RESEARCH ARTICLE

Fetuin-A in Metabolic syndrome: A systematic review and meta-analysis

Xiongfeng Pan¹, Shi Wu Wen^{2,3}, Prince L. Bestman⁴, Atipatsa C. Kaminga⁵, Kwabena Acheampong⁶, Aizhong Liu¹*

 Department of Epidemiology and Health Statistics, Xiangya School of Public Health, Central South University, Changsha, China, 2 Department of Obstetrics and Gynaecology, University of Ottawa, Ottawa, Ontario, Canada, 3 Ottawa Hospital Research Institute, Ottawa, Ontario, Canada, 4 Association of Liberian Anaesthetist, Monrovia, Liberia, 5 Department of Mathematics and Statistics, Mzuzu University, Mzuzu, Malawi, 6 Department of Public, School of Postgraduate Studies, Adventist University of Africa, Nairobi, Kenya

* lazroy@live.cn

Abstract

Objective

Fetuin-A has been associated with the progression of metabolic syndrome, but previous studies found inconsistent results on the relationship between metabolic syndrome and the concentration of fetuin-A. The aim of this study was to perform a meta-analysis to summarize previous findings on this relationship.

Method

This study was registered with the International Prospective Register of Systematic Reviews PROSPERO (CRD42019129566). Studies examining the relationship between metabolic syndrome and the concentration of circulating fetuin-A were identified using a systematic search in the electronic databases of Embase, PubMed, Web of Science, and Cochrane Library before March 2019. A random effects model was used to summarize the effect size of the association in terms of the standardized mean difference (SMD).

Results

Fourteen eligible studies compared fetuin-A concentrations between 4,551 metabolic syndrome patients and 8,805 controls. The circulating fetuin-A concentration was significantly higher in the metabolic syndrome patients than in the controls (SMD = 0.65, 95% confidence interval (CI): 0.48 to 0.83, Z = 7.18, p<0.001). Besides, circulating fetuin-A was a risk factor for metabolic syndrome (odds ratio 1.23, 95% CI: 1.08 to 1.40).

Conclusion

These findings suggest that fetuin-A may be an important indicator for metabolic syndrome, in which case this may lead to new perspectives in early diagnosis, identification of novel biomarkers, and providing novel targets for pharmacological interventions.



G OPEN ACCESS

Citation: Pan X, Wen SW, Bestman PL, Kaminga AC, Acheampong K, Liu A (2020) Fetuin-A in Metabolic syndrome: A systematic review and meta-analysis. PLoS ONE 15(3): e0229776. https://doi.org/10.1371/journal.pone.0229776

Editor: Omid Beiki, Karolinska Institutet, SWEDEN

Received: September 6, 2019

Accepted: February 13, 2020

Published: March 5, 2020

Copyright: © 2020 Pan et al. This is an open access article distributed under the terms of the <u>Creative</u> Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: This study was funded by the Canadian Institutes of Health Research [CIHR grant # FDN-148438]. Funders play no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

1. Introduction

Metabolic syndrome (MetS) is a new epidemic worldwide and it is associated with an increased risk for developing other chronic disease such as diabetes mellitus [1]. The cardinal features of MetS include the following medical conditions: central obesity, hypertension, glucose intolerance, hyper-triglyceridemia, and low serum high-density lipoprotein (HDL) [2]. It has been proposed recently that biomarkers such as circulating fetuin-A may have prognostic value for MetS [3]. Specifically, circulating fetuin-A is produced from adipose and hepatic tissue, secreted by paracrine, autocrine, and endocrine glands, also referred to as α 2-Heremans-Schmid glyco-protein. Therefore, it has been widely recognized as a multifunctional molecule that participates in many metabolic processes, including energy expenditure, appetite control, insulin resistance, and regulation of adipogenesis [4].

Several hypothetical mechanisms that play major roles in patients with MetS are being revealed. For example, it has been proposed that circulating fetuin-A modulates the insulin sensitivity and insulin resistance in organs such as adipose, liver, and other tissues [5]. According to existing data, a high calorie diet leads to free fatty acid excess and insulin resistance, hence fueling circulating fetuin-A synthesis and steatosis. Besides, circulating fetuin-A sends the chemo-attractant signals that induce macrophage secretion by hepatocytes and adipocytes, which is infiltrated into adipose tissue. Thus, the activated macrophage increases the expression of inflammatory cytokines, such as tumor necrosis factor (TNF- α) and interleukin-6 (IL-6), which contribute to subsequent steatosis and glucose metabolism disorder [6] (Fig 1). On the other hand, circulating fetuin-A inhibits the activity of insulin receptors and leads to insulin resistance. These effects are mediated through the phosphatidylinositide 3-kinase (PI3K) and Akt signaling pathways [7]. Meanwhile, circulating fetuin-A inhibits the activity of glucose transporter type 4 (GLUT4) translocation, and glucose uptake is impaired, as shown in Fig 1 (drawn by the KA).

Taken together, the preceding hypotheses suggest that circulating fetuin-A is a complex metabolic phase reactant with a debated role in MetS. However, studies investigating the association between circulating fetuin-A and MetS found inconsistent results [8, 9]. Therefore, this study aimed to conduct a comprehensive meta-analysis on the relationship between circulating fetuin-A and MetS, and to quantify the strength of this relationship. Moreover, this study conducted meta-regression to explore the potential influencing factors affecting this relationship.

2. Methods

2.1. Search strategy

This meta-analysis was conducted in accordance with the rules and regulations for meta-analysis in the Cochrane Handbook version 5.1.0. In addition, it was registered with the International Prospective Register of Systematic Reviews PROSPERO (CRD42019129566). The results were reported according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) checklist [10]. With the help of experienced librarians, the researchers identified English articles, which were published before March 2019, from the electronic databases of Web of science, Cochrane library, Embase, and PubMed, using predefined search strategies. Detailed search strategies are described in the S1 File.

2.2. Eligibility criteria

Studies meeting the following criteria were included in this meta-analysis: (1) case-control studies; (2) studies which reported diagnostic criteria for MetS; (3) studies that provided mean



Fig 1. Summarizes the hypothesis mechanism process of fetuin-A on glucolipid metabolism signaling cascade in MetS. Fig 1 Schematic representation of the effects of fetuin-A on glucolipid metabolism signaling cascade in MetS that explain positive effects of these factors on glucolipid control. MetS, Metabolic syndrome; Akt: protein kinase b; IL -6: interleukin-6; TNF- α : Tumor Necrosis Factor; AMPK: AMP activated protein-kinase; GLUT: glucose transporter; IRS: insulin receptor substrate; mTOR: mammalian target of rapamicin; APN, adiponectin; PI3K: phosphatidylinositol 3-kinase; mTOR, mechanistic target of rapamycin; NF- κ B, nuclear factor κ B; ERK 1/2, Extracellular signal-regulated kinases 1 and 2; SREBP- 1C, sterol regulatory element binding protein-1c. Fig 1 is drawn by the KA, without any copyright disputes.

https://doi.org/10.1371/journal.pone.0229776.g001

and standard deviation (SD) or odds ratio (OR) of circulating fetuin-A, or these could be obtained from authors as required; (4) studies were peer-reviewed before publication; and (5) studies were published in English. On the other hand, studies meeting the following criteria were excluded: (1) reviews or case reports; (2) MetS was studied in combination with other diseases; (3) the circulating fetuin-A was pharmacologically challenged before circulating fetuin-A measurement; (4) studies of animals; and (5) grey literature (unpublished literature). After independently screening articles, two reviewers [KA and AK] selected eligible studies and submitted them to the third reviewer [AL] who made the final decision.

