Age-dependent therapeutic effects of liver X receptor-α activation in murine polymicrobial sepsis



Innate Immunity 2015, Vol. 21(6) 609–618 © The Author(s) 2015 Reprints and permissions: sagepub.co.uk/journalsPermissions.nav DOI: 10.1177/1753425915569367 ini.sagepub.com



Gabriela Botez¹, Giovanna Piraino¹, Paul W Hake, John R Ledford¹, Michael O'Connor¹, James A Cook² and Basilia Zingarelli¹

Abstract

The severity of sepsis is significantly affected by advanced age; however, age-dependent molecular mechanisms of this susceptibility are unknown. Nuclear liver X receptor- α (LXR α) is a regulator of lipid metabolism with associated antiinflammatory properties. Here, we investigated the role of LXR α in age-dependent lung injury and outcome of sepsis. Male C57BL/6, LXR α -deficient (LXR $\alpha^{-/-}$) and wild type (WT) (LXR $\alpha^{+/+}$) mice of different ages were subjected to sepsis by cecal ligation and puncture (CLP). In pharmacological studies, treatment with the LXR α ligand T0901317 reduced lung neutrophil infiltration in C57BL/6 mice aged from 1 to 8 mo when compared with vehicle-treated animals subjected to CLP. The LXR α ligand improved survival in young mice (2–3 mo old) but did not affect survival or neutrophil infiltration in mature adult mice (11–13 mo old). Immunoblotting revealed an age-dependent decrease of lung LXR α levels. Young LXR $\alpha^{-/-}$ mice (2–3 mo old) exhibited earlier mortality than age-matched WT mice after CLP. Lung damage and neutrophil infiltration, lung activation of the pro-inflammatory NF- κ B and plasma IL-6 levels were higher in LXR $\alpha^{-/-}$ mice 18 h after CLP compared with LXR $\alpha^{+/+}$ mice. This study suggests that the anti-inflammatory properties of LXR α in sepsis are age-dependent and severely compromised in mature adult animals.

Keywords

Liver X receptor- α , T0901317, sepsis, lung injury, NF- κ B, age, neutrophil infiltration

Date received: 30 July 2014; revised: 29 November 2014; accepted: 5 January 2015

Introduction

Sepsis is a systemic response to infection characterized by hemodynamic and metabolic derangements that may result in septic shock, multiple organ system failure and death. Although it occurs in people of all ages, the incidence and mortality of sepsis is disproportionately increased in elderly adults.¹ Currently, the treatment of sepsis is centered on general organ supportive measures, ventilator strategies and antibiotic therapy to control the infectious focus. However, these approaches are not always efficacious in reducing the high mortality.² Furthermore age-dependent pathophysiological mechanisms of the systemic inflammatory process, innate immune dysfunction and organ damage are yet to be identified.

The liver X receptor (LXR) is a member of the nuclear receptor superfamily, which is involved in cholesterol, fatty acid and Glc homeostasis, and activated by oxidized forms of cholesterol (oxysterols) and intermediate products of the cholesterol biosynthetic pathway, as well as by synthetic ligands, such as the T0901317.^{3–5} Two LXR subtypes have been identified, LXR α and LXR β , with different tissue distribution. LXR α is expressed in lung, macrophages, liver, spleen, kidney, adipose tissue and small intestine, whereas LXR β is ubiquitously expressed.^{6,7} In addition

Corresponding author:

Email: basilia.zingarelli@cchmc.org

¹Division of Critical Care Medicine, Cincinnati Children's Hospital Medical Center, Department of Pediatrics, College of Medicine, University of Cincinnati, Cincinnati, OH, USA

²Department of Neurosciences, Medical University of South Carolina, Charleston, SC, USA

Basilia Zingarelli, Cincinnati Children's Hospital Medical Center, Division of Critical Care Medicine, 3333 Burnet Avenue, Cincinnati, OH 45229, USA.

