

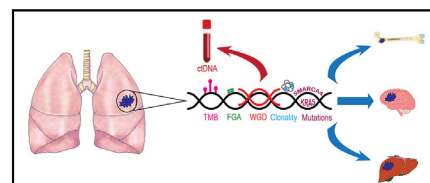
## Genomic profiling and metastatic risk in early-stage non-small cell lung cancer



Cameron N. Fick, MD,<sup>a</sup> Elizabeth G. Dunne, MD,<sup>a</sup> Manendra B. Lankadasari, PhD,<sup>a,b</sup> Brooke Mastrogiacomo, MS,<sup>a,c</sup> Tetsuhiko Asao, MD, PhD,<sup>a</sup> Stijn Vanstraelen, MD,<sup>a</sup> Yuan Liu, MD, PhD,<sup>a,b</sup> Francisco Sanchez-Vega, PhD,<sup>c</sup> and David R. Jones, MD<sup>a,b</sup>

Next-generation sequencing (NGS) and its applications have provided tremendous insight into the biologic landscape of a variety of solid tumors, including non-small cell lung cancer (NSCLC). The genomic characterization of NSCLC has not only altered treatment paradigms but has also aided investigations into tumor biology and the risk of developing metastatic disease. Complete surgical resection with or without chemotherapy and radiation has been the cornerstone of treatment for stage I-III NSCLC; however, up to 50% of patients with operable, early-stage NSCLC are at risk of metastasis after surgical resection.<sup>1,2</sup> Thus, recent investigations have applied the knowledge gained in the stage IV NSCLC setting to earlier-stage disease. Studies are now focused on leveraging tumor genomic data (DNA and RNA) to elucidate the biologic mechanisms associated with disease recurrence and to potentially identify patients with a higher risk of recurrence for additional adjuvant therapies.

Diagnostic molecular pathologic analysis and related tumor genomic and transcriptomic analyses are presently being applied to several solid tumors. An obvious example is the use of Oncotype Dx, a 21-gene expression assay for patients with ER-positive/HER2-negative stage I-III breast cancer. Oncotype Dx tests for genes related to cell proliferation, metastasis, HER2 expression, and sex hormone production.<sup>3,4</sup> Current National Comprehensive Cancer Network (NCCN) guidelines recommend Oncotype Dx as the preferred panel for prognosis and prediction of benefit from adjuvant chemotherapy.<sup>5</sup> The broad applicability of



Next-generation sequencing can be used to assess risk of metastasis in NSCLC.

### CENTRAL MESSAGE

Genomic profiling in patients with operable, early-stage NSCLC can enhance our understanding of tumor biology and metastatic risk.

gene assays such as Oncotype Dx in NSCLC has been limited by the substantial histologic and molecular heterogeneity of NSCLC. In this report, we outline the current role of genomics in the management of NSCLC, including risk-stratification of patients, and provide foundational knowledge for thoracic surgeons caring for patients with NSCLC.

### NSCLC HETEROGENEITY CONTRIBUTES TO METASTATIC RISK

Recent publications by the TRACERx Lung consortium have highlighted the presence of significant intratumoral subclonal somatic copy number alterations (SCNA) and mutations using whole-exome sequencing data on surgically resected NSCLC.<sup>6,7</sup> In a cohort of treatment-naïve patients with stage I-III NSCLC who underwent complete surgical resection, tumors with a greater proportion of subclonal mutations were associated with an increased risk of recurrence or death.<sup>6</sup> Subclonal whole-genome doubling, recent subclonal expansion, and a high level of SCNA intratumor heterogeneity (ITH) were also predictive of relapse; however, only high SCNA ITH was independently prognostic of early (<12 months after surgery) and extrathoracic recurrences.<sup>7</sup> Wu and colleagues<sup>8</sup> demonstrated similar findings in a spatial analysis of the ITH of lung adenocarcinoma in which they identified 2 distinct patterns—clustered and random geographic diversification—using both proteomic and genomic data. When clinicopathologic features

<sup>a</sup>Thoracic Service, Department of Surgery, Memorial Sloan Kettering Cancer Center, New York, NY; <sup>b</sup>Druckenmiller Center for Lung Cancer Research, Memorial Sloan Kettering Cancer Center, New York, NY; <sup>c</sup>Computational Oncology Service, Department of Epidemiology and Biostatistics, Memorial Sloan Kettering Cancer Center, New York, NY

This study was supported by the National Institutes of Health/National Cancer Institute (R01CA217169 and R01CA240472 to David R. Jones and Cancer Center Support Grant P30 CA008748 to Memorial Sloan Kettering Cancer Center).

Received for publication Aug 7, 2023; revisions received Oct 2, 2023; accepted for publication Oct 11, 2023; available ahead of print Oct 30, 2023.

Address for reprints: David R. Jones, MD, Thoracic Surgery Service, Druckenmiller Chair of Lung Cancer Research, Department of Surgery, Memorial Sloan Kettering Cancer Center, 1275 York Ave, Box 7, New York, NY 10065 (E-mail: [jonesd2@mskcc.org](mailto:jonesd2@mskcc.org)).

JTCVS Open 2023;16:9-16  
2666-2736

Copyright © 2023 The Author(s). Published by Elsevier Inc. on behalf of The American Association for Thoracic Surgery. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

<https://doi.org/10.1016/j.jtc.2023.10.016>

were controlled for, the random proteomic geographic diversification pattern was associated with a greater risk of recurrence or death. These studies, which used NGS, offer important therapeutic insights into the mechanisms of resistance and vulnerability in NSCLC tumors, and future studies will continue to improve our appreciation of the diverse intratumoral environment.

