

# Genetic causes of primary immunodeficiency in the Jordanian population

LOIY OBEIDAT<sup>1,2</sup>, MARWAN ABU-HALAWEH<sup>3</sup>, RAED ALZYLOUD<sup>4</sup>,  
EMAN ALBSOUL<sup>5</sup> and APOSTOLOS ZARAVINOS<sup>1,2</sup>

<sup>1</sup>Department of Life Sciences, School of Sciences, European University Cyprus, Nicosia 2404, Cyprus; <sup>2</sup>Basic and Translational Research Center (BTCRC), Cancer Genetics, Genomics and Systems Biology Laboratory, Nicosia 2404, Cyprus; <sup>3</sup>Primary Immunodeficiency Department, Queen Rania Al Abdullah Hospital for Children, King Hussein Medical Center, Queen Rania Al Abdullah Hospital for Children, Amman, Jordan; <sup>4</sup>Section of Immunology, Allergy and Rheumatology, Queen Rania Children's Hospital, Queen Rania Al Abdullah Hospital For Children, Amman 11855, Jordan; <sup>5</sup>Genetics Laboratory, Philadelphia University, Amman 19392, Jordan

Received April 23, 2024; Accepted July 10, 2024

DOI: 10.3892/br.2024.1848

**Abstract.** Inborn errors of immunity (IEI) represents a heterogeneous collection of >480 immune system anomalies, leading to severe infections, autoimmune disorders and malignancies. While these conditions are rare globally, their prevalence is notably higher in the Jordanian population, attributed to elevated rates of consanguinity. The intricate nature of IEI has driven the adoption of genomic technologies for the identification of associated genetic defects. In the present study, whole-exome sequencing was performed on nine Jordanian IEI patient samples, confirming germline single-nucleotide variations (SNVs) in 14 genes through Sanger sequencing. Of note, signal transducer and activator of transcription 1 (*STAT1*), elastase, neutrophil expressed (*ELANE*) and interferon induced with helicase c domain 1 (*IFIH1*) harbored mutations that were previously unreported in the Jordanian IEI population. In addition, mutations in capping protein regulator and myosin 1 linker 2 (c.3683C>T), TNF $\alpha$ -induced protein 3-interacting protein 1 (*TNIP1*) (c.460C>G) and *STAT1* (c.1061T>C) were confirmed, marking their association with Jordanian IEI. For robustness, the genomic databases Ensemble, Genome AD and ClinVar were used to confirm the SNVs' associations with IEI. Kyoto Encyclopedia of Genes and Genomes pathway analysis also showed involvement of the IL-17 signaling pathway (including IL-17 receptor A), T-helper type 17 cell differentiation (including *STAT1*), the JAK-STAT signaling pathway (including *STAT2* and tyrosine kinase 2),

neutrophil extracellular trap formation (including *ELANE*), cocaine addiction [G protein signaling modulator 1 (*GPSM1*)] and cytokine-cytokine receptor interaction (IL-17 receptor C). In summary, exome sequencing identified a likely causative genetic defect in *ELANE* (PID-28), *STAT1* (PID-30) and *IFIH1* (PID-33). The present findings reveal the association of novel *STAT1*, *ELANE* mutations with the clinical phenotype of the patients, as well as known mutations in *NLRP12*, *GPSM1* and *TNIP1*, in addition to novel *ELANE*, *STAT1* and *IFIH1* mutations associated in the context of Jordanian IEI.

## Introduction

Primary immunodeficiency disorders (PID), also termed as inborn errors of immunity (IEI), encompass a heterogeneous array of conditions characterized by compromised immune system function involving essential components, such as neutrophils, macrophages, dendritic cells (DC), complement proteins and natural killer cells, as well as T and B lymphocytes (1).

To date, a variety of >400 disorders have been listed under the term IEI and many of them have been described only in a small number of patients and families (2-4). While PIDs are primarily hereditary, acquired deficiencies have also been identified (5). It is crucial to differentiate IEI from secondary immunodeficiencies, which arise more frequently due to various factors such as infections, medications, metabolic disease and environmental conditions. By contrast, IEI stem from genetic anomalies resulting in impaired immune cell functionality (5).

Immunoglobulin A deficiency is the most common IEI, estimated to affect 1 in 300 to 1 in 500 individuals, whereas other IEI are rarer, with an approximate prevalence of 1 in 1,200 live births (6). The clinical indications of IEI are highly diverse and patients have a higher infection susceptibility. IEI may be diagnosed as a normal infection (ears, lungs and the sinuses), although they may be undetected if misdiagnosed (7). The diagnosis of such disorders requires specialized testing,

---

*Correspondence to:* Professor Apostolos Zaravinos, Department of Life Sciences, School of Sciences, European University Cyprus, 6 Diogenous Str., Egkomi, Nicosia 2404, Cyprus  
E-mail: a.zaravinos@euc.ac.cy

*Key words:* primary immunodeficiencies, whole exome sequencing, genetic diagnosis, primary immunodeficiency disorders

accurate and timely diagnosis, as well as a relatively high suspicion (8). An experienced clinical immunologist in the evaluation of immunodeficiencies should be consulted for an accurate and early diagnosis, as well as precise treatment, which could prevent the rate of morbidity related to such disorders and it should improve the patient outcomes (9). Furthermore, Amaya-Urbe *et al* (10) found that there is a high prevalence of autoimmune disease (AD) among cases of IEI, indicating a link between the occurrence of both conditions. The authors found that a defect in autoimmune regulator and T-regulatory cells of the central and peripheral tolerance is associated with AD development. In addition, the study found that chronic and recurrent infection through molecular mimicry, as well as activation of super antigens, are vital for the progression of AD, which are characteristics of IEI (10).

The basic classification of IEI depends upon the type of immune system defects; they are broadly classified as adaptive or innate immunity disorders, respectively (1). The advent of whole-exome sequencing (WES) has significantly contributed to identifying novel causative genes in IEI cases (11). Particularly in regions with elevated rates of parental consanguinity, such as Jordan, where ~40% of marriages are consanguineous, WES has proven effective in identifying disease-associated genes and variants. The prevalence of consanguinity before 1980 in Amman was ~30%, leading to a heightened predisposition to IEI (according to [www.consang.net](http://www.consang.net)). However, identifying new mutations requires robust evidence of their clinical relevance, necessitating functional genomic studies. These investigations delve into the impact of novel mutations on gene transcription, translation, transcriptional regulation and protein structure and function, ultimately shedding light on the precise effects of these mutations on proteins. Thus, genetic testing is a must when a family history of IEI has been detected in one of the family members (12). The present study, for the first time, investigated a cohort of Jordanian patients in whom signal transducer and activator of transcription 1 (*STAT1*), elastase, neutrophil expressed (*ELANE*) and interferon induced with helicase C domain1 (*IFIH1*) mutations were identified, and a new association with variants in NLR family, pyrin domain-containing 12 (*NLRP12*), G-protein-signaling modulator 1 (*GPSM1*) and TNF $\alpha$ -induced protein 3-interacting protein 1 (*TNIP1*) was discovered.

## Patients and methods

**Patient samples.** Blood samples of members from 9 families (total of 37 samples) were collected at the Primary Immunodeficiency Department at Queen Rania Al Abdullah Hospital for Children, King Hussein Medical Center (Amman, Jordan). All patients with IEI and their family members, for whom immunologic or genetic testing was carried out, signed a consent form. Genomic DNA was extracted using the DNeasy blood & tissue kit (Qiagen, Inc.) and stored at -80°C. The quality and quantity of the DNA samples was measured using a NanoPhotometer® N50 spectrophotometer (iMPLEN).

Each patient had been noted to have recurrent infection, fever, sinusitis, lymphadenopathy and a family history of IEI or autoimmunity, and had undergone extensive lab tests, including complete blood count (CBC) and flow cytometry to

determine any immune malfunction. Patients with abnormal immune results, such as agammaglobulinemia, low lymphocyte count or high immunoglobulin count, were considered patients with IEI. The samples used in the present study were chosen at random with no set criteria or procedure. The probands and their families presented at the Department of Immunology at Queen Rania Children's Hospital in September 2021. By October 2021, the blood samples had been drawn at the Department of Immunology and the consent forms had been signed.

**Patients and pedigrees.** A total of 37 IEI samples from 9 Jordanian families (patients and their father, mother and/or siblings), who had poorer response to conventional treatments and were in need of an alternative therapeutic strategy were chosen randomly for this study. A total of four patients were from consanguineous families (patient ID-26, ID-32, ID-34 and ID-35], while the remaining probands had unrelated parents. In addition, three patients had a positive family history of PID (ID-26, ID-29 and ID-32), while one (ID-29) had a strong family history of familial Mediterranean fever (FMF) and three others (ID-28, ID-30 and ID-33) had a negative family history. The father of proband PID-31 had chronic kidney disease and vision problems.

Proband ID-26 was a 2.5-year-old male who, at the age of 14 months, had developed lymphoproliferation, autoimmune cytopenia and a decreased level of consciousness. The patient had negative culture yields, Epstein-Barr virus viremia and was treated with high-dose intravenous immunoglobulin, rapamycin. The laboratory results showed that the patient had hypogammaglobulinemia (after rituximab). In addition, the proband had a negative next-generation sequencing (NGS) result for the immunodeficiency panel and an elevated double-negative T-cell level.

Patient ID-28 was a one-year-old male with recurrent admissions with fever and severe neutropenia. Once admitted with a submental abscess, the patient required surgical drainage. The patient suffered from severe neutropenia. The proband had no indication of maturation arrest in a bone marrow study performed at the age of 3 months. The patient also had normal immunoglobulin levels and a normal lymphocyte subset.

Patient ID-29 was a 12-year-old female diagnosed with refractory Juvenile dermatomyositis since the age of 6 years. The patient did not have any history of breakthrough infections. The proband had hypergammaglobulinemia and steroid-induced lymphopenia, and also had a heterozygous M694V mutation in the familial Mediterranean fever gene.

Patient ID-30 was a 7-year-old male with recurrent admission with cough and fever since the early infancy period. The patient suffered from chronic mycosis of the skin, developmental delay and hypothyroidism. The proband's immunoglobulin levels were normal, as well as the dihydrorhodamine test and CBC.

Patient ID-31 was a 3-year-old female diagnosed with biliary stasis at the age of 2 months. The patient had a history of recurrent fever since the early infantile period and a negative culture yield. Furthermore, the patient had a history of short liver synovitis, microscopic hematuria and prompt response to steroids. The laboratory analysis results showed

that the patient suffered from neutrophilic leukocytosis. Inflammatory markers were also high, as well as the urine levels of mevalonate kinase. The patient's immunoglobulin levels were normal.

Patient ID-32 was a 6-month-old male patient with a history of chronic cough, failure to thrive and chronic diarrhea. Lab tests showed decreased naïve T cells, no lymphopenia, a reduced number of switched memory B cells and normal immunoglobulin levels. The family history indicated positivity for similar illness (the proband's brother) and the parents are consanguineous.