2.3. Data extraction

Using a standardized data extraction form, methodological and outcome variables of interest were collected from each study as follows: (1) the first author's last name and year of publication; (2) characteristics of subjects such as Body Mass Index (BMI), mean age (mean, SD), and gender; (3) region of study; (4) laboratory measures such as fasting plasma glucose (FPG), diastolic and systolic blood pressure, HDL, and triglycerides; (5) biological sample characteristics such as material of sample, and mean concentration of circulating fetuin-A (mean, SD) or OR associated with the concentration of circulating fetuin-A; and (6) assay methods and storage temperatures. Two reviewers [BP and AK] used EpiData 3.0 and Excel 2007 to organize and save the extracted data. Any discrepancies were resolved by consensus.

2.4. Quality evaluation

To assess the quality of the eligible studies for this meta-analysis, the Newcastle-Ottawa Scale (NOS) was used [11]. This is a nine-star rating system designed for non-randomized studies. The NOS contains three domains and eight items. The three domains consist of the following broad perspectives: (1) Selection; (2) Comparability; and (3) Outcome. According to the NOS criteria, the studies were rated low, moderate, and high quality in accordance with the scores, 0-3, 4-6 and 7-9, respectively.

2.5. Statistical analysis

The 'meta' and 'metafor' packages in R software (version R 3.4.3) were used to perform metaanalysis. Given the variation in the populations and criteria used to define outcomes, a random effects model was used to pool the estimates from the eligible studies [12]. Using this model, the 95% confidence intervals (CIs) and corresponding odds ratios (ORs) were merged to compare the impact of internal exposure as regards the risk of major outcomes. In addition, the standardized mean differences (SMDs) and corresponding 95% CIs were merged into a single standardized mean difference (SMD) and corresponding 95% confidence interval (CI), as Cohen's d, which were used to evaluate the strength of the relationship between MetS and the concentration of circulating fetuin-A. The SMD was considered to be low if lower than 0.5, moderate if between 0.5 and 0.8, or high if greater than 0.8 [13]. For all the meta-analyses, the level of heterogeneity was assessed using Cochrane Q test and measured by I^2 statistic. The heterogeneity was considered high when the I^2 was greater than 75%, moderate when the I^2 was 25%~75% and low when the I^2 was less than 25% [14].

When heterogeneity was high, meta-regression analysis was conducted to explore the source of heterogeneity [15]. The following variables were considered for the meta-regression analysis: material (Plasma = 0, Serum = 1), region (Other = 0, Asia = 1), NOS (Other = 0, High = 1), gender (Female = 0, Male = 1), BMI ($<30 = 0, \geq 30 = 1$), age ($<18 = 0, \geq 18 = 1$), FPG (<100mg/dL = $0,\geq 100$ mg/dL = 1), systolic blood pressure (<130mmHg = $0,\geq 130$ mmHg = 1), diastolic blood pressure (<85mmHg = $0,\geq 85$ mmHg = 1), HDL (≥ 40 mg/dL = 0, <40mg/dL = 1), and triglycerides (<150mg/dL = $0,\geq 150$ mg/dL = 1). We used random-effect restricted cubic splines, with three knots at the 25%, 50% and 75% percentiles of the distribution, to examine a potential non-linear dose-response relationship between circulating fetuin-A concentrations and risk of MetS. Sensitivity analysis was performed to test whether exclusion of individual studies had a significant impact on the overall outcomes. Subgroup analyses were conducted to explore the impact of characteristics of patients on the outcomes. Finally, publication bias was assessed using the Egger funnel plot and Egger's linear regression test, when the number of studies reporting meta-analysis results was 10 or more [16]. In all the statistical tests, *p* values were calculated as two-sided and considered significant if less than 0.05.

3. Results

3.1. Literature search

Fig 2 shows the selection process of the eligible articles for this meta-analysis. Initially, a total of 1,390 articles were identified from the four electronic databases as follows: 336 from Embase, 25 from the Cochrane Library, 653 from Web of Science and 376 from PubMed. After excluding the duplicates, 1,207 articles were retained. Also, after reviewing the titles and abstracts of the 1,207 articles, 978 articles were excluded because they did not meet the inclusion criteria. A full text review of 229 articles further excluded 98 for being unrelated studies, 29 for not reporting data on circulating fetuin-A, 63 for not reporting mean and SD of the concentration of circulating fetuin-A, 13 for not comparing MetS patients with a control group, 6 for being reviews and 6 for not reporting results of control groups. Finally, a total of 14 articles met the inclusion criteria and were included in the final analysis.

3.2. Characteristics of eligible studies

MetS was defined when 3 or more of the following were satisfied: hypertriglyceridemia (\geq 150 mg/dL) or taking lipid-lowering drugs; elevated BP (\geq 85 mm Hg diastolic, \geq 130 mm Hg systolic) or taking antihypertensive drugs; low high-density lipoprotein cholesterol (<50 mg/dL in women, <40 mg/dL in men); hyperglycemia (fasting glucose \geq 100 mg/dL) or taking hypoglycemic agents drugs or insulin; and increased waist circumference (\geq 88 cm for women, \geq 102 cm for men). Table 1 presents the characteristics of the 14 eligible studies, which include subjects' characteristics such as BMI, age, gender, and region; subjects' laboratory characteristics such as FPG, diastolic blood pressure, systolic blood pressure, HDL, and triglycerides; and circulating fetuin-A sample characteristics such as assay methods and storage temperatures. Altogether, these studies compared circulating fetuin-A concentrations between 4,551 MetS patients and 8,805 controls. The NOS scores of these studies varied between 5 and 8, with 8 studies graded as high quality and 6 as moderate quality.



Fig 2. Flowchart of study selection. Showing the process by which relevant studies were retrieved from the databases, assessed, and selected, or excluded. Preferred reporting items for systematic reviews and meta-analyses (PRISMA) diagram for study search.

https://doi.org/10.1371/journal.pone.0229776.g002

3.3. Overall comparison

Fig 3 presents the forest plot of the results of the SMDs in relation to meta-analysis using the random-effects model. Circulating fetuin-A concentrations were significantly higher in the MetS patients than in the controls (SMD = 0.65, 95% CI: 0.48 to 0.83, Z = 7.18, p<0.001). However, heterogeneity was considerable (I^2 = 92.3%).

