

RESEARCH ARTICLE

# Genetic factors have a major effect on growth, number of vertebrae and otolith shape in Atlantic herring (*Clupea harengus*)

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## Abstract

Atlantic herring, *Clupea harengus*, have complex population structures. Mixing of populations is known, but the extent of connectivity is still unclear. Phenotypic plasticity results in divergent phenotypes in response to environmental factors. A marked salinity gradient occurs from Atlantic Ocean (salinity 35) into the Baltic Sea (salinity range 2–12). Herring from both habitats display phenotypic and genetic variability. To explore how genetic factors and salinity influence phenotypic traits like growth, number of vertebrae and otolith shape an experimental population consisting of Atlantic purebreds and Atlantic/Baltic F1 hybrids were incubated and co-reared at two different salinities, 16 and 35, for three years. The F1-generation was repeatedly sampled to evaluate temporal variation. A von Bertalanffy growth model indicated that reared Atlantic purebreds had a higher maximum length (26.2 cm) than Atlantic/Baltic hybrids (24.8 cm) at salinity 35, but not at salinity 16 (25.0 and 24.8 cm, respectively). In contrast, Atlantic/Baltic hybrids achieved larger size-at-age than the wild caught Baltic parental group. Mean vertebral counts and otolith aspect ratios were higher for reared Atlantic purebreds than Atlantic/Baltic hybrids, consistent with the differences between parental groups. There were no significant differences in vertebral counts and otolith aspect ratios between herring with the same genotype but raised in different salinities. A Canonical Analysis of Principal Coordinates was applied to analyze the variation in wavelet coefficients that described otolith shape. The first discriminating axis identified the differences between Atlantic purebreds and Atlantic/Baltic hybrids, while the second axis represented salinity differences. Assigning otoliths based on genetic groups (Atlantic purebreds vs. Atlantic/Baltic hybrids) yielded higher classification success (~90%) than based on salinities (16 vs. 35; ~60%). Our results demonstrate that otolith shape and vertebral counts have a significant genetic component and are therefore useful for studies on population dynamics and connectivity.

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## Introduction

Phenotypic plasticity is the ability to display different phenotypes in response to environmental factors [1]. It has become increasingly important as the basic concept to discriminate marine fish populations [2] since individuals of a population are assumed to live under specific environmental conditions. Traditionally, fish populations have been identified based on phenotypic traits, although the relative importance of genetic and environmental factors on the determination of those phenotypic traits is generally unclear [3–5]. Therefore, the genetic and/or environmental mechanisms regulating phenotypic traits used for identification of fish populations need to be clarified and defined.

As one of the ecologically and commercially most important fish species in the northeastern Atlantic, herring (*Clupea harengus*) has been a key species for studies of population structure. Iles and Sinclair [6] proposed that Atlantic herring have complex population structure and much effort has been spent to resolve this structure. Phenotypic traits like growth [7, 8], numbers of vertebrae [9, 10], otolith microstructure [11, 12], as well as otolith shape [13–15], have been used to investigate the population structure of herring. Genetic studies have also become important during recent years. While no or very limited genetic differentiation was initially found between populations [16, 17], genome-wide analyses revealed clear genetic differentiation among Atlantic herring populations, but primarily at loci underlying ecological adaptation [18, 19].

Environmental factors have a strong influence on many phenotypic traits in herring. For example, temperature affects growth [20] and otolith microstructure [21, 22], as well as the number of vertebrae and can act in combination with salinity [23, 24]. While growth and otolith microstructure can vary over time with temperature, the number of vertebrae will be determined once during metamorphosis based on the experienced environmental conditions [25]. In herring, there is also a co-variability between genetic differentiation and salinity [19, 26, 27].

A strong salinity gradient, highly correlated with the genetic differentiation of herring in this area [19], occurs throughout the Baltic Sea from the inner Bothnian Bay (salinity <6) to the opening near the fully marine North Sea/Atlantic Ocean (salinity 35). Further, this salinity gradient is associated with differences in phenotypic traits of herring inhabiting these two environments. Beside the salinity, the temperature is often examined as the main factor determining variation in phenotypic traits. However, the average temperature difference between the Atlantic and Baltic (S5 Fig) is relatively minor compared to the vast salinity variation and subject also to marked seasonal variations. In addition, the lower average temperature in the Baltic contradicts the common assumption of a negative correlation between the number of vertebrae and temperature [23].

To understand or resolve the genetic and environmental influences on phenotypic traits, we used offspring of two herring populations (Atlantic vs. Baltic) that were genetically different and living in contrasting salinities in a common garden rearing experiment. Common garden experiments are designed to rear offspring from different populations under identical environmental conditions. Both Atlantic purebreds and Atlantic/Baltic hybrids were reared under controlled conditions with fixed salinities of either 16 or 35. Our main objectives were to explore genetic and salinity influences on phenotypic traits like growth, number of vertebrae and otolith shape. Further, the experiment was conducted over a 3-year period to evaluate potential temporal variation in growth and otolith shape. Temporal variation in the number of vertebrae was tested to determine if it was subjected to selection.

## Material and methods

Spring spawning herring caught by gillnets on 21 May 2013 in the Atlantic, approximately 12 km west of Bergen, Norway (60°34'11.2"N 5°0'18.9"E) and Baltic, approximately 80 km north-east of Uppsala, Sweden (60°38'52.0"N 17°48'44.2"E) were used as parental fish in this study. Half of the eggs from one Atlantic female were fertilized and incubated on the day of capture with sperm of one Atlantic male; the other half was fertilized with sperm of one Baltic male. The Atlantic herring were 5 years of age, 30.5 cm (female) and 32.5 cm (male) in total length with 57 vertebrae. The Baltic male was 8 years of age, 20.5 cm in total length with 55 vertebrae. The age of herring was determined by counts of winter rings from otoliths. The experimental setup, including only one mother, was designed to avoid any environmental maternal effects. The parental herring were from a subset of samples representing typical Atlantic and Baltic populations that exhibited huge phenotypic differences between groups (S6 Fig) and have been genetically characterized confirming population-specific differences [19]. S1 Appendix provides further details about the parental groups.

The fertilization and rearing experiment was conducted under common garden conditions at salinities 16 and 35, with values fluctuating during incubation between 15–17 and 34–35, respectively. Water temperatures varied with seasons with an average of  $9.12 \pm 0.73^\circ\text{C}$  and  $9.04 \pm 0.71^\circ\text{C}$  at salinity 16 and 35, respectively (S5 Fig) and the light intensities fluctuated according to the seasonal and daily cycle in Bergen (60°N). Fifty percent hatching, defined as day 0, occurred on 5 June 2013. Atlantic purebreds and Atlantic/Baltic F1 hybrids, hereafter called purebreds and hybrids, were co-reared at salinity 16 and 35 in two replicated 1 m circular tanks, including in total 1000 larvae at an initial purebred/hybrid ratio of 1:2. Herring larvae were fed in excess, firstly with live natural zooplankton and cultured rotifers [28] and later with *Artemia* spp. (23 days post hatching = DPH), until feeding on formulated feed started (71 DPH). On 3 October (120 DPH), juveniles were transferred into two 3 m circular tanks, one with salinity 16 and one with salinity 35, where the herring were reared further for nearly 3 years until their first maturity. This experimental setup generated four groups (*Pop*, in statistical models) which can be distinguished genetically into purebreds and hybrids, as well as by salinity: H16 = hybrid at 16, H35 = hybrid at 35, P16 = purebred at 16 and P35 = purebred at 35.