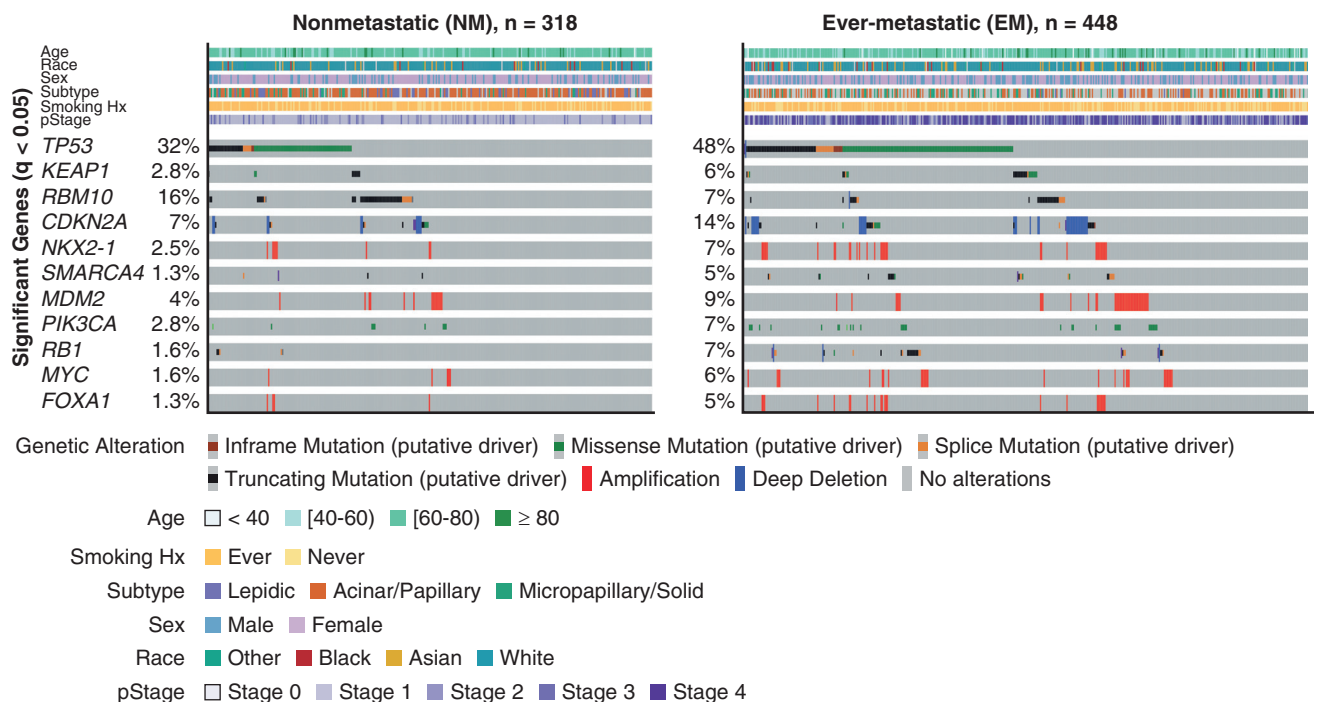
In addition to molecular heterogeneity, NSCLC displays considerable histologic heterogeneity. The most common histologic type of NSCLC is adenocarcinoma, and further division into predominant subtypes (ie, lepidic, acinar, papillary, micropapillary, and solid) has provided unique genomic insights into the risk of metastasis for these patients.<sup>9</sup> Caso and colleagues<sup>10</sup> described changes at the gene and pathway levels that are associated with histologic subtype. A greater proportion of lepidic-predominant tumors had alterations in *EGFR*, *RBM10*, and *TERT*, compared with other subtypes, whereas the more-aggressive histologic subtypes (micropapillary and solid) had enrichment of *BRAF*, *TP53*, *SETD2*, *MGA*, and *SMARCA4* mutations. The presence of an aggressive histologic subtype was also associated with greater chromosomal instability, and in the TRACERx cohort, greater chromosomal instability was associated with shorter disease-free survival (DFS) and an increased risk of metastasis.<sup>6</sup> Oncogenic pathway alterations also vary according to histologic subtype, with less-invasive lepidic-predominant tumors associated with RTK/RAS pathway changes and

more-invasive micropapillary- and solid-predominant tumors associated with Myc, p53, and Wnt pathway changes.<sup>11</sup>

### GENOMIC CHANGES ASSOCIATED WITH LUNG ADENOCARCINOMA METASTASIS

Several genes and oncogenic pathways have been associated with metastasis in early-stage lung adenocarcinoma. Lengel and colleagues<sup>12</sup> performed NGS on 766 primary lung adenocarcinoma samples and compared surgically resected primary tumors that metastasized with those that did not (Figure 1). Alterations in several genes, including *TP53*, *KEAP1*, *CDKN2A*, *MDM2*, *PIK3CA*, *NKX2-1*, *RB1*, *MYC*, *SMARCA4*, and *FOXA1*, were more common in primary lung adenocarcinomas that metastasized. Other studies have confirmed some of these findings in similar cohorts of patients.<sup>10,13,14</sup> Conversely, *RBM10* is more common in lung adenocarcinoma that does not metastasize.<sup>12</sup> In surgically resected tumors that metastasize, specific genomic alterations also influence survival; *MDM2*, *MYC*, *SMARCA4*, and *TP53* are associated with shorter metastasis-free survival, whereas *EGFR* and *NF1* are associated with longer survival after relapse.

