenic agents: Mechanistic insights, current progress, and clinical challenges. *Cancer Commun (Lond)*. 2021;41(9):830-850.
- 385. Heinhuis KM, Ros W, Kok M, Steeghs N, Beijnen JH, Schellens JHM. Enhancing antitumor response by combining immune checkpoint inhibitors with chemotherapy in solid tumors. *Ann Oncol.* 2019;30(2):219-235.
- 386. Yi M, Jiao D, Qin S, Chu Q, Wu K, Li A. Synergistic effect of immune checkpoint blockade and anti-angiogenesis in cancer treatment. *Mol Cancer*. 2019;18(1):60.
- 387. Paz-Ares L, Luft A, Vicente D, et al. Pembrolizumab plus chemotherapy for squamous non-small-cell lung cancer. N Engl J Med. 2018;379(21):2040-2051.
- 388. Gandhi L, Rodriguez-Abreu D, Gadgeel S, et al. Pembrolizumab plus chemotherapy in metastatic non-small-cell lung cancer. *N Engl J Med*. 2018;378(22):2078-2092.
- 389. Motzer R, Alekseev B, Rha SY, et al. Lenvatinib plus pembrolizumab or everolimus for qdvanced renal cell carcinoma. *N Engl J Med.* 2021;384(14):1289-1300.
- Rini BI, Plimack ER, Stus V, et al. Pembrolizumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. N Engl J Med. 2019;380(12):1116-1127.
- 391. Escudier B. Combination therapy as first-line treatment in metastatic renal-cell carcinoma. *N Engl J Med.* 2019;380(12):1176-1178.
- 392. Theelen WSME, Chen D, Verma V, et al. Pembrolizumab with or without radiotherapy for metastatic non-small-cell lung

cancer: a pooled analysis of two randomised trials. *Lancet Respir Med.* 2021;9(5):467-475.

- 393. Reno TA, Kim JY, Raz DJ. Triptolide inhibits lung cancer cell migration, invasion, and metastasis. *Ann Thorac Surg.* 2015;100(5):1817-1824, discussion 1824–1825.
- 394. Zhao M, Sun Y, Gao Z, et al. Gigantol attenuates the metastasis of human bladder cancer cells, possibly through Wnt/EMT signaling. Onco Targets Ther. 2020;13:11337-11346.
- 395. Ouanouki A, Lamy S, Annabi B. Anthocyanidins inhibit epithelial-mesenchymal transition through a TGFbeta/Smad2 signaling pathway in glioblastoma cells. *Mol Carcinog.* 2017;56(3):1088-1099.
- 396. Ning N, Liu S, Liu X, et al. Curcumol inhibits the proliferation and metastasis of melanoma via the miR-152-3p/PI3K/AKT and ERK/NF-kappaB signaling pathways. J Cancer. 2020;11(7):1679-1692.
- 397. Ning L, Ma H, Jiang Z, et al. Curcumol suppresses breast cancer cell metastasis by inhibiting MMP-9 Via JNK1/2 and AKTdependent NF-kappaB signaling pathways. *Integr Cancer Ther.* 2016;15(2):216-225.
- 398. Zhang P, Zhai Y, Cai Y, Zhao Y, Li Y. Nanomedicine-based immunotherapy for the treatment of cancer metastasis. *Adv Mater*. 2019;31(49):e1904156.
- 399. Liang C, Xu L, Song G, Liu Z. Emerging nanomedicine approaches fighting tumor metastasis: animal models, metastasis-targeted drug delivery, phototherapy, and immunotherapy. *Chem Soc Rev.* 2016;45(22):6250-6269.
- 400. Hu C, Liu X, Ran W, et al. Regulating cancer associated fibroblasts with losartan-loaded injectable peptide hydrogel to potentiate chemotherapy in inhibiting growth and lung metastasis of triple negative breast cancer. *Biomaterials*. 2017;144:60-72.
- 401. He X, Yu H, Bao X, et al. PH-responsive wormlike micelles with sequential metastasis targeting inhibit lung metastasis of breast cancer. *Adv Healthc Mater*. 2016;5(4):439-448.
- 402. Xiao J, Duan X, Yin Q, et al. The inhibition of metastasis and growth of breast cancer by blocking the NF-κB signaling pathway using bioreducible PEI-based/p65 shRNA complex nanoparticles. *Biomaterials*. 2013;34(21):5381-5390.
- 403. Xu P, Yu H, Zhang Z, et al. Hydrogen-bonded and reductionresponsive micelles loading atorvastatin for therapy of breast cancer metastasis. *Biomaterials*. 2014;35(26):7574-7587.
- 404. Zetter BR. Angiogenesis and tumor metastasis. *Annu Rev Med.* 1998;49:407-424.
- 405. Aiello NM, Kang Y. Context-dependent EMT programs in cancer metastasis. *J Exp Med*. 2019;216(5):1016-1026.
- 406. Janssen LME, Ramsay EE, Logsdon CD, Overwijk WW. The immune system in cancer metastasis: friend or foe? J Immunother Cancer. 2017;5(1):79.

**How to cite this article:** Liu M, Yang J, Xu B, Zhang X. Tumor metastasis: Mechanistic Insights and therapeutic interventions. *MedComm*. 2021;2:587–617. https://doi.org/10.1002/mco2.100