During the three years, we sampled 690 otoliths out of 950 herring (Table 1). All sampled fish were measured to the nearest mm and fin clipped for DNA analysis, whereas the number of vertebrae was counted only for some samples ( $n = 522$ , Table 1). The DNA samples were used to identify post-mortem Atlantic/Baltic hybrids and Atlantic purebreds by genotyping a diagnostic SNP using a Custom TaqMan® Assay Design Tool where the Baltic male was homozygous C (cytosine), while the Atlantic male and female were homozygous T (thymine) at a specific SNP locus (scaffold95\_175856\_SNP00029). The following TaqMan probe was used to distinguish between hybrids and purebreds:

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GCTTCTTTGTCAGCAGAGGCACAGACATTT [T/C] GTTACATAAGGAGATGCTGTGTCAGGTCAG
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where T/C, among brackets is the SNP assayed.

A digital image of each otolith was captured using a Leica MZ95 stereomicroscope and reflected light with a Nikon digital sight DS-U1 microscope camera using the software NIS-elements F (Version 2.3). Following the method by Libungan et al. [29], otolith images were read into the R software [30], and otolith shape outlines were collected from the images using the shapeR package [31]. A discrete wavelet transformation to equally spaced radii from the otolith centroid to the otolith outline was conducted to obtain wavelet coefficients (unitless). Hereafter, otolith shape refers to variation in the wavelet coefficients representing the otolith shape outline. An analysis of covariance (ANCOVA) was performed individually for each sample

**Table 1. Total numbers of analyzed herring and otoliths (in brackets).**

DPH	H16	P16	H35	P35	Sample
187	85 (73)	14 (13)	69 (61)	30 (26)	1*
297	36 (31)	4 (2)	31 (24)	19 (18)	2*
482	0	0	76 (24)	37 (19)	
524	0	0	56 (0)	34 (0)	
531	0	0	17 (0)	8 (0)	
618	27 (23)	3 (2)	16 (14)	14 (12)	3*
702	0	0	10 (8)	10 (8)	
861	11 (8)	1 (1)	19 (13)	12 (11)	4
960	7 (3)	1 (1)	23 (22)	9 (9)	4*
1055	0	0	16 (15)	14 (14)	
1079	0	0	31 (31)	8 (8)	
1098	33 (33)	5 (4)	38 (37)	14 (13)	5*
1106	23 (22)	8 (7)	18 (18)	12 (12)	5*
1120	17 (17)	4 (4)	16 (15)	14 (14)	5*
<b>Total</b>	<b>240 (210)</b>	<b>39 (34)</b>	<b>436 (282)</b>	<b>235 (164)</b>	

Samples from different sampling days (DPH = days post hatching) that were combined for otolith analyses were marked with identical numbers in the rightmost column.

\* Number of vertebrae was also counted. H16 = hybrids at salinity 16, P16 = purebreds at salinity 16, H35 = hybrids at salinity 35, P35 = purebreds at salinity 35.

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(Table 1) to determine the effect of fish length on the wavelet coefficients, as well as otolith length and width. Coefficients which showed an interaction between the four herring groups and total length were excluded from the analysis (S2 Table). In this study, fish length could also be used as a proxy for the growth rate, because all fish from a sample had the same age (days post hatching, DPH). Further, the remaining coefficients, as well as otolith length and width, were adjusted for allometric relationships with fish length applying the normalization technique of Leonart et al. [32].

For all model fittings, full and complex models were used as starting references and simplified in cases of non-significance. Length-at-age data, used as a proxy for somatic growth of individual herring, were fitted to the von Bertalanffy growth model [33]:

$$TL_t = L_{\infty Pop}(1 - e^{-K(t-t_0)})$$

where  $TL_t$  is the average length at age  $t$ ,  $L_{\infty}$  is the asymptotic maximum length of each of the four herring groups ( $Pop$ ),  $K$  is the von Bertalanffy growth rate coefficient, i.e., the rate at which length approaches the maximum length asymptote and  $t_0$  is the intercept on the time axis.

The aspect ratio (otolith length/width) was calculated for comparison with the parental groups. Further ANOVA tests were used to evaluate the significance of genetic origin ( $Gen$ ) and salinity ( $Sal$ ) on otolith width ( $OW$ ) or length ( $OL$ ) at given age classes ( $DPH$  as factor):

$$OW \text{ or } OL = \alpha + \beta_1 \times DPH + \beta_2 \times Gen + \beta_3 \times Sal$$

Differences in the otolith aspect ratio ( $AR$ ) and the mean number of vertebrae ( $VS$ ) among

genetic origin (*Gen*) and salinity (*Sal*) were tested using a one-way ANOVA:

$$AR = \alpha + \beta_1 \times Gen + \beta_2 \times Sal$$

$$VS = \alpha + \beta_1 \times Gen$$

The initial starting model also included, in both cases, the age (*DPH*) as a predictor variable, but this was removed due to non-significance. Since the number of vertebrae is fixed during metamorphosis, temporal variation would indicate some kind of selection, either through sampling or mortality. Significant differences among the four herring groups were identified using Tukey-HSD tests. A significance level  $\alpha = 0.05$  was applied for all analyses and statistical tests.

For a subset of the sampling days, otoliths had been obtained from fish in both salinities. Only these otoliths were used for the otolith shape analyses to allow for comparisons across salinities. In some cases (sample 4 and 5), adjacent sampling days (*DPH*) were combined due to low numbers (Table 1). Those samples were taken within 100 days of each other, and none of the analyzed characteristics differed. The temporal development of otolith shape outlines and general differences among the groups were examined visually by plotting the mean otolith shape outline of each group reconstructed of the wavelet coefficients (S1 Fig). To investigate which region of the otolith shape outline contributed most to the differences between the four groups, mean wavelet coefficients and their standard deviation were plotted against the angle of the outline. Thereby, each mean wavelet coefficient indicates the variation of the otolith shape outline within their predefined region. Further, the correlation within each group along the outline was estimated with an intraclass correlation (*ICC*). Consequently, a combination of a high mean wavelet coefficient ( $>0.25$ ) and a high *ICC* indicated the region along the otolith shape outline that differed most.

For statistical analysis to demonstrate the variation in otolith shape represented by wavelet coefficients, Canonical Analysis of Principal Coordinates (*CAP*) [34] followed by ANOVA-like permutation tests were applied with 2000 permutations used to assess the significance of constraints. The *CAP* and ANOVA-like permutation tests were only applied to otoliths from herring of age 187 *DPH* and 1108 *DPH* (Table 1) because these samples provided enough otoliths from all groups to ensure reliable results. Finally, the ordinations of group averages were examined with the shape descriptors along the first two canonical axes. Using the *CAP* and the ANOVA-like permutation tests, otolith shape was compared among the four herring groups with an overall test, as well as by applying comparison between salinity and genetic groups. In addition, salinity effects on the otolith shape were investigated in the absence of any genetic differences by comparing hybrids originating from salinity 16 and 35 in isolation and purebreds in isolation. The genetic signal was examined in the same way in the absence of any salinity differences.