In the non-linear dose–response relationship, there was a slight trend towards an increment in the risk of MetS with an increment in the circulating fetuin-A concentration, although the increment was not statistically significant (p = 0.06). There was high heterogeneity ($I^2 =$

Study	Material	Country	NOS	Male gender,n (%)	BMI	Mean Age	FPG	SBP	DBP	HDL	Triglycerides	Methods	Frozen
(Can et al., 2016) [<u>34]</u>	Serum	Turkey	8	18(42)	32.94 ±5.45	14.70 ±1.15	88.89 ±10.76	122.50 ±19.29	75.14 ±13.55	36.50 ±6.47	184.89±80.63	ELISA	-80°C
(Huddam, Azak, Kocak, Bayraktar, & Sezer, 2013) [35]	Serum	Turkey	7	8(32)	34.56 ±2.63	49.20 ±8.60	115.96 ±3.62	130.48 ±22.60	78.52 ±16.90	45.32 ±8.81	188.64±75.31	ELISA	-80°C
(Nagwa Abdallah Ismail et al., 2012) [<u>36]</u>	Serum	Egypt	6	6(41)	33.17 ±3.15	11.28 ±3.71	89.67 ±13.51	110.56 ±16.29	73.89 ±11.40	35.75 ±6.62	151.83±62.53	ELISA	NR
(Nagwa Abdallah Ismail et al., 2012) [<u>36]</u>	Serum	Egypt	6	14(40)	35.63 ±2.66	27.60 ±7.90	96.61 ±13.29	119.17 ±15.74	78.33 ±9.24	42.44 ±9.75	118.50±44.71	ELISA	NR
(Ix et al., 2006) [4]	Serum	USA	8	151(84)	27.00 ±5.00	69.00 ±12.00	81.00 ±29.00	86.00 ±15.00	58.00 ±18.00	33.50 ±1.40	99.30±23.50	ELISA	-70°C
(Jialal et al., 2015) [9]	Serum	USA	6	7(25)	36.0 ±6.0	51.00 ±11.00	100.00 ±10.00	132.00 ±12.00	83.00 ±9.00	41.00 ±9.00	132.00 ±103.00	ELISA	NR
(Ju et al., 2017) [23]	Serum	China	5	217(100)	28.58 ±3.16	48.57 ±12.25	95.83 ±18.4	137.84 ±18.25	86.47 ±13.19	27.7 ±9.52	173.03±12.08	ELISA	NR
(Ju et al., 2017) [23]	Serum	China	5	200(0)	27.84 ±3.22	48.28 ±11.74	95.36 ±17.7	134.88 ±19.29	85.16 ±12.93	33.25 ±5.49	112.35±11.9	ELISA	NR
(Kaess et al., 2012) [<u>37</u>]	Serum	USA	8	1705(47)	26.80 ±5.50	40.00 ±8.70	92.00 ±8.70	117.00 ±14.00	75.00 ±10.00	55.00 ±16.00	91.55±64.37	ELISA	-80°C
(Kasabri et al., 2018) [8]	Serum	Jordan	8	11(37)	31.20 ±7.20	46.00 ±3.78	86.00 ±7.98	135.00 ±13.00	84.00 ±23.00	38.00 ±5.00	199.00±26.00	ELISA	-80°C
(Koch et al., 2013) [<u>38]</u>	Serum	Germany	8	139(58)	25.90 ±2.33	65.00 ±5.80	84.50 ±7.70	140.00 ±120.00	80.00 ±7.80	33.00 ±7.00	93.00±56.00	ELISA	-80°C
(Obuchi et al., 2014) [<u>39</u>]	Plasma	Japan	5	43(26)	24.00 ±3.00	66.50 ±9.20	103.70 ±21.80	136.80 ±18.30	83.40 ±11.00	54.90 ±15.80	113.60±82.90	ELISA	NR
(Roos et al., 2010) [40]	Plasma	Germany	8	103(87)	27.00 ±3.40	60.70 ±6.90	105.70 ±27.30	120.90 ±16.10	72.90 ±9.00	36.88 ±9.10	140.00 ±110.00	ELISA	-80°C
(Weghuber, Mangge, Hochbrugger, & Stulnig, 2014) [41]	Plasma	Austria	5	36(66)	28.30 ±2.30	44.00 ±8.00	81.50 ±6.30	137.00 ±23.00	88.00 ±9.80	39.00 ±6.00	123.00±32.00	NR	NR
(Weghuber et al., 2014) [41]	Plasma	Austria	5	30(43)	29.20 ±3.50	13.00 ±2.90	71.20 ±5.50	85.00 ±13.00	61.00 ±11.00	31.00 ±5.00	91.00±16.00	NR	NR
(Xu et al., 2011) [42]	Serum	China	6	1367(56)	25.60 ±3.60	61.50 ±9.60	108.70 ±16.30	141.00 ±22.00	80.00 ±10.00	35.00 ±6.00	115.00±32.00	ELISA	NR
(Zachariah et al., 2017) [43]	Plasma	USA	8	378(50)	28.40 ±5.20	40.50 ±9.40	103.60 ±27.60	122.20 ±13.30	79.00 ±9.40	46.50 ±13.40	149.20±135.5	ELISA	-80°C

Table 1. Characteristics of the studies included for the meta-analysis of fetuin-A and MetS.

NOS, Newcastle-Ottawa Scale; BMI, Body Mass Index; ELISA, enzyme linked immunosorbent assay; NR, not report; FPG, fasting plasma glucose; DBP, diastolic blood pressure; SBP, systolic blood pressure; TG, Triglycerides; MetS, Metabolic syndrome; HDL, high-density lipoprotein. MetS was defined as the presence of 3 or more of the following: hypertriglyceridemia (\geq 150 mg/dL)or taking lipid-lowering drugs; elevated BP (\geq 85 mm Hg diastolic, \geq 130 mm Hg systolic) or taking antihypertensive drugs; low high-density lipoprotein cholesterol (<50 mg/dL in women, <40 mg/dL in men); hyperglycemia (fasting glucose \geq 100 mg/dL) or taking hypoglycemic agents drugs or insulin; and increased waist circumference (\geq 88 cm for women, \geq 102 cm for men).

https://doi.org/10.1371/journal.pone.0229776.t001

71.7%) in the analysis of non-linear dose–response relationship between circulating fetuin-A concentration and the risk of MetS (Fig 4). The results of meta-regression analysis are shown in Table 2. The estimated amount of residual heterogeneity of meta-regression analysis (SE) was 0.1870, with $I^2 = 86.54\%$. Sample material, region, NOS, gender, BMI, age, FPG, systolic blood pressure (SBP), diastolic blood pressure (DBP), and triglycerides reporting were not significantly different. However, after introducing HDL into the meta-regression analysis model, results showed that sources of heterogeneity could be explained by HDL as the difference was significant. (b = -0.95, 95%CI: -1.83 to -0.07, p = 0.034).