Patient ID-33 was a 4-year-old female diagnosed with secondary alveolar proteinosis at the age 8 months, improved on antimicrobials and systemic steroids also, and was recurrently admitted with chest infections. The patient had a negative culture yield, developmental delay, as well as epilepsy disorder at the age of 2 years.

Patient ID-34 was a 14-year-old male diagnosed with unclassified arthritis at the age of 11 years, managed by anti-TNF. The patient had been diagnosed with esophageal candidiasis and at the age of 14 years, he had a prolonged fever. The initial lab results showed normal immunodeficiency work-up and the patient also had a lymph node biopsy with atypical lymphoblasts. In addition, the patient had a positive family history of malignancy.

Patient ID-35 was a 9-month-old male diagnosed with complicated meningitis at the age of 7 months. In addition, the patient has a polio vaccine-associated paralysis and the candidate's diagnosis was based on major histocompatibility complex II deficiency. The patient has agammaglobulinemia and absent CD4 lymphocytes. The proband also had a positive family history of IEI with an affected brother. A summary of the cases is provided in Table I.

Pedigree charts were constructed using Progeny Genetics (<https://pedigree.progenygenetics.com/>), to reveal the occurrence and appearance of the PID phenotypes of particular patients and their ancestors from one generation to the next.

**WES.** Genomic DNA from each sample (3 µg) was randomly fragmented using the Covaris Acoustic System with the following settings: Frequency, 500 kHz; temperature, 4°C; and duration, 5 min. The fragmented DNA was then linked to adapters. The DNA was purified using Agencourt AMPure SPRI beads (Beckman Coulter, Inc.) according to the manufacturer's protocol, extracted, and amplified by ligation-mediated PCR. For the ligation-mediated PCR, KAPA HiFi HotStart DNA Polymerase (KAPA Biosystems) was used. The thermocycling conditions were as follows: Initial denaturation at 98°C for 2 min; 15 cycles of denaturation at 98°C for 20 sec, annealing at 60°C for 30 sec and elongation at 72°C for 1 min; and final extension at 72°C for 5 min. It was then hybridized to Agilent's SureSelect Human All Exon V8 (Agilent Technologies, Inc.) for enrichment and the captured PCR products were subjected to quantitative PCR to estimate the magnitude of enrichment. Each captured library was subsequently sequenced using an Illumina NovaSeq sequencer (Illumina, Inc.). The sequenced reads were aligned to the human genome reference [University of California Santa Cruz (UCSC) hg38; <https://genome.ucsc.edu/>]

using SOAP aligner (soap2.21) and duplicated reads were filtered out. SOAPsnp (v.1.03) was used to assemble the consensus sequence and call genotypes in target regions, and integrative genomic viewer (IGV) 2.17.4 (<https://igv.org>) was utilized to visually inspect all reported variants. Patients born in consanguineous families were expected to show an autosomal recessive (AR) inheritance pattern. Therefore, the synonymous mutations were filtered out and the variants with a >0.5% frequency in dbSNP143 (<https://vatlab.github.io/vat-docs/applications/annotation/variants/db SNP/>), 1000 genomes ([internationalgenome.org](http://internationalgenome.org)) and HapMap ([coriell.org](http://coriell.org)) were excluded. The new single nucleotide polymorphisms (SNPs) detected were confirmed using Sanger sequencing. For this purpose, exon-specific PCR primers were designed using Primer3 (<https://primer3.ut.ee/>), the annealing location of which was tested using *in-silico* PCR (<https://genome.ucsc.edu/cgi-bin/hgPcr>). Subsequently, the genomic DNA was amplified using FastGene Taq DNA polymerase (5 U/µl; 0.5 µl), 10 µM forward primer (1 µl), 10 µM reverse primer (1 µl), 10 µM dNTP mix (1 µl), 10X Buffer A (15 mM) (5 µl), 100 ng of genomic DNA, with the mixture topped up to 50 µl with nuclease-free water, in a BioRad (T100) thermal cycler (Bio-Rad Laboratories, Inc.) The thermocycling program was as follows: Initial denaturation at 95°C for 5 min, 35 cycles of 95°C for 15 sec, 60°C for 30 sec and 72°C for 1 min, followed by a final extension at 72°C for 5 min. The PCR products were electrophoresed on a 2% agarose gel and purified with a QIAquick gel extraction kit (Qiagen, Inc.). They were then Sanger sequenced at Macrogen Europe BV and the sequencing electrophoregrams were analyzed using Lasergene17 (DNASStar, Inc.).

**Pathway analysis.** The genes harboring non-silent mutations were assigned to gene sets using the Molecular Signatures Database Human Collection [MSigDB 2023.1.Hs (<https://www.gsea-msigdb.org/gsea/msigdb/collections.jsp>)], where the 'canonical pathways' were explored. Only 1 nonsynonymous mutation per gene was accounted for in each IEI sample.

**Databases.** ClinVar (<https://www.ncbi.nlm.nih.gov/clinvar/>) was used to confirm the SNVs' associations with IEI. The Online Mendelian Inheritance in Man (OMIM) database (<https://www.omim.org/>) was used to acquire information on the genes of interest and their relationship with the phenotype.

## Results

**WES results.** WES was performed and the candidate genes were selected according to the patients' symptoms and diagnosis. Nearly 200 genes were chosen from a PID gene panel and visualized for each patient in order to cover all the possible disease-causing genes. Only deleterious and damaging SNPs were considered for downstream analysis, while benign and likely benign SNPs were excluded.

The detected SNPs were in relevant genes to each patient's symptoms. These were heterozygous in the following genes: Interleukin 17 receptor A (*IL17RA*) (c.1345C>G; p.Arg449Gly), *STAT1* (c.1061T>C; p.Leu354Pro),

Table I. Clinical history, immunological laboratory findings and the mutated genes found for each patient with inborn errors of immunity in the cohort.

Patient	Patient's history	Immunological Lab findings	Mutated genes identified
ID-26	<ul style="list-style-type: none"> <li>• 2.5-year-old male patient</li> <li>• Developed lymphoproliferation, autoimmune cytopenia and decreased level of consciousness at the age of 14 months</li> <li>• Negative culture yields, EBV viremia</li> <li>• Managed by high dose IVIG, rapamycin</li> <li>• One-year-old male patient</li> <li>• Recurrent admissions with fever, severe neutropenia</li> <li>• Once admitted with submental abscess, required surgical drainage</li> </ul>	<ul style="list-style-type: none"> <li>• Hypogammaglobulinemia (after rituximab)</li> <li>• Negative NGS for immunodeficiency panel</li> <li>• High DNT</li> </ul>	<ul style="list-style-type: none"> <li>STAT2</li> <li>PIK3R4 (novel)</li> </ul>
ID-28	<ul style="list-style-type: none"> <li>• 12-year-old female patient</li> <li>• Diagnosis of refractory juvenile dermatomyositis since the age of 6 years</li> <li>• No history of breakthrough infections</li> </ul>	<ul style="list-style-type: none"> <li>• Severe neutropenia</li> <li>• No maturation arrest on bone marrow study performed at the age of 3 months</li> <li>• Normal IG level, normal lymphocyte subset</li> <li>• Hypergammaglobulinemia</li> <li>• Steroid-induced lymphopenia</li> <li>• Heterozygous mutation for FMF m694</li> </ul>	<ul style="list-style-type: none"> <li>ELANE</li> <li>IL17RA (novel)</li> </ul>
ID-30	<ul style="list-style-type: none"> <li>• 7-year-old male patient</li> <li>• Recurrent admission with cough and fever since early infancy period</li> <li>• Chronic mycosis of the skin</li> <li>• Developmental delay, hypothyroidism</li> </ul>	<ul style="list-style-type: none"> <li>• Normal immunoglobulin level</li> <li>• Normal DHR</li> <li>• Normal CBC</li> </ul>	<ul style="list-style-type: none"> <li>STAT1 (novel)</li> <li>CARMIL2 (novel)</li> </ul>
ID-31	<ul style="list-style-type: none"> <li>• 3-year-old female patient</li> <li>• Diagnosis of biliary stasis at the age of 2 months, recurrent history of fever since early infantile period, negative culture yield</li> <li>• History of short liver synovitis</li> <li>• History of microscopic hematuria</li> <li>• Prompt response to steroid</li> </ul>	<ul style="list-style-type: none"> <li>• Neutrophilic leukocytosis</li> <li>• High inflammatory markers</li> <li>• High MVK in urine</li> <li>• Normal immunoglobulin level</li> </ul>	<ul style="list-style-type: none"> <li>TNIP1 (novel)</li> </ul>
ID-32	<ul style="list-style-type: none"> <li>• 6-month-old male patient</li> <li>• History of chronic cough</li> <li>• Failure to thrive</li> <li>• Chronic diarrhea</li> </ul>	<ul style="list-style-type: none"> <li>• No lymphopenia</li> <li>• Decreased naïve T cells, reduced number of switched memory B cells</li> <li>• Normal immunoglobulin level</li> </ul>	<ul style="list-style-type: none"> <li>GPSM1</li> <li>NLRP12 (deleterious)</li> </ul>
ID-33	<ul style="list-style-type: none"> <li>• 4-year-old female patient</li> <li>• Diagnosed with secondary alveolar proteinosis at the age of 8 months, improved on antimicrobials and systemic steroids</li> <li>• Recurrent admissions with chest infection</li> <li>• Negative culture yields</li> <li>• Developmental delay, epilepsy disorder at the age of 2 years</li> </ul>	<ul style="list-style-type: none"> <li>• No lymphopenia</li> <li>• Transient hypogammaglobulinemia, improved after the age of 2 years</li> <li>• Low CD4 count</li> </ul>	<ul style="list-style-type: none"> <li>IFIH1</li> <li>MBL2 (pathogenic)</li> </ul>

Table I. Continued.