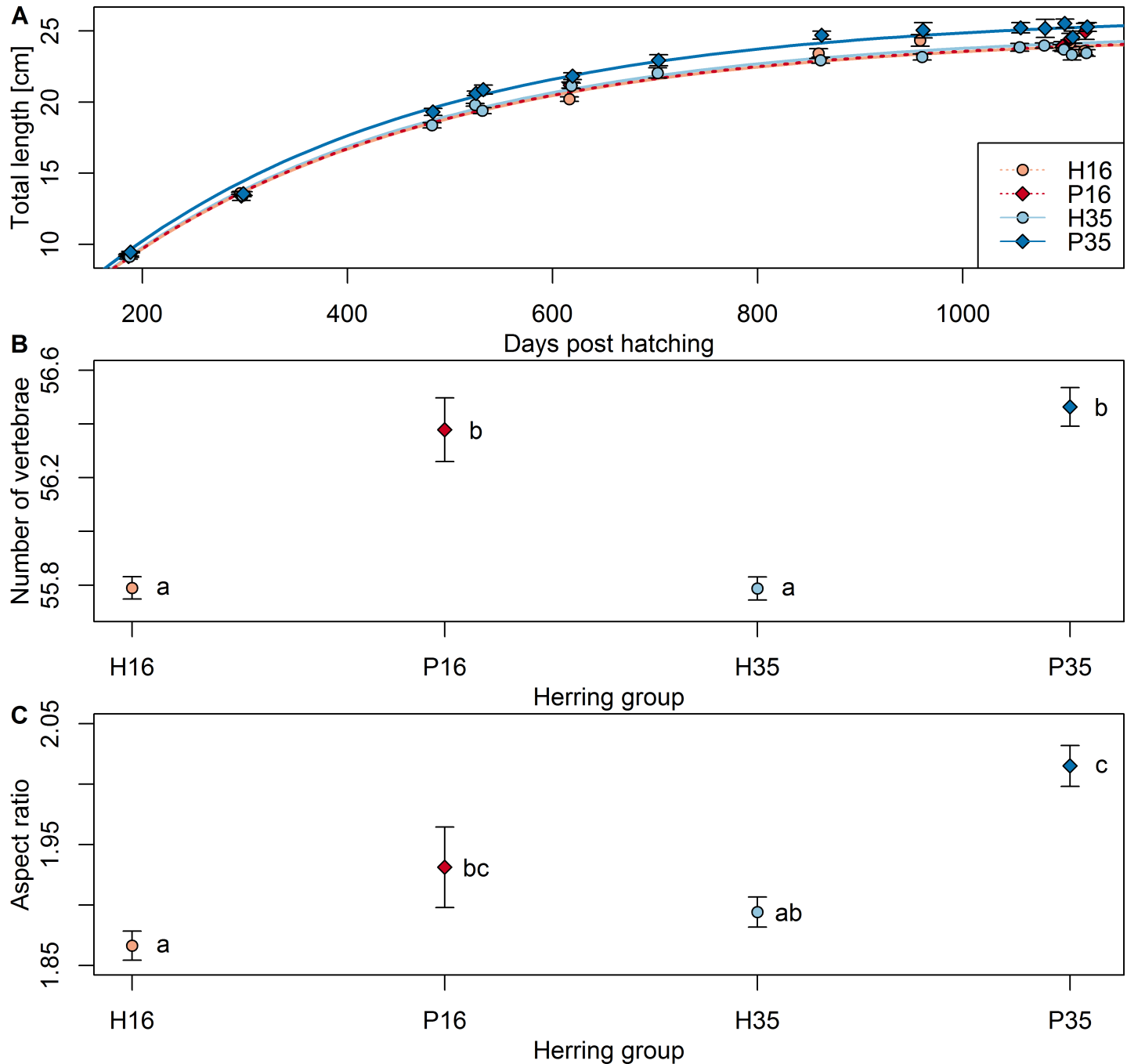
To validate if otolith shape analysis can be used for assigning individual herring to a given group, we applied a linear discriminant analysis (*LDA*) to the standardized wavelet coefficients of sample 1 and 5. The classification success into salinities, genetics, as well as the four groups, was estimated using the leave-one-out cross-validation [35]. Thus, each otolith was removed individually from the dataset and assigned to one of the predefined groups.

## Ethics statement

Herring samples of the parental Atlantic population were caught with permission of the Directorate of Fisheries, Bergen, Norway. The parental Baltic herring were purchased from a local commercial fisherman. The common garden experiment and rearing of the F1-generation was approved by the Norwegian national animal ethics committee (Forsøksdyrutvalget–FOTS ID-5072).

### Results

Somatic growth of herring reared under common garden conditions over three years only differed in the maximum asymptotic length among the four herring groups (Fig 1A). Atlantic purebreds reared at salinity 35 ( $L_{\infty} = 26.2$  cm) were larger (ANOVA:  $F = 194.5$ ,  $d.f. = 944$ ,



**Fig 1. Comparison of phenotypic traits among the four herring groups.** (A) length-at-age and von Bertalanffy growth models, (B) number of vertebrae, and (C) otolith aspect ratio were compared among H16 = hybrids at salinity 16, P16 = purebreds at salinity 16, H35 = hybrids at salinity 35, P35 = purebreds at salinity 35. Mean values and 1\*SE are shown. Letters indicated posterior Tukey-HSD test results of all pair-wise comparisons. Groups which do not share a letter are significantly different to each other.

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**Table 2. Results from the ANOVA tests investigating the effects of age (days post hatching), salinity (16 vs. 35) and genetics (purebred vs. hybrid) on otolith width and length.**

Variable	Otolith width				Otolith length			
	d.f.	MS	F	p	d.f.	MS	F	p
Days post hatching	11	23.46	3357.3	<0.001	11	105.09	4185.4	<0.001
Salinity	1	0.00	0.3	0.55	1	0.02	0.7	0.41
Genetics	1	0.10	13.6	<0.001	1	0.69	27.5	<0.001
Residuals	673	0.01			673	0.03		

