

Video Article

Assessment of Pulmonary Capillary Blood Volume, Membrane Diffusing Capacity, and Intrapulmonary Arteriovenous Anastomoses During Exercise

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Abstract

Exercise is a stress to the pulmonary vasculature. With incremental exercise, the pulmonary diffusing capacity (DL_{CO}) must increase to meet the increased oxygen demand; otherwise, a diffusion limitation may occur. The increase in DL_{CO} with exercise is due to increased capillary blood volume (Vc) and membrane diffusing capacity (Dm). Vc and Dm increase secondary to the recruitment and distension of pulmonary capillaries, increasing the surface area for gas exchange and decreasing pulmonary vascular resistance, thereby attenuating the increase in pulmonary arterial pressure. At the same time, the recruitment of intrapulmonary arteriovenous anastomoses (IPAVA) during exercise may contribute to gas exchange impairment and/or prevent large increases in pulmonary artery pressure.

We describe two techniques to evaluate pulmonary diffusion and circulation at rest and during exercise. The first technique uses multiple-fraction of inspired oxygen (F_{iO_2}) DL_{CO} breath holds to determine Vc and Dm at rest and during exercise. Additionally, echocardiography with intravenous agitated saline contrast is used to assess IPAVAs recruitment.

Representative data showed that the DL_{CO} , Vc, and Dm increased with exercise intensity. Echocardiographic data showed no IPAVA recruitment at rest, while contrast bubbles were seen in the left ventricle with exercise, suggesting exercise-induced IPAVA recruitment.

The evaluation of pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA recruitment using echocardiographic methods is useful to characterize the ability of the lung vasculature to adapt to the stress of exercise in health as well as in diseased groups, such as those with pulmonary arterial hypertension and chronic obstructive pulmonary disease.

Video Link

The video component of this article can be found at <https://www.jove.com/video/54949/>

Introduction

During exercise, cardiac output can increase up to six-fold above resting values¹. Given that the lungs are the only organ to receive 100% of the cardiac output, exercise presents a considerable stress to the pulmonary system. With incremental exercise, pulmonary diffusing capacity (DL_{CO}) must increase to meet the increased oxygen demand². From rest to peak exercise, DL_{CO} can increase to up to 150% of resting values without reaching an upper limit with respect to cardiac output^{3,4,5}. The increase in diffusing capacity occurs as a result of increases in membrane diffusing capacity (Dm) and capillary blood volume (Vc), secondary to the recruitment and distension of pulmonary capillaries⁵.

Roughton and Forster (1957) developed a technique to partition Dm and Vc⁷ by modulating the fraction of inspired oxygen (F_{iO_2}) during a standard diffusion capacity for carbon monoxide test (DL_{CO}). Oxygen and carbon monoxide (CO) competitively bind to heme sites on hemoglobin, such that increasing F_{iO_2} will decrease the DL_{CO} ^{8,9}. By modulating the F_{iO_2} during a standard DL_{CO} maneuver, this relationship can be exploited to measure Vc and Dm⁷. We have recently adapted this technique to be used during exercise⁵. Similar to previous work, we have found that DL_{CO} continuously increases up to peak exercise secondarily to increases in both Vc and Dm⁵. Interestingly, we have found that in endurance-trained athletes who have a greater oxygen consumption and thus a greater need for diffusing capacity, there is an increase in the DL_{CO} at peak exercise, secondary to an increased Dm, and not Vc, suggesting a potential adaptation in the pulmonary membrane of the athlete⁵.

The increases in Vc and Dm during exercise are accomplished by an increase in pulmonary artery pressure, which results in the recruitment and distension of pulmonary capillaries previously hypo-perfused at rest^{4,10}. This results in an increase in the cross-sectional area of the pulmonary capillary network, thereby decreasing pulmonary vascular resistance and attenuating the increase in pulmonary artery pressure.