Patient	Patient's history	Immunological Lab findings	Mutated genes identified
ID-34	<ul style="list-style-type: none"> <li>• 14-year-old male patient</li> <li>• Diagnosed with unclassified arthritis at the age of 11 years, managed by anti-TNF</li> <li>• Diagnosed with esophageal candidiasis</li> <li>• Prolonged fever, disseminated intravascular coagulation at the age of 14 years</li> </ul>	<ul style="list-style-type: none"> <li>• Initial lab showed normal immunodeficiency work-up</li> <li>• LN biopsy with atypical lymphoblast</li> </ul>	<p>TYK2 UNG</p>
ID-35	<ul style="list-style-type: none"> <li>• 9-month-old male patient</li> <li>• Diagnosis of complicated meningitis at the age of 7 months</li> <li>• Polio vaccine-associated paralysis</li> </ul>	<ul style="list-style-type: none"> <li>• Agammaglobulinemia absent</li> <li>• CD4 lymphocyte subset</li> </ul>	<p>IL17RC CDCA7</p>

NGS, next-generation sequencing; STAT2, signal transducer and activator of transcription 2; PIK3R4, phosphoinositide 3-kinase regulatory subunit 4; ELANE, elastase, neutrophil expressed; IL-17RA, interleukin 17 receptor A; CARMIL2, capping protein regulator and myosin 1 linker 2; TNIP1, TNF $\alpha$ -induced protein 3-interacting protein 1; GPSM1, G-protein-signaling modulator 1; NLRP12, NLR family, pyrin domain-containing 12; IFIH1, interferon induced with helicase C domain 1; MBL2, mannose binding lectin 2; TYK2, tyrosine kinase 2; UNG, uracil DNA glycosylase; CDCA7, cell division cycle associated 7; EBV, Epstein-Barr virus; IG, immunoglobulin; DHR, dihydrorhodamine flow cytometric Test; CBC, complete blood count; MVK, mevalonate kinase.

phosphoinositide 3-kinase regulatory subunit 4 (*PIK3R4*) (c.574C>T; p.Leu192Phe), *TNIP1* (c.460C>G; p.Asp154His), *STAT2* (c.1466C>G; p.Pro489Arg), *ELANE* (c.659G>A; p.Arg220Gln), capping protein regulator and myosin 1 linker 2 (*CARMIL2*) (c.3683C>T; p.Pro1228Leu), *NLRP12* (c.2591T>A; p.Leu863Gln), mannose binding lectin 2 (*MBL2*) (c.154C>T; p.Arg52Cys), *TYK2* (c.1342C>T; p.Arg448Trp), uracil DNA glycosylase (*UNG*) (c.262C>T; p.Arg88Cys) and cell division cycle associated 7 (c.679C>T; p.Arg227Cys). Furthermore, two homozygous SNPs were detected, one in *GPSM1* (c.1283C>A, stop gained; p.Ser428Ter) and another in *IFIH1* (c.2891G>A; p.Cys964Tyr). In addition, a CTC/CT frameshift mutation (L/X) was detected in *GPSM1* in patient PID-32 and a TCTGGTCTT in-frame insertion (p.V42VWSF) in *IL17RC* in patient PID-35. Of note, five novel SNPs were identified in *CARMIL2*, *TNIP1*, *STAT1*, *IL17RA* and *PIK3R4*. Of these, c.1061T>C in *STAT1* was *de novo* and further confirmed using Sanger sequencing (Figs. 1-5 and Tables II and III).

Kyoto Encyclopedia of Genes and Genomes (KEGG; <https://www.genome.jp/kegg/pathway.html>) pathway analysis also showed involvement of the IL-17 signaling pathway [IL-17 receptor A (*IL17RA*), c.1345C>G], T helper type 17 (Th17) cell differentiation (*STAT1*, c.1061T>C), the JAK-STAT signaling pathway [*STAT1* and tyrosine kinase 2 (*TYK2*)], neutrophil extracellular trap (NET) formation (*ELANE*, c.659G>A), cocaine addiction (*GPSM1*, c.1283C>A) and cytokine-cytokine receptor interaction (*IL17RC*, TCTGGTCTT in-frame insertion) (Fig. 6).

## Discussion

Primary immunodeficiencies encompass a diverse spectrum of disorders that necessitate precise diagnostic strategies. The emergence of NGS techniques has revolutionized clinical immunology. These methodologies not only aid in pinpointing the causative genetic variations but also inform therapeutic interventions, tailoring treatments to individual patient's genetic profiles (13,14).

A salient factor influencing the incidence of novel mutations is consanguinity. In Jordan, consanguinity is more prevalent in rural areas (~40%) than in its capital, Amman, where these levels are ~25.5%. The historical trend of consanguinity in Amman also seems to have influenced the prevalence of PID. Marriages before 1980 had a consanguinity rate of ~30%. (15)

Consanguinity does not seem to be a relevant bias in the present cohort of patients with PID. A novel homozygous mutation of uncertain significance was found in *GPSM1*, only in patient ID-32. In addition, the homozygous *IFIH1* variant was found in a family where consanguinity was not previously reported. In all other patients, only heterozygous variants/mutations were found.

Arunachalam *et al* (14) elucidated that PIDs are typically monogenic, arising from an SNV. By contrast, polygenic diseases often manifest as autoimmune conditions such as rheumatoid arthritis or systemic lupus erythematosus. Of note, certain genetic variations that are associated with PIDs also appear in autoimmune diseases, underscoring the interwoven nature of immunological disorders (14,15).

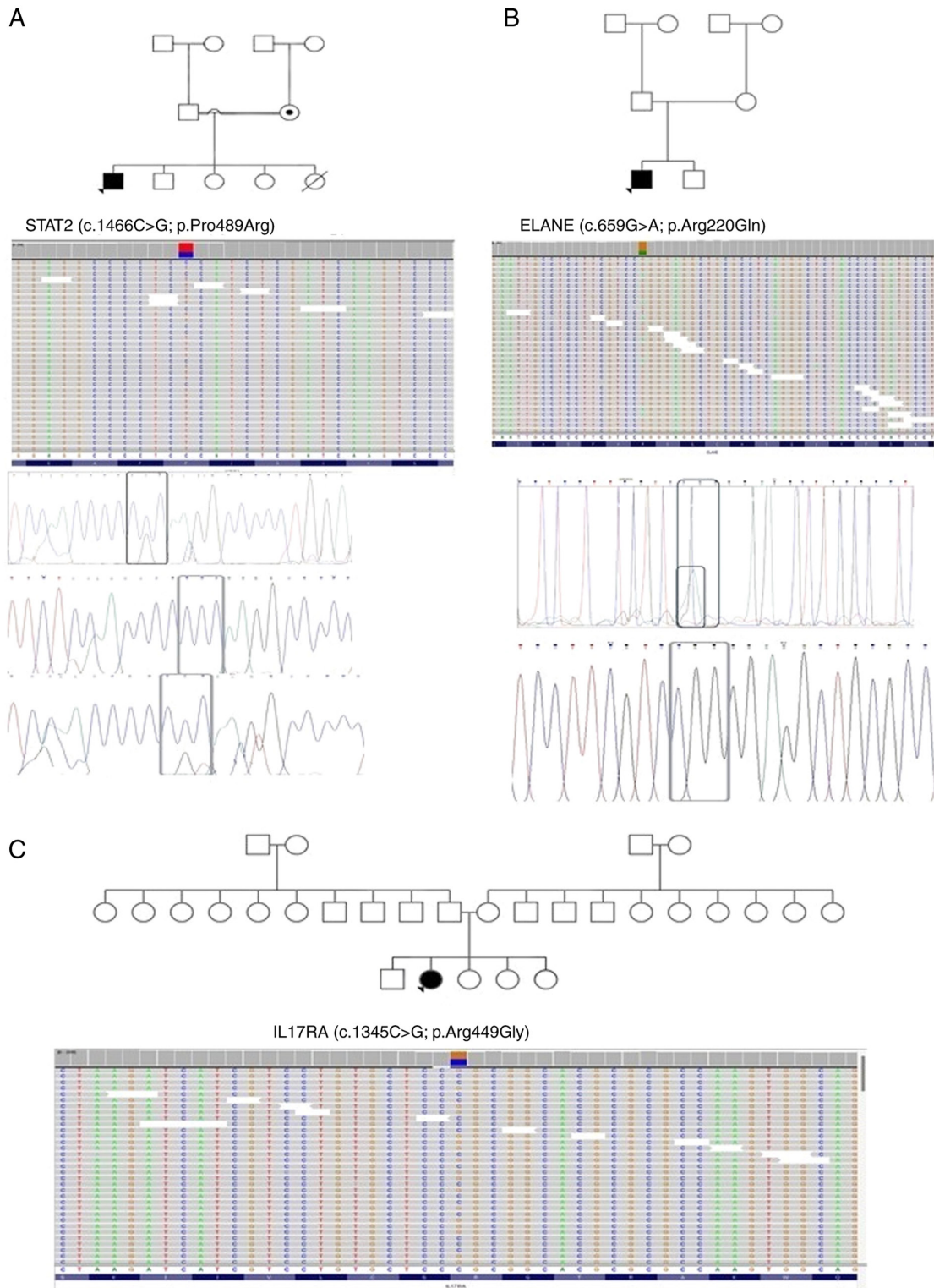


Figure 1. Pedigrees of patients (A) ID-26, (B) ID-28 and (C) ID-29, along with the single nucleotide polymorphisms detected in STAT2, ELANE and IL17RA, respectively, through exome-sequencing and Sanger sequencing. Proband is indicated with arrowheads. Circles with a black dot indicate the mother or sister being a carrier of the corresponding SNV. A double line between parents represents consanguinity. A square with a black dot indicates the father or brother being a carrier of the corresponding SNV. A circle/square with a strike-through line indicates a deceased subject. The different colors in the next-generation sequencing reads from IGV depict the different nucleotides. Red is for 'T', blue is for 'C', green is for 'A' and brown is for 'G'. STAT2, signal transducer and activator of transcription 2; ELANE, elastase, neutrophil expressed; IL17RA, interleukin 17 receptor A; SNV, single nucleotide variant.

It is true that several studies have shed light on the genetic landscape of PIDs, but with different results. For instance, Arunachalam *et al* (14) identified numerous genetic variants across different PID subtypes. In addition, studies from

Aghamohammadi *et al* (16,17), Abolhassani *et al* (18,19) and Kilic *et al* (20) provided valuable insight into the distribution of PID subtypes in Iranian and Turkish populations, respectively.