d.f. = degrees of freedom, MS = mean square, F = F-value, p = p-value

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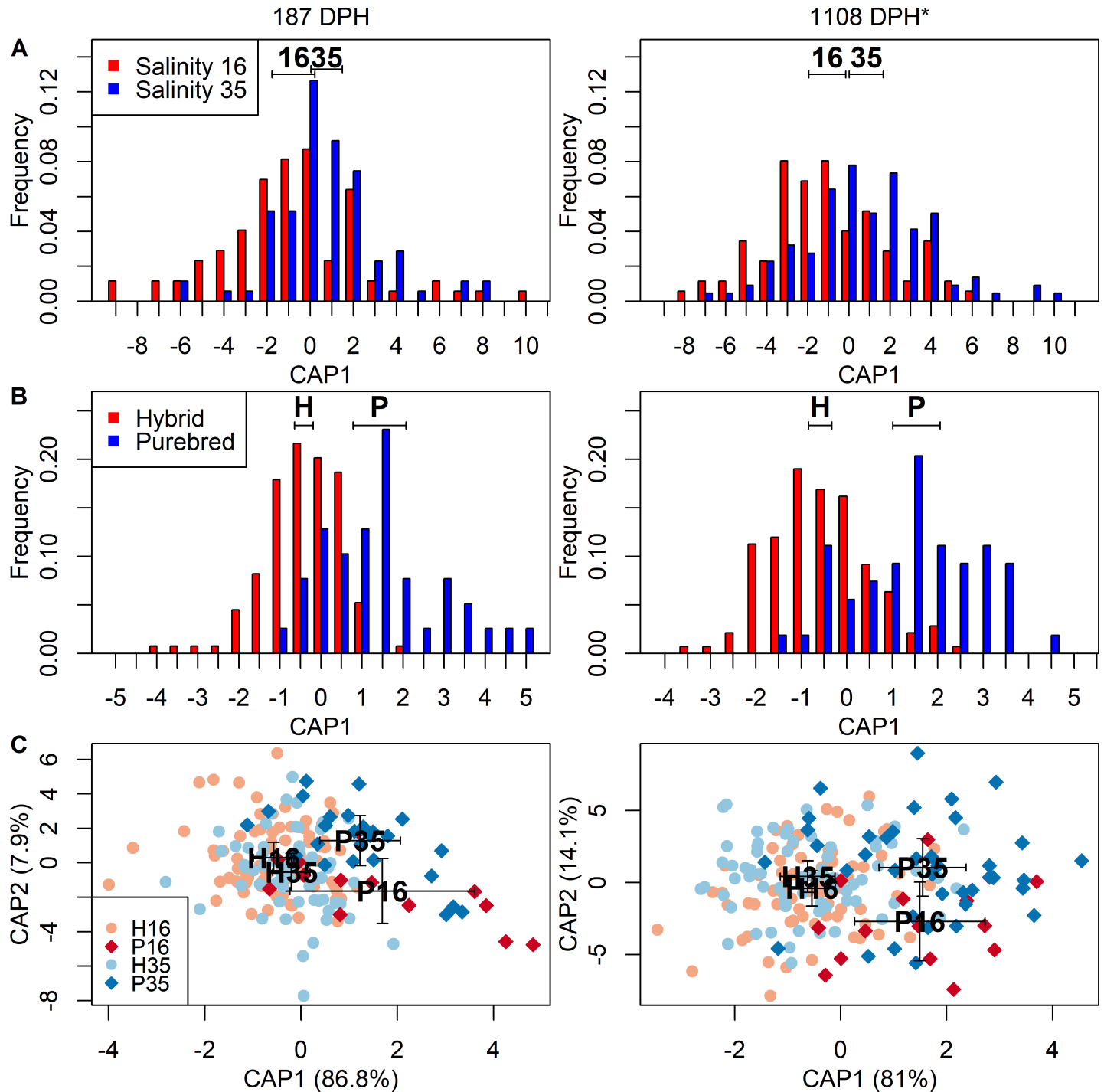
$p < 0.001$ ) than the other three groups (purebreds at salinity 16 ( $L_{\infty} = 25.0$  cm), Atlantic/Baltic hybrids at salinity 35 ( $L_{\infty} = 24.8$  cm) and 16 ( $L_{\infty} = 24.8$  cm)), which did not differ from each other (ANOVA:  $F = 0.69$ ,  $d.f. = 712$ ,  $p > 0.05$ ).

For the number of vertebrae, only a genetic effect could be demonstrated (Fig 1B; ANOVA:  $F = 109.1$ ,  $d.f. = 520$ ,  $r^2 = 0.17$ ,  $p < 0.001$ ). Mean vertebral counts were higher for Atlantic purebreds compared to Atlantic/Baltic hybrids irrespective of salinity (Tukey-HSD tests:  $p < 0.001$ ). The number of vertebrae did not differ over time within each group, indicating that no selection in terms of sampling or mortality had occurred for this trait (ANOVA:  $F = 0.97$ ,  $d.f. = 517$ ,  $p > 0.05$ ).

In general, otoliths of hybrids were shorter but wider compared with purebreds, but there were no differences between otoliths of herring originating from different salinities within each genetic group (Table 2). The otolith aspect ratio was higher for purebreds than hybrids (Fig 1C; ANOVA:  $F = 5.9$ ,  $d.f. = 560$ ,  $r^2 = 0.08$ ,  $p < 0.001$ ), and higher for herring reared at salinity 35 (ANOVA:  $F = 2.5$ ,  $p < 0.02$ ). Purebreds at salinity 16 had a higher aspect ratio than hybrids at salinity 16, but the ratio was not significantly different from hybrids at salinity 35 (Tukey-HSD tests:  $p > 0.05$ ). The aspect ratio did not vary significantly between samples of different ages (ANOVA:  $F = 1.1$ ,  $d.f. = 7$ ,  $p > 0.05$ ). Additional results of the development of the otolith shape outline can be found in the S1 Appendix.

For the following analyses, only otoliths from herring at age 187 DPH and 1108 DPH were used due to a sufficient sample size. Significant differences in otolith shape, represented by wavelet coefficients, were observed between fish from the two salinities, between hybrids and purebreds, as well as among all four groups combined within both ages (Fig 2, Table 3). Comparing the salinity effect separated for hybrids and purebreds demonstrated significant differences (ANOVA:  $F = 2.0$ ,  $d.f. = 1$ ,  $p = 0.049$ ) for hybrids from 187 DPH and purebreds from 1108 DPH, but not for purebreds from 187 DPH and hybrids from 1108 DPH (S3 Fig, S3 Table). While separating herring based on the salinity, otolith shape was always significantly different between hybrids and purebreds originating either from salinity 16 or 35 at both sampling days (S4 Fig). Genetic differences had a higher impact on the otolith shape than salinity differences, as indicated by higher F-values (Table 3). CAP and ANOVA-like permutation tests combining both parameters revealed clear differences among the four groups. The first canonical axis explained most of the variation between genetic groups, both for otoliths from 187 DPH (CAP1: 86.8%) and 1108 DPH (CAP1: 81.0%, Fig 2). Differences between the two salinities were explained by the second canonical axis, but only for otoliths from older herring (CAP2: 14.1%). The otolith shape of younger herring also varied along the second canonical axis, but not significantly and without any distinct pattern.

Otoliths from herring at age 187 DPH and 1108 DPH were classified based on their otolith shape. Classification success of otolith shape varied depending on the classifying character



**Fig 2. Canonical analysis of principal (CAP) scores of herring otolith shapes on discriminating axes.** Scores of the first axis are shown for (A) salinity and (B) genetics, scores of the first and second axis for (C) all four groups. H16 = hybrids at salinity 16, P16 = purebreds at salinity 16, H35 = hybrids at salinity 35, P35 = purebreds at salinity 35. Black bold letters represent the mean canonical value for each character  $\pm 1^*SE$ . Individual fish are represented by frequencies (A, B) or symbols (C). \* Mean day post hatching (DPH) for combined samples.

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(Table 4). Otoliths of both ages had comparable results for each classifying character. Assigning otoliths based on salinities or the four groups had both a success rate between 50–60%,



**Table 3. Results from ANOVA-like permutation tests comparing the otolith shape (represented by wavelet coefficients) among salinities and genetic groups, as well as the four herring groups in the present study.**

Variable	187 days post hatching				1108 days post hatching			
	d.f.	Var	F	p	d.f.	Var	F	p
Salinity	1	0.77	2.2	<0.05	1	1.87	2.3	0.03
Residuals	171	60.31			194	155.67		
Genetics	1	5.09	15.5	<0.001	1	11.89	15.8	<0.001
Residuals	171	55.99			194	145.65		
Genetics	1	5.09	15.7	<0.001	1	11.89	16.0	<0.001
Salinity	1	0.41	1.3	0.23	1	1.54	2.1	0.03
Genetics*Salinity	1	0.72	2.2	0.06	1	1.28	1.7	0.08
Residuals	169	54.87			192	142.83		

d.f. = degrees of freedom, Var = variance, F = F-value, p = p-value.