Studies using agitated saline contrast echocardiography have shown evidence of intrapulmonary arteriovenous anastomoses (IPAVA) recruitment during exercise^{11,12,13,14}. The significance of IPAVA recruitment is not yet clear, and while some studies suggest that they may contribute to gas exchange impairment^{12,14} and may serve to unload the right ventricle^{11,12}, the topic remains controversial^{15,16}. Further, while the exact mechanism of IPAVA recruitment is not known, we have found that increasing cardiac output, as well as exogenous dopamine, causes IPAVA recruitment at rest¹⁷. An acutely-increasing pulmonary artery pressure¹⁸ or dopamine blockade does not appear to significantly affect IPAVA recruitment during exercise¹¹. There is speculation that these larger-diameter IPAVA vessels may help to protect the pulmonary capillaries from the large increases in pulmonary artery pressure by reducing pulmonary vascular resistance^{12,17,19,20,21}.

When combined with the evaluation of Vc and Dm, agitated saline contrast echocardiography is a valuable tool to examine the adaptation of the pulmonary circulation to the stress of exercise^{22,23}.

Protocol

This protocol follows the guidelines of the human research ethics board at the University of Alberta and conforms to the standards set by the latest revision of the *Declaration of Helsinki*.

1. Graded Exercise Test (VO_{2peak})

1. Obtain written, informed consent from the subject. Have the subject read and answer the questions listed on the Physical Activity Readiness Questionnaire+ (PAR-Q+) to determine their readiness for exercise²⁴.
2. Adjust the seat height of the cycle ergometer in accordance to subject preference. Place four electrocardiogram (ECG) electrodes on the back of the patient according to standard 3-lead ECG placement, with modified limb leads to measure the heart rate (HR)²⁵.
3. Insert the mouthpiece into the subject's mouth to measure the exhaled gas and ventilation throughout the test using a metabolic measurement system²⁵.
NOTE: The metabolic system will measure real-time oxygen consumption (VO₂), carbon dioxide production (VCO₂), ventilation (V_E), heart rate (HR), and end tidal CO₂ (P_{ET}CO₂).
4. Following 2 min of collection of baseline data, instruct the subject to start cycling with an initial workload of 50 watt, to maintain a consistent cadence of ≥60 RPM. Increase the workload in 25 W steps every 2 min, until the subject reaches volitional exhaustion or requests to stop the test²⁵.

2. Multiple Fraction of Inspired Oxygen (F_IO₂) Diffusing Capacity (DL_{CO}) Method⁷

1. Calculate the workloads corresponding to 30%, 50%, 70%, and 90% of the VO_{2peak} using the peak VO₂ obtained in the graded exercise test. At least 48 h after the graded exercise test, have the subject return to the laboratory for DLCO maneuvers.
2. Do not exceed 12 DLco tests per day, as carboxyhemoglobin (COHb) build-up can occur with repeated testing⁵. Therefore, perform testing on multiple days based on the number of exercise workloads to be conducted and the quality of the DLCO data.
3. Prepare pre-breathing gases by attaching a tank of 100% O₂ gas and a tank of medical-grade air (21% O₂ and 79% N₂) to an air blender system. Fill two 60 L non-diffusing Douglas bags, one containing 40% O₂, and one containing 60% O₂, using the air blender system.
4. Set up two large-bore, three-way stopcock valves that will allow for the modulation of inhaled gas mixtures. These will be referred to as the "pre-breath valves."
5. Connect the Douglas bags to the valve system using flexible, non-compressible tubing. Connect the valve system to a two-way, T-shaped non-rebreathing valve connected to the test gas intake assembly of the mass flow sensor of the metabolic measurement system.
6. For resting measurements, have the subject seated upright, with both feet flat on the floor. For exercise trials, ensure that the subject is in a steady state by monitoring HR using the ECG (HR ± 3 bpm for steady state).
NOTE: Steady state may not be reached at 90% of the VO_{2peak}; thus, begin the measurement once the subject has reached the HR equivalent to 90% of the VO_{2peak} on the graded exercise test.
7. Collect a single drop of capillary blood via a finger prick and analyze it for hemoglobin concentration. Then, adjust all subsequent DL_{CO} for [Hb] using the following equation²⁶:
$$DL_{CO_{adj}} = DL_{CO} \times \frac{10.22 + [Hb]}{1.7 \times [Hb]}$$
8. Select an F_IO₂ (21%, 40%, or 60%) at random by switching the pre-breathe valves to the desired orientation. Choose the corresponding F_IO₂-DL_{CO} gas by turning the DL_{CO} gas valve selector (see **Figure 1C**).
9. Instruct the subject to affix the nose clips and to breathe normally into the mouthpiece for five breaths from the Douglas bag corresponding to the respective F_IO₂.
10. Instruct the subject to expire to residual volume. When the lung volume plateaus at residual volume, have the subject inhale the DL_{CO} gas mixture to total lung capacity and hold their breath for 6 s before exhaling to residual volume.
11. Monitor the methane tracing during the exhalation to ensure that the slope is horizontal, as this indicates that the CO test gas is well equilibrated in the lung.
NOTE: Alveolar volume (V_A) and breath hold time are calculated automatically and reported by the metabolic measurement system.
12. Ensure that the V_A for each DLco maneuver is within 5% of previous trials. Similarly, breath hold time should be 6.0 ± 0.3 s. If not, repeat the maneuver.
13. Wait 4 min to allow residual carbon monoxide to wash out, and then repeat steps 2.8 - 2.11 for each remaining F_IO₂ at rest.