Certain pathway alterations also increase a patient’s risk of NSCLC metastasis, including alterations in the p53, PI3K, cell cycle, and transforming growth factor-beta pathways.<sup>12</sup> Changes in the Wnt signaling pathway have also been associated with poor outcomes in lung adenocarcinoma without



**FIGURE 1.** Oncoprint comparing nonmetastatic (NM) and ever-metastatic (EM) primary lung adenocarcinoma tumors. The oncoprint displays clinical features and genes altered at significantly different frequencies between primary lung adenocarcinomas that metastasized (EM) and those that did not (NM). *Hx*, History; *pStage*, pathologic stage. (From Lengel and colleagues.<sup>12</sup> Reprinted with permission).

a clear oncogenic driver.<sup>15</sup> Cui and colleagues<sup>15</sup> showed that differential expression of 4 Wnt pathway genes (*CTNNB1*, *SOB9*, *DVL3*, and *WNT2B*) can be used to partition patients into low- and high-risk groups with respect to overall survival. This finding is supported by the work of Kim and colleagues,<sup>13</sup> who identified *CTNNB1* as an independent predictor of recurrence in early-stage lung adenocarcinoma. Finally, the total number of oncogenic pathways altered is also important. An increased number of pathways altered is not only associated with high-risk clinicopathologic features—such as maximum standardized uptake value on positron emission tomography, aggressive histologic subtype, and lymph node involvement—but is also an independent predictor of worse DFS.<sup>16</sup>

There has been a focus on driver alterations in NSCLC, and targeted therapies for an increasing number of these mutations have been developed.<sup>13</sup> However, activating mutations in *EGFR* and their association with prognosis in early-stage lung adenocarcinoma is unclear. Kim and colleagues<sup>13</sup> found that the presence of an *EGFR* mutation portended a better prognosis, whereas others have found no association between *EGFR* mutation and presence or absence of metastasis.<sup>12,13</sup> Deng and colleagues<sup>17</sup> reported that *EGFR* mutations were associated with metastatic organotropism to the brain and bone in a cohort of 1531 patients who underwent NGS; however, Lengel and colleagues<sup>12</sup> did not find a link between *EGFR* alterations and metastatic organotropism. The diverging results on the association between *EGFR* and metastasis may be attributable to the differing frequencies of *EGFR* mutations in Asian and North American populations.<sup>18</sup> *KRAS* mutations have also been implicated in recurrence of early-stage NSCLC. *KRAS* G12C alterations are associated with shorter recurrence-free survival (RFS), compared with other *KRAS* mutations, in surgically resected lung adenocarcinoma.<sup>19</sup> Furthermore, co-mutation of any *KRAS* mutation with *STK11*, *ATM*, or *LRR1Q3* was associated with shorter RFS, compared with *KRAS* mutation alone.<sup>14</sup> *KRAS* alterations have also been associated with early metastasis, which is defined as a recurrence within 2 years of surgical resection.<sup>17</sup>

Finally, individual genomic alterations are not the sole determinant of prognosis in lung adenocarcinoma, as other

genomic parameters play a major role. Multiple studies, including a recent report of TRACERx data, found that high tumor mutation burden, fraction of genome altered, and whole-genome doubling were all more common in primary lung adenocarcinoma tumors that metastasized than in those that did not.<sup>6,12</sup> Tumor mutation burden is an important biomarker, as studies have demonstrated its ability to predict response to immunotherapy in NSCLC.<sup>20</sup> These observations strongly suggest that NGS panels should investigate changes at the chromosomal level in addition to individual gene alterations.<sup>21</sup>

