Table S1. Summary of PET biodegradation assay 1. A PET biodegradation assay (see Methods) was performed using 17 test conditions and 2 unsupplemented YSV control conditions [(pos) = with *P. sakaiensis*; (neg) = no bacteria]. Chemical concentrations in each supplemented test condition are given as percentages (w/v) for all but sodium phosphate pH 7 and sodium nitrite (mM). PET strip masses were measured weekly for 5 weeks. The percent biodegradation is the total PET mass degraded by *P. sakaiensis* after 5 weeks. Fold-differences in % biodegradation

(fold-diff) were calculated relative to the unsupplemented positive control. An average biodegradation rate and standard deviation (mg·cm⁻²·day⁻¹) was calculated from the weekly biodegradation rates for each condition. Finally, weekly changes in biodegradation rate were calculated and summed for the PET biodegradation dynamics supported by each condition.

										weekly	change i	n biodegi	weekly change in biodegradation rate (%)	ate (%)
condition #	chem1	[chem1]	chem2	[chem2]	% biodegradation	fold-diff	 biodegradation rate>	stdev	fold-diff	1-2	2-3	3-4	4-5	mns
1	gm802	0.39		1	12.60	2.13	0.093	0.023	2.15	-0.15	-24.89	-23.92	17.96	-31.00
2	gm802	0.78		1	12.62	2.14	0.093	0.018	2.14	-14.41	-12.37	-23.92	41.56	-9.15
3	gm802	12.50		!	9.74	1.65	0.070	890'0	1.61	33.14	25.19	-111.41	-194.37	-247.46
4	GABA	0.03		1	3.81	0.64	0.027	0.027	0.63	-14.41	-41.58	-100.00	0.00	-155.99
2	sodium phosphate	10.00	ethylene glycol	0.78	11.60	1.96	0.088	0.040	2.03	128.23	53.35	14.12	7.85	203.55
9	sodium phosphate	5.00	ethylene glycol	0.78	6.29	1.06	0.047	0.029	1.08	185.29	-12.37	-77.18	88.74	184.49
7	sodium phosphate	10.00	GABA	0.01	10.03	1.70	0.074	0.032	1.71	14.12	2.23	-67.40	41.56	-9.49
8	gm802	6.25	GABA	0.02	3.75	0.64	0.027	0.057	0.62	-57.21	-129.21	14.12	-100.00	-272.30
6	gm802	0.20	sodium phosphate	5.00	14.59	2.47	0.108	0.019	2.50	-0.15	12.67	-36.60	13.24	-10.84
10	gm802	0.20	sodium phosphate	2.50	11.27	1.91	0.084	0.018	1.93	14.12	2.23	-34.79	-5.63	-24.07
11	gm802	0.78	L-serine	0.03	10.24	1.73	0.075	0.035	1.72	-57.21	-41.58	185.29	13.24	99.75
12	gm802	0.78	L-serine	0.02	12.01	2.03	0.090	200'0	2.09	-4.90	5.16	14.12	-5.63	8.74
13	gm802	0.39	L-serine	0.05	11.00	1.86	0.083	0.020	1.92	-4.90	22.68	-51.09	88.74	55.43
14	gm802	0.39	L-serine	0.01	10.17	1.72	0.074	0.032	1.72	-28.68	-29.90	42.65	-62.25	-78.18
15	gm802	0.39	GABA	0.03	11.54	1.95	0.086	0.021	1.99	-23.92	75.26	-42.94	41.56	49.95
16	gm802	0.78	GABA	0.02	12.27	2.08	0.090	0.017	2.08	-28.68	-12.37	14.12	13.24	-13.69
17	gm802	0.20	GABA	0.01	11.42	1.93	0.083	0.021	1.92	-42.94	31.44	-23.92	17.96	-17.46
18	gm802	0.78	ethylene glycol	0.13	10.86	1.84	0.080	0.016	1.86	-4.90	22.68	-34.79	-5.63	-22.64
19	gm802	0.39	ethylene glycol	0.13	13.47	2.28	0.099	0.014	2.29	-14.41	-12.37	-4.90	32.12	0.43
20 (pos)	-				5.91	1.00	0.043	0.011	1.00	-42.94	31.44	14.12	-37.09	-34.47
21 (neg)	1		1		0.00		0.000	0.000						

Table S2. Summary of PET biodegradation assay 2. A PET biodegradation assay (see Methods) was performed using 19 test conditions and 2 unsupplemented YSV control conditions

[(pos) = with P. sakaiensis; (neg) = no bacteria]. Chemical concentrations in each supplemented test condition are given as percentages (w/v) for all but sodium phosphate pH 7 (mM). PET strip masses were measured weekly for 5 weeks. The percent biodegradation is the total PET mass degraded by P. sakaiensis after 5 weeks. Fold-differences in % biodegradation (fold-diff) were calculated relative to the unsupplemented positive control. An average biodegradation rate and standard deviation (mg·cm⁻²·day⁻¹) was calculated from the weekly biodegradation rates for each condition. Finally, weekly changes in biodegradation rate were calculated and summed for the PET biodegradation dynamics supported by each condition.