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whereas based on the genetic groups ~90% of the otoliths were classified correctly. Splitting the results into individual comparisons (e.g., hybrids and purebreds when comparing the salinity effect) gave similar classification successes as for combined samples (Table 4).

### Discussion

This study provides the strongest evidence reported so far that the number of vertebrae and otolith shape (represented by wavelet coefficients) in Atlantic herring have a clear genetic basis and genetics had a more profound effect on these phenotypes than salinity. In general, this study confirms the genetic regulation of otolith shape [36]. The clearly demonstrated genetic effects based on phenotypic data from offspring were consistent with the phenotypic difference between the parental populations (Atlantic vs. Baltic herring). Further, temporal variations over a 3-year period in these traits were not evident, indicating that selection did not occur. The demonstrated differences in otolith aspect ratio or shape were genetically affected and independent of growth rate variations because the allometric relationship was removed by scaling the otoliths according to Leonart et al. [32].

**Table 4. Overall classification success (bold) of otoliths into salinity, genetic groups and the four groups based on a linear discriminant analysis.** The overall groups were evaluated separately (e.g. only hybrids were assigned to salinity) and their classification success presented. The analyses were conducted independently for otoliths from herring at different ages (187 and 1108 days post hatching = DPH).

	Classification success	
	187 DPH	1108 DPH
<b>Salinity</b>	<b>56.7%</b>	<b>60.7%</b>
Hybrid	58.2%	61.3%
Purebred	46.2%	55.6%
<b>Genetics</b>	<b>91.9%</b>	<b>87.2%</b>
Salinity 16	81.4%	77.0%
Salinity 35	81.6%	78.9%
<b>All groups</b>	<b>54.9%</b>	<b>49.5%</b>
Hybrid 16	58.9%	44.4%
Hybrid 35	54.1%	52.9%
Purebred 16	23.1%	33.3%
Purebred 35	61.5%	59.0%

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So far, most studies on population discrimination in fish have largely been based on phenotypic traits without knowledge of the specific genetic background influencing these traits (see references in [37]). Yet, studies combining genetic and environmental impacts are essential to understand population structures in marine fish. In this study, the genetic effect was the main factor underlying the observed phenotypic variation, supporting the use of genetic markers for population discrimination. Additional factors, such as temperature, could not be assessed with the current experimental design of common garden conditions. Further, phenotypes are not as informative as direct genetic data because a lack of phenotypic differences does not prove a lack of genetic differentiation.

Despite the large salinity differences experienced throughout the entire lifecycle of the experimental fish, salinity had only a minor impact on the phenotypic variation. However, long-term differences in population-specific salinity environments are not only considered to be associated with genetic distinctness [38–40] but can also lead to ecological speciation [41, 42]. Further, some fish species demonstrate higher growth at intermediate salinities than at a fully marine salinity [43, 44]. However, within this study Atlantic purebreds were growth retarded at salinity 16 compared with salinity 35, whereas the Atlantic/Baltic hybrids grew equally well in both salinities. This indicates the adaptation of Atlantic purebreds to high salinity. On the other hand, hybrids clearly outgrew the wild caught Baltic parental group within two years and had a much larger size-at-age (S6 and S7 Figs). It is possible that the genetic influence of the Atlantic parent contributed to this difference in growth conditions, but the captive F1-hybrids also grew at strikingly different environmental conditions compared with their wild-caught Baltic herring parent. In herring, some of the loci underlying genetic differentiation between Atlantic and Baltic herring show a strong correlation with average salinity conditions experienced by the different populations [18, 19]. However, there are also many other variables, for example in nutrition and temperature [21, 45], which could affect the growth of captive Atlantic/Baltic hybrids and wild Baltic herring. Herring in this study were fed in excess, and the water temperatures were generally higher than in the Baltic Sea (S5 Fig) which most likely promoted higher growth of Atlantic/Baltic hybrids compared to the Baltic parental group.

Besides salinity, temperature is known to have a high impact on phenotypic traits, like growth [46] and number of vertebrae [47]. Temperature has been demonstrated to be the major determinant of otolith growth, and therefore differences in otolith shape are essentially influenced by temperature [48, 49]. Further, environmental factors such as temperature and feeding conditions have impacts on otolith shape differences [14], even in the absence of genetic differences [50, 51]. However, in some cases, temperature does not affect otolith shape [52], or the genotype still has a higher impact than temperature on other phenotypic traits like growth [53, 54] or number of vertebrae [55]. Countergradient variation, i.e., the inverse relationship between environmental conditions and individual growth response [56], can maintain morphological similarity across populations and compensate for the effect of temperature [57].

Common garden experiments are ideally suited to dissect the relative importance of genetic and environmental factors affecting phenotypic traits [58, 59] and can play an essential role in resolving population structure [54]. Further, the exact knowledge of genetic and environmental origin is an enormous benefit, in contrast to natural samples where the origin can only be assumed. This knowledge was used for an individual assignment of otoliths, which achieved highest classification success when assigning the otoliths to their genetic origin (87.2–91.9%): this is comparable to other classification studies among populations using otolith shape [29, 60, 61]. Only a minor part of the otolith shape variation could be explained by salinity differences in the absence of genetic variation. Still, those differences are relevant for resolving the

stock structure of herring, because otolith shape is also used to distinguish between groups that currently cannot be separated using genetics [62].

This high classification success based on otolith shape is practically used to separate populations from mixed fisheries (see, e.g., [60, 63]). This could then be incorporated in fisheries management and assessment to allow more sustainable exploitation of the populations [64]. Also in herring, otolith shape is used to discriminate between mixing stocks with varying spawning seasons [65]. However, there are other examples where spawning components have been identified but are still assessed as a unit stock [49]. Based on our results demonstrating the effect of genetics, we encourage the establishment of otolith shape baselines. These baselines can be further used in combination with machine-learning techniques [66] to assign individuals from mixed fisheries to populations.

In conclusion, our study revealed that the variation in several phenotypic traits, specifically growth, otolith shape and the number of vertebrae, was primarily controlled by genetic factors, while salinity played a minor role. To the best of our knowledge, this is the first time that hybrids of two herring populations were reared under common garden conditions until maturity. Finally, our results show that some of the phenotypic traits included in this study provide information to distinguish genetically differentiated herring. These phenotypic traits can be further used to study population dynamics and connectivity because they are to a large extent genetically determined. However, other factors, which were excluded due to the experimental design, might outplay the genetic response demonstrated within this study.

## Supporting information

### **S1 Appendix. Further details on the otolith shape outline development, as well as the parental group.**

(PDF)

### **S1 Table. Total numbers of analyzed parental fish and otoliths (in brackets) for each sample and parental group.**

(PDF)

### **S2 Table. Number (N) and wavelet coefficients that were removed by adjusting otolith shape for allometric relationships with fish length individually for each sample.**

(PDF)

### **S3 Table. Results from ANOVA like permutation tests comparing the otolith shape among salinities and genetic groups in isolation.**

(PDF)

### **S1 Fig. An otolith shape outline example with lines indicating length and width going through the center of gravity.** PoR = postrostrum, PaR = pararostrum, EMi = excisura minor, EMa = excisura major, R = rostrum, AR = antirostrum.

