



REVIEW

The Role of Novel Antibiotics in the Management of Diabetic Foot Infection

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ABSTRACT

Diabetic foot infection is a frequent and potentially life-threatening complication of diabetes mellitus. Antibiotic treatment is the cornerstone of management of diabetic foot infection but the rising prevalence of antibiotic resistance has resulted in increasing rates of treatment failure. In this context, the development of several novel antibiotics might represent a useful tool in severe diabetic foot infections caused by multidrug-resistant bacteria. In the present review, we summarize the safety and efficacy of novel antibiotics in patients with diabetic foot infection. Relevant data are limited, and randomized controlled studies that evaluated the role of these agents in this field are lacking. Until more robust data are available, cefiderocol and dalbavancin, which

have been studied more extensively in patients with bone infections, might be attractive options in carefully selected patients with severe diabetic foot infection.

Keywords: Diabetes mellitus; Diabetic foot infection; Antibiotics; Dalbavancin; Oritavancin; Tedizolid; Ceftaroline; Ceftazidime/avibactam; Meropenem/vaborbactam; Imipenem-cilastatin/relebactam; Ceftolozane/azobactam; Plazomicin; Cefiderocol

Key Summary Points

The selection of the appropriate antibiotic in patients with diabetic foot infection is complicated by the rising prevalence of multidrug-resistant bacteria.

Among novel antibiotics, cefiderocol and dalbavancin might be attractive options in carefully selected patients with severe diabetic foot infection.

Collections of appropriate specimens to identify the causative pathogen(s) is a sine qua non for the successful treatment of diabetic foot infection.

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INTRODUCTION

Diabetic foot infection is a leading cause of non-traumatic lower limb amputation [1]. It is also associated with increased risk for cardiovascular and all-cause mortality in patients with diabetes mellitus [2]. According to current guidelines, the management of severe diabetic foot infection consists of assessment for need for surgical treatment, evaluation for peripheral arterial disease and urgent management if present and administration of empiric broad-spectrum antibiotic treatment targeting both gram-positive and -negative bacteria, including anaerobes [3]. The antibiotic therapy should then be adjusted based on both the clinical response and culture findings [3]. The most common pathogens causing diabetic foot infection are *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Escherichia coli*, whereas almost one fifth of cases are polymicrobial [4, 5]. However, the presence of antibiotic-resistant pathogens is becoming more frequent in diabetic foot infection [6–8]. Indeed, almost 20% of bacteria causing diabetic foot infection are multidrug resistant, particularly *S. aureus* and *P. aeruginosa* [4, 9]. Accordingly, the incidence of failure of antibiotic treatment is also rising [10]. Overuse of broad-spectrum antibiotics is a major cause of the emergence of resistant bacteria [11]. An earlier meta-analysis did not identify clear differences in resolution of diabetic foot infection among conventional carbapenems, antipseudomonal penicillins and cephalosporins [12]. In this context, recently approved antibiotics might have a role in the management of difficult-to-treat diabetic foot infections.

In the present review, we summarize the role of novel antibiotics in the management of these patients. This article is based on previously conducted studies and does not contain any new studies with human participants or animals performed by any of the authors.

CEPHALOSPORINS

Ceftaroline

Ceftaroline is a fifth-generation cephalosporin and is approved for the treatment of acute bacterial skin and skin-structure infections (ABSSSI) (with additional approval for *S. aureus* bacteremia associated with ABSSSI) and community-acquired pneumonia [13]. Ceftaroline exhibits activity against methicillin-resistant *S. aureus* (MRSA) through binding penicillin-binding protein 2a (PBP2a), but also against many gram-positive and -negative pathogens [*Streptococcus pneumoniae*, *Haemophilus influenzae*, *Moraxella catarrhalis*, coagulase-negative *Staphylococcus* (CoNS), *Klebsiella spp.*, *E. coli*], with the exception of non-fermentative (*P. aeruginosa*, *Acinetobacter baumannii*) and multidrug-resistant gram-negative bacilli [extended spectrum β -lactamase (ESBL) or *Klebsiella pneumoniae* carbapenemase (KPC) producing isolates] [13–15]. Ceftaroline is used at a standard intravenous dose of 600 mg every 12 h if creatinine clearance is > 50 ml/min and the dose may be increased to 600 mg every 8 h in the case of complicated skin and soft tissue infections (SSTIs) caused by *S. aureus* with a minimal inhibitory concentration (MIC) 2 or 4 mg/l to ceftaroline [15]. Dosage adjustment is required for altered kidney function with creatinine clearance ≤ 50 ml/min. Ceftaroline is well tolerated, and its most common adverse reactions are gastrointestinal (diarrhea, nausea), headache, pruritus and *Clostridium difficile* infection, even though there are some post-market reports of myelotoxicity related to prolonged (> 7 days) ceftaroline exposure [13–15].

Approval of ceftaroline was based on two clinical studies, CANVAS 1 and 2, which excluded patients with diabetic foot infection and therefore; the use of ceftaroline for treatment of diabetic foot infection is off-label [16]. However, increasing data support the potential role of ceftaroline for diabetic foot infection as monotherapy or in combination with other antibiotics for wider gram-negative and anaerobic coverage. Retrospective data from the Clinical Assessment Program and Teflaro[®]

Utilization Registry (CAPTURE) registry ($n = 201$ patients with diabetic foot infection) showed clinical success in 81% of patients who received ceftaroline for a mean duration of 6.1 days, either as monotherapy (65%) or in combination with other antibiotics [16]. Interestingly, the presence of comorbidities, the type of pathogen and the need for surgical intervention did not affect the outcome [14]. Of note, in the CAPTURE registry, there were higher MRSA isolation rates (28%) than in other studies [16]. In another retrospective cohort of 223 patients with diabetic foot infection, the use of ceftaroline was associated with lower 90-day hospital re-admission and mortality compared with daptomycin [17]. There are also data from the CAPTURE registry indicating clinical success with ceftaroline in gram-positive osteomyelitis in patients with diabetes [18]. In this registry, 150 patients with osteomyelitis (59.3% with diabetes or peripheral arterial disease) due to gram-positive bacteria received ceftaroline, and clinical success rates were very high and similar in the total population and in patients with diabetes (92.7 and 91.0%, respectively) as well as in those who received ceftaroline monotherapy or in combination with other antibiotics (91.0 and 96.0%, respectively) [18]. Moreover, only 1.3% of patients discontinued ceftaroline because of adverse effects [18].