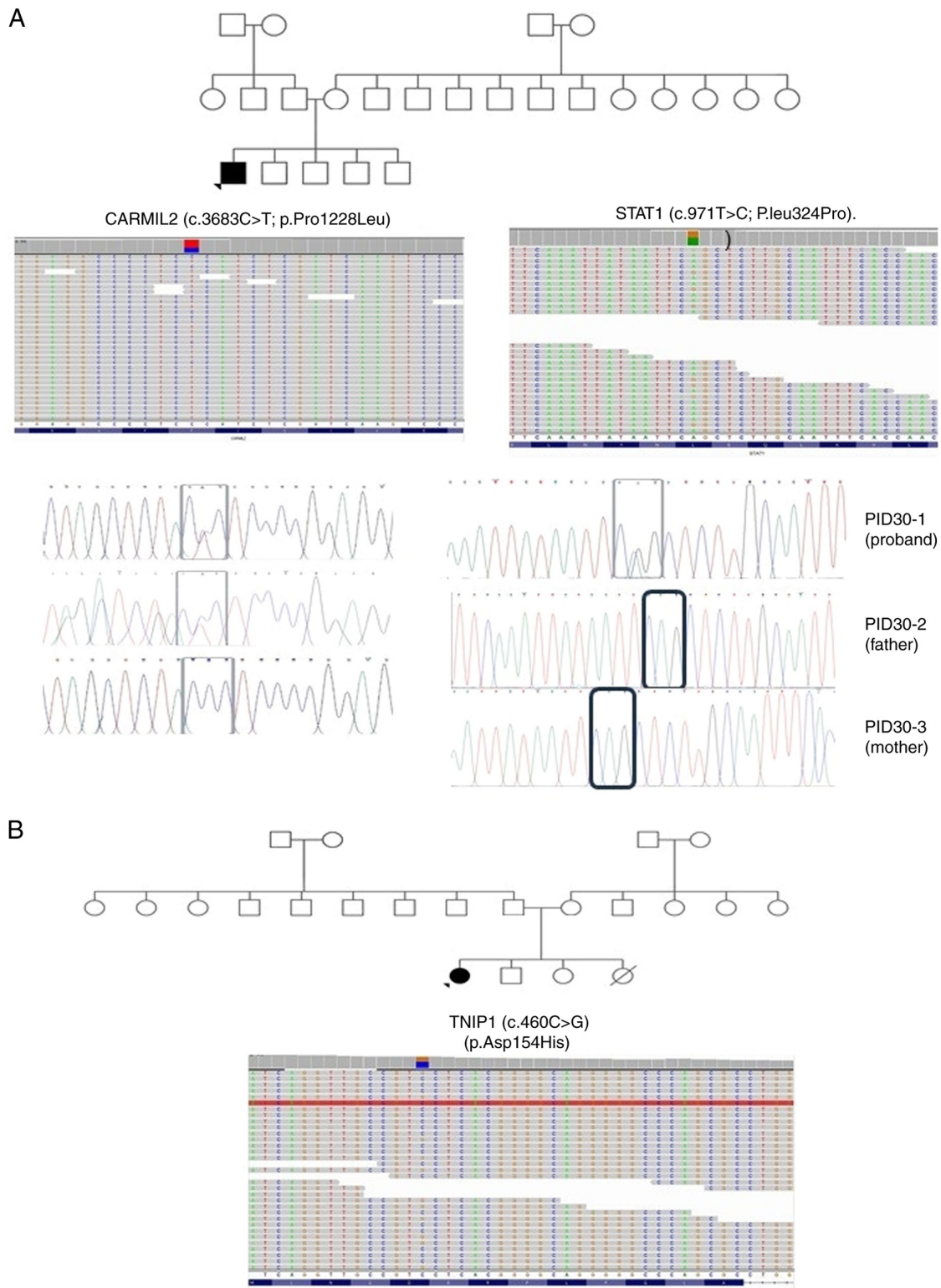


Figure 2. Pedigrees of patients (A) ID-30 and (B) ID-31, along with the single nucleotide polymorphisms detected in (A) CARMIL2 and STAT1 and (B) TNIP1 through exome-sequencing and Sanger sequencing. Proband is indicated with arrowheads. Circles with a black dot indicate the mother or sister being a carrier of the corresponding SNV. A double line between parents represents consanguinity. A square with a black dot indicates the father or brother being a carrier of the corresponding SNV. A circle/square with a strike-through line indicates a deceased subject. The different colors in the next-generation sequencing reads from IGV depict the different nucleotides. Red is for 'T', blue is for 'C', green is for 'A' and brown is for 'G'. CARMIL2, capping protein regulator and myosin 1 linker 2; STAT1, signal transducer and activator of transcription 1; TNIP1, TNF $\alpha$ -induced protein 3-interacting protein 1; SNV, single nucleotide variant.

In the present study, the genetic underpinnings of PIDs in 9 Jordanian patients were explored, identifying mutations in key immunological genes, including *STAT1/2*, *MBL2*,

*TYK2*, *UNG*, *TNIP1*, *CARMIL2* and *ELANE*, as well as a 9-bp insertion in *IL17RC*, and one frameshift and nonsense mutation in the same exon in *GPSM1*. Of note, a *STAT2*

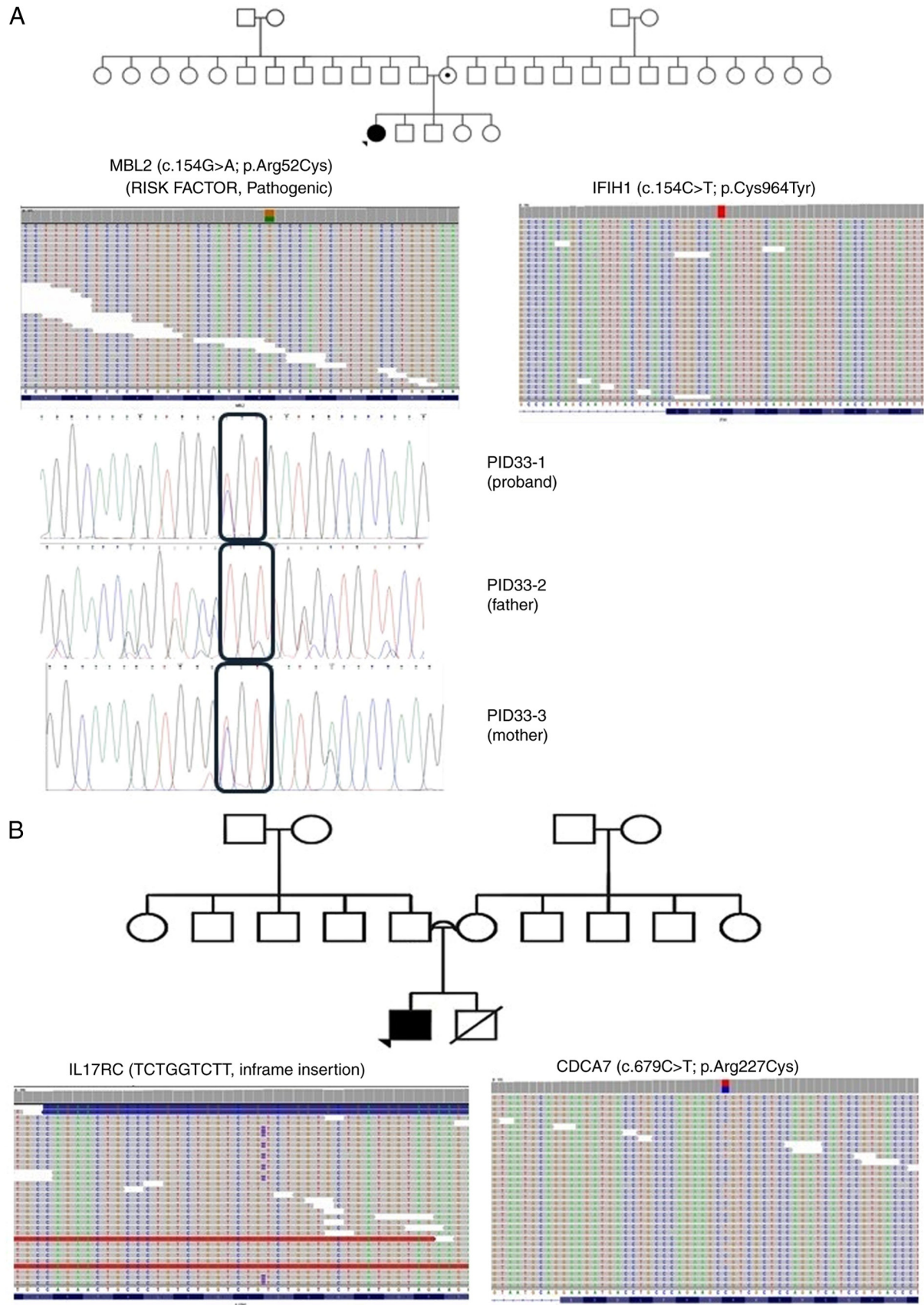


Figure 3. Pedigrees of patients (A) ID-33 and (B) ID-35, along with the single nucleotide polymorphisms detected in (A) MBL2 and IFIH1 and (B) IL17RC and CDCA7 through exome-sequencing and Sanger sequencing. Proband is indicated with arrowheads. Circles with a black dot indicate the mother or sister being a carrier of the corresponding SNV. A double line between parents represents consanguinity. A square with a black dot indicates the father or brother being a carrier of the corresponding SNV. A circle/square with a strike-through line indicates a deceased subject. The different colors in the next-generation sequencing reads from IGV depict the different nucleotides. Red is for 'T', blue is for 'C', green is for 'A' and brown is for 'G'. MBL2, mannose binding lectin 2; IFIH1, interferon induced with helicase C domain1; IL17RC, interleukin 17 receptor C; CDCA7, cell division cycle associated 7; cell division cycle associated 7; SNV, single nucleotide variant.