			MetS		С	ontrol		Sta	ndar	dised	I Mea	an				
Study	Total	Mean	SD	Total	Mean	SD			Diff	eren	ce		:	SMD	95%-CI	Weight
Can 2016	43	43.64	14.00	43	33.12	10.55				_				0.84	[0.40; 1.28]	5.3%
Huddam 2013A	25	65.10	12.38	25	58.12	8.63					—			0.64	[0.07; 1.21]	4.4%
Huddam 2013B	25	71.37	12.96	25	58.12	8.63				-		_		1.18	[0.58; 1.79]	4.2%
Ismail 2012A	14	158.93	55.85	25	96.93	35.85					-			1.38	[0.65; 2.11]	3.4%
Ismail 2012B	36	226.39	69.49	25	152.43	50.75						_		1.17	[0.61; 1.72]	4.5%
Jialal 2015	28	163.00	63.00	25	131.00	35.00					—			0.61	[0.06; 1.16]	4.5%
Ju 2017A	217	296.59	94.62	509	245.01	97.28				-+				0.53	[0.37; 0.70]	7.5%
Ju 2017B	200	300.72	94.76	410	269.28	75.29				-+-				0.38	[0.21; 0.55]	7.4%
Kasabri 2018	30	57.84	43.45	30	67.56	55.57							-	-0.19	[-0.70; 0.31]	4.8%
Koch 2013	234	330.90	28.37	307	301.00	24.93								1.13	[0.94; 1.31]	7.3%
Obuchi 2014	191	300.85	25.53	468	255.31	33.62					+			1.44	[1.26; 1.63]	7.3%
Roos 2010	229	69.70	23.10	820	67.00	19.50				+				0.13	[-0.01; 0.28]	7.6%
Weghuber 2014A	30	71.26	27.05	25	65.55	14.04				-∤∎-	_			0.25	[-0.28; 0.79]	4.7%
Weghuber 2014B	39	86.47	32.72	32	71.14	17.65					⊢			0.56	[0.08; 1.04]	5.1%
Xu 2011	2460	322.79	37.53	3009	307.32	35.31				+				0.43	[0.37; 0.48]	7.9%
Zachariah 2017A	36	455.60	56.70	1560	439.40	37.80					-			0.42	[0.09; 0.75]	6.3%
Zachariah 2017B	714	490.20	37.35	1467	458.80	59.80				-+				0.59	[0.50; 0.68]	7.8%
Random effects model	4551			8805						•				0.65	[0.48; 0.83]	100.0%
Heterogeneity: $I^2 = 92\%$, τ^2	$^{2} = 0.10$	040, <i>p</i> < 0	0.01				1	I			I					
						-	-3	-2	-1	0	1	2	3			
						Increa	ased	in co	ntrols	In	creas	sed in	Met	tS		

Fig 3. Forest plot of fetuin-A between MetS participants and controls. Study effect sizes of fetuin-A concentration differences between MetS and controls. Each data marker represents a study, and the size of the data marker is proportional to the total number of individuals in that study. The summary effect size for each fetuin-A concentration is denoted by a diamond. MetS, Metabolic syndrome; SMD, standardized mean difference.

https://doi.org/10.1371/journal.pone.0229776.g003

The results of the analysis of dose-response relationship between HDL and SMD are shown in Fig 5. There was a positive dose-response linear relationship between HDL and SMD (b = 0.02, p = 0.014). Moreover, subgroup analyses were conducted to explore the impact of HDL, which showed that circulating fetuin-A concentrations were significantly higher in the MetS patients than in the controls for the two subgroups, high HDL (\geq 40mg/dL) concentrations subgroup (k = 7, SMD = 0.86, 95%CI: 0.48 to 1.25, Z = 4.39, p<0.001) and low HDL (<40mg/dL) concentrations subgroup (k = 10, SMD = 0.52, 95%CI: 0.32 to 0.72, Z = 5.01, p<0.001), but with high heterogeneity (I^2 = 91.9% and 89.9%, respectively).

Table 3 presents the subgroup analyses for circulating fetuin-A concentrations between the MetS patients and the controls. Thus, most of the subgroup analysis results are consistent with the overall meta-analysis results, suggesting that these results are relatively stable. However, it is worth noting that age and DBP significantly explained the source of heterogeneity in relation to the outcomes. For example, circulating fetuin-A concentrations were significantly higher in the MetS patients younger than 18 years than in the control groups, but still with residual heterogeneity (I² = 41.5%). Besides, the concentration of circulating fetuin-A was significantly higher in the MetS patients with DBP \geq 85mmHg than in the control group, and most of the heterogeneity was explained ($I^2 = 7.7\%$).

Sensitivity analysis showed that the SMD and corresponding 95% CI changed little after each individual study was excluded sequentially, indicating that the overall results were relatively stable. The Egger funnel plot for circulating fetuin-A concentrations was symmetrical, and the Egger's test did not reject the hypothesis that there was no publication bias (t = 1.21, p = 0.245).



Fig 4. Non-linea dose-response relationship between the risk of MetS and fetuin-A concentrations. Odds ratio (OR;—) and the corresponding 95% CI (---) were summarized for thenon-linear dose-response relationship between fetuin-A concentrations with MetS risk. MetS, Metabolic syndrome; OR, odds ratio.

https://doi.org/10.1371/journal.pone.0229776.g004

Finally, all the results had moderate quality evidence. In this regard, methodological issues might have limited the overall quality of evidence. For example, the evidence of the difference

Tabl	le 2.	Meta-	regression	of	the	studie	s for	the	fetuin	-A and	d MetS.
------	-------	-------	------------	----	-----	--------	-------	-----	--------	--------	---------

	Estimate	Standard error	Z value	p value		95% CI
Intrcpt	1.8402	0.6119	3.0073	0.0026	0.6409	3.0396
Material (Plasma = 0, Serum = 1)	0.2277	0.4958	0.4593	0.6460	-0.7441	1.1995
Region (Other = 0, Asia = 1)	0.2645	0.3707	0.7137	0.4754	-0.4620	0.9910
NOS (Other = 0 , High = 1)	-0.0641	0.3412	-0.1877	0.8511	-0.7328	0.6047
Gender (Female = 0, Male = 1)	0.3276	0.5227	0.6267	0.5308	-0.6969	1.3520
BMI (<30 = 0, ≥30 = 1)	-0.2076	0.7624	-0.2722	0.7854	-1.7019	1.2868
Age (<18 = 0, \geq 18 = 1)	-1.0068	0.8061	-1.2489	0.2117	-2.5868	0.5732
FPG (<100mg/dL = 0,≥100mg/dL = 1)	-0.3789	0.4631	-0.8183	0.4132	-1.2865	0.5287
SBP (<130mmHg = 0,≥130mmHg = 1)	0.4204	0.4892	0.8594	0.3901	-0.5383	1.3791
DBP (<85mmHg = 0,≥85mmHg = 1)	-0.3779	0.6094	-0.6201	0.5352	-1.5723	0.8165
HDL (\geq 40mg/dL = 0, <40mg/dL = 1)	-0.9491	0.4482	-2.1177	0.0342	-1.8275	-0.0707
$TG (<150mg/dL = 0, \ge 150mg/dL = 1)$	-0.2494	0.5934	-0.4203	0.6743	-1.4124	0.9136

NOS, Newcastle-Ottawa Scale; BMI, Body Mass Index; ELISA, enzyme linked immunosorbent assay; NR, not report; FPG, fasting plasma glucose; DBP, diastolic blood pressure; SBP, systolic blood pressure; TG, Triglycerides; MetS, Metabolic syndrome; HDL, high-density lipoprotein.

https://doi.org/10.1371/journal.pone.0229776.t002



Fig 5. Dose-response relationships between HDL value index and SMD value outcomes based on data from each studies. Each data point overlaps into a circle. The size of the circle corresponds to the inverse variance weight of the SMD effect in the analysis.MetS, Metabolic syndrome; SMD, standardized mean difference.