to its cholesterol-regulating properties, LXRa has been reported to control transcriptional programs involved in the inflammatory response. For example, in LPSstimulated murine macrophages, LXRa activation inhibits the expression of pro-inflammatory cytokines and inducible nitric oxide synthetase.^{8,9} In vivo experimental studies have confirmed the anti-inflammatory actions of LXRa in models of LPS-induced lung injury and carrageenan-induced pleurisy.^{10,11} Our laboratory has also demonstrated that treatment of rats with the LXRa synthetic agonist T0901317 reduced plasma cytokines and chemokines, lung neutrophil infiltration and histological damage after hemorrhagic shock.¹² These in vitro and in vivo experimental studies have demonstrated that a plausible anti-inflammatory mechanism for LXR α is through inhibition of NF- κ B,^{8,9} a transcription factor central to many pro-inflammatory and immune genes involved in sepsis.¹³ However, in these in vivo studies the anti-inflammatory effects of LXRa activation have been determined in young rodents only. The ability of LXRa to modulate inflammation in the complex event of an infectious disease setting in aged organisms remains to be determined.

Based on these previous findings, we investigated the biological effects of LXR^{\alpha} modulation on the inflammatory response of polymicrobial sepsis in mice of different ages. Specifically, in *in vivo* pharmacological studies, we demonstrated that treatment with the LXRa ligand T0901317 ameliorated the extent of the sepsis-induced inflammatory response in mice of younger age (1-8 mo old) but not in mature, adult mice (11-13 mo old), which instead exhibited a decrease of the receptor levels in the lung. We also demonstrated that LXR α gene deletion in young septic mice (2–3 mo old) was associated with increased susceptibility to sepsis-induced inflammation, lung injury and earlier mortality. Thus, our data provide evidence that LXRa is an important modulator of innate immune response in sepsis; however, its function declines with age, limiting its therapeutic potentials.

Materials and methods

Murine model of polymicrobial sepsis

The investigation conformed to the National Institutes of Health Guide for the Care and Use of Laboratory Animals and commenced with the approval of the Institutional Animal Care and Use Committee. In pharmacological studies, male C57BL/6 mice (Charles River Laboratories, Wilmington MA, USA) were used at an age of between 1 and 1.5 mo (very young group), 2 and 3 mo (young group), 6 and 8 mo (adult group) or between 11 and 13 mo (mature adult group). In complementary studies, male wild type (WT) (LXR $\alpha^{+/+}$) and LXR α -deficient (LXR $\alpha^{-/-}$) mice, both on a mixed-strain background (C57BL/6:129SvEv), were used at an

age of between 2 and 3 mo (young group) or between 11 and 13 mo (mature adult group). Polymicrobial sepsis was induced by an established model of cecal ligation and puncture (CLP).¹⁴ Mice were anesthetized with pentobarbital (40 mg/kg) i.p. After opening the abdomen, the cecum was exteriorized and ligated by a 6-0 silk ligature at its base without obstructing intestinal continuity. The cecum was punctured twice with a 22-G needle and squeezed to excrete small amount of fecal material into the peritoneal cavity. The cecum was then returned into the peritoneal cavity and the abdominal incision was closed with a 6-0 silk ligature suture. After the surgical procedure, mice were resuscitated with 0.6 ml normal saline solution (s.c.) to replace the fluid and blood loss during operation. In pharmacological studies, C57BL/6 mice received vehicle or the LXRa ligand T0901317 (30 mg/kg i.p.) at 1, 6 and 12 h after CLP. In time-course studies, mice (n = 4-19) were sacrificed at 3, 6 and 18h after CLP. Blood and plasma samples, peritoneal fluid, lung, liver and spleen were collected for histological and biochemical studies described below. In survival studies, mice (n = 12-20)in each group) were monitored for mortality rates for 72 h.

Histopathological analysis

Lungs were fixed in 4% paraformaldehyde and embedded in paraffin. Sections were stained with hematoxylin and eosin, and evaluated by five independent observers who were unaware of the experimental protocol. Specifically, lung injury was analyzed by a semiquantitative score based on the following histologic features: (a) alveolar congestion; (b) hemorrhage; (c) infiltration of neutrophils in airspace or vessel wall; and (d) thickness of alveolar wall/hyaline membrane formation. Each feature was graded from 0 to 4 (i.e. absent, minimal, mild, significant, or severe). The four variables were summed to represent the lung injury score (total score, 0–16).