- At least 48 h later, repeat steps 2.9 - 2.15 during steady state at each exercise intensity (30%, 50%, 70%, and 90% of the VO_{2peak}) for each $F_I O_2$. Reduce the workload between the breath holds at 90% of the VO_{2peak} workload to recover the subject.
- Wait 2 min between DLco tests during exercise to clear alveolar CO during exercise. Do not exceed 12 DLco tests per day to avoid carboxyhemoglobin (COHb) build-up⁵.

3. Calculating Pulmonary Capillary Blood Volume and Membrane Diffusing Capacity

- Calculate the alveolar partial pressure of O_2 ($P_A O_2$) using the following equation

$$P_A O_2 = F_I O_2 (P_{BAR} - P_{H_2O}) - P_a CO_2 \times \frac{(1 - F_I O_2)}{RER}$$

NOTE: $F_I O_2$ is the fraction of inspired O_2 , P_{BAR} is the atmospheric pressure, P_{H_2O} is the water vapor pressure, $P_a CO_2$ is the pressure of arterial CO_2 , and RER is the respiratory exchange ratio.

- Estimate the RER and $P_a CO_2$ using the measured 30-s average $P_{ET} CO_2$ and RER for the respective exercise intensity from the data obtained in the previous graded exercise test.
- Calculate θ_{CO} using the following equation⁷. $\frac{1}{\theta_{CO}} = 0.0058 \times P_A O_2 + 0.73$
- Graph the relationship between $1/DL_{COadj}$ and $1/\theta_{CO}$ for each $F_I O_2$ and calculate the regression equation.
NOTE: The minimum acceptable r^2 value is 0.95, and DL_{CO} maneuvers should be repeated when r^2 values are outside of this range²¹.

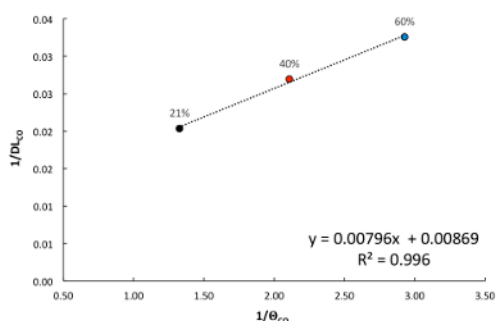


Figure 2: Representative Graph of $1/DL_{CO}$ versus $1/\theta_{CO}$ at Peak Exercise. The relationship between $1/DL_{CO}$ and $1/\theta_{CO}$ is plotted for three breath holds at the various $F_I O_2$ (21%, 40%, and 60%). The calculation of V_c and D_m are derived from the regression equation for the relationship above. The inverse of the slope ($1/0.00796$) of the line gives the value for V_c (125.5 mL), and the inverse of the y-intercept ($1/0.00869$) gives the value for D_m ($115.0 \text{ mL} \cdot \text{min}^{-1} \cdot \text{mmHg}^{-1}$). [Please click here to view a larger version of this figure.](#)