https://doi.org/10.1371/journal.pone.0229776.g005

	Subgroup	SMD	95%-CI	p value	I^2
Material	Plasma	0.5800	[0.1725; 0.9874]	0.64	95.9%
	Serum	0.6889	[0.4757; 0.9020]		87.6%
Region	Asia	0.7502	[0.4697; 1.0307]	0.31	93.3%
	Other	0.5377	[0.2437; 0.8316]		91.7%
NOS	High	0.5849	[0.2904; 0.8794]	0.51	91.7%
	Other	0.7240	[0.4398; 1.0081]		93.4%
Gender	Female	0.7943	[0.4121; 1.1765]	0.20	90.2%
	Male	0.5166	[0.3261; 0.7071]		92.5%
BMI	<30	0.6007	[0.3890; 0.8125]	0.43	95.1%
	\geq 30	0.7816	[0.3900; 1.1733]		71.2%
Age	<18	0.8547	[0.4540; 1.2554]	0.30	41.5%
	≥ 18	0.6187	[0.4259; 0.8115]		93.5%
FPG	<100mg/dL	0.6546	[0.3744; 0.9348]	1.00	86.3%
	≥100mg/dL	0.6557	[0.3974; 0.9140]		95.0%
SBP	<130mmHg	0.6311	[0.3644; 0.8978]	0.90	85.5%
	≥130mmHg	0.6561	[0.3762; 0.9360]		94.5%
DBP	<85mmHg	0.7159	[0.4988; 0.9330]	0.04	93.6%
	≥85mmHg	0.4510	[0.3288; 0.5731]		7.7%
HDL	<40mg/dL	0.5183	[0.3154; 0.7213]	0.12	89.9%
	≥40mg/dL	0.8627	[0.4776; 1.2478]		91.9%
TG	<150mg/dL	0.6471	[0.4175; 0.8766]	0.92	95.2%
	≥150mg/dL	0.6679	[0.3598; 0.9760]		68.6%

Table 3. Subgroup analysis of the studies for the fetuin-A and MetS.

MetS, Metabolic syndrome; SMD, standardized mean difference; NOS, Newcastle-Ottawa Scale; BMI, Body Mass Index; ELISA, enzyme linked immunosorbent assay; NR, not report; FPG, fasting plasma glucose; DBP, diastolic blood pressure; SBP, systolic blood pressure; TG, Triglycerides; MetS, Metabolic syndrome; HDL, high-density lipoprotein.

https://doi.org/10.1371/journal.pone.0229776.t003

in the circulating fetuin-A concentrations between the MetS patients and the controls was downgraded by one level because all plausible residual confounding would reduce the demonstrated effect; and the evidence of circulating fetuin-A concentrations, as a risk of Mets, was downgraded by one level because publication bias was suspected, and all plausible residual confounding would reduce the demonstrated effect, but with dose response gradient.

4. Discussion

The present meta-analysis revealed that there might be a relationship between circulating fetuin-A and MetS. That is, the circulating fetuin-A concentration in MetS patients was significantly higher than that in the control group. Therefore, subjects with high circulating fetuin-A concentration may have increased risk of developing MetS. Moreover, this study found that there was a slight trend towards an increment in the risk of MetS with an increment in the circulating fetuin-A concentration. Nevertheless, heterogeneity in the meta-regression, in relation to the foregoing results, was significantly high and substantially explained by HDL. Also, from the subsequent linear regression model test, a positive linear correlation was observed between HDL and SMD, indicating that HDL may play an important regulating role between circulating fetuin-A and MetS. This study also analyzed and summarized data from different age populations in order to see whether circulating fetuin-A concentrations could be affected by age differences [17].

The results suggested that the concentrations of circulating fetuin-A in the MetS patients was higher than that in the control group for all the age groups considered. However, metaanalysis with respect to the adult subgroup (18 years or older) was associated with significantly higher heterogeneity. Thus, it may be suggested that the activity of circulating fetuin-A, or the molecular mechanisms of circulating fetuin-A bioconversion were impaired in the adult MetS patients. Further investigations are necessary to understand the molecular mechanisms that account for the changes of circulating fetuin-A concentrations for different age groups among the MetS patients.

Furthermore, subgroup analysis showed that, for MetS patients with DBP≥85mmHg, the concentration of circulating fetuin-A was significantly higher than that of the control group, and this subgroup explained most of the heterogeneity. Considering that there were only three studies reporting DBP≥85mmHg, which represents limited available data thus far as regards MetS patients with DBP≥85mmHg, future research should expand the sample size to verify whether DBP plays a regulatory role in the MetS patients and circulating fetuin-A.

Many published reviews have suggested that circulating fetuin-A may play a role in the risk of type 2 diabetes mellitus, cardiovascular disease, and nonalcoholic fatty liver disease [18, 19]. For example, a meta-analysis on the association between type 2 diabetes mellitus and circulating fetuin-A showed that circulating fetuin-A concentrations in type 2 diabetes mellitus patients were significantly higher than in the controls [19]. This is in agreement with the findings of this study, suggesting that circulating fetuin-A may play a role in the disorder of glucose and lipid metabolism in humans with MetS. Although MetS share some features with type 2 diabetes mellitus or cardiovascular diseases, the role of circulating fetuin-A in MetS may be more complex [6].

For instance, free fatty acids and circulating fetuin-A play important roles in the interaction and metabolic processes between the liver and adipose tissues. According to some experiments, free fatty acids can increase the activity of nuclear factor κ B (NF- κ B) by restraining the AMP protein-kinase (AMPK), which in turn is a consequence of ERK 1/2 activation [20, 21]. Likewise, NF- κ B up-regulates circulating fetuin-A gene expression and circulating fetuin-A mRNA expression, which may directly stimulate mechanistic target of rapamycin (mTOR) phosphorylation and sterol regulatory element binding protein-1c (SREBP-1C) expression [20, 22]. Moreover, available data indicated that up-regulation of lipogenic enzymes by SREBP-1C is beneficial to the accumulation of triacylglycerol. On the other hand, circulating fetuin-A inhibits the activity of insulin receptors and leads to insulin resistance. These effects are mediated through the PI3K and Akt signaling pathway [7].

Evidence from animal experiments has shown that elevated circulating fetuin-A concentrations could lead to impaired glucose control due to migration and activation of macrophage, impaired signaling of insulin receptors, triglyceride accumulation in hepatocytes, and dysfunction of adipocytes [23]. When the concentrations of glucose and free fatty acids in the blood of a MetS patient are increased, they can stimulate the binding of NF- κ B to the circulating fetuin-A promoter, and then activate the expression of circulating fetuin-A mRNA to further promote circulating fetuin-A protein synthesis and secretion [20]. However, the inhibitory effect of adiponectin in this process has received attention recently. It is widely accepted that the adiponectin inhibits the expression of circulating fetuin-A induced by glycolipid disorder through AMPK pathway [24]. Therefore, the common hypoadiponectinemia in MetS patients may be another cause of increased circulating fetuin-A.