(TIF)

**S2 Fig. (A) Mean otolith shape outline reconstructed from wavelet coefficients (unitless) and (B) their differences at respective otolith angle.** Data are shown for each sampling date and the four herring groups (H16 = hybrids at salinity 16, P16 = purebreds at salinity 16, H35 = hybrids at salinity 35, P35 = purebreds at salinity 35). The mean and standard deviation (SD) of the wavelet coefficients represent otolith shape outline variation among all groups and the intraclass correlation (ICC, black solid line) represents the variation within each group. \* Mean day post hatching (DPH) for combined samples. Note that the otolith outline for P16 at

279, 618 and 910 DPH is based on  $N = 2$ .  
(TIF)

**S3 Fig. Canonical analysis of principal (CAP) scores of herring otolith shapes indicating differences for salinity separated by the genetic groups (Hybrids and purebreds).** Data given for the samples A) 187 and B) 1098 days post hatching. Black bold letters represent the mean canonical value for each character  $\pm 1*SE$ . Individual fish are represented by frequencies.  
(TIF)

**S4 Fig. Canonical analysis of principal (CAP) scores of herring otolith shapes indicating differences for genetics separated by salinity (16 and 35).** Data given for the samples A) 187 and B) 1098 days post hatching. Black bold letters represent the mean canonical value for each character  $\pm 1*SE$ . Individual fish are represented by frequencies.  
(TIF)

**S5 Fig. Daily water temperatures Atlantic purebred and Atlantic/Baltic hybrids were reared at their entire life in either salinity 16 (light blue) or salinity 35 (dark blue).** Water temperatures of the Atlantic (light red) were measured at stationary hydrographic stations in Ytre Utsira and Sognesjøen. Daily temperatures were combined for both stations and average for depths from 20–120 meters. Water temperatures of the Baltic (dark red) were extracted from <https://sharkweb.smhi.se/> and restricted to the area 16–23° E and 56.5–62° N. Daily temperatures were combined all stations within the area and average for depths from 20–50 meters. Mean $\pm$ SD are given in the legend and lines represent a running mean.  
(TIF)

**S6 Fig. Weight-at-length data of the parental groups.** Individuals used as parents for the F1-generation are marked (Atlantic male, Atlantic female, Baltic male).  
(TIF)

**S7 Fig. Comparison of mean length (left), number of vertebrae (middle), and otolith aspect ratio (otolith length/otolith width, right) among the parental fish.** Mean values and  $1*SE$  are shown.  
(TIF)

**S8 Fig. Average otolith shape outline for parental groups.** The shown outline does not correspond to the actual size and ratio of the original otoliths.  
(TIF)

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## References

1. Via S, Gomulkiewicz R, De Jong G, Scheiner SM, Schlichting CD, Van Tienderen PH. Adaptive phenotypic plasticity: consensus and controversy. *Trends in Ecology & Evolution*. 1995; 10(5):212–7. [https://doi.org/10.1016/S0169-5347\(00\)89061-8](https://doi.org/10.1016/S0169-5347(00)89061-8)
2. Grabowski TB, Young SP, Libungan LA, Steinarsson A, Marteinsdóttir G. Evidence of phenotypic plasticity and local adaptation in metabolic rates between components of the Icelandic cod (*Gadus morhua* L.) stock. *Environmental Biology of Fishes*. 2009; 86(3):361–70. <https://doi.org/10.1007/s10641-009-9534-z>
3. Swain DP, Foote CJ. Stocks and chameleons: the use of phenotypic variation in stock identification. *Fisheries Research*. 1999; 43(1–3):113–28. [https://doi.org/10.1016/S0165-7836\(99\)00069-7](https://doi.org/10.1016/S0165-7836(99)00069-7)
4. Mitchell-Olds T, Willis JH, Goldstein DB. Which evolutionary processes influence natural genetic variation for phenotypic traits? *Nature Reviews Genetics*. 2007; 8(11):845–56. <https://doi.org/10.1038/nrg2207> PMID: 17943192
5. Barrett RDH, Hoekstra HE. Molecular spandrels: tests of adaptation at the genetic level. *Nature Reviews Genetics*. 2011; 12(11):767–80. <https://doi.org/10.1038/nrg3015> PMID: 22005986
6. Iles TD, Sinclair M. Atlantic herring: stock discreteness and abundance. *Science*. 1982; 215(4533):627–33. <https://doi.org/10.1126/science.215.4533.627> PMID: 17842372
7. Gröhsler T, Oeberst R, Schaber M, Larson N, Kornilovs G. Discrimination of western Baltic spring-spawning and central Baltic herring (*Clupea harengus* L.) based on growth vs. natural tag information. *ICES Journal of Marine Science*. 2013; 70(6):1108–17. <https://doi.org/10.1093/icesjms/fst064>
8. Clausen LAW, Stæhr KJ, Rindorf A, Mosegaard H. Effect of spatial differences in growth on distribution of seasonally co-occurring herring *Clupea harengus* stocks. *Journal of Fish Biology*. 2015; 86(1):228–47. <https://doi.org/10.1111/jfb.12571> PMID: 25470075
9. Berg F, Slotte A, Johannessen A, Kvamme C, Clausen LAW, Nash RDM. Comparative biology and population mixing among local, coastal and offshore Atlantic herring (*Clupea harengus*) in the North Sea, Skagerrak, Kattegat and western Baltic. *PLoS ONE*. 2017; 12(10):e0187374. <https://doi.org/10.1371/journal.pone.0187374> PMID: 29084258
10. Hulme TJ. The use of vertebral counts to discriminate between North Sea herring stocks. *ICES Journal of Marine Science*. 1995; 52(5):775–9. <https://doi.org/10.1006/jmsc.1995.0074>
11. Clausen LAW, Bekkevold D, Hatfield EMC, Mosegaard H. Application and validation of otolith microstructure as a stock identification method in mixed Atlantic herring (*Clupea harengus*) stocks in the North Sea and western Baltic. *ICES Journal of Marine Science*. 2007; 64(2):377–85. <https://doi.org/10.1093/icesjms/fsl036>
12. Brophy D, Danilowicz BS. Tracing populations of Atlantic herring (*Clupea harengus* L.) in the Irish and Celtic Seas using otolith microstructure. *ICES Journal of Marine Science*. 2002; 59(6):1305–13. <https://doi.org/10.1006/jmsc.2002.1304>
13. Libungan LA, Slotte A, Husebø Å, Godiksen JA, Pålsson S. Latitudinal gradient in otolith shape among local populations of Atlantic herring (*Clupea harengus* L.) in Norway. *PLoS ONE*. 2015; 10(6):e0130847. <https://doi.org/10.1371/journal.pone.0130847> PMID: 26101885
14. Burke N, Brophy D, King PA. Shape analysis of otolith annuli in Atlantic herring (*Clupea harengus*); a new method for tracking fish populations. *Fisheries Research*. 2008; 91:133–43. <https://doi.org/10.1016/j.fishres.2007.11.013>
15. Bird JL, Eppler DT, Checkley DM Jr. Comparisons of herring otoliths using fourier series shape analysis. *Canadian Journal of Fisheries and Aquatic Sciences*. 1986; 43(6):1228–34. <https://doi.org/10.1139/f86-152>
16. Dahle G, Eriksen AG. Spring and autumn spawners of herring (*Clupea harengus*) in the North Sea, Skagerrak and Kattegat; population genetic analysis. *Fisheries Research*. 1990; 9(2):131–41. [https://doi.org/10.1016/0165-7836\(90\)90060-9](https://doi.org/10.1016/0165-7836(90)90060-9)
17. Ryman N, Lagercrantz U, Andersson L, Chakraborty R, Rosenberg R. Lack of correspondence between genetic and morphologic variability patterns in Atlantic herring (*Clupea harengus*). *Heredity*. 1984; 53(3):687–704. <https://doi.org/10.1038/hdy.1984.127>