Ceftazidime/Avibactam

The combination of ceftazidime, a third generation cephalosporin, and avibactam, a non- β -lactam inhibitor of β -lactamases, is effective against a spectrum of gram-positive cocci and gram-negative bacilli, including *P. aeruginosa*, KPC, ESBL and oxacillinase (OXA) 48, but not against *Acinetobacter* and Ambler class B metallo- β -lactamases (MBL), while it has poor activity against anaerobic bacteria [19]. It is approved for complicated urinary tract infections (cUTIs) and complicated intra-abdominal infections (cIAI) [20, 21]. Notably, case series suggest that ceftazidime/avibactam is effective in the treatment of bone and joint infection [22]. Moreover, current guidelines recommend the use of ceftazidime monotherapy for

moderately severe or severe infections to cover gram-negative bacteria and *Pseudomonas* [23]. Therefore, ceftazidime/avibactam might have a role in the management of diabetic foot infection caused by carbapenem-resistant *Enterobacteriaceae* (CRE) and carbapenem-resistant *P. aeruginosa* [24]. However, ceftazidime/avibactam is affected by mechanisms of resistance such as efflux pumps and porin channel mutations that are commonly used by multidrug resistant (MDR) *Pseudomonas* and is less effective in vitro than ceftolozane/tazobactam against MDR *Pseudomonas* [20].

Ceftolozane-Tazobactam

Ceftolozane is a fifth-generation cephalosporin and is effective against *P. aeruginosa* [25]. Its combination with tazobactam was first approved in 2015 for the management of complicated cUTIs and cIAIs [20]. Ceftolozane/tazobactam is effective against gram-negative bacilli, including ESBL *Enterobacteriaceae* and MDR *P. aeruginosa*, but has minimal activity against gram-positive bacteria, particularly *Streptococcus* and *Enterococcus* species [20]. Therefore, ceftolozane/tazobactam might have a role in diabetic foot infection caused by MDR gram-negative bacteria [24, 25]. A case of a patient with diabetic foot infection due to MDR *P. aeruginosa* who was successfully managed with ceftolozane-tazobactam in combination with fosfomycin was recently reported [26].

Cefiderocol

Cefiderocol is a novel parenteral siderophore cephalosporin with unique mode of action, mediated through its catechol side chain [27]. By forming a chelated complex with ferric iron, cefiderocol enters the outer lipopolysaccharide membrane of gram-negative bacteria through the bacterial iron transport system. Like other cephalosporins, it binds to penicillin-binding proteins (PBPs) and inhibits bacterial cell wall synthesis [28].

Cefiderocol is resistant to hydrolysis by all classes of β -lactamases, including KPC, OXA and MBL [29, 30]. As a result, its antibacterial

spectrum includes resistant *Enterobacterales*, *P. aeruginosa*, *A. baumannii* and *Stenotrophomonas maltophilia*, while it has poor gram-positive and anaerobic activity [31–33]. The successful evasion of all carbapenem-resistance mechanisms renders ceftiderocol a particularly attractive choice for the treatment of infections due to MBL-producing pathogens and *A. baumannii*, where antimicrobial options are limited.

The adverse effect profile of ceftiderocol is comparatively favorable and similar to other β -lactams. The most frequently reported adverse events are diarrhea, rash, infusion site reactions, pyrexia and elevation in liver function tests [34]. In the CREDIBLE-CR study, rates of drug-related acute kidney injury were lower in the ceftiderocol arm than in the best available therapy arm [35]. These results are appealing because alternative therapies for MBL-producing bacteria and extensively drug-resistant (XDR) *A. baumannii*, such as polymyxins and aminoglycosides, are associated with increased risk for nephrotoxicity.

To date, ceftiderocol has been granted authorization for the treatment of cUTIs, hospital-acquired pneumonia (HAP) and ventilator-associated bacterial pneumonia (VAP) with limited or no alternative treatment options [26]. However, this agent has also been successfully employed for compassionate use in other indications [36, 37]. In the CREDIBLE-CR study, ceftiderocol showed comparable clinical and microbiological efficacy with best available treatment in patients with nosocomial pneumonia, bloodstream infections or cUTIs caused by carbapenem-resistant gram-negative bacteria. However, ceftiderocol carries a Food and Drug Administration label warning, in the light of CREDIBLE-CR results demonstrating an increased all-cause mortality in the ceftiderocol arm compared with the best available therapy arm, with no clear ceftiderocol-related toxicity or other explanation for this finding [26]. Interestingly, ceftiderocol is the first novel antimicrobial agent that is expected to be approved for the treatment of infections caused by carbapenem-resistant *A. baumannii* [32].

Favorable experience with the use of ceftiderocol in the treatment of osteomyelitis has been recently reported, potentially extending its

utility in diabetic foot infections, where the incidence of multidrug-resistant pathogens is rising [6, 7]. Chavda et al. described the successful use of ceftiderocol for the treatment of osteomyelitis due to a pan-resistant *P. aeruginosa*. More specifically, 28 days of ceftiderocol treatment in combination with an oral fluoroquinolone, in combination with surgical debridement, resulted in clinical improvement and avoidance of amputation [38]. Alamarat et al. also reported the case of successful management of osteomyelitis caused by XDR *P. aeruginosa* and ESBL *K. pneumoniae* in a pediatric patient, with prolonged administration of ceftiderocol [39]. Similarly, Dagher et al. efficiently treated a case of extensively drug-resistant *A. baumannii* osteomyelitis with ceftiderocol in combination with surgical debridement, where alternative antimicrobial options did not exist [40].

Notably, iron is known to be utilized in the formation of biofilm, and siderophore production has been shown to be increased in biofilms. Thus, ceftiderocol with its unique exploitation of the bacterial iron transport system might prove to be particularly efficacious against infections characterized by biofilm formation, such as in the case of bone infections [41, 42].

CARBAPENEMS

The combination of meropenem, a wide spectrum carbapenem, and vaborbactam, a boronic β -lactamase inhibitor of class A and C, such as KPC, ESBL and ampicillinase C (AmpC), is approved for the management of UTIs [20, 43]. However, meropenem/vaborbactam is not active against OXA-type carbapenemases and MBL (class B) or MDR *P. aeruginosa* and *Acinetobacter* [20]. This agent was shown to be as effective as piperacillin-tazobactam in the management of UTIs and as effective as the best available therapy in a variety of infections caused (or suspected to be caused) by CRE, including cUTIs, cIAI, HAP and VAP [21]. Therefore, meropenem/vaborbactam might be an option in diabetic foot infection caused by CRE producing KPC but has not been studied in these patients [24].

The combination of imipenem, a wide-spectrum carbapenem, with cilastatin, which delays the degradation of imipenem and prolongs its antibiotic effect, and with relebactam, a novel β -lactamase inhibitor that provides effectiveness against class A and C β -lactamases, but not class B, was shown to be more effective than imipenem/cilastatin in patients with cUTIs and cIAs [20, 21]. This agent is also effective against carbapenemase-resistant *P. aeruginosa* and might be a suitable option for the treatment of diabetic foot infection caused by this pathogen or by KPC-producing CRE, even though relevant studies are lacking [24].