										We	weekly change in biodegradation rate (%)	n biodegra	lation rate	(%)
condition #	chem1	[chem1]	chem2	[chem2]	[chem2] % biodegradation fold-diff		 biodegradation rate>		stdev fold-diff	1-2	2-3	3-4	4-5	uns
1a	sodium phosphate	10.00	GABA	0.008	13.43	1.78	0.093	0.015	1.79	-30.33	43.52	-16.39	-10.30	-13.49
;	mnipos			0			000			i i			L C	
qŢ	phosphate	10.00	GABA	0.008	11.52	1.53	0.080	0.015	1.52	-75.68	56.98	-4.45	-72.75	1.61
2a	gm802	0.39	GABA	0.031	10.33	1.37	0.071	0.030	1.36	-30.33	7.64	-25.68	-55.15	-103.51
2b	gm802	0.39	GABA	0.031	8.98	1.19	0.063	0.022	1.20	-25.68	34.55	-25.68	-55.15	-71.96
33	sodium	10.00	ethylene	0 780	92 11	1 56	080 0	0.017	1 52	30 35	25 58	75 02-	25.58	70.14
1	sodium		ethylene						1					
3b	phosphate	10.00	glycol	0.780	15.58	2.07	0.105	0.045	2.02	11.48	124.26	11.48	-19.27	127.95
4a	gm802	0.78	L-serine	0.016	11.72	1.55	0.083	0.020	1.58	67.220	19.603	-30.325	7.643	64.141
4b	8m802	0.78	L-serine	0.016	25.59	3.39	0.172	0.075	3.29	178.70	61.46	-7.10	-22.26	210.81
5a	8m802	0.78	GABA	0.016	11.71	1.55	080'0	0.021	1.53	67.22	19.60	-30.33	-10.30	46.20
2b	gm802	0.78	GABA	0.016	13.14	1.74	0.091	0.038	1.74	95.09	40.96	-49.33	-10.30	76.43
6а	gm802	0.39			22.43	2.97	0.154	0.070	2.95	178.70	61.46	-25.68	-25.25	189.24
q9	gm802	0.39	-		19.27	2.56	0.137	0.023	2.62	11.48	12.13	22.63	-10.30	35.94
7а	gm802	0.78			12.92	1.71	0.091	0.011	1.74	11.48	19.60	-16.39	-10.30	4.40
7b	gm802	0.78	-		12.16	1.61	0.085	0.015	1.63	11.48	4.65	-36.30	56.98	36.81
8	gm807	0 30	ethylene	0.125	18 46	2.45	0.129	0.068	2 47	150.83	59 47	-23.36	-59 23	177.77
	0		ethylene											
gp	gm802	0.39	glycol	0.125	22.93	3.04	0.156	0.047	2.99	53.28	38.63	-40.98	-0.33	50.60
66 0	gm802	0.20	sodium	5.000	15.27	2.02	0.108	0.044	2.07	95.09	66.59	-39.97	-10.30	111.41
			sodium											
q6	gm802	0.20	phosphate	5.000	12.56	1.67	0.085	0.021	1.63	39.35	25.58	-4.45	19.60	80.09
10a (pos)			-		92'8	1.16	0.059	0.014	1.14	11.48	34.55	-44.26	19.60	21.38
10b (pos)	-		-		6.32	0.84	0.045	0.017	0.86	-16.39	19.60	-72.13	258.81	189.89
11 (neg)	1		ı		0.00		0.000	0.000						

Table S3. Summary of PET biodegradation assay 3. A PET biodegradation assay (see Methods) was performed using 9 test conditions and 2 unsupplemented YSV control conditions [(pos) = with *P. sakaiensis*; (neg) = no bacteria]. Conditions 1-10 were done in duplicate (a & b). Chemical concentrations in each supplemented test condition are given as percentages (w/v) for all but sodium phosphate pH 7 (mM). PET strip masses were measured weekly for 5 weeks. The

percent biodegradation is the total PET mass degraded by *P. sakaiensis* after 5 weeks. Fold-differences in % biodegradation (fold-diff) were calculated relative to the unsupplemented positive control. An average biodegradation rate and standard deviation (mg·cm⁻²·day⁻¹) was calculated from the weekly biodegradation rates for each condition. Finally, weekly changes in biodegradation rate were calculated and summed for the PET biodegradation dynamics supported by each condition.