18. Lamichhaney S, Martinez Barrio A, Rafati N, Sundström G, Rubin C-J, Gilbert ER, et al. Population-scale sequencing reveals genetic differentiation due to local adaptation in Atlantic herring. *Proceedings of the National Academy of Sciences*. 2012; 109(47):19345–50. <https://doi.org/10.1073/pnas.1216128109> PMID: 23134729
19. Martinez Barrio A, Lamichhaney S, Fan G, Rafati N, Pettersson M, Zhang H, et al. The genetic basis for ecological adaptation of the Atlantic herring revealed by genome sequencing. *eLife*. 2016; 5:e12081. <https://doi.org/10.7554/eLife.12081> PMID: 27138043
20. Blaxter JHS. Herring rearing II—The effect of temperature and other factors on development. *Mar Res*. 1956; 5:1–19.
21. Folkvord A, Johannessen A, Moksness E. Temperature-dependent otolith growth in Norwegian spring-spawning herring (*Clupea harengus* L.) larvae. *Sarsia*. 2004; 89(5):297–310. <https://doi.org/10.1080/00364820410002532>
22. Berg F, Husebø Å, Godiksen JA, Slotte A, Folkvord A. Spawning time of Atlantic herring (*Clupea harengus*) populations within a restricted area reflects their otolith growth at the larval stage. *Fisheries Research*. 2017; 194:68–75. <https://doi.org/10.1016/j.fishres.2017.05.009>
23. Täning ÅV. Experimental study of meristic characters in fishes. *Biological Reviews*. 1952; 27(2):169–93. <https://doi.org/10.1111/j.1469-185X.1952.tb01392.x>
24. Lindsey CC. Factors controlling meristic variation. In: Hoar WS, Randall DJ, editors. *Fish Physiology*. Volume 11, Part B. San Diego: Academic Press; 1988. p. 197–274.
25. Fjelldal PG, Totland GK, Hansen T, Kryvi H, Wang X, Søndergaard JL, et al. Regional changes in vertebral morphology during ontogeny reflect the life history of Atlantic cod (*Gadus morhua* L.). *Journal of Anatomy*. 2013; 222(6):615–24. <https://doi.org/10.1111/joa.12049> PMID: 23587057
26. Bekkevold D, André C, Dahlgren TG, Clausen LAW, Torstensen E, Mosegaard H, et al. Environmental correlates of population differentiation in Atlantic herring. *Evolution*. 2005; 59(12):2656–68. <https://doi.org/10.1111/j.0014-3820.2005.tb00977.x> PMID: 16526512
27. Jørgensen HBH, Hansen MM, Bekkevold D, Ruzzante DE, Loeschcke V. Marine landscapes and population genetic structure of herring (*Clupea harengus* L.) in the Baltic Sea. *Molecular Ecology*. 2005; 14(10):3219–34. <https://doi.org/10.1111/j.1365-294X.2005.02658.x> PMID: 16101787
28. Folkvord A, Høie H, Johannessen A, Solbakken T. Effects of prey concentration, light regime, and parental origin on growth and survival of herring larvae under controlled experimental conditions. *ICES Journal of Marine Science*. 2009; 66(8):1702–9. <https://doi.org/10.1093/icesjms/fsp072>
29. Libungan LA, Óskarsson GJ, Slotte A, Jacobsen JA, Pálsson S. Otolith shape: a population marker for Atlantic herring *Clupea harengus*. *Journal of Fish Biology*. 2015; 86(4):1377–95. <https://doi.org/10.1111/jfb.12647> PMID: 25846860
30. R Core Team. R: A language and environment for statistical computing (version 3.4.1). R Foundation for Statistical Computing, Vienna, Austria (<http://www.R-project.org>). 2017.
31. Libungan LA, Pálsson S. ShapeR: An R package to study otolith shape variation among fish populations. *PLoS ONE*. 2015; 10(3):e0121102. <https://doi.org/10.1371/journal.pone.0121102> PMID: 25803855
32. Leonart J, Salat J, Torres GJ. Removing allometric effects of body size in morphological analysis. *Journal of Theoretical Biology*. 2000; 205:85–93. <https://doi.org/10.1006/jtbi.2000.2043> PMID: 10860702
33. Lv Bertalanffy. Untersuchungen über die Gesetzmäßigkeit des Wachstums. *Wilhelm Roux' Archiv für Entwicklungsmechanik der Organismen*. 1934; 131(4):613–52. <https://doi.org/10.1007/BF00650112> PMID: 28353935
34. Anderson MJ, Willis TJ. Canonical analysis of principal coordinates: a useful method of constrained ordination for ecology. *Ecology*. 2003; 84(2):511–25. [https://doi.org/10.1890/0012-9658\(2003\)084\[0511:CAOPCA\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2003)084[0511:CAOPCA]2.0.CO;2)
35. Geisser S. The Predictive Sample Reuse Method with Applications. *Journal of the American Statistical Association*. 1975; 70(350):320–8. <https://doi.org/10.2307/2285815>
36. Söllner C, Burghammer M, Busch-Nentwich E, Berger J, Schwarz H, Riekel C, et al. Control of crystal size and lattice formation by starmaker in otolith biomineralization. *Science*. 2003; 302(5643):282–6. <https://doi.org/10.1126/science.1088443> PMID: 14551434
37. Cadrin SX, Kerr LA, Mariani S. *Stock Identification Methods*. 2nd ed. San Diego: Academic Press; 2014. 566 p.
38. Teacher AGF, André C, Jonsson PR, Merilä J. Oceanographic connectivity and environmental correlates of genetic structuring in Atlantic herring in the Baltic Sea. *Evolutionary Applications*. 2013; 6(3):549–67. <https://doi.org/10.1111/eva.12042> PMID: 23745145