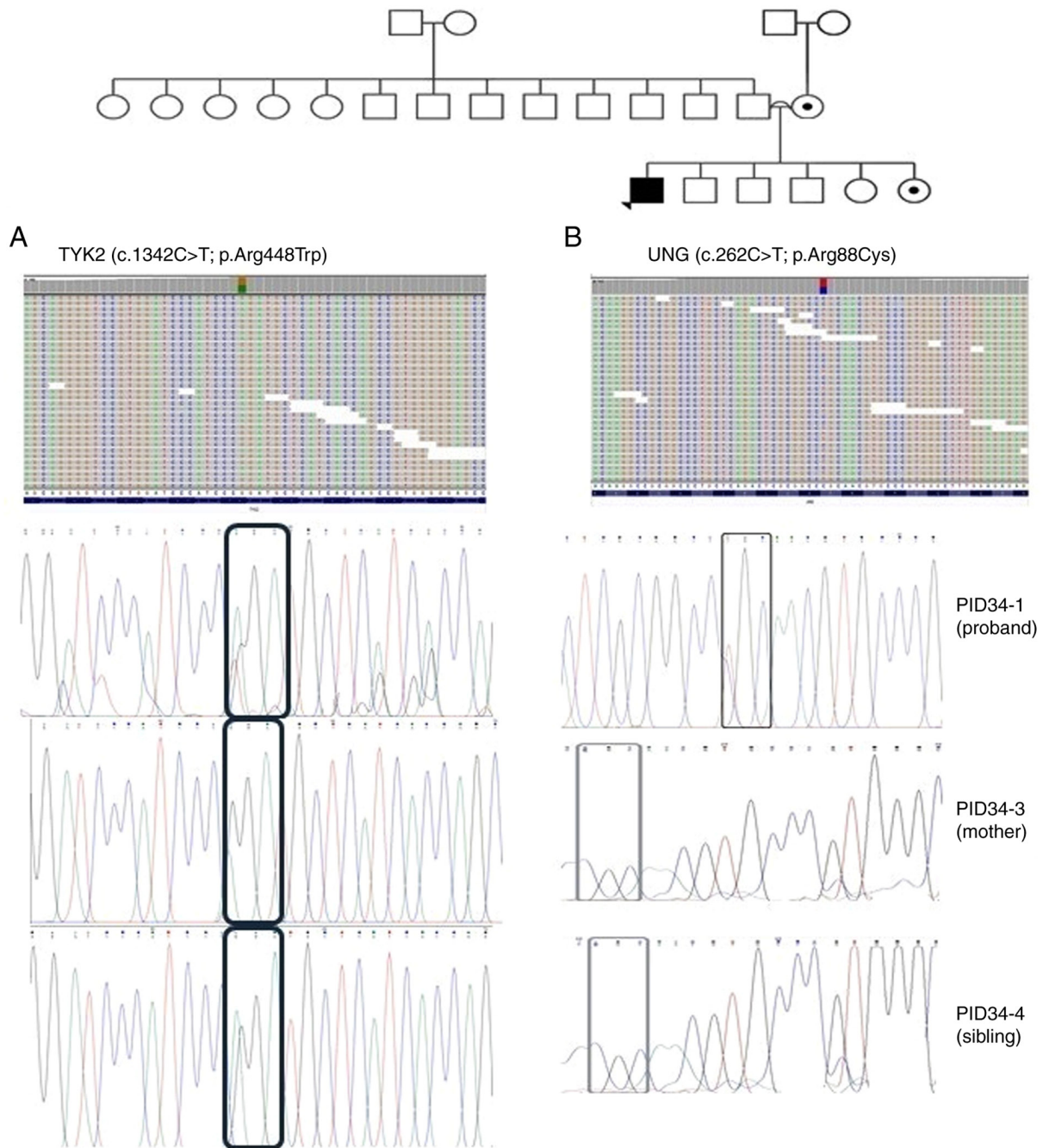


Figure 4. Pedigree of patient ID-34, along with the single nucleotide polymorphisms detected in TYK2 (A) and UNG (B) through exome-sequencing and Sanger sequencing. Probands are indicated with arrowheads. Circles with a black dot indicate the mother or sister being a carrier of the corresponding SNV. A double line between parents represents consanguinity. A square with a black dot indicates the father or brother being a carrier of the corresponding SNV. A circle/square with a strike-through line indicates a deceased subject. The different colors in the next-generation sequencing reads from IGV depict the different nucleotides. Red is for 'T', blue is for 'C', green is for 'A' and brown is for 'G'. TYK2, tyrosine kinase 2; UNG, uracil DNA glycosylase; SNV, single nucleotide variant.

mutation was found in patient ID-26 suffering from hypogammaglobulinemia (after treatment with rituximab). However, STAT2 heterozygosity may have no impact on the patient; thus, it should be investigated along with PIK3R4 in order to determine their effect on the patient.

Furthermore, mutations in TYK2 and UNG were found in patient ID-34, who had been diagnosed with unclassified arthritis and esophageal candidiasis.

Mutations in STAT1 have been reported in different ethnicities, affecting critical cellular processes, such as

STAT1-mediated responses to IFNG and IL27 (21,22) In the present cohort, patient ID-30 had a heterozygous SNV in the CARMIL2 and STAT1 genes. CARMIL2 is associated with immunodeficiency-58 (IMD58). This is an AR primary immunologic disorder distinguished by early-onset skin lesions, such as eczematous dermatitis, infectious abscesses and warts, as well as recurrent respiratory infections or allergies, among others. CARMIL2 deficiency, as described by Wang *et al* (23), presents with a unique set of clinical features, with compromised T-cell subsets and antibody

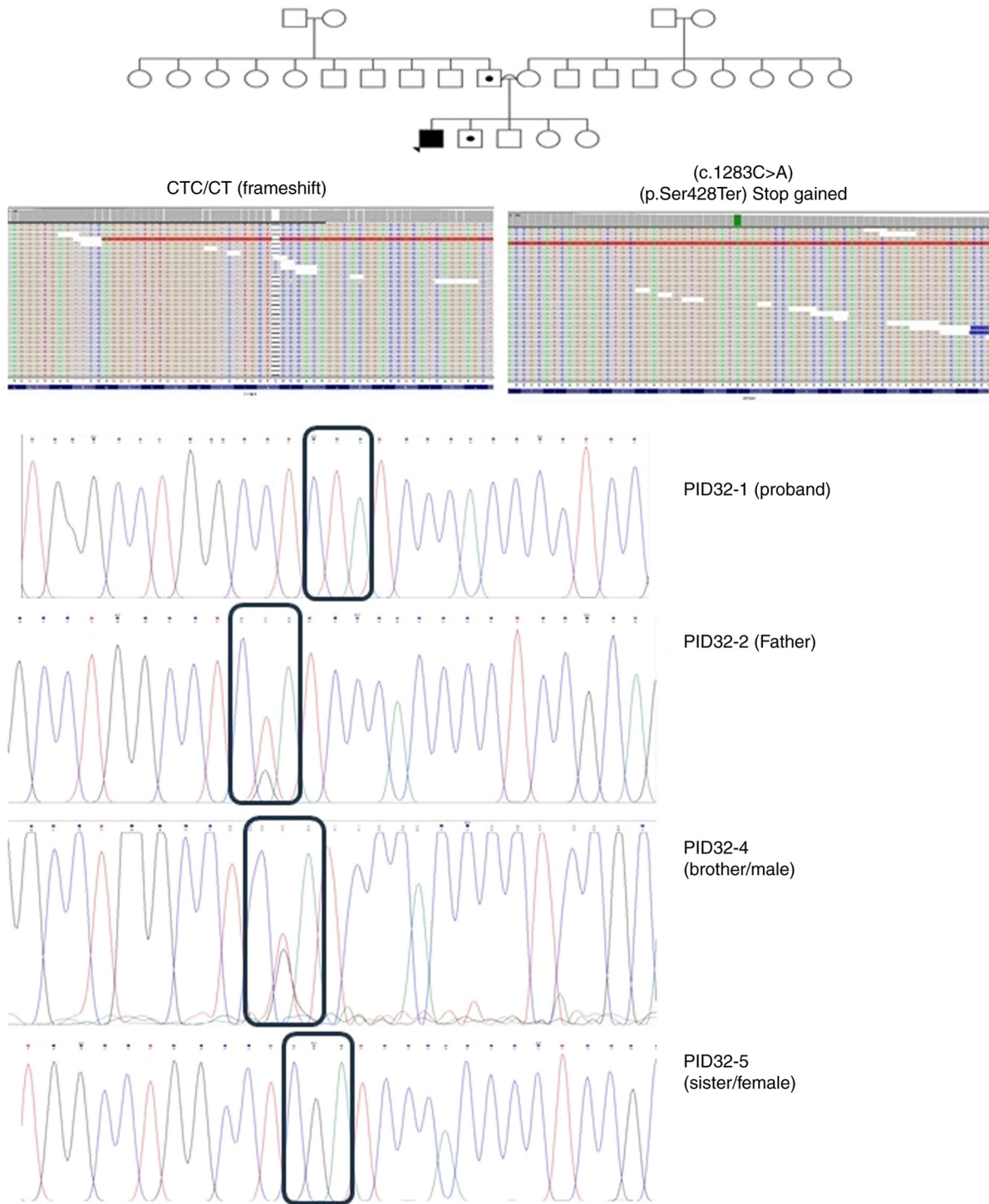


Figure 5. Pedigree of patient ID-32, along with the single nucleotide polymorphisms detected in GPSM1 (frameshift and stop gained), through exome-sequencing and Sanger sequencing. Proband is indicated with arrowheads. Circles with a black dot indicate the mother or sister being a carrier of the corresponding SNV. A double line between parents represents consanguinity. A square with a black dot indicates the father or brother being a carrier of the corresponding SNV. A circle/square with a strike-through line indicates a deceased subject. The different colors in the next-generation sequencing reads from integrative genomic viewer depict the different nucleotides. Red is for 'T', blue is for 'C', green is for 'A' and brown is for 'G'. GPSM1, G-protein-signaling modulator 1; SNV, single nucleotide variant.

responses. STAT1 deficiency, on the other hand, is associated with three distinct phenotypes depending on whether it's loss of function (LOF) AD/AR or gain of function (GOF) AD, while LOF mutations correlate with compromised Th1 immunity. GOF mutations are associated with impaired Th17 immunity-in the case of patient ID-30, GOF mutations were

more likely suggested based on the patient's clinical presentation. IMD31C is an immunologic dysregulation disorder with highly variable manifestations caused by AD GOF mutations in STAT1. Most patients present with chronic mucocutaneous candidiasis in infancy or early childhood (24). Other highly variable characteristics include persistent infections,

**Table II. Variants for each gene, mode of inheritance, CDS and protein change, consanguinity and family history for each patient with inborn errors of immunity.**

Patient	Gene	Zygoty	Inheritance	CDS change	Protein change	Consanguinity	Family history
ID-26	i) STAT2	i) HET	i) AR	i) c.1345C>G	i) p.Arg449Gly	Consanguineous	• Positive family history of similar phenotype
	ii) PIK3R4	ii) HET	ii) -	ii) c.574C>T	ii) p.Leu192Phe	Consanguineous	• Sister died due to uncontrolled autoimmunity
ID-28	ELANE	HET	AD	c.659G>A	p.Arg220Gln	Non-consanguineous	Negative family history
ID-29	IL17RA	HET	AR	c.1345C>G	p.Arg449Gly	Non-consanguineous	• Strong family history of FMF
ID-30	i) STAT1	i) HET	i) AR/AD	i) c.1061T>C	i) p.Leu354Pro	Non-consanguineous	• Brother diagnosed with sacroiliitis
	ii) CARMIL2	ii) HET	ii) AR	ii) c.3683C>T	ii) p.Pro1228Leu	Non-consanguineous	Negative family history
ID-31	TNIP1	HET	-	c.460C>G	p.Asp154His	Non-consanguineous	Father has chronic kidney disease and vision problems
ID-32	i) GPSM1	i) HOM	i) -	i) c.1283C>A	i) p.Ser428Ter	Consanguineous	Positive family history of similar illness (proband's brother)
	ii) NLRP12	ii) HET	ii) AD	ii) c.2591T>A	ii) p.Leu863Gln	Consanguineous	
ID-33	i) IFIH1	i) HOM	i) AR	i) c.2891G>A	i) p.Cys964Tyr	Consanguineous	Negative family history
	ii) MBL2	ii) HET	ii) AD	ii) c.154C>T	ii) p.Arg52Cys	Consanguineous	
ID-34	i) TYK2	i) HET	i) AR	i) c.1342C>T	i) p.Arg448Trp	Consanguineous	• Positive family history of malignancy (father, uncle and brother)
	ii) UNG	ii) HET	ii) AR	ii) c.262C>T	ii) p.Arg88Cys	Non-consanguineous	• Proband's aunt died due to SLE
ID-35	i) IL17-RC	i) HET	i) AR	i) TCTGGTCIT	i) p.V42VWSF	Non-consanguineous	Positive family history (proband's brother)
	ii) CDCA7	ii) HET	ii) AR	ii) c.679C>T	ii) p.Arg227Cys	Non-consanguineous	

CDS, coding sequence; AR, autosomal recessive; AD, autosomal dominant; HOM, homozygous; HET, heterozygous; STAT2, signal transducer and activator of transcription 2; PIK3R4, phosphoinositide 3-kinase regulatory subunit 4; ELANE, elastase, neutrophil expressed; IL-17RA, interleukin 17 receptor A; CARMIL2, capping protein regulator and myosin 1 linker2; TNIP1, TNF $\alpha$ -induced protein 3-interacting protein 1; GPSM1, G-protein-signaling modulator 1; NLRP12, NLR family, pyrin domain-containing 12; IFIH1, interferon induced with helicase C domain 1; MBL2, mannose binding lectin 2; TYK2, tyrosine kinase 2; UNG, uracil DNA glycosylase; CDCA7, cell division cycle associated 7.