AMINOGLYCOSIDES

Plazomicin is a next-generation aminoglycoside that displays dose-dependent bactericidal activity against *Enterobacteriaceae* including strains that produce ESBL as well as some carbapenemases such as KPC and OXA-48 [44–47]. However, it lacks activity against many New Delhi metallo- β -lactamase (NDM)-harboring strains, its activity toward *P. aeruginosa* and *A. baumannii* is variable and similar to other aminoglycosides, and it is not active against anaerobes [48–50]. Interestingly, plazomicin shows activity against *S. aureus* and CoNS, including methicillin-resistant strains [49], which are frequently encountered in polymicrobial diabetic foot infections. Plazomicin is labeled for the treatment of cUTI, including pyelonephritis, and is reserved for patients with limited or no alternative treatment options [44]. Its main side effects are nephrotoxicity and ototoxicity, similar to other members of this class of antibiotics [44].

In the EPIC trial, plazomicin demonstrated non-inferiority compared to meropenem in composite clinical and microbiological cure at the test-of-cure visit in patients with cUTI and acute pyelonephritis caused by *Enterobacteriaceae*, including MDR pathogens [51]. In the Combating Antibiotic-Resistant *Enterobacteriaceae* (CARE) trial, plazomicin-based were compared with colistin-based combinations in patients with CRE-related bloodstream infections, HAP and VAP. Unfortunately, the study

was terminated prematurely because of difficulties in enrollment, but preliminary findings showed lower all-cause mortality and less serious adverse events, including nephrotoxicity, in the plazomicin arm [52].

To the best of our knowledge, there are no studies evaluating the use of plazomicin in the treatment of SSTIs and bone infections, including diabetic foot infections. However, pharmacokinetic studies demonstrated that aminoglycosides effectively penetrate into bone and joint tissues, despite their hydrophilicity [53]. Moreover, plazomicin was found to be active and superior, compared with the older aminoglycosides, against CRE isolates from SSTIs [54].

LIPOGLYCOPEPTIDES

Dalbavancin

Dalbavancin is a novel long-acting lipoglycopeptide and is active against gram-positive pathogens, including MRSA, coagulase-negative staphylococci, streptococci and vancomycin-susceptible enterococci (VSE), but lacks efficacy against vancomycin-resistant enterococci (VRE) or staphylococci strains [14, 55]. Given the limited availability of direct test susceptibility to dalbavancin, in vitro susceptibility test to vancomycin is acceptable as a surrogate for susceptibility to dalbavancin [56]. Dalbavancin has a long plasma half-life of approximately 14 days mainly due to 93% protein binding, a predominantly non-renal clearance and a good tissue penetration in different sites of infection, with notable activity against bacterial biofilm [55, 57]. Importantly, no dosage adjustment is needed in patients with mild/moderate renal impairment or mild hepatic impairment [55].

Dalbavancin has currently received approval for the treatment of ABSSSI administered as a single intravenous dose 1500 mg or a 1000 mg dose followed by 500 mg 1 week later [55, 57]. In the DISCOVER 1 and 2 double-blind, multicenter, randomized trials ($n = 1301$ patients with ABSSSI, among which 13% had diabetes mellitus), once-weekly intravenous dalbavancin was non-inferior to twice-daily intravenous

vancomycin followed by oral linezolid [58]. Moreover, adverse events were less frequent in the dalbavancin group [58]. In another randomized, double-blind study, a single 1500 mg dose of dalbavancin was non-inferior to a two-dose regimen (1000 mg followed by 500 mg one week later) in patients with ABSSSI [59].

Given its unique pharmacokinetic and pharmacodynamic properties, there is interest in the use of dalbavancin beyond approved indications, including for outpatient parenteral antimicrobial therapy of infections caused by gram-positive bacteria, such as osteomyelitis, which is a frequent cause of “difficult-to-treat” diabetic foot infections. Indeed, in a randomized trial in 80 patients with osteomyelitis, treatment with dalbavancin (1500 mg administered on day 1 and 8) resulted in similar clinical cure rates compared with standard of care (97 and 88%, respectively), with no difference in tolerability between the two groups [60]. However, the duration of hospitalization was almost half in patients treated with dalbavancin (15.8 days vs. 33.3 days in the standard of care arm) [60]. Notably, 6% of patients in the dalbavancin group presented with diabetic foot infection [60]. Several observational studies also showed the efficacy and safety of dalbavancin in the treatment of bone and joint infections [57, 61–67]. In a recent retrospective study in 22 patients with diabetic foot infection who were treated with dalbavancin as a second choice therapy, the cure rate was 87% [68]. A case of successful treatment with dalbavancin of diabetic foot osteomyelitis caused by *Enterococcus faecium* was recently reported [70].

Oritavancin

Oritavancin is another novel synthetic lipoglycopeptide, similar in structure to vancomycin, and is approved for the management of ABSSSI. Oritavancin exhibits bactericidal activity against gram-positive pathogens, including MRSA, and is also active in vitro against VRE and vancomycin-resistant *S. aureus*, unlike dalbavancin [69]. Similar to dalbavancin, oritavancin has a long plasma half-life of 8–10 days, a high protein binding of 90% and slow kidney

clearance, allowing for single-dose treatment [69]. No adjustment is needed in patients with mild or moderate renal insufficiency. However, oritavancin exerts weak inhibitory activity on the cytochrome P450, which can potentially lead to drug–drug interactions, especially with warfarin, and may artificially prolong activated partial thromboplastin time for up to 5 days [14, 69].

Two large, randomized, controlled trials (RCTs), SOLO-1 and -2, demonstrated that a single 1200 mg dose of oritavancin was non-inferior to twice-daily vancomycin for 7–10 days for the treatment of ABSSSI caused by gram-positive pathogens [71, 72]. No significant differences in safety were noted [67, 68]. The most frequently reported adverse effects in the oritavancin group were nausea, headache, vomiting and diarrhea [71, 72]. In addition, transient transaminase elevations were noted in 2% of the patients but without signs of drug-induced liver injury [71, 72]. Of note, in the SOLO-2 study, five cases of osteomyelitis were reported as adverse events in the oritavancin group; therefore, osteomyelitis is listed as a warning for oritavancin, even though these events occurred within the first 9 days after drug initiation, suggesting that it may have been already present [69, 72]. However, retrospective data, case series and real-world data suggested that oritavancin is both safe and effective in the management of osteomyelitis [73–77]. Therefore, the role of this agent in the management of diabetic foot infection is currently unclear.

OXAZOLIDINONES

Tedizolid

Tedizolid is a second-generation oxazolidinone and is active against gram-positive microorganisms, including linezolid-resistant MRSA and VRE, as well as *Nocardia spp.*, *Mycobacterium tuberculosis* and non-tuberculous mycobacteria [78]. Tedizolid has oral bioavailability > 90% and can be administered either intravenously or per os at a dose of 200 mg once daily because of a half-life of 12 h, which is longer than linezolid

[78]. It does not require dose modification in patients with renal or hepatic impairment [78].