39. DeFaveri J, Jonsson PR, Merilä J. Heterogeneous genomic differentiation in marine threespine sticklebacks: adaptation along an environmental gradient. *Evolution*. 2013; 67(9):2530–46. <https://doi.org/10.1111/evo.12097> PMID: 24033165
40. Berg PR, Jentoft S, Star B, Ring KH, Knutsen H, Lien S, et al. Adaptation to low salinity promotes genomic divergence in Atlantic cod (*Gadus morhua* L.). *Genome Biology and Evolution*. 2015; 7(6):1644–63. <https://doi.org/10.1093/gbe/evv093> PMID: 25994933
41. Momigliano P, Jokinen H, Fraimout A, Florin A-B, Norkko A, Merilä J. Extraordinarily rapid speciation in a marine fish. *Proceedings of the National Academy of Sciences*. 2017; 114(23):6074–9. <https://doi.org/10.1073/pnas.1615109114> PMID: 28533412
42. Erlandsson J, Östman Ö, Florin A-B, Pekcan-Hekim Z. Spatial structure of body size of European flounder (*Platichthys flesus* L.) in the Baltic Sea. *Fisheries Research*. 2017; 189(Supplement C):1–9. <https://doi.org/10.1016/j.fishres.2017.01.001>
43. Imsland AK, Foss A, Gunnarsson S, Berntssen MHG, FitzGerald R, Bonga SW, et al. The interaction of temperature and salinity on growth and food conversion in juvenile turbot (*Scophthalmus maximus*). *Aquaculture*. 2001; 198(3):353–67. [https://doi.org/10.1016/S0044-8486\(01\)00507-5](https://doi.org/10.1016/S0044-8486(01)00507-5)
44. Bœuf G, Payan P. How should salinity influence fish growth? *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*. 2001; 130(4):411–23. [https://doi.org/10.1016/S1532-0456\(01\)00268-X](https://doi.org/10.1016/S1532-0456(01)00268-X)
45. Werner RG, Blaxter JHS. Growth and survival of larval herring (*Clupea harengus*) in relation to prey density. *Canadian Journal of Fisheries and Aquatic Sciences*. 1980; 37(7):1063–9. <https://doi.org/10.1139/f80-138>
46. Morrongiello JR, Thresher RE. A statistical framework to explore ontogenetic growth variation among individuals and populations: a marine fish example. *Ecological Monographs*. 2015; 85(1):93–115. <https://doi.org/10.1890/13-2355.1>
47. Reimchen TE, Cox KD. Differential temperature preferences of vertebral phenotypes in *Gasterosteus*. *Canadian Journal of Zoology*. 2015; 94(1):1–5. <https://doi.org/10.1139/cjz-2015-0193>
48. Campana SE, Casselman JM. Stock discrimination using otolith shape analysis. *Canadian Journal of Fisheries and Aquatic Sciences*. 1993; 50(5):1062–83. <https://doi.org/10.1139/f93-123>
49. Begg GA, Overholtz WJ, Munroe NJ. The use of internal otolith morphometrics for identification of haddock (*Melanogrammus aeglefinus*) stocks on Georges Bank. *Fishery Bulletin*. 2001; 99:1–14.
50. Cardinale M, Doering-Arjes P, Kastowsky M, Mosegaard H. Effects of sex, stock, and environment on the shape of known-age Atlantic cod (*Gadus morhua*) otoliths. *Canadian Journal of Fisheries and Aquatic Sciences*. 2004; 61(2):158–67. <https://doi.org/10.1139/f03-151>
51. Simoneau M, Casselman JM, Fortin R. Determining the effect of negative allometry (length/height relationship) on variation in otolith shape in lake trout (*Salvelinus namaycush*), using Fourier-series analysis. *Canadian Journal of Zoology*. 2000; 78(9):1597–603. <https://doi.org/10.1139/z00-093>
52. Hüsey K. Otolith shape in juvenile cod (*Gadus morhua*): Ontogenetic and environmental effects. *Journal of Experimental Marine Biology and Ecology*. 2008; 364(1):35–41. <https://doi.org/10.1016/j.jembe.2008.06.026>
53. Jensen LF, Hansen MM, Pertoldi C, Holdensgaard G, Mensberg K-LD, Loeschcke V. Local adaptation in brown trout early life-history traits: implications for climate change adaptability. *Proceedings of the Royal Society B: Biological Sciences*. 2008; 275(1653):2859–68. <https://doi.org/10.1098/rspb.2008.0870> PMID: 18755673
54. Hutchings JA, Swain DP, Rowe S, Eddington JD, Puvanendran V, Brown JA. Genetic variation in life-history reaction norms in a marine fish. *Proceedings of the Royal Society B: Biological Sciences*. 2007; 274(1619):1693–9. <https://doi.org/10.1098/rspb.2007.0263> PMID: 17490948
55. Løken S, Pedersen T. Effect of parent type and temperature on vertebrae number in juvenile cod, *Gadus morhua* (L.), in Northern Norway. *Sarsia*. 1996; 80(4):294–8. <https://doi.org/10.1080/00364827.1996.10413603>
56. Conover DO, Duffy TA, Hice LA. The covariance between genetic and environmental influences across ecological gradients. *Annals of the New York Academy of Sciences*. 2009; 1168(1):100–29. <https://doi.org/10.1111/j.1749-6632.2009.04575.x> PMID: 19566705
57. Marcil J, Swain DP, Hutchings JA. Countergradient variation in body shape between two populations of Atlantic cod (*Gadus morhua*). *Proceedings of the Royal Society B: Biological Sciences*. 2006; 273(1583):217–23. <https://doi.org/10.1098/rspb.2005.3306> PMID: 16555790
58. de Villemereuil P, Gaggiotti OE, Mouterde M, Till-Bottraud I. Common garden experiments in the genomic era: new perspectives and opportunities. *Heredity*. 2016; 116(3):249–54. <https://doi.org/10.1038/hdy.2015.93> PMID: 26486610



59. Conover DO, Baumann H. The role of experiments in understanding fishery-induced evolution. *Evolutionary Applications*. 2009; 2(3):276–90. <https://doi.org/10.1111/j.1752-4571.2009.00079.x> PMID: 25567880
60. Brophy D, Haynes P, Arrizabalaga H, Fraile I, Fromentin JM, Garibaldi F, et al. Otolith shape variation provides a marker of stock origin for north Atlantic bluefin tuna (*Thunnus thynnus*). *Marine and Freshwater Research*. 2015; 67(7):1023–36. <https://doi.org/10.1071/MF15086>
61. Hüseyin K, Mosegaard H, Albertsen CM, Nielsen EE, Hemmer-Hansen J, Eero M. Evaluation of otolith shape as a tool for stock discrimination in marine fishes using Baltic Sea cod as a case study. *Fisheries Research*. 2016; 174:210–8. <https://doi.org/10.1016/j.fishres.2015.10.010>
62. DeVries DA, Grimes CB, Prager MH. Using otolith shape analysis to distinguish eastern Gulf of Mexico and Atlantic Ocean stocks of king mackerel. *Fisheries Research*. 2002; 57(1):51–62. [https://doi.org/10.1016/S0165-7836\(01\)00332-0](https://doi.org/10.1016/S0165-7836(01)00332-0)
63. Shepard KE, Patterson WF, De Vries DA. Trends in Atlantic contribution to mixed-stock king mackerel landings in south Florida inferred from otolith shape analysis. *Marine and Coastal Fisheries*. 2010; 2(1):195–204. <https://doi.org/10.1577/C09-014.1>
64. Schindler DE, Hilborn R, Chasco B, Boatright CP, Quinn TP, Rogers LA, et al. Population diversity and the portfolio effect in an exploited species. *Nature*. 2010; 465(7298):609–12. <https://doi.org/10.1038/nature09060> PMID: 20520713
65. ICES. Report of Herring Assessment Working Group for the South of 62° N (HAWG). ICES CM 2016/ACOM:07. 2016.
66. Chen K-Y, Marschall EA, Sovic MG, Fries AC, Gibbs HL, Ludsin SA. assignPOP: An R package for population assignment using genetic, non-genetic, or integrated data in a machine-learning framework. *Methods in Ecology and Evolution*. 2017:n/a-n/a. <https://doi.org/10.1111/2041-210X.12897>