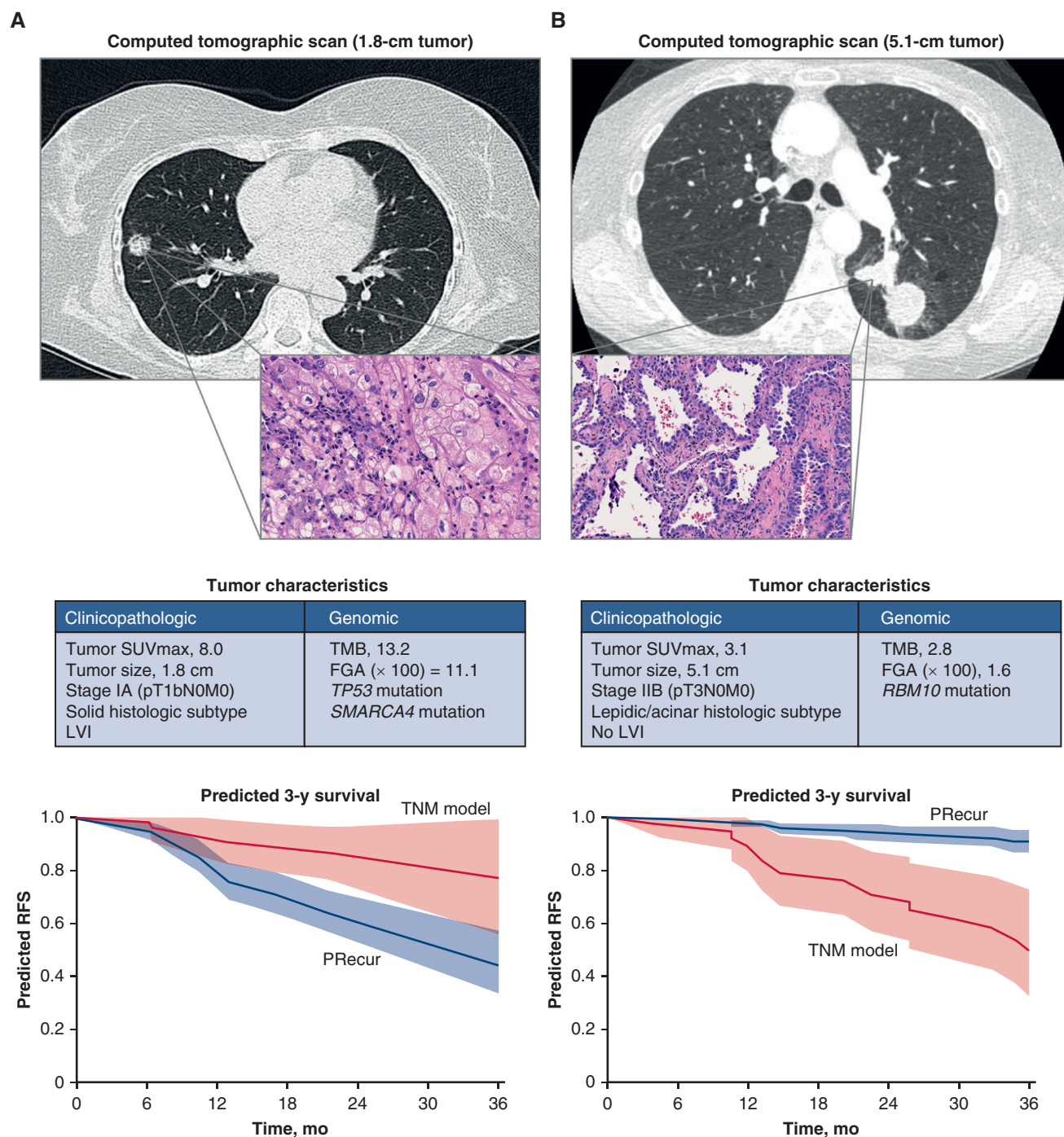
### VALIDATED MODELS PREDICTING RECURRENCE IN LUNG ADENOCARCINOMA

The identification of significant clinical, pathologic, and genomic features that influence metastasis and survival is an important aspect of properly risk-stratifying patients and provides useful information for decisions regarding treatment. Yet these factors in isolation are often inadequate, and models incorporating multiple variables to predict patient outcomes are more clinically applicable. In a cohort of patients with surgically resected early-stage lung adenocarcinoma (75% pathologic stage I), a multivariable analysis including clinicopathologic and genomic variables showed that fraction of genome altered and alterations in *SMARCA4* and *TP53* were associated with worse RFS.<sup>22</sup> Based on association analyses, Jones and colleagues created a recurrence prediction model, PRrecur, using a machine-learning framework (Table 1).<sup>22,23,E1</sup> When PRrecur was used to stratify patients on the basis of predicted RFS, it outperformed the standard prognostic tumor, node and metastasis model (concordance probability estimate, 0.73 vs 0.61;  $P < .001$ ) (Figure 2).<sup>22</sup> Interestingly, the PRrecur model correctly classified 83% of patients with stage I who developed a recurrence.

In a similar cohort of patients with nonsquamous NSCLC who underwent surgical resection, a smaller, 14-gene assay (Encore Clinical) identified patients at high risk of recurrence (Table 1).<sup>23</sup> This assay has been validated in both North American and Asian populations, and it outperformed tumor, node and metastasis staging and the NCCN high-risk guidelines for prediction of recurrence.<sup>24,25</sup> Furthermore, in a prospective cohort of 100 patients with

**TABLE 1. Genes included in select risk prediction models**

Prediction model	Genes included
Kratz et al <sup>23</sup>	<i>BAG1, BRCA1, CDC6, CD2AP1, ERBB3, FUT3, IL11, LCK, RND3, SH3BGR, WNT3A, ESD, TBP, YAP1</i>
ORACLE <sup>E1</sup>	<i>ANLN, ASPM, CDCA4, ERRF1, FURIN, GOLGA8A, ITGA6, JAG1, LRP12, MAFF, MRPS17, PLK1, PNP, PPP1R13L, PRKCA, PTTG1, PYGB, RPP25, SPEP1, SCL46A3, SNX7, TPBG, XBP1</i>
PRrecur <sup>22</sup>	<i>ALK, APC, ARD11A, ARID2, ATM, B2M, BAP1, BCOR, BRAF, BRCA2, CDK4, CDKN2A, CDKN2B, CTNNB1, EGFR, ERBB2, FAT1, FOXA1, GLI1, GNAS, KEAP1, KIT, KRAS, MDM2, MED12, MET, MGA, MYC, NF1, NF2, NKX2.1, NTRK1, PIK3CA, PIK3R1, PTPRD, PTPRT, RB1, RBM10, RET, ROS1, SETD2, SMAD4, SMARCA4, STK11, TERT, TP53, U2AF1</i>