Myeloperoxidase activity

Myeloperoxidase (MPO) activity was measured as an indicator of neutrophil infiltration in lung tissue after septic shock. Tissues were homogenized in a solution containing 0.5% hexa-decyl-trimethyl-ammonium bromide dissolved in 10 mM potassium phosphate buffer (pH 7.0) and centrifuged for $30 \min at 4000 g$ at 4°C. An aliquot of the supernatant was allowed to react with a solution of tetra-methyl-benzidine (1.6 mM) and hydrogen peroxide (0.1 mM). The rate of change in absorbance was measured by spectrophotometry at 650 nm. MPO activity was defined as the quantity of enzyme degrading 1 µmol of hydrogen peroxide/min at 37°C and expressed in units per 100 mg mass of tissue.

Cytosol and nuclear protein extraction

Lungs were homogenized using a Polytron homogenizer (Brinkman Instruments, West Orange, NY, USA) in a buffer containing 0.32 M sucrose, 10 mM Tris-HCl (pH 7.4), 1mM EGTA, 2mM EDTA, 5mM NaN₃, 10 mM β -mercaptoethanol, $20 \mu \text{M}$ leupeptin, $0.15 \mu \text{M}$ pepstatin A, 0.2 mM phenylmethanesulfonyl fluoride, 50 mM NaF, 1 mM sodium orthovanadate and 0.4 nM microcystin. Samples were centrifuged at 1000 g for 10 min at 4°C and the supernatants collected as cytosol extracts. The pellets were then solubilized in Triton buffer (1% Triton X-100, 250 mM NaCl, 50 mM Tris HCl at pH 7.5, 3mM EGTA, 3mM EDTA, phenylmethanesulfonyl fluoride, 0.1 mM 0.1 mM sodium orthovanadate, 10% glycerol, 2 mM pnitrophenyl phosphate, 0.5% NP-40 and 46 µM aprotinin). The lysates were centrifuged at 15,000 g for 30 min at 4°C and the supernatant collected as nuclear extracts.

Western blot analysis

Cytosol and nuclear content of LXR α was determined by immunoblot analysis on nitrocellulose membranes using primary Abs against LXR α and secondary peroxidase-conjugated Ab. Membranes were also reprobed with primary Ab against β -actin to ensure equal loading samples. Immunoreaction was visualized by chemiluminescence. Densitometric analysis of blots was performed using Quantity One (Bio-Rad Laboratories, Des Plaines, IL, USA).

Measurement of NF-kB DNA binding activity

NF- κ B DNA binding activity was analyzed by a TransAM Transcription Factor assay kit specific for the activated form of p65 (Active Motif, Carlsbad, CA, USA) using the NF- κ B consensus site (5'–GGG ACTTTCC-3') according to the manufacturer's protocol.

Bacterial colony counts

Bacterial clearance was indirectly assessed by counting bacterial colonies in blood and peritoneal fluid, and homogenized samples of spleen, liver and lung at 18 h after CLP surgery. Serial dilutions of samples were cultured on 5% sheep blood trypticase soy agar plates (BBL Stacker Plate TSA II, BD Biosciences, Sparks, MD, USA) and incubated at 37°C in aerobic conditions for 24 h. Colony forming units (CFU) were counted and log-transformed to obtain a normal distribution. Results were expressed as log CFU/ml for blood, log CFU/g for tissues and log CFU/ mouse for peritoneal lavage fluid.

Plasma levels of cytokines and adipokines

Plasma levels of IL-6 and TNF- α were evaluated by a commercially available solid-phase ELISA kit (R&D Systems, Minneapolis, MN, USA). Plasma levels of resistin and leptin were evaluated by a commercially available multiplex array system (Linco-Research, St. Charles, MO, USA) using the protocols recommended by the manufacturer.

