





Complete Genome Sequence of a Rodent Torque Teno Virus in Hainan Island, China

Yue Wu, a.b.c Shanshan Wang, a.b.c Yunchun Chen, d Lihua Li, Wengi Wang, G Huanhuan Zhou, G You Zhang, a.b.c Lei Zhang, e Xiuji Cui,^c Gang Lv,^{a,b,c} Feifei Yin^{a,b,c}

^aHainan Medical University–The University of Hong Kong Joint Laboratory of Tropical Infectious Diseases, Hainan Medical University, Haikou, Hainan, China

bKey Laboratory of Translation Medicine Tropical Diseases, Hainan Medical University, Haikou, Hainan, China

^cDepartment of Pathogen Biology, Hainan Medical University, Haikou, Hainan, China

dHaikou Branch, Yueyang Hospital of Integrated Traditional Chinese and Western Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai, China

eCore Facility and Technical Support of Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan, Hubei, China

ABSTRACT Torque teno virus (TTV) has been reported in a wide range of mammals. In this study, we sequenced and analyzed the complete genome of a genetic variant of a rodent TTV, RoTTV3-HMU1 (Hainan Medical University 1). The virus was found in a rat (Rattus norvegicus) in a residential area of Hainan Island, China.

orque teno virus (TTV) is a nonenveloped single-stranded DNA virus (1). TTV is found widely distributed throughout the world and infects an extensive range of mammals (1-7). The possible role of TTV in disease has not been fully elucidated. Rodent TTV (RoTTV) was initially identified in the United Kingdom in populations of wild rodents in 2014, and the genotypes are RoTTV1 and RoTTV2 (8). The same research team further detected RoTTV2 in commonly used laboratory rats (9). Recently, a novel genotype of RoTTV, RoTTV3, was found in murine rodents and house shrews in 6 regions of 4 provinces in China (10).

In this study, a genetic variant of RoTTV3, RoTTV3-HMU1 (Hainan Medical University 1), was identified in a rat (Rattus norvegicus) captured in a residential area of Haikou City, Hainan Province, southern China. Viral DNA was extracted from the liver tissue with the QIAamp MinElute virus spin kit (Qiagen), and sequence-independent amplification of viral nucleic acids was performed as described previously (11). The amplicons in the 250 to 500-bp range were purified with a gel extraction kit (Tiangen). Five hundred nanograms of DNA were fragmented with Covaris S2 shearing and subjected to high-throughput paired-end 2 × 100-bp sequencing on an Illumina HiSeq 2000 instrument. After cleaning with Trimmomatic using standard parameters, reads were de novo assembled with Trinity version 2.0.6 (12, 13, 14). The contigs were compared with the NCBI nucleotide database, and 5 contigs ($N_{50} = 361$) were matched to RoTTV with a coverage of 0.68. Primers were designed to cover the genome by PCR amplification and Sanger sequencing. The genome was annotated with the NCBI ORFfinder and FGENESV0 (15, 16). Phylogenetic analysis was implemented with the neighbor-joining method in the MEGA6 software package (17).

The complete genome of RoTTV3-HMU1 is 2,570 nucleotides (nt) long with a G+C content of 46.93%. The genomic organization of RoTTV3-HMU1 is consistent with that of other RoTTVs, and the genome size and length of the open reading frames (ORFs) showed close similarity to those of RoTTV3. RoTTV3-HMU1 encoded 3 unidirectional overlapping ORFs. ORF1, ORF2, and ORF3 encoded proteins of 574, 79, and 98 amino acids (aa), respectively. The genome of the RoTTV3-HMU1 virus was most closely related

Received 31 July 2018 Accepted 17 October 2018 **Published** 15 November 2018

Citation Wu Y, Wang S, Chen Y, Li L, Wang W, Zhou H, Zhang Y, Zhang L, Cui X, Lv G, Yin F. 2018. Complete genome sequence of a rodent torque teno virus in Hainan Island, China. Microbiol Resour Announc 7:e01074-18. https://doi.org/10.1128/MRA.01074-18.

Editor Iddo Friedberg, Iowa State University

Copyright © 2018 Wu et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Feifei Yin. yinfeifeiff@163.com.



to RN_2_Se15 (GenBank accession no. KM668486), with an identity of 95% at nucleotide level. They were also closely related at their ORF encoded proteins (ORF1, 97%; ORF2, 96%; ORF3, 89%). Phylogenetic analysis based on both ORF1 and the total genome sequence placed RoTTV3-HMU1 in the clad RoTTV3 of the RoTTV.

Hainan Island is isolated from mainland China by sea, but the same RoTTV genotype was identified on both the island and the mainland. The detection of RoTTV3-HMU1 may contribute to a better understanding of the origin and evolution of RoTTV.

Data availability. The genomic sequence of RoTTV3-HMU1 has been deposited in GenBank under accession no. MF688246. This whole-genome shotgun project and the assembly reads have been deposited in GenBank under the accession no. SRP158097 and SRR7700917, respectively.