Table III. SNP and protein change for each detected gene per patient with inborn errors of immunity, along with the SIFT, PolyPhen-2, CADD score, ClinVar and OMIM phenotype.

Patient	Gene ID	SNP change	Protein change	SIFT	PolyPhen-2	CADD score	ClinVar	OMIM phenotype
ID-26	STAT2	c.1466C>G	p.Pro489Arg	0 (deleterious)	0.966 (damaging)	22.5	Uncertain significance	Immunodeficiency 44
	PIK3R4	c.574C>T	p.Leu192Phe	0 (deleterious)	0.959 (damaging)	26.9	-	-
ID-28	ELANE	c.659G>A	p.Arg220Gln	0.56 (tolerated)	0.96 (probably damaging)	21.8	Pathogenic	Severe congenital neutropenia
ID-29	IL17RA	c.1345C>G	p.Arg449Gly	0 (deleterious)	0.997 (probably damaging)	25.8	-	IMD-51
ID-30	STAT1	c.1061T>C	p.Leu354Pro	0 (deleterious)	0.989 (probably damaging)	25.2	-	IMD 31 A,B,C
	CARMIL2	c.3683C>T	p.Pro1228Leu	0 (deleterious)	0.995 (probably damaging)	25.0	-	IMD 58
ID-31	TNIP1	c.460C>G	p.Asp154His	0.01 (deleterious)	0.996 (possibly damaging)	24.2	-	-
ID-32	GPSM1	CTC/CT frameshift c.1283C>A (Stop gained)	L/X p.Ser428Ter	-	-	0.07 0.22	-	-
	NLRP12	c.2591T>A	p.Leu863Gln	0 (deleterious)	0.934 (possibly damaging)	24.3	-	Familial cold autoinflammatory syndrome 2
ID-33	IFIH1	c.2891G>A	p.Cys964Tyr	0 (deleterious)	1 (probably damaging)	26.1	-	Immunodeficiency 95
	MBL2	c.154C>T	p.Arg52Cys	0 (deleterious)	0.997 (Probably damaging)	24.3	Risk factor (pathogenic)	Chronic infection (due to MBL2 deficiency)
ID-34	TYK2	c.1342C>T	p.Arg448Trp	0.01 (deleterious)	0.462 (possibly damaging)	24.8	Uncertain significance	Immunodeficiency 35
	UNG	c.262C>T	p.Arg88Cys	0 (deleterious)	0.714 (possibly damaging)	31.0	Uncertain significance; conflicting interpretations of pathogenicity	Immunodeficiency with hyper IgM, type 5
ID-35	CDCA7	c.679C>T	p.Arg227Cys	0.01 (deleterious)	0.899 (possibly damaging)	23.6	-	ICF3
	IL17RC	TCTGGTCTT (inframe insertion)	p. v42VWSF	-	-	-	-	Candidiasis

SNP, single nucleotide polymorphism; ClinVar, Clinical Variance; OMIM, Online Mendelian Inheritance in Man; STAT2, signal transducer and activator of transcription 2; PIK3R4, phosphoinositide 3-kinase regulatory subunit 4; ELANE, elastase, neutrophil expressed; IL-17RA, interleukin 17 receptor A; CARMIL2, capping protein regulator and myosin 1 linker 2; TNIP1, TNF $\alpha$ -induced protein 3-interacting protein 1; GPSM1, G-protein-signaling modulator 1; NLRP12, NLR family, pyrin domain-containing 12; IFIH1, interferon induced with helicase C domain 1; MBL2, mannose binding lectin 2; TYK2, tyrosine kinase 2; UNG, uracil DNA glycosylase; CDCA7, cell division cycle associated 7; IMD 51, immunodeficiency 51; ICF3, immunodeficiency-centromeric instability-facial anomalies syndrome-3.

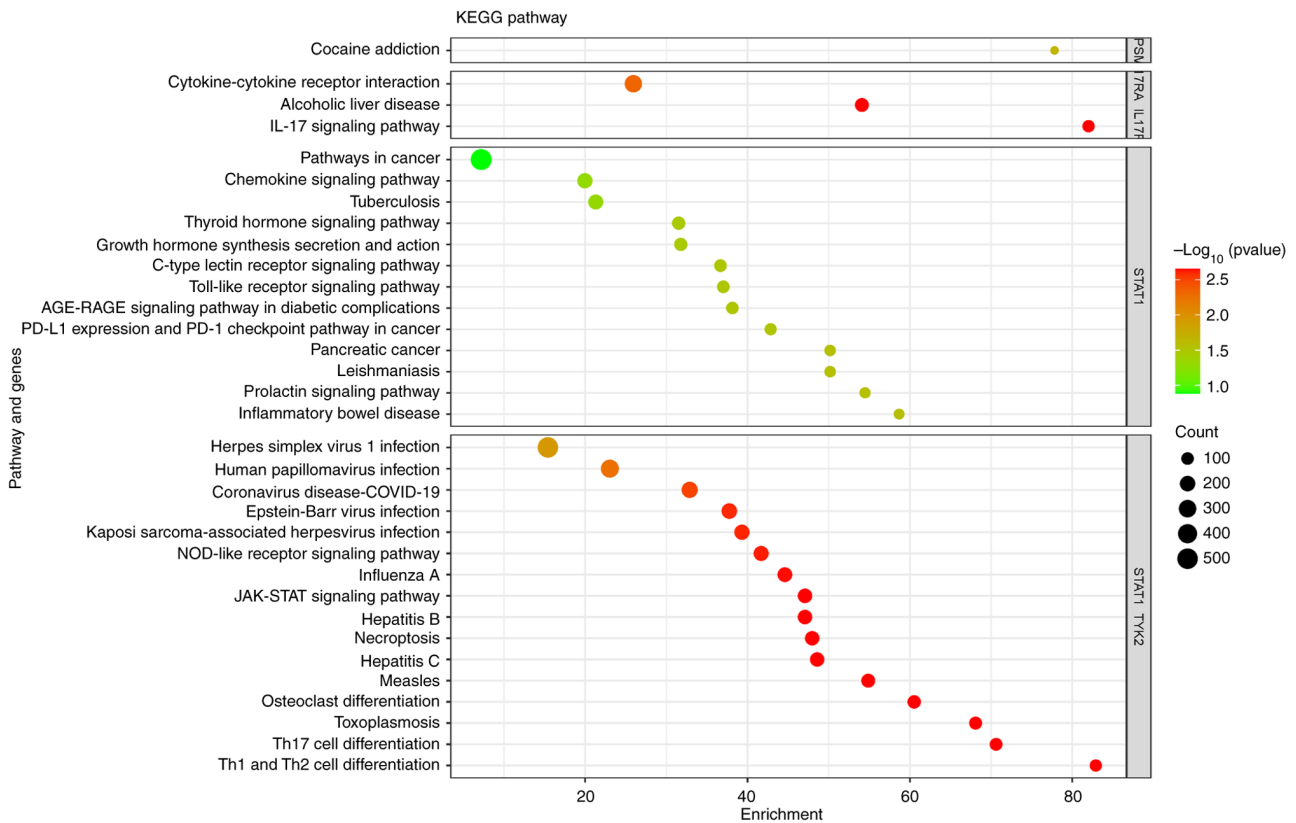


Figure 6. Bubble plot of KEGG pathway enrichment analysis of the genes with the detected single nucleotide polymorphisms in the Jordanian cohort with inborn errors of immunity. 'Count' indicates the number of genes enriched in each pathway. KEGG, Kyoto Encyclopedia of Genes and Genomes.

enteropathy with villous atrophy and autoimmune disorders such as hypothyroidism and diabetes mellitus. In addition, *IMD31C* is inherited in an AD manner (25-27). Both genes (*STAT1*, *CARMIL2*) could be relevant to the patient's disease, *STAT1* function (e.g. phosphorylation) should be investigated, in addition to *IL-17A* production testing post-stimulation by phorbol 12-myristate 13-acetate and ionomycin, are useful to activate transcription factors for intracellular signaling and production of cytokines of many different immune cell types in the presence of Brefeldin, as well as the enumeration of Th17 lymphocytes, feasible through C-C motif chemokine receptor 6 and C-X-C motif chemokine receptor 3 markers, to ascertain the diagnosis. In addition, *CARMIL2* expression should be detected in the patient. Similarly, mutations in *IL17RA* and *TYK2* have been associated with characteristic clinical phenotypes and cellular defects (28-30). In the present study, a new *IL17RA* mutation was detected in a patient suffering from hypergammaglobulinemia and steroid-induced lymphopenia, namely ID-29. Two new mutations, one in *STAT1* and another in *CARMIL2*, were also detected in a patient suffering from chronic mycosis of the skin, developmental delay and hypothyroidism, namely ID-30. In addition, a new *TNIP1* mutation was found in ID-31, a patient diagnosed with biliary stasis at the age of 2 months, with are current history of fever since the early infantile period, and a history of short liver synovitis and microscopic hematuria. Furthermore, studies have emphasized the clinical significance of mutations in genes such as *UNG*, *MBL2* and *IL17RC* (14,31-35).

Furthermore, studies reported the existence of *NLRP12* deficiency in Chinese patients, being a compelling case of the diverse clinical manifestations associated with *PID* mutations, ranging from erythema nodosa to systemic symptoms such as fever and lymphadenopathy (36-38).

*ELANE* encodes leukocyte elastase, being produced by neutrophil precursors, and its mutations have been associated with different immunodeficiencies, such as congenital neutropenia (14,39,40). In the present study, *ELANE* mutation was found in patient ID-28, who suffered from severe neutropenia. The patient was diagnosed with no maturation arrest in the bone marrow study performed at the age of 3 months.

*IFIH1* (*IMD95*) encodes an RIG-I-like receptor involved in the sensing of viral RNA (41,42). *IFIH1* variants are also related to hepatitis C virus clearance (43). Heterozygous gain-of-function *IFIH1* mutations have also been associated with autoimmune diseases (44). Furthermore, rare gain-of-function mutations in *IFIH1* significantly increase the production of type I IFN, leading to Aicardi-Goutières or Singleton-Merten syndromes (45,46). Nonetheless, *IMD95* is an AR disorder characterized primarily by recurrent and severe viral respiratory infections beginning in infancy or early childhood (47). Infections such as human rhinovirus which is the most common viral infectious agent in humans and is the predominant cause of the common cold and respiratory syncytial virus, a contagious virus that causes infections of the respiratory tract, frequently necessitate hospitalization or respiratory support. Immunologic testing is usually normal, though some minor abnormalities may be detected. The

disorder is caused by a loss of the innate immune system's ability to sense viral genetic information, resulting in a lack of interferon production, poor response to viral and immunologic stimulation and failure to control viral replication (48). The patient in whom the IFIH1 mutation was detected had a low CD4 count. She was diagnosed with secondary alveolar proteinosis at the age of 8 months, had recurrent admissions with chest infection, as well as developmental delay and epilepsy disorder at the age of 2 years, which were associated with the patient's symptoms and initial diagnosis (15).