- Calculate V_c by taking the inverse of the slope of the regression equation between $1/DL_{CO}$ and $1/\theta_{CO}$. Calculate D_m by taking the inverse of the y-intercept of the equation.

4. Intrapulmonary Arteriovenous Anastomosis Recruitment

- On a separate day from the DL_{CO} data collection, insert a 20-gauge intravenous (IV) catheter into an antecubital vein and attach it to a three-way stopcock via a 6-in IV extension tube for the injection of agitated saline for contrast echocardiography^{11,17}.



Figure 3: Agitated Saline Contrast Setup. An IV catheter is placed in the antecubital space and is connected to a three-way stopcock via a 6-in extension. Two 10 mL syringes are attached to the stopcock to create the contrast solution, which contains 10 mL of saline and 0.5 mL of room air. [Please click here to view a larger version of this figure.](#)

2. Connect two 10 mL syringes to the three-way stopcock. Combine 10 mL of 0.9% sterile saline with 0.5 mL of air, and forcefully agitate it through the three-way stopcock, back and forth between the two syringes, to form fine, suspended bubbles until the sonographer is ready for contrast.
3. Have an experienced sonographer or cardiologist obtain a standard apical four-chamber view of the heart. At rest, have the echocardiographer evaluate the intra-atrial septum and ventricular septum for an intra-cardiac shunt with standard echocardiographic and color Doppler imaging.
 1. If no intra-cardiac shunt is detected, instruct the subject to perform a Valsalva maneuver during the contrast injection to evaluate for a patent foramen ovale (PFO)^{11,17}. Repeat the measurement during non-Valsalva.
4. Inject the contrast while the sonographer maintains the four-chamber view. Record 15 cardiac cycles following the detection of contrast in the right ventricle.
5. Repeat the contrast-enhanced imaging during steady-state exercise at 30%, 50%, and 70% of the VO_{2peak} . As steady state cannot be reached at 90% of the VO_{2peak} , begin the imaging once the target HR, identified by the HR at 90% of the VO_{2peak} during the graded exercise test, is reached.

NOTE: The time between exercise intensities depends on the clearance of contrast from both ventricles, ≥ 2 min.
6. Have an echocardiographer who is blinded to experimental conditions interpret the agitated saline contrast echocardiograms according to a previously-described scoring system^{17,27}.

NOTE: Scoring is based on the maximum number of contrast bubbles visible within the left ventricle (LV) in a single echocardiographic frame, as follows: no contrast bubbles in the LV = 0, ≤ 3 bubbles = 1, 4 - 12 bubbles = 2, > 12 bubbles = 3.

NOTE: The appearance of contrast in the left ventricle after five cardiac cycles suggests an IPAVA. An intracardiac shunt is graded by the appearance of contrast in less than five cardiac cycles²⁷.