**FIGURE 2.** Computational machine-learning prediction model (PRecur) for relapse-free survival applied to 2 patient scenarios. A, Patient with a small, 1.8-cm tumor (pT1bN0M0, stage IA). Three-year RFS curves were predicted by PRecur versus the TNM model for all patients with pT1bN0M0 in the study cohort (n = 136). B, Patient with a large, 5.1-cm tumor (pT3N0M0, stage IIB). Three-year RFS curves were predicted by PRecur versus the TNM model for all patients with pT3N0M0 in the study cohort (n = 53). *SUVmax*, Maximum standardized uptake value; *TMB*, tumor mutation burden; *FGA*, fraction of genome altered; *LVI*, lymphovascular invasion; *RFS*, recurrence-free survival; *TNM*, tumor, node and metastasis. (From Jones and colleagues.<sup>22</sup> Reprinted with permission).

TABLE 2. Genes associated with recurrence in non-small cell lung cancer

Histologic type	Genes associated with worse prognosis	Genes associated with improved prognosis
Adenocarcinoma <sup>10,12-14,17,19</sup>	<i>TP53, KEAP1, CDKN2A, MDM2, PIK3CA, NKX2-1, RB1, MYC, SMARCA4, FOXA1, CTNBN1, ALK/RET/ROS1, KRAS G12C</i>	<i>RBM10</i>
Squamous cell carcinoma <sup>E6-E8</sup>	<i>TP53, BCL6, ARID1A</i>	<i>PIK3CA</i>

stage I-IIA disease, 5-year DFS was 92% among molecularly high-risk patients who received adjuvant chemotherapy, compared with 49% among molecularly high-risk patients who did not receive adjuvant chemotherapy.<sup>24</sup> Based on these results, panels like this may be able to identify early-stage patients who would benefit from adjuvant therapy.

Given the complexity of ITH in NSCLC, Biswas and colleagues<sup>E1</sup> tested previously published RNA sequencing (RNA-seq)-based and microarray-based prognostic assays on multiple tumor regions in 48 patients. They recognized that whether a patient was classified as low risk or high risk on RNA-seq prognostic models was dependent on the tumor region analyzed for a given patient, with discordance rates of 29% to 43%. When microarray-based prognostic models were used, median discordance was 50% among 9 models, which means that half of patients could be misclassified by these models, depending on the area of the tumor selected for analysis. Therefore, using RNA-seq data to identify genes associated with intratumoral homogeneity but high variability between tumors, the authors developed a 23-gene prognostic signature, ORACLE (Table 1). ORACLE had a dramatically lower rate of discordance of 11%. Furthermore, this model has been predictive of overall survival in several external validation cohorts.<sup>E2,E3</sup>

Last, recent work by Sorin and colleagues<sup>E4</sup> has highlighted the predictive role that the tumor immune microenvironment plays with respect to patient outcomes. They used highly multiplexed imaging mass cytometry to perform spatial analysis of immune cells and their activation states and, using deep learning, identified patients who will have recurrence after surgical resection. While this method is not widely available at this time, it is a tremendous advancement that could serve as a valuable tool for future investigations.

## GENOMIC PROFILING IN LUNG SQUAMOUS CELL CARCINOMA

NGS has identified far fewer distinct genomic profiles in lung squamous cell carcinoma than in lung adenocarcinoma. Genomic alterations in lung squamous cell carcinoma appear to resemble alterations in squamous cell carcinomas of other solid organs, rather than alterations in lung adenocarcinoma.<sup>E5</sup> Sanchez-Vega and colleagues<sup>11</sup> analyzed the oncogenic signaling pathways altered in 502 lung adenocarcinoma and 464 lung squamous cell

carcinoma samples from The Cancer Genome Atlas (TCGA). Squamous cell carcinoma samples more frequently had alterations in the p53 (86% vs 61%) and PI3K (68% vs 38%) pathways and less frequently had alterations in the RTK/RAS pathway (54% vs 77%), compared with adenocarcinoma samples. Compared with genomic alterations in lung adenocarcinoma, alterations in squamous cell carcinoma have been investigated less frequently; however, several of these commonly altered genes have been associated with recurrence in patients with squamous cell carcinoma. Alterations in *TP53* have been associated with an increased risk of recurrence, whereas *PIK3CA* mutations exhibit increased time to recurrence and better overall survival.<sup>E6,E7</sup> *BCL6* and *ARID1A* alterations have also been shown to be associated with shorter DFS and overall survival, respectively, in patients with lung squamous cell carcinoma.<sup>E8</sup>

The tumor immune microenvironment appears to play a significant role in recurrence of early-stage squamous cell carcinoma, as Fan and colleagues uncovered differentially expressed immune-related genes.<sup>E9</sup> They developed a signature comprising 17 immune-related genes to predict overall survival in patients with early-stage lung squamous cell carcinoma; this gene signature has been externally validated. A summary of genes associated with both lung adenocarcinoma and squamous cell carcinoma is shown in Table 2.