It has been proven from many clinical studies that circulating fetuin-A concentration in individuals with centripetal obesity is usually elevated. Given that centripetal obesity is closely related to insulin resistance and dyslipidemia, it is considered as a major sign of MetS [6]. The expansion of adipose tissue helps several pro-inflammatory mediators, such as macrophages and inflammatory cytokines (IL-6, TNF- α), to move into fatty tissue, where they interfere with the mechanisms by which adipose tissue responds to insulin action [25]. This leads to insulin resistance and insulin-mediated anti-lipolysis damage, which in turn lead to increased release of circulating fetuin-A from adipose tissue, excessive circulating fetuin-A uptake by the liver, and movement of circulating fetuin-A through the energy sensor AMPK, which is pivotal to directing hepatocytes to potentially deleterious pathways that lead to triglyceridemia [26]. Additionally, in a 1-year longitudinal follow-up study, gastric bypass surgery among morbidly obese patients reduced the concentration of circulating fetuin-A [27]. Another longitudinal study revealed that physical exercise intervention and weight loss practices were associated with significant reductions in fetoprotein levels [28].

The hypothesis of an association between MetS and circulating fetuin-A was supported not only by clinical and experimental studies, but also genetic epidemiological studies. For instance, it is widely accepted that the AHSG gene is located on chromosome 3q27, a region that has been identified as a MetS susceptibility locus. Thus, AHSG gene variant associated with lower circulating fetuin-A was more common in normal-weight men than in their overweight counterparts [5]. Therefore, the link between circulating fetuin-A and MetS appears to be at least partially mediated by genetics [29]. Other studies have shown that a single nucleotide polymorphism (SNP) of AHSG gene was associated with the prevalence of type 2 diabetes mellitus, and circulating fetuin-A concentration was negatively correlated with insulin secretion [30].

In this study, it was also shown that HDL played an important role in the relationship between MetS and circulating fetuin-A, which may be due to the fact that HDL is a possible component of dyslipidemia in MetS. However, it was further shown, through subgroup analysis, that there was significant heterogeneity among groups with different HDL concentrations and this was contrary to the results of the meta-regression analysis of HDL. The preceding contradiction may be caused by treating HDL differently in the subgroup and meta-regression analyses. For example, HDL was considered as a categorical variable in the subgroup analysis, whereas it was considered as a continuous variable in the meta-regression analysis. That is, taking HDL as a continuous variable might have increased the statistical efficiency of metaregression analysis than considering it as a categorical variable in the subgroup analysis.

Thus, the results of subgroup analysis due to HDL should be seen as exploratory and interpreted cautiously. Nevertheless, recent studies also revealed that there was a correlation between HDL and circulating fetuin-A [31]. Moreover, experimental studies have shown that HDL can improve the lipid and glucose metabolism by activating the AMPK/SIRT1 signaling pathway [32]. Meanwhile, HDL can also promote glucose uptake in the adipocytes and glycogen synthesis through enhancing GLUT4 by mechanisms involving PI3K/Akt via AMPK signaling pathways [33]. In addition, AMPK pathway is the main pathway that inhibits circulating fetuin-A. Therefore, the common low levels of HDL in patients with MetS may be another reason for higher levels of circulating fetuin-A in this population.

4.1. Limitations

This meta-analysis and meta-regression are subject to several limitations. First, although there was a dose-response relationship between circulating fetuin-A and the risk of MetS, all the eligible studies analyzed were case-control studies, which could not make causality inference. In future, prospective cohort studies should be considered to explore the link between circulating fetuin-A and MetS. Second, majority of the eligible studies did not adjust for the possible confounding variables. Third, heterogeneity was high when synthesizing evidence on the relationship between the concentration of circulating fetuin-A and MetS. Therefore, future studies should consider investigating the association between the concentration of circulating fetuin-A and MetS in homogeneous populations using same methods. Last, only articles published in English were included. Therefore, this meta-analysis may be more prone to bias since some negative results are generally reported in regional journals in native languages.

5. Conclusions

Circulating fetuin-A concentration in the MetS patients was significantly higher than that in the control groups, and there was a slight trend towards an increment in the risk of MetS with an increment in the circulating fetuin-A concentration. Also, HDL may play a possible regulating role between circulating fetuin-A and MetS. Therefore, these results provide a rationale for evaluating circulating fetuin-A to see if it could affect the pathophysiological process of the risk of MetS. Specifically, this evaluation might lead to new perspectives in the early diagnosis, identification of novel biomarkers, and discovery of novel targets for pharmacological interventions.

Supporting information

S1 File. Search strategies. Details of search strategy. (DOC)

S1 Table. PRISMA checklist. (DOC)

Acknowledgments

We are grateful to Central South University Library for the assistance during literature search. Furthermore, we sincerely appreciate the hard work of the distinguished editors and reviewers, who contributed tremendously in improving the clarity of this manuscript.

Author Contributions

Conceptualization: Xiongfeng Pan, Shi Wu Wen, Aizhong Liu.

Data curation: Prince L. Bestman, Atipatsa C. Kaminga, Kwabena Acheampong.

Formal analysis: Prince L. Bestman, Atipatsa C. Kaminga, Kwabena Acheampong.

Funding acquisition: Shi Wu Wen.

Methodology: Prince L. Bestman, Atipatsa C. Kaminga, Kwabena Acheampong.

Software: Prince L. Bestman, Atipatsa C. Kaminga, Aizhong Liu.

Validation: Xiongfeng Pan, Aizhong Liu.

Visualization: Aizhong Liu.

Writing – original draft: Xiongfeng Pan, Shi Wu Wen, Atipatsa C. Kaminga, Kwabena Acheampong, Aizhong Liu.

Writing - review & editing: Xiongfeng Pan, Shi Wu Wen, Aizhong Liu.