Materials

The LXR α agonist T0901317 was obtained from Cayman Chemical (Ann Arbor, MI, USA). The primary Ab directed at LXR α was obtained from Abcam (Cambridge, MA, USA). The secondary Abs and β -actin were obtained from Santa Cruz Biotechnology (Santa Cruz, CA, USA). All other chemicals were obtained from Sigma-Aldrich (St. Louis, MO, USA).

Data analysis

Statistical analysis was performed using SigmaStat for Windows Version 3.10 (SysStat Software, San Jose, CA, USA). Data in the figures and text are expressed as means \pm SEM or median with 25th and 75th percentiles of *n* observations (n = 4-19 animals for each group). The results were examined by ANOVA followed by the Student–Newman–Keuls's correction post hoc *t*-test. Statistical analysis of damage scores was performed using the Mann–Whitney Rank Sum test; when normality and equal variance passed, data were further analyzed by *t*-test. The Gehan–Breslow and log-rank tests were used to compare differences in survival rates. A value of P < 0.05 was considered significant.

Results

Pharmacological activation of $LXR\alpha$ improves survival in young but not mature adult mice

In order to achieve a similar magnitude of organ inflammatory response and mortality between different groups of age and to evaluate the potential therapeutic efficacy of LXR α activation, we used a severe model of CLP (22-G needle) without antibiotics to produce high mortality during the early phase. With this procedure, the onset of clinical symptoms in C57BL/6 mice, such as piloerection and inactivity, were observed by 6–12 h. When survival curves were monitored for 3d after CLP, mortality rates were similar between vehicletreated young (2-3 mo old) and mature adult (11-13 mo old) groups (Figure 1). Treatment with the LXRa ligand T0901317 improved survival in young mice (P = 0.049); however, it did not modify survival rate in mature adult mice when compared with agematched vehicle-treated mice (P = 0.941) (Figure 1).

infiltration in mature neutrophil adult mice (11–13 mo) when compared with age-matched vehicletreated mice, thus suggesting the potential of nonspecific toxic effects (Figure 2).

Age-dependent down-regulation of LXR α in the lung

As pharmacological activation of LXRa afforded protection in young and adult animals only, but not in the mature adult groups, we investigated whether sepsis was associated with age-dependent changes in LXRa levels. By Western blot analysis, we observed that, when compared with basal levels of age-matched sham animals, cytosol and nuclear content of LXRa in lungs was reduced at 6h after CLP in both young and mature adult groups. In sham mature adult mice, however, total levels of protein were lower when compared with the young group (Figure 3). At 18 h after CLP, there was a partial replenishment of this receptor in both young and mature adult mice; however, total levels of LXR α were lower, although not statistically different, in mature adult groups when compared with young mice. In addition, mature adult mice exhibited retention of the receptor in the cytosol when compared with young mice (Figure 3).

Genetic ablation of LXR α exacerbates early mortality rate in young mice

To confirm the protective role of LXR α in sepsis, we performed additional survival studies in genetically LXRa-deficient mice and their WT controls. When analysis of survival rate was focused only on the initial period of sepsis, we observed that death occurred earlier in young $LXR\alpha^{-/-}$ mice and in both mature adult LXR $\alpha^{+/+}$ and mature adult LXR $\alpha^{-/-}$ mice when compared with young LXR $\alpha^{+/+}$ WT animals (P=0.044 at 36 h after CLP) (Figure 4). However, at 72 h after CLP, mortality rates (approximately 90-100%) were similar between the young (2-3 mo old) and mature adult (11-13 mo old) groups of both genotypes (data not shown).