## USING CIRCULATING TUMOR DNA (ctDNA) TO ASSESS METASTATIC RISK

When considering metastasis in NSCLC, providers face the challenge of risk-stratifying patients with early-stage NSCLC and how to accurately surveil them postoperatively. At present, the NCCN guidelines recommend that all patients undergo a history, physical examination, and computed tomography of the chest with or without contrast every 6 months for 2 to 3 years, and a low-dose noncontrast computed tomography of the chest annually thereafter.<sup>E10</sup> One recent development that provides an assessment of a patient's status in real time is ctDNA. LUNGCA-1, a prospective multicenter study of perioperative ctDNA in patients with NSCLC undergoing surgery, demonstrated that a high concentration of ctDNA before resection was a negative predictor of RFS.<sup>E11</sup> Conversely, in the CheckMate 816 cohort, ctDNA clearance before the last cycle of nivolumab and chemotherapy was associated with pathologic complete

response, compared with those who remained ctDNA positive (46% vs 0%); pathologic complete response was associated with better event-free survival.<sup>E12</sup> In a similar cohort of patients, Abbosh and colleagues<sup>E13</sup> found that the presence of preoperative ctDNA was more common in patients with nonadenocarcinoma lung cancer (compared with lung adenocarcinoma [92% vs 42%]), more smoking pack-years, and clinically occult mediastinal disease in lung adenocarcinoma. A landmark analysis of these patients 120 days postoperatively revealed that 93% of patients with minimal residual disease (MRD) detected by the presence of ctDNA developed recurrence. They also used ctDNA to determine that clonal expansion predicted metastasis.

Currently, there is no standard ctDNA assay, and the basis for detection in these assays can be either tumor-specific or tumor-independent (ie, epigenetic features), with variable limits of detection.<sup>E14</sup> While tumor-independent methods have the benefit of lower costs and do not require tissue acquisition, tumor-specific approaches have better limits of detection and sensitivity. In the MRD setting, sensitivity remains an issue, as a landmark analysis of a variety of ctDNA assays across multiple solid tumors revealed a median sensitivity of only 56%.<sup>E15</sup> Chaudhuri and colleagues<sup>E16</sup> improved on the previous methodology with the use of a tumor-informed NGS platform (CAPP-seq) that detects commonly mutated lung cancer genes at a lower limit of detection of 0.002%. In a cohort that included patients with stage I-III NSCLC treated with curative intent and healthy controls, receiver operating characteristic analysis revealed an area under the curve of 0.97 and maximal sensitivity and specificity of 93% and 96%, respectively. Similarly, Kurtz and colleagues<sup>E17</sup> greatly improved the limit of detection of ctDNA and thus assay sensitivity using an alternative method called phased variant enrichment and detection sequencing (PhasED-seq), which detects multiple somatic mutations in individual DNA fragments. Although these assays are not widely available in clinical practice, these 2 studies offer the most promising ctDNA assays that should be incorporated in future trials.

Concerns have also been raised regarding the predictive value of MRD ctDNA assays in the context of recurrence. The IMpower010 trial, which investigated atezolizumab versus best supportive care (BSC) after chemotherapy in patients with resected stage IB-IIIa NSCLC, used a different commercially available ctDNA assay (Signatera; Natera) to detect MRD postoperatively.<sup>E18</sup> Patients who were ctDNA-negative had better DFS than ctDNA-positive patients, regardless of which treatment they received (atezolizumab vs BSC). These results demonstrate the strong positive predictive value of these assays. However, in the BSC arm, nearly two-thirds of recurrences occurred in patients who were ctDNA negative, highlighting the low negative predictive value of the current ctDNA platforms. The

IMpower010 authors also found that ctDNA positivity increased with stage (IB = 9%, II = 14%, IIIa = 29%). The ctDNA assays presented here are only three of several platforms available. Thus, ctDNA is an exciting development in the fight against metastasis in NSCLC, and subsequent studies will need to build on the current assays to improve the applicability of this biomarker in the clinical setting.

## CONCLUSIONS

Tumor genomic profiling is now a standard-of-care tool for early-stage NSCLC and has several clinical implications. Models incorporating tumor genomic alterations serve as important predictors of metastasis and assist in our ability to risk-stratify patients. However, genomic analyses have also shown that patients with NSCLC are a diverse group who cannot be separated in neatly organized silos. Currently, clinicians caring for these patients face the challenge of synthesizing all the data that genomics has provided to make treatment decisions. Strategies incorporating the complexity of ITH in NSCLC into these prediction models will strengthen their utility and clinical applicability. As researchers better understand genetic alterations in NSCLC, additional treatments tailored to the individual patient will become available. Further, ctDNA assays that detect ctDNA preoperatively or the presence of MRD postoperatively represent a promising development for the identification of patients with early-stage disease who may benefit from neoadjuvant or adjuvant therapy. Overall, genomic profiling has the potential to move us away from a one-size-fits-all treatment approach to a personalized approach that offers patients a greater benefit to survival.

## Conflict of Interest Statement

David R. Jones serves as a consultant or speaker for or has received grant support from Merck, AstraZeneca, Genentech, More Health, and DAVA Oncology. All other authors reported no conflicts of interest.

The *Journal* policy requires editors and reviewers to disclose conflicts of interest and to decline handling or reviewing manuscripts for which they may have a conflict of interest. The editors and reviewers of this article have no conflicts of interest.

## References

1. Brandt WS, Yan W, Zhou J, Tan KS, Montecalvo J, Park BJ, et al. Outcomes after neoadjuvant or adjuvant chemotherapy for cT2-4N0-1 non-small cell lung cancer: a propensity-matched analysis. *J Thorac Cardiovasc Surg*. 2019;157:743-53.e743.
2. Potter AL, Costantino CL, Suliman RA, Haridas CS, Senthil P, Kumar A, et al. Recurrence after complete resection for non-small cell lung cancer in the National Lung Screening Trial. *Ann Thorac Surg*. 2023;116:684-92.
3. Paik S, Shak S, Tang G, Kim C, Baker J, Cronin M, et al. A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. *N Engl J Med*. 2004;351:2817-26.