Familial cold autoinflammatory syndrome-2 is an AD autoinflammatory disorder that causes episodic and recurring rash, urticaria, arthralgia, myalgia and headaches. In most patients, these episodes are accompanied by fever and serologic signs of inflammation. The majority of patients, but not all, report that exposure to colds triggers their episodes. Other symptoms may include abdominal pain, thoracic pain and sensorineural deafness. The age of onset varies, ranging from infancy to middle age, and the severity and clinical manifestations are diverse (49). In the present study, patient ID-32 had a mutation in NLRP12 and according to various databases, the variant found is pathogenic. The patient had a history of chronic cough, failure to thrive and chronic diarrhea, which explains the symptoms. Furthermore, a mutation in GPSM1 was detected for the same patient; he had decreased naïve T cells and a reduced number of switched memory B cells.

Despite the high rate of consanguinity (4 out of 9 patients), only two homozygous variants were identified, with only one potentially linked to an IEI. This can be explained by considering several factors related to genetic diversity, the complexity of inheritance patterns and the nature of the diseases involved. IEIs are often genetically heterogeneous, meaning they can be caused by mutations in different genes. Despite the high rate of consanguinity, the patients may carry mutations in different genes that lead to similar clinical phenotypes. In addition, not all individuals with a pathogenic variant will necessarily manifest the disease (incomplete penetrance) and the severity of the disease can vary among those who do (variable expressivity) (38,50). This could result in only some of the patients showing symptoms of an IEI despite having related genetic backgrounds. Certain genetic conditions are caused by having two different mutations in a gene (one on each allele) rather than two identical mutations (homozygosity). In consanguineous populations, there may still be a presence of compound heterozygous variants that are not immediately apparent as homozygous but that lead to disease (51). Besides homozygous variants, other mechanisms, such as copy number variations, epigenetic changes or mutations in regulatory regions may be responsible for IEIs and may not be easily detected by standard genetic screening focused on homozygous variants (52-54). Finally, IEIs may be influenced by multiple genes and environmental factors. Therefore, not all individuals with consanguinity and clinical symptoms will have a clear, single-gene cause that can be identified as homozygous mutations.

The IL-17 family of cytokines plays crucial roles in both acute and chronic inflammatory responses and IL-17A, the hallmark cytokine of the Th17 cells, has important roles in

protecting the host against extracellular pathogens, but also promotes inflammatory pathology in autoimmune disease (55).

The mutation in *STAT1* affects the differentiation of Th17 cells, which are a subset of CD4+ T cells involved in epithelial cell- and neutrophil-mediated immune responses against extracellular microbes and in the pathogenesis of autoimmune diseases. The same mutation along with the c.1342C>T mutation in *TYK2* also affects the JAK-STAT signaling pathway. This is the principal signaling mechanism for a wide array of cytokines and growth factors. Following the binding of cytokines to their cognate receptor, STATs are activated by members of the JAK family of tyrosine kinases. Once activated, they dimerize and translocate to the nucleus and modulate the expression of target genes. In addition to the activation of STATs, JAKs mediate the recruitment of other molecules such as the MAPKs and PI3K. These molecules process downstream signals via the Ras/Raf/MAPK and PI3K pathways, which results in the activation of additional transcription factors.

Our analysis also hints towards a deregulation in the NET formation through the mutation in *ELANE*. Neutrophils play a central role in innate immune defense. One of the mechanisms of neutrophil action is the formation of NETs, the extracellular structures composed of chromatin coated with histones, proteases and granular and cytosolic proteins that help catch and kill microorganisms.

Finally, two more pathways affected by mutations in *GPSM1* and *IL17RC* were those of cocaine addiction and cytokine-cytokine receptor interaction, respectively.

The present study has certain limitations, which should be acknowledged. It focuses on the Jordanian population, thus limiting the generalizability of the present findings. Furthermore, the sample size was relatively small (nine families), potentially affecting the statistical power of the study. In addition, the present study lacks functional validation of the main findings. However, it still provides a direction for future research.

In conclusion, the intricate tapestry of primary immunodeficiencies is gradually being unraveled with the aid of advanced genetic techniques. Understanding the genetic basis of these disorders among consanguineous Jordanian families holds promise for the development of personalized therapeutic interventions and improved patient outcomes.

### Acknowledgements

The authors would like to thank all the physicians involved in the care of the patients described but were not included as authors: Dr. Marwan Abu-Halaweh (Genetics Laboratory, Philadelphia University, Amman, Jordan) for managing the patient samples and Ms. Eman Al-Bsoul (Genetics Laboratory, Philadelphia University, Amman, Jordan) for providing help with the DNA extraction and the patients' physician, Dr. Raed Al-Zyoud (MD; Consultant Paediatric Allergist, Immunologist and Rheumatologist at Royal Medical services, Amman, Jordan) for providing the medical history of the patients.

### Funding

No funding was received.

### Availability of data and materials

The data generated in the present study may be requested from the corresponding author. All sequencing data have been uploaded to the Figshare repository (10.6084/m9.figshare.26183549).

### Authors' contributions

LO, AZ and MAH proposed and designed the study. LO and AZ carried out the molecular genetic studies. LO, RA and AZ analyzed the results, performed immunological description of the patients and statistical analysis. RA, EA and MAH prepared patient samples and extracted DNA for genetic studies. AZ and LO drafted the manuscript. AZ edited the manuscript. All authors have read and agreed to the final version of the manuscript. LO and AZ confirm the authenticity of all the raw data.

### Ethics approval and consent to participate

The study was conducted in accordance with the Declaration of Helsinki and approved by the Ethical Committee of the Jordanian Royal Medical Services (11/11/2021), and was supervised by Dr Raed Al-Zyoud (Consultant Paediatric Allergist, Immunologist and Rheumatologist at Royal Medical services, Amman, Jordan). Written informed consent to participate in the study was obtained from all subjects and family members involved in the study. Parents/guardians provided consent for minor probands.

### Patient consent for publication

The patients and their family members provided written consent for their medical data to be published. The parents/guardians provided consent for minor probands.

### Competing interests

The authors declare that they have no competing interests.

### References

- Geha RS, Notarangelo LD, Casanova JL, Chapel H, Conley ME, Fischer A, Hammarström L, Nonoyama S, Ochs HD, Puck JM, *et al*: Primary immunodeficiency diseases: An update from the International Union of Immunological Societies Primary Immunodeficiency Diseases Classification Committee. *J Allergy Clin Immunol* 120: 776-794, 2007.
- Tangye SG, Al-Herz W, Bousfiha A, Cunningham-Rundles C, Franco JL, Holland SM, Klein C, Morio T, Oksenhendler E, Picard C, *et al*: Human inborn errors of immunity: 2022 update on the classification from the International Union of Immunological Societies expert committee. *J Clin Immunol* 42: 1473-1507, 2022.
- Tangye SG, Al-Herz W, Bousfiha A, Cunningham-Rundles C, Franco JL, Holland SM, Klein C, Morio T, Oksenhendler E, Picard C, *et al*: The ever-increasing array of novel inborn errors of immunity: An interim update by the IUIS committee. *J Clin Immunol* 41: 666-679, 2021.
- Erdős M, Mironska K, Kareva L, Stavric K, Hasani A, Lányi Á, Kállai J and Maródi L: A novel mutation in SLC39A7 identified in a patient with autosomal recessive agammaglobulinemia. The impact of the J Project. *Pediatr Allergy Immunol* 33: e13805, 2022.
- Notarangelo LD: Primary immunodeficiencies. *J Allergy Clin Immunol* 125 (2 Suppl 2): S182-S194, 2010.
- Boyle JM and Buckley RH: Population prevalence of diagnosed primary immunodeficiency diseases in the United States. *J Clin Immunol* 27: 497-502, 2007.
- Bonilla FA, Khan DA, Ballas ZK, Chinen J, Frank MM, Hsu JT, Keller M, Kobrynski LJ, Komarow HD, Mazer B, *et al*: Practice parameter for the diagnosis and management of primary immunodeficiency. *J Allergy Clin Immunol* 136: 1186-1205.e1-78, 2015.
- Oliveira JB and Fleisher TA: Laboratory evaluation of primary immunodeficiencies. *J Allergy Clin Immunol* 125 (2 Suppl 2): S297-S305, 2010.
- Shehata N, Palda V, Bowen T, Haddad E, Issekutz TB, Mazer B, Schellenberg R, Warrington R, Easton D, Anderson D and Hume H: The use of immunoglobulin therapy for patients with primary immune deficiency: An evidence-based practice guideline. *Transfus Med Rev* 24 (Suppl 1): S28-S50, 2010.
- Amaya-Uribe L, Rojas M, Azizi G, Anaya JM and Gershwin ME: Primary immunodeficiency and autoimmunity: A comprehensive review. *J Autoimmun* 99: 52-72, 2019.
- Lam HY, Clark MJ, Chen R, Chen R, Natsoulis G, O'Huallachain M, Dewey FE, Habegger L, Ashley EA, Gerstein MB, *et al*: Performance comparison of whole-genome sequencing platforms. *Nat Biotechnol* 30: 78-82, 2011.
- El Hawary RE, Meshal SS, Abd Elaziz DS, Alkady R, Lotfy S, Eldash A, Erfan A, Chohayeb EA, Saad MM, Darwish RK, *et al*: Genetic testing in Egyptian patients with inborn errors of immunity: A single-center experience. *J Clin Immunol* 42: 1051-1070, 2022.
- Rios JJ and Delgado MR: Using whole-exome sequencing to identify variants inherited from mosaic parents. *Eur J Hum Genet* 23: 547-550, 2015.
- Arunachalam AK, Maddali M, Aboobacker FN, Korula A, George B, Mathews V and Edison ES: Primary immunodeficiencies in India: Molecular diagnosis and the role of next-generation sequencing. *J Clin Immunol* 41: 393-413, 2021.
- Lamborn I, Jing H, Zhang Y, Drutman SB, Abbott JK, Munir S, Bade S, Murdock HM, Santos CP, Brock LG, *et al*: Recurrent rhinovirus infections in a child with inherited MDA5 deficiency. *J Exp Med* 214: 1949-1972, 2017.
- Aghamohammadi A, Rezaei N, Yazdani R, Delavari S, Kutukculer N, Topyildiz E, Ozen A, Baris S, Karakoc-Aydiner E, Kilic SS, *et al*: Consensus Middle East and North Africa Registry on inborn errors of immunity. *J Clin Immunol* 41: 1339-1351, 2021.
- Aghamohammadi A, Mohammadinejad P, Abolhassani H, Mirminachi B, Movahedi M, Gharagozlou M, Parvaneh N, Zeiaee V, Mirsaeed-Ghazi B, Chavoshzadeh Z, *et al*: Primary immunodeficiency disorders in Iran: Update and new insights from the third report of the national registry. *J Clin Immunol* 34: 478-490, 2014.
- Abolhassani H, Kiaee F, Tavakol M, Chavoshzadeh Z, Mahdavi SA, Momen T, Yazdani R, Azizi G, Habibi S, Gharagozlou M, *et al*: Fourth update on the Iranian National registry of primary immunodeficiencies: Integration of molecular diagnosis. *J Clin Immunol* 38: 816-832, 2018.
- Abolhassani H, Azizi G, Sharifi L, Yazdani R, Mohsenzadegan M, Delavari S, Sohani M, Shirmast P, Chavoshzadeh Z, Mahdavi SA, *et al*: Global systematic review of primary immunodeficiency registries. *Expert Rev Clin Immunol* 16: 717-732, 2020.
- Kilic SS, Ozel M, Hafizoglu D, Karaca NE, Aksu G and Kutukculer N: The prevalences (Correction) and patient characteristics of primary immunodeficiency diseases in Turkey-two centers study. *J Clin Immunol* 33: 74-83, 2013.
- Cheng J, Myers TG, Levinger C, Kumar P, Kumar J, Goshu BA, Bosque A and Catalfamo M: IL-27 induces IFN/STAT1-dependent genes and enhances function of TIGIT+ HIVGag-specific T cells. *iScience* 25: 103588, 2022.
- Kamiya S, Owaki T, Morishima N, Fukai F, Mizuguchi J and Yoshimoto T: An indispensable role for STAT1 in IL-27-induced T-bet expression but not proliferation of naive CD4+ T cells. *J Immunol* 173: 3871-3877, 2004.
- Wang Y, Ma CS, Ling Y, Bousfiha A, Camcioglu Y, Jacquot S, Payne K, Crestani E, Roncagalli R, Belkadi A, *et al*: Dual T cell- and B cell-intrinsic deficiency in humans with biallelic RLTFR mutations. *J Exp Med* 213: 2413-2435, 2016.
- Tolomeo M, Cavalli A and Cascio A: STAT1 and its crucial role in the control of viral infections. *Int J Mol Sci* 23: 4095, 2022.