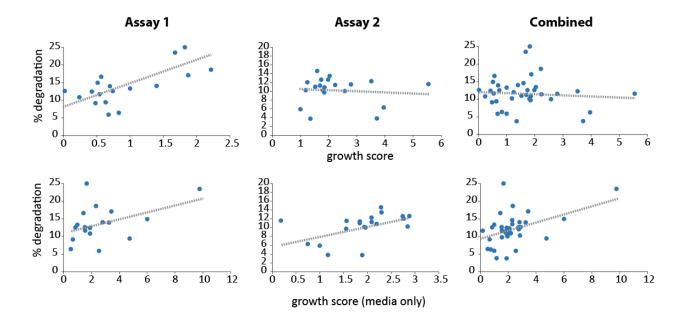


Fig S1. The extent of PET biodegradation after 5 weeks correlates with growth scores. Percent biodegradation after 5 weeks vs. growth scores measured after 2 days for biodegradation assay 1 (left plots), assay 2 (middle plots), and combined data (right plots). Two different growth scores were calculated: 1) from A600 measurements of each well of the 24-well plates (= growth score); and 2) from A600 measurements of 120 microL media taken from each well of the 24-well plates and transferred to a 96-well plate (= media only growth score). A600 measurements from media only are specific for planktonic bacterial growth, while A600 measurements from 24-well plates are affected by variable light scattering from partly degraded PET, the thickness of the PET-bound biofilm, and the presence of planktonic bacteria. (Linear fits, clockwise starting from the top left: 1) y = 5.48x + 8.80; $R^2 = 0.45$; 2) y = -0.23x + 10.77; $R^2 = 0.01$; 3) y = 0.30x + 12.42; $R^2 = 0.01$; 4) y = 1.28x + 9.04; $R^2 = 0.23$; 5) y = 2.08x + 6.32; $R^2 = 0.24$; and 6) y = 0.98x + 11.18; $R^2 = 0.19$).

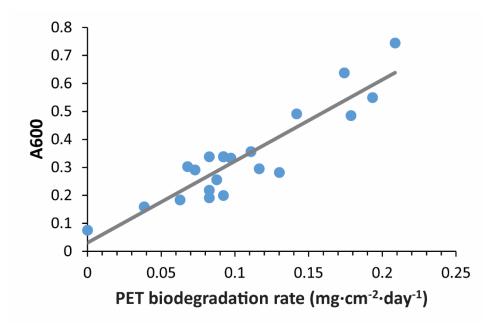


Fig S2. Microbial growth strongly correlates with PET biodegradation rates. Average weekly culture media A600 vs. average weekly PET biodegradation rate measured for biodegradation assay 3. Linear fit: y = 2.91x + 0.03; $R^2 = 0.83$

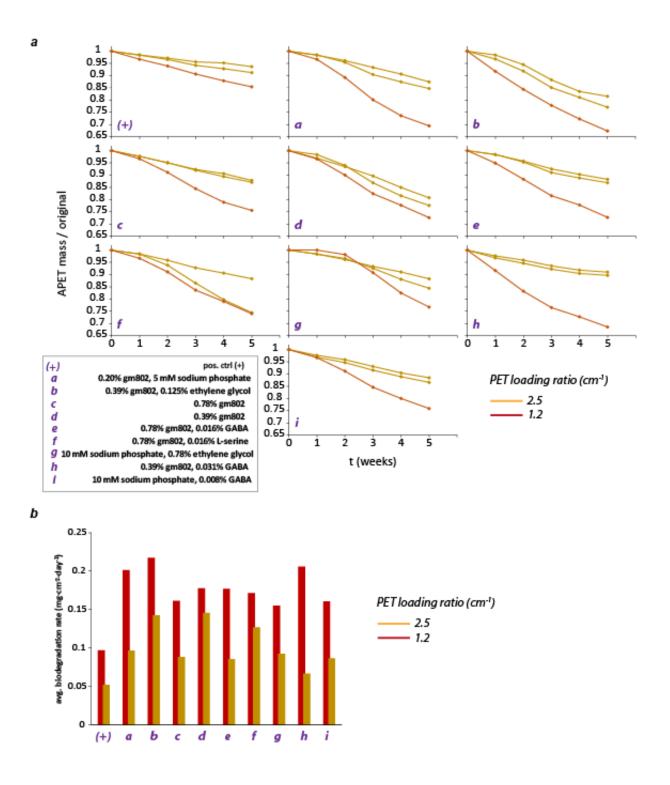


Fig S3. Lowering the PET loading ratio increases biodegradation by *P. sakaiensis*. (A) PET biodegradation plots showing total PET mass over time for 10 different culture conditions ([+] and a-i), each at two different PET loading ratios (2.5 cm⁻¹ in yellow and 1.2cm⁻¹ in red). Assays with PET loading ratio of 2.5 cm⁻¹ were done in duplicate. (B) Average weekly PET biodegradation rates increase with decreasing PET loading ratio.