References

- Lent-Schochet D, McLaughlin M, Ramakrishnan N, Jialal I. Exploratory metabolomics of metabolic syndrome: A status report. World journal of diabetes. 2019; 10(1):23–36. Epub 2019/01/31. https://doi.org/ 10.4239/wjd.v10.i1.23 PMID: 30697368; PubMed Central PMCID: PMC6347655.
- Rubio-Ruiz ME, Guarner-Lans V, Perez-Torres I, Soto ME. Mechanisms Underlying Metabolic Syndrome-Related Sarcopenia and Possible Therapeutic Measures. Int J Mol Sci. 2019; 20(3). Epub 2019/ 02/06. https://doi.org/10.3390/ijms20030647 PMID: <u>30717377</u>; PubMed Central PMCID: PMC6387003.
- Brandenburg V, Ix J, Westenfeld R, Whooley M, Shlipak M, Ketteler M. The association between fetuin-A in serum and the metabolic syndrome: Data from the heart-and-soul study. Medizinische Klinik. 2006; 101(4):A12–A. WOS:000237562000026.
- Ix JH, Shlipak MG, Brandenburg VM, Ali S, Ketteler M, Whooley MA. Association between human fetuin-A and the metabolic syndrome: data from the Heart and Soul Study. Circulation. 2006; 113 (14):1760–7. Epub 2006/03/29. https://doi.org/10.1161/CIRCULATIONAHA.105.588723 PMID: 16567568; PubMed Central PMCID: PMC2776669.
- Axelsson J, Wang X, Ketteler M, Qureshi AR, Heimburger O, Barany P, et al. Is fetuin-A/alpha2-Heremans-Schmid glycoprotein associated with the metabolic syndrome in patients with chronic kidney disease? Am J Nephrol. 2008; 28(4):669–76. Epub 2008/03/14. https://doi.org/10.1159/000121358 PMID: 18337634.
- Trepanowski JF, Mey J, Varady KA. Fetuin-A: a novel link between obesity and related complications. International journal of obesity (2005). 2015; 39(5):734–41. Epub 2014/12/04. <u>https://doi.org/10.1038/ijo.2014.203</u> PMID: 25468829.
- Siegel-Axel DI, Ullrich S, Stefan N, Rittig K, Gerst F, Klingler C, et al. Fetuin-A influences vascular cell growth and production of proinflammatory and angiogenic proteins by human perivascular fat cells. Diabetologia. 2014; 57(5):1057–66. Epub 2014/02/05. https://doi.org/10.1007/s00125-014-3177-0 PMID: 24493202.
- Kasabri V, Shawakri S, Akour A, Naffa R, Khawaja N, Al-Sarraf I, et al. Cross-sectional correlates of increased IL-18 but reduced fetuin-A and oxytocin with adiposity and blood indices in metabolic syndrome patients with and without prediabetes. Therapeutic Advances in Endocrinology and Metabolism. 2018; 9(12):329–38. <u>https://doi.org/10.1177/2042018818788802</u> WOS:000452260400001. PMID: 30515292
- Jialal I, Devaraj S, Bettaieb A, Haj F, Adams-Huet B. Increased adipose tissue secretion of Fetuin-A, lipopolysaccharide-binding protein and high-mobility group box protein 1 in metabolic syndrome. Atherosclerosis. 2015; 241(1):130–7. https://doi.org/10.1016/j.atherosclerosis.2015.04.814 PMID: 25978344
- Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and metaanalyses: the PRISMA statement. BMJ (Clinical research ed). 2009; 339:b2535. Epub 2009/07/23. https://doi.org/10.1136/bmj.b2535 PMID: 19622551; PubMed Central PMCID: PMC2714657.

- Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. Eur J Epidemiol. 2010; 25(9):603–5. Epub 2010/07/24. https://doi. org/10.1007/s10654-010-9491-z PMID: 20652370.
- Pan X, Kaminga AC, Wen SW, Liu A. Catecholamines in Post-traumatic Stress Disorder: A Systematic Review and Meta-Analysis. Front Mol Neurosci. 2018; 11:450. Epub 2018/12/20. https://doi.org/10. 3389/fnmol.2018.00450 PMID: 30564100; PubMed Central PMCID: PMC6288600.
- Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. BMJ (Clinical research ed). 2003; 327(7414):557–60. Epub 2003/09/06. https://doi.org/10.1136/bmj.327. 7414.557 PMID: 12958120; PubMed Central PMCID: PMC192859.
- DerSimonian R, Laird N. Meta-analysis in clinical trials revisited. Contemp Clin Trials. 2015; 45(Pt A):139–45. Epub 2015/09/08. https://doi.org/10.1016/j.cct.2015.09.002 PMID: 26343745; PubMed Central PMCID: PMC4639420.
- Pan X, Wang Z, Wu X, Wen SW, Liu A. Salivary cortisol in post-traumatic stress disorder: a systematic review and meta-analysis. BMC Psychiatry. 2018; 18(1):324. Epub 2018/10/07. https://doi.org/10.1186/ s12888-018-1910-9 PMID: 30290789; PubMed Central PMCID: PMC6173866.
- Egger M, Davey Smith G, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. BMJ (Clinical research ed). 1997; 315(7109):629–34. Epub 1997/10/06. https://doi.org/10.1136/ bmj.315.7109.629 PMID: 9310563; PubMed Central PMCID: PMC2127453.
- Marhaug G, Shah V, Shroff R, Varsani H, Wedderburn LR, Pilkington CA, et al. Age-dependent inhibition of ectopic calcification: a possible role for fetuin-A and osteopontin in patients with juvenile dermatomyositis with calcinosis. Rheumatology (Oxford, England). 2008; 47(7):1031–7. Epub 2008/05/02. https://doi.org/10.1093/rheumatology/ken136 PMID: 18448482.
- Ramirez-Velez R, Garcia-Hermoso A, Hackney AC, Izquierdo M. Effects of exercise training on Fetuina in obese, type 2 diabetes and cardiovascular disease in adults and elderly: a systematic review and Meta-analysis. Lipids Health Dis. 2019; 18(1):23. Epub 2019/01/24. https://doi.org/10.1186/s12944-019-0962-2 PMID: 30670052; PubMed Central PMCID: PMC6343360.
- Roshanzamir F, Miraghajani M, Rouhani MH, Mansourian M, Ghiasvand R, Safavi SM. The association between circulating fetuin-A levels and type 2 diabetes mellitus risk: systematic review and meta-analysis of observational studies. J Endocrinol Invest. 2018; 41(1):33–47. Epub 2017/06/24. <u>https://doi.org/ 10.1007/s40618-017-0697-8 PMID: 28643299</u>.
- Dasgupta S, Bhattacharya S, Biswas A, Majumdar SS, Mukhopadhyay S, Ray S, et al. NF-kappaB mediates lipid-induced fetuin-A expression in hepatocytes that impairs adipocyte function effecting insulin resistance. Biochem J. 2010; 429(3):451–62. Epub 2010/05/21. <u>https://doi.org/10.1042/BJ20100330</u> PMID: 20482516.
- Ou HY, Wu HT, Hung HC, Yang YC, Wu JS, Chang CJ. Endoplasmic reticulum stress induces the expression of fetuin-A to develop insulin resistance. Endocrinology. 2012; 153(7):2974–84. Epub 2012/ 05/24. https://doi.org/10.1210/en.2011-2043 PMID: 22619360.
- Jung TW, Youn BS, Choi HY, Lee SY, Hong HC, Yang SJ, et al. Salsalate and adiponectin ameliorate hepatic steatosis by inhibition of the hepatokine fetuin-A. Biochem Pharmacol. 2013; 86(7):960–9. Epub 2013/08/21. https://doi.org/10.1016/j.bcp.2013.07.034 PMID: 23948064.
- Ju H, Zhou Z, Sun M, Chen H. Association of fetuin-A to adiponectin ratio with metabolic syndrome: a cross-sectional study. Endocrine. 2017; 58(1):190–3. https://doi.org/10.1007/s12020-017-1383-5 PMID: 28779425
- Stefan N, Sun Q, Fritsche A, Machann J, Schick F, Gerst F, et al. Impact of the adipokine adiponectin and the hepatokine fetuin-A on the development of type 2 diabetes: prospective cohort- and cross-sectional phenotyping studies. PLoS One. 2014; 9(3):e92238. Epub 2014/03/20. https://doi.org/10.1371/ journal.pone.0092238 PMID: 24643166; PubMed Central PMCID: PMC3958485.
- EI-Deeb TS, Bakkar SM, Eltoony L, Zakhary MM, Kamel AA, Nafee AM, et al. The adipokine Chemerin and Fetuin-A Serum Levels in Type 2 Diabetes Mellitus: Relation to Obesity and Inflammatory Markers. The Egyptian journal of immunology. 2018; 25(1):191–202. Epub 2018/09/23. PMID: 30243011.
- 26. Hennige AM, Staiger H, Wicke C, Machicao F, Fritsche A, Haring HU, et al. Fetuin-A induces cytokine expression and suppresses adiponectin production. PLoS One. 2008; 3(3):e1765. Epub 2008/03/13. https://doi.org/10.1371/journal.pone.0001765 PMID: 18335040; PubMed Central PMCID: PMC2258416.
- Huang HH, Yeh C, Chen JC, Lee TH, Chen SC, Lee WJ, et al. Does bariatric surgery influence plasma levels of fetuin-A and leukocyte cell-derived chemotaxin-2 in patients with type 2 diabetes mellitus? PeerJ. 2018; 6:e4884. Epub 2018/06/19. <u>https://doi.org/10.7717/peerj.4884</u> PMID: 29910974; PubMed Central PMCID: PMC6003398.
- Blumenthal JB, Gitterman A, Ryan AS, Prior SJ. Effects of Exercise Training and Weight Loss on Plasma Fetuin-A Levels and Insulin Sensitivity in Overweight Older Men. Journal of diabetes research.