4. Kalinsky K, Barlow WE, Gralow JR, Meric-Bernstam F, Albain KS, Hayes DF, et al. 21-Gene assay to inform chemotherapy benefit in node-positive breast cancer. *N Engl J Med*. 2021;385:2336-47.
5. National Comprehensive Cancer Network. Breast cancer. Version 4.2023. Accessed August 3, 2023. <https://www.nccn.org/guidelines/guidelines-detail?category=1&id=1419>
6. Jamal-Hanjani M, Wilson GA, McGranahan N, Birkbak NJ, Watkins TBK, Veeriah S, et al. Tracking the evolution of non-small-cell lung cancer. *N Engl J Med*. 2017;376:2109-21.
7. Frankell AM, Dietzen M, Al Bakir M, Lim EL, Karasaki T, Ward S, et al. The evolution of lung cancer and impact of subclonal selection in TRACERx. *Nature*. 2023;616:525-33.
8. Wu HJ, Temko D, Maliga Z, Moreira AL, Sei E, Minussi DC, et al. Spatial intratumor heterogeneity is associated with survival of lung adenocarcinoma patients. *Cell Genom*. 2022;2:100165.
9. Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisinger KR, Yatabe Y, et al. International Association for the Study of Lung Cancer/American Thoracic Society/European Respiratory Society international multidisciplinary classification of lung adenocarcinoma. *J Thorac Oncol*. 2011;6:244-85.
10. Caso R, Sanchez-Vega F, Tan KS, Mastrogiacomo B, Zhou J, Jones GD, et al. The underlying tumor genomics of predominant histologic subtypes in lung adenocarcinoma. *J Thorac Oncol*. 2020;15:1844-56.
11. Sanchez-Vega F, Mina M, Armenia J, Chatila WK, Luna A, La KC, et al. Oncogenic signaling pathways in The Cancer Genome Atlas. *Cell*. 2018;173:321-37.e310.
12. Lengel HB, Mastrogiacomo B, Connolly JG, Tan KS, Liu Y, Fick CN, et al. Genomic mapping of metastatic organotropism in lung adenocarcinoma. *Cancer Cell*. 2023;41:970-85.e973.
13. Kim IA, Hur JY, Kim HJ, Park JH, Hwang JJ, Lee SA, et al. Targeted next-generation sequencing analysis for recurrence in early-stage lung adenocarcinoma. *Ann Surg Oncol*. 2021;28:3983-93.
14. Kadara H, Choi M, Zhang J, Parra ER, Rodriguez-Canales J, Gaffney SG, et al. Whole-exome sequencing and immune profiling of early-stage lung adenocarcinoma with fully annotated clinical follow-up. *Ann Oncol*. 2017;28:75-82.
15. Cui Y, Fang W, Li C, Tang K, Zhang J, Lei Y, et al. Development and validation of a novel signature to predict overall survival in "driver gene-negative" lung adenocarcinoma (LUAD): results of a multicenter study. *Clin Cancer Res*. 2019;25:1546-56.
16. Zhou J, Sanchez-Vega F, Caso R, Tan KS, Brandt WS, Jones GD, et al. Analysis of tumor genomic pathway alterations using broad-panel next-generation sequencing in surgically resected lung adenocarcinoma. *Clin Cancer Res*. 2019;25:7475-84.
17. Deng C, Zhang Y, Fu F, Ma X, Wen Z, Ma Z, et al. Genetic-pathological prediction for timing and site-specific recurrence pattern in resected lung adenocarcinoma. *Eur J Cardio Thorac Surg*. 2021;60:1223-31.
18. Zhang YL, Yuan JQ, Wang KF, Fu XH, Han XR, Threapleton D, et al. The prevalence of EGFR mutation in patients with non-small cell lung cancer: a systematic review and meta-analysis. *Oncotarget*. 2016;7:78985-93.
19. Jones GD, Caso R, Tan KS, Mastrogiacomo B, Sanchez-Vega F, Liu Y, et al. KRAS (G12C) mutation is associated with increased risk of recurrence in surgically resected lung adenocarcinoma. *Clin Cancer Res*. 2021;27:2604-12.
20. Hellmann MD, Ciuleanu T-E, Pluzanski A, Lee JS, Otterson GA, Audigier-Valette C, et al. Nivolumab plus ipilimumab in lung cancer with a high tumor mutational burden. *N Engl J Med*. 2018;378:2093-104.
21. Leader AM, Grout JA, Maier BB, Nabet BY, Park MD, Tabachnikova A, et al. Single-cell analysis of human non-small cell lung cancer lesions refines tumor classification and patient stratification. *Cancer Cell*. 2021;39:1594-609.e1512.
22. Jones GD, Brandt WS, Shen R, Sanchez-Vega F, Tan KS, Martin A, et al. A Genomic-pathologic annotated risk model to predict recurrence in early-stage lung adenocarcinoma. *JAMA Surg*. 2021;156:e205601.
23. Kratz JR, He J, Van Den Eeden SK, Zhu ZH, Gao W, Pham PT, et al. A practical molecular assay to predict survival in resected non-squamous, non-small-cell lung cancer: development and international validation studies. *Lancet*. 2012;379:823-32.
24. Woodard GA, Wang SX, Kratz JR, Zoon-Besselink CT, Chiang CY, Gubens MA, et al. Adjuvant chemotherapy guided by molecular profiling and improved outcomes in early stage, non-small-cell lung cancer. *Clin Lung Cancer*. 2018;19:58-64.
25. Haro GJ, Sheu B, Cook NR, Woodard GA, Mann MJ, Kratz JR. Comparison of conventional TNM and novel TNMB staging systems for non-small cell lung cancer. *JAMA Netw Open*. 2019;2:e1917062.