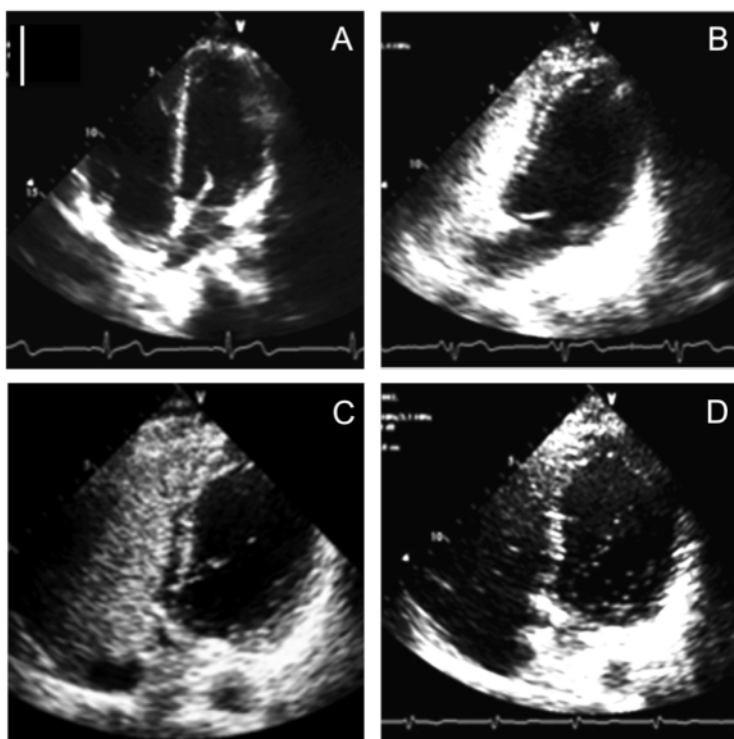


Figure 4: Representative Images for IPAVA Scoring. The scale is 5 cm (solid white line). (A) Pre contrast injection. (B) IPAVA score = 0. (C) IPAVA score = 1. (D) IPAVA score = 3. [Please click here to view a larger version of this figure.](#)

Representative Results

The effect of increasing exercise intensity on oxygen consumption, diffusing capacity, pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA score is shown in **Table 1**. VO_2 , DL_{CO} , V_c , and D_m increase in response to increasing power output.

Figure 2 shows a representative calculation of V_c and D_m using the multiple $F_I O_2$ - DL_{CO} technique during exercise. DL_{CO} decreases with increasing $F_I O_2$, and this relationship is exploited to partition V_c and D_m . Calculating the inverse of the slope of $1/DL_{CO}$ versus $1/\theta_{CO}$ results in the V_c , and the inverse of the y-intercept yields the value for the D_m . As expected, both the V_c and D_m increase during exercise compared to resting values.

The results show that these techniques can be used to assess the pulmonary vasculature response during exercise. The multiple- $F_{I}O_2$ DL_{CO} and agitated saline contrast echocardiography method provides investigators with more insight into the contributions of pulmonary capillary and membrane recruitment to the overall diffusion capacity and could supplement traditional pulmonary function testing in the clinical setting. Failure to increase Vc or Dm during exercise would lead to a diffusion limitation and hypoxemia. For example, a low DL_{CO} secondary to a low Vc would indicate changes to the pulmonary capillaries; similarly, a decreased Dm would indicate changes to the pulmonary membrane.

Figure 4 shows representative tracings of four-chamber contrast echocardiographs. With increasing exercise intensity, the IPAVA score increases from 0 (*i.e.*, no evidence of IPAVAs) at rest to 3 at the highest exercise intensity (**Table 1**). Previous work has shown that exercise increases the IPAVA score^{11,12,14}, but there is no consensus as to how these IPAVAs are recruited. There is evidence that IPAVAs can be recruited pharmacologically at rest with dopamine^{17,28}, as well as by increasing cardiac output with dobutamine^{17,28} and epinephrine²⁸. Inotropes such as dopamine and epinephrine are of particular interest, as they increase endogenously during exercise²⁹. Furthermore, there is some evidence that IPAVA recruitment may be important to exercise hemodynamics, in that the absence of IPAVAs appears to result in greater pulmonary artery pressure, decreased cardiac output, and decreased peak power output¹². Thus, this technique may be used in studies examining individuals with pulmonary artery hypertension.