Tedizolid is approved for the treatment of ABSSSI, based on evidence from two RCTs, ESTABLISH 1 and 2 [78]. In these studies, tedizolid 200 mg once daily for 6 days was non-inferior to linezolid 600 mg twice daily for 10 days for the treatment of ABSSSI, suspected or documented to be due to a gram-positive pathogen [79, 80]. The 6-day tedizolid course was well tolerated with less frequent gastrointestinal adverse events (nausea, diarrhea, vomiting) than linezolid as well as thrombocytopenia [79, 80]. Retrospective studies suggest that long-term tedizolid treatment is also well tolerated and is less frequently associated with hematological toxicity than linezolid [81, 82]. In a meta-analysis of 15 studies, tedizolid had superior clinical efficacy than vancomycin in ABSSSI caused by MRSA [83].

Since patients with diabetic foot infection were excluded from these two large trials, use of tedizolid in serious diabetic foot infections

remains off-label but promising. According to a recent study that compared tedizolid pharmacokinetics and tissue penetration between diabetic patients with wound infections and healthy volunteers, the penetration into and exposure to tissue were similar in both groups, supporting the need for further evaluation of tedizolid use in diabetic foot infection [84]. Tedizolid appears promising in the treatment of more severe gram-positive infections, such as osteomyelitis, with some clinical trials underway, whereas data from smaller retrospective studies suggest good outcomes in osteoarticular and diabetic foot infections, with a cure rate of 83%, comparable to that of linezolid, and with a better safety profile [82, 85, 86].

CONCLUSION

Several antibiotics have been developed in the last decade, and some of them appear to hold promise as therapeutic options in patients with diabetic foot infection (Table 1). However, there

Table 1 Common pathogens in diabetic foot infection, antibiotics they are resistant to and novel antibiotics that might be useful in their management

Pathogen	Current treatment options with rising rates of resistance	Novel antibiotics that could be used for treatment
<i>Staphylococcus aureus</i>	Methicillin	Ceftaroline
	Vancomycin	Dalbavancin
	Daptomycin	Oritavancin Tedizolid
<i>Pseudomonas aeruginosa</i>	Piperacillin/tazobactam	Ceftazidime/avibactam
	Meropenem	Ceftolozane/tazobactam Cefiderocol Imipenem/cilastatin/relebactam
<i>Enterobacteriaceae</i>	Piperacillin/tazobactam	Ceftazidime/avibactam
	Meropenem	Ceftolozane/tazobactam Cefiderocol Meropenem/vaborbactam Imipenem/cilastatin/relebactam Plazomicin

are very limited data regarding the safety of these novel antibiotics in patients with diabetic foot infection, and randomized controlled studies in this field are lacking. Until more robust data are available, cefiderocol and dalbavancin, which have been studied more extensively in patients with bone infections, might be attractive options in carefully selected patients with severe diabetic foot infection. To avoid the emergence of resistance to these novel antibiotics, their use should be limited to patients with severe diabetic foot infection caused by bacteria susceptible only to these antibiotics and no other treatment options.

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REFERENCES

1. Saeed K, Esposito S, Akram A, Ascione T, Bal AM, Bassetti M, Carnelutti A, Chan M, Davis J, Dryden M, Farhan MFM, Fernando S, Gottlieb T, Gould I, Yildiz M, Lye DC, Pagliano P, Poole S, Pottinger PS, Spera AM, Unal S, Yalcin AN, International Society of Antimicrobial Chemotherapy. Hot topics in diabetic foot infection. *Int J Antimicrob Agents*. 2020;55: 105942. <https://doi.org/10.1016/j.ijantimicag.2020.105942>.
2. Brownrigg JR, Davey J, Holt PJ, Davis WA, Thompson MM, Ray KK, Hinchliffe RJ. The association of ulceration of the foot with cardiovascular and all-cause mortality in patients with diabetes: a meta-analysis. *Diabetologia*. 2012;55:2906–12. <https://doi.org/10.1007/s00125-012-2673-3>.
3. Schaper NC, van Netten JJ, Apelqvist J, Bus SA, Hinchliffe RJ, Lipsky BA, IWGDF Editorial Board. Practical Guidelines on the prevention and management of diabetic foot disease (IWGDF 2019 update). *Diabetes Metab Res Rev*. 2020;36(Suppl 1): e3266. <https://doi.org/10.1002/dmrr.3266>.
4. Du F, Ma J, Gong H, Bista R, Zha P, Ren Y, Gao Y, Chen D, Ran X, Wang C. Microbial infection and antibiotic susceptibility of diabetic foot ulcer in China: literature review. *Front Endocrinol (Lausanne)*. 2022;13: 881659. <https://doi.org/10.3389/fendo.2022.881659>.
5. Ramirez-Acuña JM, Cardenas-Cadena SA, Marquez-Salas PA, Garza-Veloz I, Perez-Favila A, Cid-Baez MA, Flores-Morales V, Martinez-Fierro ML. Diabetic foot ulcers: current advances in antimicrobial therapies and emerging treatments. *Antibiotics (Basel)*. 2019;8:193. <https://doi.org/10.3390/antibiotics8040193>.
6. Yan X, Song JF, Zhang L, Li X. Analysis of risk factors for multidrug-resistant organisms in diabetic foot infection. *BMC Endocr Disord*. 2022;22:46. <https://doi.org/10.1186/s12902-022-00957-0>.