ACKNOWLEDGMENTS

This study was funded by the National Natural Science Foundation of China (grant no. 31460017, 81860367, 81672072, and 81760378), the Higher Educational Scientific Research Project of Hainan Province, the Ministry of Education (grant no. Hnky2017ZD-16 and Hnkyzx2014-08), and the Key Research and Development Plan of Hainan Province, Ministry of Science and Technology (grant no. ZDYF2017091).

REFERENCES

- 1. Hino S, Miyata H. 2007. Torque teno virus (TTV): current status. Rev Med Virol 17:45–57. https://doi.org/10.1002/rmv.524.
- Abe K, Inami T, Ishikawa K, Nakamura S, Goto S. 2000. TT virus infection in nonhuman primates and characterization of the viral genome: identification of simian TT virus isolates. J Virol 74:1549–1553. https://doi .org/10.1128/JVI.74.3.1549-1553.2000.
- Leary TP, Erker JC, Chalmers ML, Desai SM, Mushahwar IK. 1999. Improved detection systems for TT virus reveal high prevalence in humans, non-human primates and farm animals. J Gen Virol 80:2115–2120. https://doi.org/10.1099/0022-1317-80-8-2115.
- Martínez L, Kekarainen T, Sibila M, Ruiz-Fons F, Vidal D, Gortázar C, Segalés J. 2006. Torque teno virus (TTV) is highly prevalent in the European wild boar (Sus scrofa). Vet Microbiol 118:223–229. https://doi.org/10.1016/j.vetmic.2006.07.022.
- Ng TFF, Suedmeyer WK, Wheeler E, Gulland F, Breitbart M. 2009. Novel anellovirus discovered from a mortality event of captive California sea lions. J Gen Virol 90:1256–1261. https://doi.org/10.1099/vir.0.008987-0.
- Okamoto H, Takahashi M, Nishizawa T, Tawara A, Fukai K, Muramatsu U, Naito Y, Yoshikawa A. 2002. Genomic characterization of TT viruses (TTVs) in pigs, cats and dogs and their relatedness with species-specific TTVs in primates and tupaias. J Gen Virol 83:1291–1297. https://doi.org/ 10.1099/0022-1317-83-6-1291.
- Spandole S, Cimponeriu D, Berca LM, Mihăescu G. 2015. Human anelloviruses: an update of molecular, epidemiological and clinical aspects. Arch Virol 160:893–908. https://doi.org/10.1007/s00705-015-2363-9.
- Nishiyama S, Dutia BM, Stewart JP, Meredith AL, Shaw DJ, Simmonds P, Sharp CP. 2014. Identification of novel anelloviruses with broad diversity in UK rodents. J Gen Virol 95:1544–1553. https://doi.org/10.1099/vir.0 .065219-0.
- Nishiyama S, Dutia BM, Sharp CP. 2015. Complete genome sequences of novel anelloviruses from laboratory rats. Genome Announc 3:e01262-14. https://doi.org/10.1128/genomeA.01262-14.

- Xiong Y-Q, Mo Y, Chen M-J, Cai W, He W-Q, Chen Q. 2018. Detection and phylogenetic analysis of torque teno virus (TTV) carried by murine rodents and house shrews in China. Virology 516:189–195. https://doi .org/10.1016/j.virol.2018.01.017.
- Ge X, Li Y, Yang X, Zhang H, Zhou P, Zhang Y, Shi Z. 2012. Metagenomic analysis of viruses from bat fecal samples reveals many novel viruses in insectivorous bats in China. J Virol 86:4620–4630. https://doi.org/10 .1128/JVI.06671-11.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, Adiconis X, Fan L, Raychowdhury R, Zeng Q, Chen Z, Mauceli E, Hacohen N, Gnirke A, Rhind N, di Palma F, Birren BW, Nusbaum C, Lindblad-Toh K, Friedman N, Regev A. 2011. Full-length transcriptome assembly from RNA-Seq data without a reference genome. Nat Biotechnol 29:644–652. https://doi.org/10.1038/nbt.1883.
- Borodina T, Adjaye J, Sultan M. 2011. A strand-specific library preparation protocol for RNA sequencing. Methods Enzymol 500:79 –98. https://doi.org/10.1016/B978-0-12-385118-5.00005-0.
- Wheeler DL, Church DM, Federhen S, Lash AE, Madden TL, Pontius JU, Schuler GD, Schriml LM, Sequeira E, Tatusova TA, Wagner L. 2003. Database resources of the National Center for Biotechnology. Nucleic Acids Res 31:28–33. https://doi.org/10.1093/nar/gkg033.
- Solovyev VV, Salamov AA. 1999. INFOGENE: a database of known gene structures and predicted genes and proteins in sequences of genome sequencing projects. Nucleic Acids Res 27:248–250. https://doi.org/10 .1093/nar/27.1.248.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. Mol Biol Evol 30:2725–2729. https://doi.org/10.1093/molbev/mst197.

mra.asm.org 2

Volume 7 Issue 19 e01074-18