Genetic ablation of LXR α exacerbates lung injury in young mice

Having determined that LXRa ablation could reduce short-term survival, we focused on organ injury changes. Given the early high mortality observed in this model of CLP, examination was limited to early time points of CLP in the young group only. In the lung of WT mice, histological examination revealed extravasation of red cells and accumulation of neutrophils into the air spaces at 18 h after CLP (Figure 5). In LXR $\alpha^{-/-}$ mice, lung injury was more severe and consisted of reduced alveolar air spaces, large hemorrhagic areas and infiltration of inflammatory cells. We also measured activity of MPO to confirm tissue

Figure 2. MPO activity in lung at 6 h after CLP. Data are means \pm SEM of 5–18 mice for each group. *P < 0.05 vs, age-matched sham mice; #P < 0.05 vs. vehicle treatment of age-matched group. Mice received T0901317 (30 mg/kg i.p.) or

2.0-3.0

6.0-8.0

11.0-13.0

Age-dependent effect of pharmacological activation of LXR α in sepsis-induced lung injury

As a serious consequence of sepsis is the occurrence of multiple organ failure, which is preceded by accumulation of neutrophils in major vital organs, we evaluated the effect of the LXRa ligand T0901317 on sepsisinduced lung neutrophil infiltration by measurement of the activity of MPO, an enzyme specific to neutrophil lysosomes. Treatment with T0901317 (30 mg/kg i.p.) reduced MPO activity in different age groups of mice (from 1 to 8 mo) compared with their age-matched vehicle-treated mice subjected to CLP (Figure 2). On the contrary, treatment with T0901317 worsened

Figure 1. Kaplan-Meier curve of survival monitored for 72 h. *P=0.049 by Gehan-Breslow analysis. Male C57BL/6 mice received T0901317 (30 mg/kg i. p.) or vehicle at 1, 6 and 12 h following CLP.

40

Hours after CLP

60

Young CLP + Vehicle (n12)

Young CLP + T0901317 (n=18)

Mature CLP + T0901317 (n=20)

80

Mature CLP + Vehicle (n=15)



1.0

0.8

0.6

0.4

0.2

0.0

0

150

100

50

0

vehicle at 1 h following CLP.

1.0-1.5

20

Survival (log rank)



Figure 3. (a) Western blot of cytosol and nuclear LXR α content and β -actin (used as loading control protein) in the lung at 0, 6 and 18 h after CLP. (b) Image analyses of LXR α content as determined by densitometry. Data are mean \pm SEM of 4–12 animals for each group and are expressed as relative intensity units. **P* < 0.05 vs. age-matched controls; #*P* < 0.05 vs. the young group; $\ddagger P < 0.05$ vs. 6 h time point of age matched-mice.

neutrophil infiltration. Lung neutrophil infiltration was similar in the two genotypes in the earlier phases of sepsis at 3 and 6 h after CLP. However, neutrophil infiltration was higher in $LXR\alpha^{-/-}$ mice when compared with WT mice at 18 h after CLP, probably signifying an inability to turn off the inflammatory process (Figure 5f). To examine the effect of genetic absence of LXR α on the capability to eliminate bacterial pathogens, we counted bacterial CFU in blood, peritoneal fluid, lung, spleen and liver at 18 h after CLP. The numbers of bacterial colonies were similar in all these compartments in $LXR\alpha^{+/+}$ and $LXR\alpha^{-/-}$ mice (Table 1).

Genetic ablation of LXR α increases systemic elevation of IL-6 in young mice

Increased early mortality of $LXR\alpha^{-/-}$ mice was associated with higher plasma levels of the pro-inflammatory IL-6 when compared with WT mice at 18 h after CLP. Plasma levels of other pro-inflammatory cytokines and adipokines, such as TNF- α , resistin and leptin, were similarly elevated in both genotypes (Figure 6).

Decrease of $LXR\alpha$ content is associated with up-regulation of NF- κ B activation in the lung following polymicrobial sepsis

To determine the inflammatory signaling pathways affected by LXR α genetic deletion, we also investigated the kinetics of NF- κ B activation. NF- κ B activity was similar in the two genotypes in the earlier phases of sepsis. However, NF- κ B activity persisted at higher levels in LXR $\alpha^{-/-}$ mice when compared with WT mice at 18 h after CLP (Figure 7a). Interestingly, by Western blot analysis, we observed that the nuclear content of LXR α decreased in a time-dependent manner up to 6 h after CLP and was then replenished at 18 h in lung nuclear extracts of WT LXR $\alpha^{+/+}$ mice (Figures 7b, c). These data are consistent with the lung injury pattern, which was different in the two genotypes at 18 h.