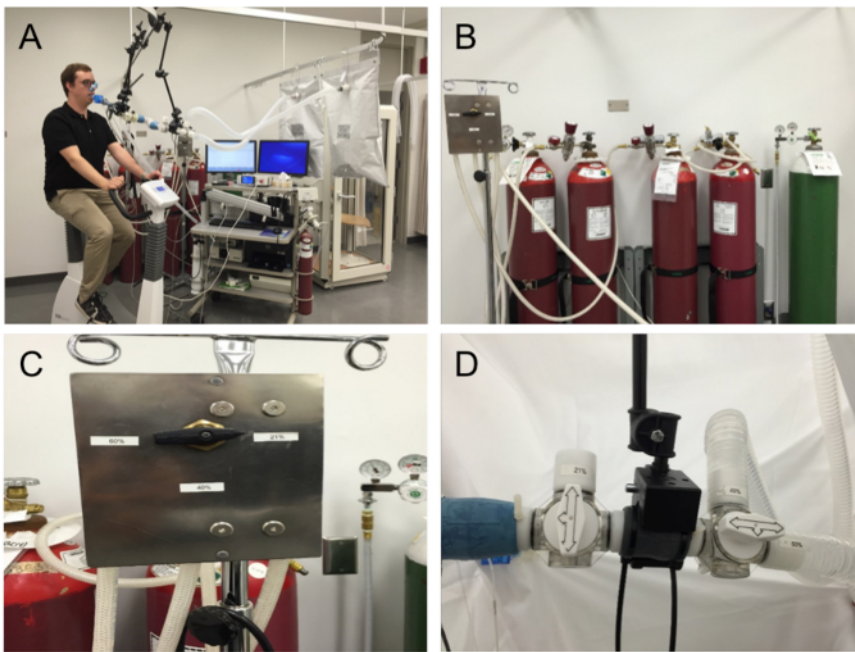


Figure 1: Multiple $F_{I}O_2$ DL_{CO} Setup. (A) Setup overview. (B) Compressed-gas cylinders containing 21%, 40%, and 60% O_2 with 0.3% CO , 0.3% methane, and balance nitrogen, as well as a supplemental oxygen compressed-gas cylinder. (C) Three-way valve selector for the three $F_{I}O_2$ DL_{CO} tanks. (D) Valve switch for three-way valves in the series for the selection of $F_{I}O_2$ for pre-breathing. [Please click here to view a larger version of this figure.](#)

| | Rest | 30% | 50% | 70% | 90% |
|---|------|------|------|-------|-------|
| VO_2 ($mL \cdot kg^{-1} \cdot min^{-1}$) | 6.0 | 17.0 | 28.3 | 40.1 | 50.9 |
| DL_{CO} ($mL \cdot min^{-1} \cdot mmHg^{-1}$) | 36.6 | 45.1 | 48.0 | 51.5 | 60.0 |
| Vc (mL) | 70.8 | 92.7 | 95.2 | 105.0 | 125.5 |
| Dm ($mL \cdot min^{-1} \cdot mmHg^{-1}$) | 75.7 | 87.7 | 96.9 | 101.2 | 115.0 |
| IPAVA Score | 0 | 1 | 1 | 2 | 3 |

Table 1: Representative Data for One Subject at Rest and During Exercise at 30, 50, 70, and 90% of the VO_{2peak} . VO_2 , volume of oxygen consumption relative to body mass; DL_{CO} , diffusing capacity for carbon monoxide; Vc, pulmonary capillary blood volume; Dm, membrane diffusing capacity; IPAVA score, scoring of contrast appearance in the left ventricle after five cardiac cycles. Data modified from Tedjasaputra *et al.* 2016.

Discussion

This method enables the evaluation of the pulmonary diffusing capacity and intrapulmonary arteriovenous anastomosis recruitment during exercise.

Critical steps within the protocol

Although the DL_{CO} breath hold is relatively simple at rest, breath holding during exercise presents a unique challenge to the subject, as it is counter-intuitive, and subjects have a high drive to breathe during exercise. Thus, a good-quality determination of V_c and D_m relies on the rapport and clear communication between the tester and the subject. The tester's technical ability can be quantified with the variability of the alveolar volume ($\pm 5\%$ of previous trials) and a breath-hold time (BHT) of 6.0 ± 0.3 s.