2017; 2017:1492581. Epub 2017/08/05. https://doi.org/10.1155/2017/1492581 PMID: 28770230; PubMed Central PMCID: PMC5523541.

- 29. Thakkinstian A, Chailurkit L, Warodomwichit D, Ratanachaiwong W, Yamwong S, Chanprasertyothin S, et al. Causal relationship between body mass index and fetuin-A level in the asian population: a bidirectional Mendelian randomization study. Clinical endocrinology. 2014; 81(2):197–203. Epub 2013/08/01. https://doi.org/10.1111/cen.12303 PMID: 23899227.
- Jensen MK, Bartz TM, Djousse L, Kizer JR, Zieman SJ, Rimm EB, et al. Genetically elevated fetuin-A levels, fasting glucose levels, and risk of type 2 diabetes: the cardiovascular health study. Diabetes Care. 2013; 36(10):3121–7. Epub 2013/06/27. https://doi.org/10.2337/dc12-2323 PMID: 23801724; PubMed Central PMCID: PMC3781539.
- Ismail NA, Ragab S, El Dayem SM, Elbaky AA, Salah N, Hamed M, et al. Fetuin-A levels in obesity: differences in relation to metabolic syndrome and correlation with clinical and laboratory variables. Arch Med Sci. 2012; 8(5):826–33. Epub 2012/11/28. https://doi.org/10.5114/aoms.2012.31616 PMID: 23185191; PubMed Central PMCID: PMC3506238.
- 32. Zang Y, Fan L, Chen J, Huang R, Qin H. Improvement of Lipid and Glucose Metabolism by Capsiate in Palmitic Acid-Treated HepG2 Cells via Activation of the AMPK/SIRT1 Signaling Pathway. Journal of agricultural and food chemistry. 2018; 66(26):6772–81. Epub 2018/06/12. <u>https://doi.org/10.1021/acs.jafc.8b01831</u> PMID: 29886733.
- Zhang Q, Zhang Y, Feng H, Guo R, Jin L, Wan R, et al. High density lipoprotein (HDL) promotes glucose uptake in adipocytes and glycogen synthesis in muscle cells. PLoS One. 2011; 6(8):e23556. Epub 2011/09/03. https://doi.org/10.1371/journal.pone.0023556 PMID: 21886796; PubMed Central PMCID: PMC3158770.
- Can U, Buyukinan M, Guzelant A, Ugur A, Karaibrahimoglu A, Yabanciun S. Investigation of the inflammatory biomarkers of metabolic syndrome in adolescents. J Pediatr Endocrinol Metab. 2016; 29 (11):1277–83. Epub 2016/10/19. https://doi.org/10.1515/jpem-2016-0136 PMID: 27754964.
- **35.** Huddam B, Azak A, Kocak G, Bayraktar N, Sezer S. The relationship between serum fetuin-A, cystatin-C levels, and microalbuminuria in patients with metabolic syndrome. J Clin Lab Anal. 2013; 27(4):317–22. Epub 2013/07/16. https://doi.org/10.1002/jcla.21605 PMID: 23852792.
- 36. Ismail NA, Ragab S, Abd El Dayem SM, Abd ElBaky A, Salah N, Hamed M, et al. Fetuin-A levels in obesity: differences in relation to metabolic syndrome and correlation with clinical and laboratory variables. Archives of Medical Science. 2012; 8(5):826–33. <u>https://doi.org/10.5114/aoms.2012.31616</u> WOS:000310935200010. PMID: 23185191
- Kaess BM, Enserro DM, McManus DD, Xanthakis V, Chen MH, Sullivan LM, et al. Cardiometabolic correlates and heritability of fetuin-A, retinol-binding protein 4, and fatty-acid binding protein 4 in the Framingham Heart Study. J Clin Endocrinol Metab. 2012; 97(10):E1943–7. Epub 2012/08/03. https://doi.org/10.1210/jc.2012-1458 PMID: 22855337; PubMed Central PMCID: PMC3674297.
- Koch M, Jacobs G, Hampe J, Rosenstiel P, Krawczak M, Nothlings U. Higher fetuin-A level is associated with coexistence of elevated alanine aminotransferase and the metabolic syndrome in the general population. Metab Syndr Relat Disord. 2013; 11(6):377–84. Epub 2013/08/27. https://doi.org/10.1089/met.2013.0078 PMID: 23971757.
- Obuchi A, Adachi H, Enomoto M, Fukami A, Kumagai E, Nakamura S, et al. High plasma fetuin-A levels are associated with metabolic syndrome among males but not females in a Japanese general population. Diabetes Research and Clinical Practice. 2014; 106(1):128–35. <u>https://doi.org/10.1016/j.diabres</u>. 2014.07.002 PMID: 25110104
- Roos M, Eynatten MV, Heemann U, Rothenbacher D, Brenner H, Breitling LPJAJoC. Serum Fetuin-A, Cardiovascular Risk Factors, and Six-Year Follow-up Outcome in Patients With Coronary Heart Disease. 2010; 105(12):1666–72. https://doi.org/10.1016/j.amjcard.2010.01.342 PMID: 20538112
- **41.** Weghuber D, Mangge H, Hochbrugger E, Stulnig TM. Impact of age and metabolic syndrome on the adipokine profile in childhood and adult obesity. Exp Clin Endocrinol Diabetes. 2014; 122(6):363–7. Epub 2014/06/19. https://doi.org/10.1055/s-0034-1375647 PMID: 24941433.
- Xu Y, Xu M, Bi Y, Song A, Huang Y, Liu Y, et al. Serum fetuin-A is correlated with metabolic syndrome in middle-aged and elderly Chinese. Atherosclerosis. 2011; 216(1):180–6. Epub 2011/02/12. <u>https://</u> doi.org/10.1016/j.atherosclerosis.2011.01.020 PMID: 21310413.
- Zachariah JP, Quiroz R, Nelson KP, Teng Z, Keaney JF Jr., Sullivan LM, et al. Prospective Relation of Circulating Adipokines to Incident Metabolic Syndrome: The Framingham Heart Study. J Am Heart Assoc. 2017; 6(7). Epub 2017/07/18. https://doi.org/10.1161/JAHA.116.004974 PMID: 28713076; PubMed Central PMCID: PMC5586264.