7. Richard JL, Sotto A, Jourdan N, Combescure C, Vannereau D, Rodier M, Lavigne JP, Nîmes University Hospital Working Group on the Diabetic Foot (GP30). Risk factors and healing impact of multidrug-resistant bacteria in diabetic foot ulcers. *Diabetes Metab.* 2008;34:363–9. <https://doi.org/10.1016/j.diabet.2008.02.005>.
8. <https://www.hmpgloballearningnetwork.com/site/podiatry/update-antibiotic-resistance-and-dfus>. Accessed 8/11/2022
9. Xie X, Bao Y, Ni L, Liu D, Niu S, Lin H, Li H, Duan C, Yan L, Huang S, Luo Z. Bacterial profile and antibiotic resistance in patients with diabetic foot ulcer in Guangzhou, Southern China: focus on the differences among different Wagner's Grades, IDSA/IWGDF grades, and ulcer types. *Int J Endocrinol.* 2017;2017:8694903. <https://doi.org/10.1155/2017/8694903>.
10. Tchero H, Kangambega P, Noubou L, Becsangele B, Fluieraru S, Teot L. Antibiotic therapy of diabetic foot infections: a systematic review of randomized controlled trials. *Wound Repair Regen.* 2018;26:381–91. <https://doi.org/10.1111/wrr.12649>.
11. Piaggese A, Apelqvist J, editors. *The diabetic foot syndrome*. Front diabetes, vol. 26. Karger: Basel; 2018. p. 167–83. <https://doi.org/10.1159/000480099>.
12. Selva-Olid A, Solà I, Barajas-Nava LA, Gianneo OD, Bonfill-Cosp X, Lipsky BA. Systemic antibiotics for treating diabetic foot infections. *Cochrane Database Syst Rev.* 2015;2015: CD009061. <https://doi.org/10.1002/14651858.CD009061.pub2>.
13. Giurazza R, Mazza MC, Andini R, Sansone P, Pace MC, Durante-Mangoni E. Emerging treatment options for multi-drug-resistant bacterial infections. *Life (Basel).* 2021;11:519. <https://doi.org/10.3390/life11060519>.
14. Abbas M, Paul M, Huttner A. New and improved? A review of novel antibiotics for gram-positive bacteria. *Clin Microbiol Infect.* 2017;23:697–703. <https://doi.org/10.1016/j.cmi.2017.06.010>.
15. Pani A, Colombo F, Agnelli F, Frantellizzi V, Baratta F, Pastori D, Scaglione F. Off-label use of ceftaroline fosamil: a systematic review. *Int J Antimicrob Agents.* 2019;54:562–71. <https://doi.org/10.1016/j.ijantimicag.2019.06.025>.
16. Lipsky BA, Cannon CM, Ramani A, Jandourek A, Calmaggi A, Friedland HD, Goldstein EJ. Ceftaroline fosamil for treatment of diabetic foot infections: the CAPTURE study experience. *Diabetes Metab Res Rev.* 2015;31:395–401. <https://doi.org/10.1002/dmrr.2624>.
17. Eaves AC, Teng C, Evoy KE, Frei CR. Retrospective cohort evaluating the comparative effectiveness of Ceftaroline and Daptomycin as first-line therapies for inpatient treatment of diabetic foot infection in the United States Veterans Health Care system. *Drugs Real World Outcomes.* 2022. <https://doi.org/10.1007/s40801-022-00319-1>. (Epub ahead of print).
18. Johnson LB, Ramani A, Guervil DJ. Use of Ceftaroline Fosamil in Osteomyelitis: CAPTURE study experience. *BMC Infect Dis.* 2019;19:183. <https://doi.org/10.1186/s12879-019-3791-z>.
19. Lagacé-Wiens P, Walkty A, Karlowsky JA. Cef-tazidime-avibactam: an evidence-based review of its pharmacology and potential use in the treatment of Gram-negative bacterial infections. *Core Evid.* 2014;9:13–25. <https://doi.org/10.2147/CE.S40698>.
20. Matlock A, Garcia JA, Moussavi K, Long B, Liang SY. Advances in novel antibiotics to treat multidrug-resistant gram-negative bacterial infections. *Intern Emerg Med.* 2021;16:2231–41. <https://doi.org/10.1007/s11739-021-02749-1>.
21. Wong D, van Duin D. Novel beta-lactamase inhibitors: unlocking their potential in therapy. *Drugs.* 2017;77:615–28. <https://doi.org/10.1007/s40265-017-0725-1>.
22. Rempenault C, Pagis V, Noussair L, Berbescu S, Duran C, Bouchand F, de Laroche M, Salomon E, Nich C, Bauer T, Rottman M, Davido B, Matt M, Dinh A. Treatment of bone and joint infections by ceftazidime/avibactam and ceftolozane/tazobactam: a cohort study. *J Glob Antimicrob Resist.* 2021;25:282–6. <https://doi.org/10.1016/j.jgar.2021.04.003>.
23. Lipsky BA, Aragón-Sánchez J, Diggle M, Embil J, Kono S, Lavery L, Senneville É, Urbančić-Rovan V, Van Asten S, International Working Group on the Diabetic Foot, Peters EJ. IWGDF guidance on the diagnosis and management of foot infections in persons with diabetes. *Diabetes Metab Res Rev.* 2016;32(Suppl 1):45–74. <https://doi.org/10.1002/dmrr.2699>.
24. Jabbour JF, Sharara SL, Kanj SS. Treatment of multidrug-resistant Gram-negative skin and soft tissue infections. *Curr Opin Infect Dis.* 2020;33:146–54. <https://doi.org/10.1097/QCO.0000000000000635>.
25. Srivastava P, Sivashanmugam K. Combinatorial drug therapy for controlling *Pseudomonas aeruginosa* and its association with chronic condition of diabetic foot ulcer. *Int J Low Extrem Wounds.* 2020;19:7–20. <https://doi.org/10.1177/1534734619873785>.
26. Wong M, Wong D, Malhotra S. Intravenous fosfomycin as salvage therapy for osteomyelitis caused