Modifications and troubleshooting

At the conclusion of a V_c/D_m measurement, the tester should quickly graph the three DL_{CO} maneuvers to determine the best-fit line of the data points; the DL_{CO} measured with 21% F_IO₂ should always be greater than that with 40%, which should be greater than that with 60%. If not, it is recommended to check if the valve switch corresponds to the correct testing gas. Similarly, check that the pre-breathing bags are filled with the correct F_IO₂ gas corresponding to the testing gas (**Figure 1B-1D**). Caution should be taken when testing a participant who is a smoker, as elevated COHb levels may underestimate DL_{CO}.

For the IPAVA recruitment assessment, the position of the subject is critical to ensure high-quality image acquisition. It is possible to replace the upright cycle ergometer with a recumbent cycle ergometer to minimize the movement of the subject. However, recumbent cycle exercise will elicit a different metabolic response for a given work rate, and thus the graded exercise test should be repeated on the recumbent cycle ergometer. Scanning of the upper chest may be uncomfortable to some women; in this case, a female sonographer is recommended. Finally, the recommended exercise protocol is designed for a young, healthy individual; accordingly, the exercise protocol can be modified for a different target population.

Limitations of the technique

The principal limitations of the multiple F_IO₂ DL_{CO} technique are the skill of the tester and the ability of the subject to follow commands and to remain calm during the breath hold, as Valsalva or Müllerian maneuvers will affect the measurements. Secondly, the number of breath holds in one session should be limited to 12, due to an increase in CO backpressure, which may affect the V_c and D_m measurement^{5,30} and pose a health risk to the subject. Depending on the research design, it may be necessary to complete the testing across multiple sessions to allow for the clearance of CO and to limit participant fatigue. With good participant coaching and good technical ability, we have determined a satisfactory coefficient of variation between trials for DL_{CO}, V_c, and D_m to be 7%, 8%, and 15%, respectively.

The multiple F_IO₂ DL_{CO} technique assumes that the alveolar O₂ is the same as the capillary O₂, and thus, caution should be exercised when interpreting the data in individuals with known gas exchange impairment.

Agitated saline contrast echocardiographic imaging is limited by the technical ability of the sonographer and the ability of the subject to minimize thoracic movement while exercising. It is also critical that the interpreter of the images be familiar with the scale for scoring IPAVA recruitment according to established procedures (**Figure 4**)²⁷. The significance of a positive saline contrast echocardiography during exercise remains a topic of debate^{15,16}, and there is some discussion that a positive agitated saline contrast in the left ventricle may be secondary to capillary distention, and not IPAVA recruitment. Ongoing work is attempting to resolve this issue.

Significance of the technique with respect to existing/alternative methods

By utilizing these physiological techniques, it is possible to assess the pulmonary vasculature during exercise in a variety of conditions, including in health, in disease, and in drug interventions. Although the quality relies with the ability of the tester, these skills are easily and quickly acquired with proper mentorship and training. The multiple F_IO₂ DL_{CO} method is considered the "gold standard" in the measurement of D_m and V_c³¹. While these measures are not calculated clinically, the values could be used to determine the mechanisms for hypoxemia and exercise intolerance, to predict patient outcomes, and to further characterize diagnosis^{31,32}. Likewise, the agitated saline echocardiography technique is the most widely-used method in determining the recruitment of IPAVAs.

Future applications or directions after mastering this technique

These techniques are applicable for use in a range of experimental conditions and interventions. We demonstrate these techniques during exercise, but they can easily be modified to measure pulmonary vascular responses during a drug infusion, such as dobutamine or dopamine, inotropes known to increase cardiac output¹⁷. Furthermore, it is possible to use these techniques in clinical populations, such as in those with heart failure³⁴ or chronic obstructive pulmonary disease (COPD), in which the DL_{CO} is lower compared to age-matched control subjects³⁵.

Disclosures

The authors declare that they have no competing financial interests.

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