Figure 4. Kaplan–Meier curve of survival monitored for 36 h. *P = 0.049 by log-rank analysis vs. young LXR $\alpha^{-/-}$, mature adult LXR $\alpha^{+/+}$ and mature adult LXR $\alpha^{-/-}$.

Discussion

Our studies demonstrate that pharmacological activation of LXR α by T0901317 reduces pulmonary neutrophil infiltration and improves survival in young mice subjected to polymicrobial sepsis. On the contrary, our studies demonstrate that mature adult mice have less total LXR α , thus losing their capability to respond to the protective effects of the LXR α ligand. Thus, our data suggest that LXR α is an important regulator of the innate immune response; however, a decrease of LXR α content in the lung may represent an additional mechanism underlying the age-dependent poor recovery in sepsis.

LXR α is a nuclear receptor with widely recognized roles in cholesterol metabolism and inflammation. In the lung, the receptor is expressed in macrophages, neutrophils, type II pneumocytes and airway smooth muscle cells.^{7,15,16} Therefore, it is biologically plausible that LXR α may play a part in the delicate pro- vs. antiinflammatory balance. Previous work has already shown that activation of LXR α attenuates inflammation in a variety of preclinical models of lung injury in young rodents, such as LPS administration,^{10,17} pneumonia,¹⁵ carrageenan-induced pleurisy¹¹ and hemorrhagic shock.¹²

Acute lung injury involves intrapulmonary neutrophil accumulation and activation with the release of cytotoxic compounds, which leads to loss of the alveolar epithelial and capillary endothelial barrier function and increase in permeability of the alveolo-capillary membrane.^{18,19} In the present study, we used complementary pharmacological gain-of-function and genetic loss-of-function approaches to demonstrate that LXRa signaling is instrumental in modulating neutrophil infiltration in sepsis-induced lung injury. Treatment of C57BL/6 mice (1–8 mo old) with the LXR α ligand T0901317 significantly reduced pulmonary tissue MPO activity compared with the vehicle-treated group. Conversely, in young LXRa-deficient animals (2–3 mo old), the degree of lung neutrophil infiltration was significantly higher than in the lungs of WT animals at a late time after sepsis and was associated with worse histological damage. These findings are consistent with our previous work demonstrating that LXRa activation leads to a reduction in lung neutrophil infiltration in a model of hemorrhagic shock,¹² as well as the work of others using models of LPS-induced lung injury,¹⁰ pneumonia¹⁵ and carrageenan-induced pleurisy.11

Early therapeutic intervention is critical for a positive outcome in septic patients.²⁰ However, the early pathophysiological events of organ damage have not been yet defined. Given the dynamic nature of the inflammatory cascade that leads to sepsis-induced lung injury, we determined the kinetics of $LXR\alpha$ levels after the induction of sepsis. We found that lung content of LXR α in LXR $\alpha^{+/+}$ mice was significantly lower at the early time points after CLP compared with basal levels and it partially recovered at 18 h later. Of more importance, the temporal partial recovery of LXR α nuclear content in LXR $\alpha^{+/+}$ mice was associated with lower pulmonary leukocyte recruitment, less lung damage and lower levels of the proinflammatory cytokine IL-6 when compared with septic LXR $\alpha^{-/-}$ mice. Interestingly, plasma levels of TNF- α , resistin and leptin were not different between the two genotypes. Thus, deficiency of $LXR\alpha^{-/-}$ seems to negatively affect the outcome of septic animals, probably by enhancing the pro-inflammatory cytokine IL-6 and lung neutrophil infiltration, while not affecting the production of the adipokines resistin and leptin. Other studies confirm that LXRa is an important contributor of the protective immune response during infections by modulating the cytokine response.²¹ Furthermore, LXRa activation has been shown to potently inhibit the release of IL-6 and the development of experimental skin fibrosis in mice.²² Taken together with these previous reports, our data suggest that LXR α is a part of the early regulatory host response to CLP-induced sepsis and may be used as a therapeutic target; its initial decrease may favor pro-inflammatory mechanisms, while its subsequent replenishment may promote anti-inflammatory mechanisms, as the body attempts to limit potentially damaging inflammation.