- by multidrug-resistant *Pseudomonas aeruginosa*. *Am J Health Syst Pharm*. 2021;78:2209–15. <https://doi.org/10.1093/ajhp/zxab294>].
27. Choi JJ, McCarthy MW. Cefiderocol: a novel siderophore cephalosporin. *Expert Opin Investig Drugs*. 2018;27:193–7. <https://doi.org/10.1080/13543784.2018.1426745>.
 28. Ito A, Sato T, Ota M, Takemura M, Nishikawa T, Toba S, Kohira N, Miyagawa S, Ishibashi N, Matsumoto S, Nakamura R, Tsuji M, Yamano Y. In vitro antibacterial properties of Cefiderocol, a novel siderophore cephalosporin, against gram-negative bacteria. *Antimicrob Agents Chemother*. 2017;62:e01454-17. <https://doi.org/10.1128/AAC.01454-17>].
 29. Kohira N, West J, Ito A, Ito-Horiyama T, Nakamura R, Sato T, Rittenhouse S, Tsuji M, Yamano Y. In vitro antimicrobial activity of a siderophore cephalosporin, S-649266, against *Enterobacteriaceae* clinical isolates including carbapenem-resistant strains. *Antimicrob Agents Chemother*. 2015;60:729–34. <https://doi.org/10.1128/AAC.01695-15>].
 30. Ito-Horiyama T, Ishii Y, Ito A, Sato T, Nakamura R, Fukuhara N, Tsuji M, Yamano Y, Yamaguchi K, Tateda K. Stability of novel siderophore cephalosporin S-649266 against clinically relevant carbapenemases. *Antimicrob Agents Chemother*. 2016;60:4384–6. <https://doi.org/10.1128/AAC.03098-15>].
 31. Ito A, Kohira N, Bouchillon SK, West J, Rittenhouse S, Sader HS, Rhomberg PR, Jones RN, Yoshizawa H, Nakamura R, Tsuji M, Yamano Y. In vitro antimicrobial activity of S-649266, a catechol-substituted siderophore cephalosporin, when tested against non-fermenting Gram-negative bacteria. *J Antimicrob Chemother*. 2016;71:670–7. <https://doi.org/10.1093/jac/dkv402>].
 32. Isler B, Doi Y, Bonomo RA, Paterson DL. New treatment options against carbapenem-resistant *Acinetobacter baumannii* infections. *Antimicrob Agents Chemother*. 2018;63: e01110-18. <https://doi.org/10.1128/AAC.01110-18>].
 33. Jacobs MR, Abdelhamed AM, Good CE, Rhoads DD, Hujer KM, Hujer AM, Domitrovic TN, Rudin SD, Richter SS, van Duin D, Kreiswirth BN, Greco C, Fouts DE, Bonomo RA. ARGONAUT-I: Activity of Cefiderocol (S-649266), a siderophore cephalosporin, against gram-negative bacteria, including carbapenem-resistant nonfermenters and *Enterobacteriaceae* with defined extended-spectrum β -lactamases and Carbapenemases. *Antimicrob Agents Chemother*. 2018;63: e01801-18. <https://doi.org/10.1128/AAC.01801-18>].
 34. Saisho Y, Katsube T, White S, Fukase H, Shimada J. Pharmacokinetics, safety, and tolerability of cefiderocol, a novel siderophore cephalosporin for gram-negative bacteria, in healthy subjects. *Antimicrob Agents Chemother*. 2018;62:e02163-17. <https://doi.org/10.1128/AAC.02163-17>.
 35. Bassetti M, Echols R, Matsunaga Y, Ariyasu M, Doi Y, Ferrer R, Lodise TP, Naas T, Niki Y, Paterson DL, Portsmouth S, Torre-Cisneros J, Toyozumi K, Wunderink RG, Nagata TD. Efficacy and safety of cefiderocol or best available therapy for the treatment of serious infections caused by carbapenem-resistant Gram-negative bacteria (CREDIBLE-CR): a randomised, open-label, multicentre, pathogen-focused, descriptive, phase 3 trial. *Lancet Infect Dis*. 2021;21:226–40. [https://doi.org/10.1016/S1473-3099\(20\)30796-9](https://doi.org/10.1016/S1473-3099(20)30796-9)].
 36. Trearichi EM, Quirino A, Scaglione V, Longhini F, Garofalo E, Bruni A, Biamonte E, Lionello R, Serapide F, Mazzitelli M, Marascio N, Matera G, Liberto MC, Navalesi P, Torti C, IMAGES Group. Successful treatment with cefiderocol for compassionate use in a critically ill patient with XDR *Acinetobacter baumannii* and KPC-producing *Klebsiella pneumoniae*: a case report. *J Antimicrob Chemother*. 2019;74:3399–401. <https://doi.org/10.1093/jac/dkz318>].
 37. Edgeworth JD, Merante D, Patel S, Young C, Jones P, Vithlani S, Wyncoll D, Roberts P, Jones A, Den Nagata T, Ariyasu M, Livermore DM, Beale R. Compassionate use of cefiderocol as adjunctive treatment of native aortic valve endocarditis due to extremely drug-resistant *Pseudomonas aeruginosa*. *Clin Infect Dis*. 2019;68:1932–4. <https://doi.org/10.1093/cid/ciy963>].
 38. Chavda A, Gilchrist M, Samarasinghe D. *Education*: a compassionate use of cefiderocol to treat osteomyelitis caused by an XDR *Pseudomonas aeruginosa*. *JAC Antimicrob Resist*. 2021;3(Suppl 1):i18–20. <https://doi.org/10.1093/jacamr/dlab054>].
 39. Alamarat ZI, Babic J, Tran TT, Wootton SH, Dinh AQ, Miller WR, Hanson B, Wanger A, Gary JL, Arias CA, Pérez N. Long-term compassionate use of cefiderocol to treat chronic osteomyelitis caused by extensively drug-resistant *Pseudomonas aeruginosa* and extended-spectrum- β -lactamase-producing *Klebsiella pneumoniae* in a pediatric patient. *Antimicrob Agents Chemother*. 2020;64:e01872-19. <https://doi.org/10.1128/AAC.01872-19>].
 40. Dagher M, Ruffin F, Marshall S, Taracila M, Bonomo RA, Reilly R, Fowler VG Jr, Thaden JT. Case report: successful rescue therapy of extensively drug-resistant *Acinetobacter baumannii* osteomyelitis with cefiderocol. *Open Forum Infect Dis*. 2020;7:ofaa150. <https://doi.org/10.1093/ofid/ofaa150>].

41. Ito A, Nishikawa T, Matsumoto S, Yoshizawa H, Sato T, Nakamura R, Tsuji M, Yamano Y. Side-phore cephalosporin cefiderocol utilizes ferric iron transporter systems for antibacterial activity against *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother*. 2016;60:7396–401. <https://doi.org/10.1128/AAC.01405-16>.
42. Kang D, Kirienko NV. Interdependence between iron acquisition and biofilm formation in *Pseudomonas aeruginosa*. *J Microbiol*. 2018;56:449–57. <https://doi.org/10.1007/s12275-018-8114-3>.
43. Thaden JT, Pogue JM, Kaye KS. Role of newer and re-emerging older agents in the treatment of infections caused by carbapenem-resistant *Enterobacteriaceae*. *Virulence*. 2017;8:403–16. <https://doi.org/10.1080/21505594.2016.1207834>.
44. Eljaaly K, Alharbi A, Alshehri S, Ortwine JK, Pogue JM. Plazomicin: a novel aminoglycoside for the treatment of resistant gram-negative bacterial infections. *Drugs*. 2019;79:243–69. <https://doi.org/10.1007/s40265-019-1054-3>.
45. Ramirez MS, Tolmasky ME. Aminoglycoside modifying enzymes. *Drug Resist Updat*. 2010;13:151–71. <https://doi.org/10.1016/j.drup.2010.08.003>.
46. Aggen JB, Armstrong ES, Goldblum AA, Dozzo P, Linsell MS, Gliedt MJ, Hildebrandt DJ, Feeney LA, Kubo A, Matias RD, Lopez S, Gomez M, Wlasichuk KB, Diokno R, Miller GH, Moser HE. Synthesis and spectrum of the neoglycoside ACHN-490. *Antimicrob Agents Chemother*. 2010;54:4636–42. <https://doi.org/10.1128/AAC.00572-10>.
47. Zhanel GG, Lawson CD, Zelenitsky S, Findlay B, Schweizer F, Adam H, Walkty A, Rubinstein E, Gin AS, Hoban DJ, Lynch JP, Karlowsky JA. Comparison of the next-generation aminoglycoside plazomicin to gentamicin, tobramycin and amikacin. *Expert Rev Anti Infect Ther*. 2012;10:459–73. <https://doi.org/10.1586/eri.12.25>.
48. Shaer KM, Zmarlicka MT, Chahine EB, Piccicacco N, Cho JC. Plazomicin: a next-generation aminoglycoside. *Pharmacotherapy*. 2019;39:77–93. <https://doi.org/10.1002/phar.2203>.
49. Walkty A, Adam H, Baxter M, Denisuk A, Lagacé-Wiens P, Karlowsky JA, Hoban DJ, Zhanel GG. In vitro activity of plazomicin against 5,015 gram-negative and gram-positive clinical isolates obtained from patients in canadian hospitals as part of the CANWARD study, 2011–2012. *Antimicrob Agents Chemother*. 2014;58:2554–63. <https://doi.org/10.1128/AAC.02744-13>.
50. Castanheira M, Deshpande LM, Woosley LN, Serio AW, Krause KM, Flamm RK. Activity of plazomicin compared with other aminoglycosides against isolates from European and adjacent countries, including *Enterobacteriaceae* molecularly characterized for aminoglycoside-modifying enzymes and other resistance mechanisms. *J Antimicrob Chemother*. 2018;73:3346–54. <https://doi.org/10.1093/jac/dky344>.
51. Wagenlehner FME, Cloutier DJ, Komirenko AS, Cebrik DS, Krause KM, Keepers TR, Connolly LE, Miller LG, Friedland I, Dwyer JP, EPIC Study Group. Once-daily plazomicin for complicated urinary tract infections. *N Engl J Med*. 2019;380:729–40. <https://doi.org/10.1056/NEJMoa1801467>.
52. McKinnell JA, Dwyer JP, Talbot GH, Connolly LE, Friedland I, Smith A, Jubb AM, Serio AW, Krause KM, Daikos GL, CARE Study Group. Plazomicin for infections caused by carbapenem-resistant *Enterobacteriaceae*. *N Engl J Med*. 2019;380:791–3. <https://doi.org/10.1056/NEJMc1807634>.
53. Thabit AK, Fatani DF, Bamakhrama MS, Barnawi OA, Basudan LO, Alhejaili SF. Antibiotic penetration into bone and joints: an updated review. *Int J Infect Dis*. 2019;81:128–36. <https://doi.org/10.1016/j.ijid.2019.02.005>.
54. Castanheira M, Deshpande LM, Hubler CM, Mendes RE, Serio AW, Krause KM, Flamm RK. Activity of plazomicin against *Enterobacteriaceae* isolates collected in the United States including isolates carrying aminoglycoside-modifying enzymes detected by whole genome sequencing. *Open Forum Infect Dis*. 2017;4(Suppl 1):S377–1377. <https://doi.org/10.1093/ofid/ofx163.931>.
55. Soriano A, Rossolini GM, Pea F. The role of dalbavancin in the treatment of acute bacterial skin and skin structure infections (ABSSSIs). *Expert Rev Anti Infect Ther*. 2020;18:415–22. <https://doi.org/10.1080/14787210.2020.1746643>.
56. Jones RN, Farrell DJ, Flamm RK, Sader HS, Dunne MW, Mendes RE. Surrogate analysis of vancomycin to predict susceptible categorization of dalbavancin. *Diagn Microbiol Infect Dis*. 2015;82:73–7. <https://doi.org/10.1016/j.diagmicrobio.2015.01.017>.
57. Gatti M, Andreoni M, Pea F, Viale P. Real-world use of dalbavancin in the era of empowerment of outpatient antimicrobial treatment: a careful appraisal beyond approved indications focusing on unmet clinical needs. *Drug Des Devel Ther*. 2021;15:3349–78. <https://doi.org/10.2147/DDDT.S313756>.
58. Boucher HW, Wilcox M, Talbot GH, Puttagunta S, Das AF, Dunne MW. Once-weekly dalbavancin versus daily conventional therapy for skin infection. *N Engl J Med*. 2014;370:2169–79. <https://doi.org/10.1056/NEJMoa1310480>.