25. Uzel G, Sampaio EP, Lawrence MG, Hsu AP, Hackett M, Dorsey MJ, Noel RJ, Verbsky JW, Freeman AF, Janssen E, *et al.*: Dominant gain-of-function STAT1 mutations in FOXP3 wild-type immune dysregulation-polyendocrinopathy-enteropathy-X-linked-like syndrome. *J Allergy Clin Immunol* 31: 1611-1623, 2013.
26. Sampaio EP, Hsu AP, Pechacek J, Bax HI, Dias DL, Paulson ML, Chandrasekaran P, Rosen LB, Carvalho DS, Ding L, *et al.*: Signal transducer and activator of transcription 1 (STAT1) gain-of-function mutations and disseminated coccidioidomycosis and histoplasmosis. *J Allergy Clin Immunol* 131: 1624-1634, 2013.
27. Tsumura M, Okada S, Sakai H, Yasunaga S, Ohtsubo M, Murata T, Obata H, Yasumi T, Kong XF, Abhyankar A, *et al.*: Dominant-negative STAT1 SH2 domain mutations in unrelated patients with Mendelian susceptibility to mycobacterial disease. *Hum Mutat* 33: 1377-1387, 2012.
28. Frede N, Rojas-Restrepo J, Caballero Garcia de Oteyza A, Buchta M, Hübscher K, Gámez-Díaz L, Proietti M, Saghafi S, Chavoshzadeh Z, Soler-Palacin P, *et al.*: Genetic analysis of a cohort of 275 patients with Hyper-IgE syndromes and/or chronic mucocutaneous candidiasis. *J Clin Immunol* 41: 1804-1838, 2021.
29. Lv G, Sun G, Wu P, Du X, Zeng T, Wen W, Zhou L, An Y, Tang X, He T, *et al.*: Novel mutations of TYK2 leading to divergent clinical phenotypes. *Pediatr Allergy Immunol* 33: e13671, 2022.
30. Levy R, Okada S, Béziat V, Moriya K, Liu C, Chai LY, Migaud M, Hauck F, Al Ali A, Cyrus C, *et al.*: Genetic, immunological, and clinical features of patients with bacterial and fungal infections due to inherited IL-17RA deficiency. *Proc Natl Acad Sci USA* 113: E8277-E8285, 2016.
31. Bonilla FA and Geha RS: Update on primary immunodeficiency diseases. *J Allergy Clin Immunol* 117: S435-S441, 2006.
32. Okada S, Puel A, Casanova JL and Kobayashi M: Chronic mucocutaneous candidiasis disease associated with inborn errors of IL-17 immunity. *Clin Transl Immunology* 5: e114, 2016.
33. Imai K, Slupphaug G, Lee WI, Revy P, Nonoyama S, Catalan N, Yel L, Forveille M, Kavli B, Krokkan HE, *et al.*: Human uracil-DNA glycosylase deficiency associated with profoundly impaired immunoglobulin class-switch recombination. *Nat Immunol* 4: 1023-1028, 2003.
34. Garcia-Laorden MI, Sole-Violan J, Rodriguez de Castro F, Aspa J, Briones ML, Garcia-Saavedra A, Rajas O, Blanquer J, Caballero-Hidalgo A, Marcos-Ramos JA, *et al.*: Mannose-binding lectin and mannose-binding lectin-associated serine protease 2 in susceptibility, severity, and outcome of pneumonia in adults. *J Allergy Clin Immunol* 122: 368-374, 374.e1-2, 2008.
35. Ling Y, Cypowyj S, Aytekin C, Galicchio M, Camcioglu Y, Nepesov S, Ikinciogullari A, Dogu F, Belkadi A, Levy R, *et al.*: Inherited IL-17RC deficiency in patients with chronic mucocutaneous candidiasis. *J Exp Med* 212: 619-631, 2015.
36. Miao J, Zhang J, Huang X, Wu N, Wu D and Shen M: NLRP12-associated autoinflammatory disease in Chinese adult patients: A single-centre study. *RMD Open* 9: e003598, 2023.
37. Wang W, Zhou Y, Zhong LQ, Li Z, Jian S, Tang XY and Song HM: The clinical phenotype and genotype of NLRP12-autoinflammatory disease: A Chinese case series with literature review. *World J Pediatr* 16: 514-519, 2020.
38. Kingdom R and Wright CF: Incomplete penetrance and variable expressivity: From clinical studies to population cohorts. *Front Genet* 25: 920390, 2022.
39. Zhang D, Su G, Hao S, Lai J and Feng S: Paediatric autoimmune diseases with ELANE mutations associated with neutropenia. *Pediatr Rheumatol Online J* 21: 41, 2023.
40. Skokowa J, Germeshausen M, Zeidler C and Welte K: Severe congenital neutropenia: Inheritance and pathophysiology. *Curr Opin Hematol* 14: 22-28, 2007.
41. Ferreira RC, Pan-Hammarström Q, Graham RR, Gateva V, Fontán G, Lee AT, Ortmann W, Urcelay E, Fernández-Arquero M, Núñez C, *et al.*: Association of IFIH1 and other autoimmunity risk alleles with selective IgA deficiency. *Nat Genet* 42: 777-780, 2010.
42. Song J, Li M, Li C, Liu K, Zhu Y and Zhang H: Friend or foe: RIG-I like receptors and diseases. *Autoimmun Rev* 21: 103161, 2022.
43. Huang P, Wu JJ, Zhang JW, Hou YQ, Zhu P, Yin R, Yu RB, Zhang Y, Yue M and Hou W: Genetic variants of IFIH1 and DHX58 affect the chronicity of hepatitis C in the Chinese Han population. *PeerJ* 11: e14740, 2023.
44. Zhang J, Liu X, Meng Y, Wu H, Wu Y, Yang B and Wang L: Autoimmune disease associated IFIH1 single nucleotide polymorphism related with IL-18 serum levels in Chinese systemic lupus erythematosus patients. *Sci Rep* 8: 9442, 2018.
45. Xiao W, Feng J, Long H, Xiao B and Luo ZH: Case report: Aicardi-Goutières syndrome and singleton-merten syndrome caused by a gain-of-function mutation in IFIH1. *Front Genet* 12: 660953, 2021.
46. Bursztejn AC, Briggs TA, del Toro Duany Y, Anderson BH, O'Sullivan J, Williams SG, Bodemer C, Freitag S, Gebhard F, Leheup B, *et al.*: Unusual cutaneous features associated with a heterozygous gain-of-function mutation in IFIH1: Overlap between Aicardi-Goutières and Singleton-Merten syndromes. *Br J Dermatol* 173: 1505-1513, 2015.
47. Asgari S, Schlapbach LJ, Anchisi S, Hammer C, Bartha I, Junier T, Mottet-Osman G, Posfay-Barbe KM, Longchamp D, Stocker M, *et al.*: Severe viral respiratory infections in children with IFIH1 loss-of-function mutations. *Proc Natl Acad Sci USA* 114: 8342-8347, 2017.
48. Cananzi M, Wohler E, Marzollo A, Colavito D, You J, Jing H, Bresolin S, Gaio P, Martin R, Mescoli C, *et al.*: IFIH1 loss-of-function variants contribute to very early-onset inflammatory bowel disease. *Hum Genet* 140: 1299-1312, 2021.
49. Shen M, Tang L, Shi X, Zeng X and Yao Q: NLRP12 autoinflammatory disease: A Chinese case series and literature review. *Clin Rheum* 36: 1661-1667, 2017.
50. Gruber C and Bogunovic D: Incomplete penetrance in primary immunodeficiency: A skeleton in the closet. *Hum Genet* 139: 745-757, 2020.
51. Erzurumluoglu AM, Shihab HA, Rodriguez S, Gaunt TR and Day IN: Importance of genetic studies in consanguineous populations for the characterization of novel human gene functions. *Ann Hum Genet* 80: 187-196, 2016.
52. Rodríguez-Ubreva J, Calvillo CL, Satter LR and Ballestar E: Interplay between epigenetic and genetic alterations in inborn errors of immunity. *Trends Immunol* 44: 902-916, 2023.
53. Peng XP, Al-Ddafari MS, Caballero-Oteyza A, El Mezouar C, Mrovecova P, Dib SE, Massen Z, Smahi MC, Faiza A, Hassaine RT, *et al.*: Next generation sequencing (NGS)-based approach to diagnosing Algerian patients with suspected inborn errors of immunity (IEIs). *Clin Immunol* 256: 109758, 2023.
54. Romano R, Cillo F, Moracas C, Pignata L, Nannola C, Toriello E, De Rosa A, Cirillo E, Coppola E, Giardino G, *et al.*: Epigenetic alterations in inborn errors of immunity. *J Clin Med* 11: 1261, 2022.
55. Isailovic N, Daigo K, Mantovani A and Selmi C: Interleukin-17 and innate immunity in infections and chronic inflammation. *J Autoimmun* 60: 1-11, 2015.



Copyright © 2024 Obeidat *et al.* This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.