**Key Words:** early-stage non-small cell lung cancer, genomics, metastasis, tumor heterogeneity, ctDNA

**E-References**

- E1. Biswas D, Birkbak NJ, Rosenthal R, Hiley CT, Lim EL, Papp K, et al. A clonal expression biomarker associates with lung cancer mortality. *Nat Med*. 2019;25:1540-8.
- E2. Djureinovic D, Hallström BM, Horie M, Mattsson JSM, La Fleur L, Fagerberg L, et al. Profiling cancer testis antigens in non-small-cell lung cancer. *JCI Insight*. 2016;1:e86837.
- E3. Okayama H, Kohno T, Ishii Y, Shimada Y, Shiraishi K, Iwakawa R, et al. Identification of genes upregulated in ALK-positive and EGFR/KRAS/ALK-negative lung adenocarcinomas. *Cancer Res*. 2012;72:100-11.
- E4. Sorin M, Rezanejad M, Karimi E, Fiset B, Desharnais L, Perus LJM, et al. Single-cell spatial landscapes of the lung tumour immune microenvironment. *Nature*. 2023;614:548-54.
- E5. Campbell JD, Alexandrov A, Kim J, Wala J, Berger AH, Pedamallu CS, et al. Distinct patterns of somatic genome alterations in lung adenocarcinomas and squamous cell carcinomas. *Nat Genet*. 2016;48:607-16.
- E6. Al Bakir M, Huebner A, Martínez-Ruiz C, Grigoriadis K, Watkins TBK, Pich O, et al. The evolution of non-small cell lung cancer metastases in TRACERx. *Nature*. 2023;616:534-42.
- E7. McGowan M, Hoven AS, Lund-Iversen M, Solberg S, Helland Å, Hirsch FR, et al. PIK3CA mutations as prognostic factor in squamous cell lung carcinoma. *Lung Cancer*. 2017;103:52-7.
- E8. Connolly JG, Tan KS, Sanchez-Vega F, Jones GD, Liu Y, Caso R, et al. A23 A Genomically adjusted clinicopathologic model predicts recurrence in resected early-stage lung squamous cell carcinoma. *J Thorac Oncol*. 2020;15:S19-20.
- E9. Fan T, Lu Z, Liu Y, Wang L, Tian H, Zheng Y, et al. A novel immune-related seventeen-gene signature for predicting early stage lung squamous cell carcinoma prognosis. *Front Immunol*. 2021;12:665407.
- E10. National Comprehensive Cancer Network. Non-Small Cell Lung Cancer; Version 3.2023. Accessed August 3, 2023. <https://www.nccn.org/guidelines/guidelines-detail?category=1&id=1450>
- E11. Xia L, Mei J, Kang R, Deng S, Chen Y, Yang Y, et al. Perioperative ctDNA-based molecular residual disease detection for non-small cell lung cancer: a prospective multicenter cohort study (LUNGCA-1). *Clin Cancer Res*. 2022;28:3308-17.
- E12. Forde PM, Spicer J, Lu S, Provencio M, Mitsudomi T, Awad MM, et al. Neoadjuvant nivolumab plus chemotherapy in resectable lung cancer. *N Engl J Med*. 2022;386:1973-85.
- E13. Abbosh C, Frankell AM, Harrison T, Kisistok J, Garnett A, Johnson L, et al. Tracking early lung cancer metastatic dissemination in TRACERx using ctDNA. *Nature*. 2023;616:553-62.
- E14. Bestvina CM, Garassino MC, Neal JW, Wakelee HA, Diehn M, Vokes EE. Early-stage lung cancer: using circulating tumor DNA to get personal. *J Clin Oncol*. 2023;JCO2300258.
- E15. Moding EJ, Nabet BY, Alizadeh AA, Diehn M. Detecting liquid remnants of solid tumors: circulating tumor DNA minimal residual disease. *Cancer Discov*. 2021;11:2968-86.
- E16. Chaudhuri AA, Chabon JJ, Lovejoy AF, Newman AM, Stehr H, Azad TD, et al. Early detection of molecular residual disease in localized lung cancer by circulating tumor DNA profiling. *Cancer Discov*. 2017;7:1394-403.
- E17. Kurtz DM, Soo J, Co Ting Keh L, Alig S, Chabon JJ, Sworder BJ, et al. Enhanced detection of minimal residual disease by targeted sequencing of phased variants in circulating tumor DNA. *Nat Biotechnol*. 2021;39:1537-47.
- E18. Zhou C, Das Thakur M, Srivastava MK, Zou W, Xu H, Ballinger M, et al. 20 IMPower010: Biomarkers of disease-free survival (DFS) in a phase III study of atezolizumab (atezo) vs best supportive care (BSC) after adjuvant chemotherapy in stage IB-IIIa NSCLC. *Ann Oncol*. 2021;32:S1374.