59. Dunne MW, Puttagunta S, Giordano P, Krievins D, Zelasky M, Baldassarre J. A randomized clinical trial of single-dose versus weekly dalbavancin for treatment of acute bacterial skin and skin structure infection. *Clin Infect Dis*. 2016;62:545–51. <https://doi.org/10.1093/cid/civ982>.
60. Rappo U, Puttagunta S, Shevchenko V, Shevchenko A, Jandourek A, Gonzalez PL, Suen A, Mas Casullo V, Melnick D, Miceli R, Kovacevic M, De Bock G, Dunne MW. Dalbavancin for the treatment of osteomyelitis in adult patients: a randomized clinical trial of efficacy and safety. *Open Forum Infect Dis*. 2018;6: ofy331. <https://doi.org/10.1093/ofid/ofy331>.
61. Bouza E, Valerio M, Soriano A, Morata L, Carus EG, Rodríguez-González C, Hidalgo-Tenorio MC, Plata A, Muñoz P, Vena A, DALBUSE Study Group (Dalbavancina: Estudio de suusoclinicoenEspaña). Dalbavancin in the treatment of different gram-positive infections: a real-life experience. *Int J Antimicrob Agents*. 2018;51:571–7. <https://doi.org/10.1016/j.ijantimicag.2017.11.008>.
62. Bai F, Aldieri C, Cattelan A, Raumer F, Di Meco E, Moioli MC, Tordato F, Morelli P, Borghi F, Rizzi M, Van Hauwermeiren E, Castelli F, Migliorino G, Menzaghi B, Rizzardini G, Saracino A, Cascio A, Puoti M, d'Arminio Monforte A, Marchetti G. Efficacy and safety of dalbavancin in the treatment of acute bacterial skin and skin structure infections (ABSSSIs) and other infections in a real-life setting: data from an Italian observational multicentric study (DALBITA study). *Expert Rev Anti Infect Ther*. 2020;18:1271–9. <https://doi.org/10.1080/14787210.2020.1798227>.
63. Núñez-Núñez M, Casas-Hidalgo I, García-Fumero R, Vallejo-Rodríguez I, Anguita-Santos F, Hernández-Quero J, Cabeza-Barrera J, Ruiz-Sancho A. Dalbavancin is a novel antimicrobial against Gram-positive pathogens: clinical experience beyond labelled indications. *Eur J Hosp Pharm*. 2020;27:310–2. <https://doi.org/10.1136/ejhpharm-2018-001711>.
64. Wunsch S, Krause R, Valentin T, Prattes J, Janata O, Lenger A, Bellmann-Weiler R, Weiss G, Zollner-Schwetz I. Multicenter clinical experience of real life Dalbavancin use in gram-positive infections. *Int J Infect Dis*. 2019;81:210–4. <https://doi.org/10.1016/j.ijid.2019.02.013>.
65. Tobudic S, Forstner C, Burgmann H, Lagler H, Steininger C, Traby L, Vossen MG, Winkler S, Thalhammer F. Real-world experience with dalbavancin therapy in gram-positive skin and soft tissue infection, bone and joint infection. *Infection*. 2019;47:1013–20. <https://doi.org/10.1007/s15010-019-01354-x>.
66. Veve MP, Patel N, Smith ZA, Yeager SD, Wright LR, Shorman MA. Comparison of dalbavancin to standard-of-care for outpatient treatment of invasive Gram-positive infections. *Int J Antimicrob Agents*. 2020;56: 106210. <https://doi.org/10.1016/j.ijantimicag.2020.106210>.
67. Wang Y, Wang J, Wang R, Li Y, Cai Y. Efficacy and safety of dalbavancin in the treatment of Gram-positive bacterial infections. *J Glob Antimicrob Resist*. 2021;24:72–80. <https://doi.org/10.1016/j.jgar.2020.11.018>.
68. Navarro-Jiménez G, Fuentes-Santos C, Moreno-Núñez L, Alfayate-García J, Campelo-Gutierrez C, Sanz-Márquez S, Pérez-Fernández E, Velasco-Arribas M, Hervás-Gómez R, Martín-Segarra O, Losa-García JE. Experience in the use of dalbavancin in diabetic foot infection. *Enferm Infecc Microbiol Clin (Engl Ed)*. 2022;40:296–301. <https://doi.org/10.1016/j.eimce.2022.03.001>.
69. Roberts KD, Sulaiman RM, Rybak MJ. Dalbavancin and Oritavancin: an innovative approach to the treatment of gram-positive infections. *Pharmacotherapy*. 2015;35:935–48. <https://doi.org/10.1002/phar.1641>.
70. Loupa CV, Lykoudi E, Meimeti E, Moisoglou I, Voyatzoglou ED, Kalantzi S, Konsta E. Successful treatment of diabetic foot osteomyelitis with dalbavancin. *Med Arch*. 2020;74:243–5. <https://doi.org/10.5455/medarh.2020.74.243-245>.
71. Corey GR, Kabler H, Mehra P, Gupta S, Overcash JS, Porwal A, Giordano P, Lucasti C, Perez A, Good S, Jiang H, Moeck G, O'Riordan W, SOLO I Investigators. Single-dose oritavancin in the treatment of acute bacterial skin infections. *N Engl J Med*. 2014;370:2180–90. <https://doi.org/10.1056/NEJMoa1310422>.
72. Corey GR, Good S, Jiang H, Moeck G, Wikler M, Green S, Manos P, Keech R, Singh R, Heller B, Bubnova N, O'Riordan W, SOLO II Investigators. Single-dose oritavancin versus 7–10 days of vancomycin in the treatment of gram-positive acute bacterial skin and skin structure infections: the SOLO II noninferiority study. *Clin Infect Dis*. 2015;60:254–62. <https://doi.org/10.1093/cid/ciu778>.
73. Scoble PJ, Reilly J, Tillotson GS. Real-world use of oritavancin for the treatment of osteomyelitis. *Drugs Real World Outcomes*. 2020;7(Suppl 1): 46–54. <https://doi.org/10.1007/s40801-020-00194-8>.
74. Chastain DB, Davis A. Treatment of chronic osteomyelitis with multidose oritavancin: a case series and literature review. *Int J Antimicrob*

- Agents. 2019;53:429–34. <https://doi.org/10.1016/j.ijantimicag.2018.11.023>].
75. Stewart CL, Turner MS, Frens JJ, Snider CB, Smith JR. Real-world experience with oritavancin therapy in invasive gram-positive infections. *Infect Dis Ther.* 2017;6:277–89. <https://doi.org/10.1007/s40121-017-0156-z>].
 76. Van Hise NW, Chundi V, Didwania V, Anderson M, McKinsey D, Roig I, Sharma A, Petrak RM. Treatment of acute osteomyelitis with once-weekly oritavancin: a two-year, multicenter retrospective study. *Drugs Real World Outcomes.* 2020;7(Suppl 1):41–5. <https://doi.org/10.1007/s40801-020-00195-7>].
 77. Redell M, Sierra-Hoffman M, Assi M, Bochan M, Chansolme D, Gandhi A, Sheridan K, Soosaipillai I, Walsh T, Massey J. The CHROME study, a real-world experience of single- and multiple-dose oritavancin for treatment of gram-positive infections. *Open Forum Infect Dis.* 2019;6: ofz479. <https://doi.org/10.1093/ofid/ofz479>.
 78. Bouza E, Muñoz P, Burillo A. The role of tedizolid in skin and soft tissue infections. *Curr Opin Infect Dis.* 2018;31:131–40. <https://doi.org/10.1097/QCO.000000000000439>].
 79. Prokocimer P, De Anda C, Fang E, Mehra P, Das A. Tedizolid phosphate vs linezolid for treatment of acute bacterial skin and skin structure infections: the ESTABLISH-1 randomized trial. *JAMA.* 2013;309:559–69. <https://doi.org/10.1001/jama.2013.241>.
 80. Moran GJ, Fang E, Corey GR, Das AF, De Anda C, Prokocimer P. Tedizolid for 6 days versus linezolid for 10 days for acute bacterial skin and skin-structure infections (ESTABLISH-2): a randomised, double-blind, phase 3, non-inferiority trial. *Lancet Infect Dis.* 2014;14:696–705. [https://doi.org/10.1016/S1473-3099\(14\)70737-6](https://doi.org/10.1016/S1473-3099(14)70737-6).
 81. Mensa Vendrell M, TasiásPitarch M, SalavertLletí M, Calabuig Muñoz E, Morata Ruiz L, Castells Lao G, López Suñé E, Mensa Pueyo J, OltraSempere MR, Pedro-BotetMontoya ML, Isernia V, Reynaga Sosa EA, Moreno Nuñez L, PasquauLiaño J, SequeraArquelladas S, Yuste Ara JR, Soriano Viladomiu A. Safety and tolerability of more than six days of tedizolid treatment. *Antimicrob Agents Chemother.* 2020;64: e00356-20. <https://doi.org/10.1128/AAC.00356-20>].
 82. Benavent E, Morata L, Escriva-Vidal F, Reynaga EA, Soldevila L, Albiach L, Pedro-Botet ML, Padullés A, Soriano A, Murillo O. Long-term use of tedizolid in osteoarticular infections: benefits among oxazolidinone drugs. *Antibiotics (Basel).* 2021;10:53. <https://doi.org/10.3390/antibiotics10010053>].
 83. McCool R, Gould IM, Eales J, Barata T, Arber M, Fleetwood K, Glanville J, Kauf TL. Systematic review and network meta-analysis of tedizolid for the treatment of acute bacterial skin and skin structure infections caused by MRSA. *BMC Infect Dis.* 2017;17:39. <https://doi.org/10.1186/s12879-016-2100-3>].
 84. Stainton SM, Monogue ML, Baummer-Carr A, Shepard AK, Nugent JF, Kuti JL, Nicolau DP. Comparative assessment of tedizolid pharmacokinetics and tissue penetration between diabetic patients with wound infections and healthy volunteers via in vivo microdialysis. *Antimicrob Agents Chemother.* 2017;62: e01880-17. <https://doi.org/10.1128/AAC.01880-17>].
 85. SalavertLletí M, García-Bustos V, Morata Ruiz L, Cabañero-Navalon MD. Tedizolid: new data and experiences for clinical practice. *Rev Esp Quimioter.* 2021;34(Suppl 1):22–5. <https://doi.org/10.37201/req/s01.06.2021>].
 86. York JA, Adams K, Cullen L, Delahay J, Ivan M, Lillie PJ, MacLachlan L, Barlow G. Tedizolid: a service evaluation in a large UK teaching hospital. *Eur J Clin Microbiol Infect Dis.* 2021;40:397–405. <https://doi.org/10.1007/s10096-020-04015-2>].