Our results are consistent with other studies, which have shown that the content of LXR α decreases after LPS-induced lung injury.¹⁰ Interestingly, a similar



Figure 5. Representative histology photomicrographs of lung sections of WT LXR $\alpha^{+/+}$ and LXR $\alpha^{-/-}$ mice subjected to polymicrobial sepsis. Lungs were harvested at 18 h after CLP and were stained with hematoxylin and eosin. Normal lung architecture in (a, b) sham mice of both genotypes and (c, d) lung derangement in septic mice at 18 h after CLP (c, d). Magnification × 100. A similar pattern was seen in 4–6 different tissue sections in each experimental group. (e) Histopathological scores of lung sections (n = 5 mice for each group). Lung injury was scored from 0 (no damage) to 16 (maximum damage). Box plots represent 25th percentile, median and 75th percentile; error bars define 10th and 90th percentiles. (f) Time-course analysis of lung MPO activity after CLP in WT LXR $\alpha^{+/+}$ mice and LXR $\alpha^{-/-}$ mice. Data represent the mean ± SEM of 4–9 animals per group. *P < 0.05 vs. LXR $\alpha^{+/+}$ mice.

dynamic of LXR α levels was observed in the livers of mice subjected to CLP,²³ supporting the notion that LXR α might be involved in regulating sepsis-related inflammation in a variety of organs. Our results are also consistent with our previous reports demonstrating that the acute phase response of septic shock, or endotoxemia, is associated with a marked decrease of other nuclear hormone receptors of the peroxisome proliferator-activated receptor (PPAR) family, such as PPAR α , PPAR γ and PPAR δ , which share with LXR α important regulatory functions in lipid and

Glc metabolism.^{24–26} Experimental *in vitro* studies further support that the inflammatory response induced by pro-inflammatory cytokines can directly induce changes in the levels of metabolic nuclear receptors including PPAR α , PPAR γ and LXR α in Hep3B human hepatoma cells.²⁷ Other studies have also demonstrated that aging is associated with reduced activity of several nuclear receptors, including LXR α , in rat brain.²⁸ Although the mechanisms of protein loss have not yet determined, it is possible that posttranslational modifications of the nuclear receptor may influence the protein stability and its dynamic changes. In fact, it has been previously demonstrated that LXR α deacetylation favors its ubiquitination and subsequent degradation, whereas specific LXR α ligands prevent protein degradation and increase the recruitment of the receptor to the target gene promoters.^{29,30}

In the present study, young LXR α -deficient mice exhibited significantly higher NF- κ B activity in the lung at 18h after CLP compared with LXR $\alpha^{+/+}$

Table 1. Tissue bacterial content in young $LXR\alpha^{+/+}$ and $LXR\alpha^{-/-}$ mice.

Tissues	LXRa ^{+/+} mice CFU (log)	LXRα ^{-/-} mice CFU (log)
Blood	$\textbf{3.48} \pm \textbf{0.34}$	$\textbf{3.08} \pm \textbf{0.44}$
Peritoneal fluid	4.73 ± 0.47	$\textbf{4.46} \pm \textbf{0.46}$
Lung	2.31 ± 0.26	2.11 ± 0.28
Spleen	$\textbf{2.96} \pm \textbf{0.29}$	2.55 ± 0.23
Liver	$\textbf{2.18} \pm \textbf{0.25}$	1.95 ± 0.26

Tissue samples were harvested at 18h after CLP surgery. CFU were counted and log-transformed to obtain a normal distribution. Data \pm SEM (n = 12 mice for each group) were expressed as log CFU/ml for blood, log CFU/mouse for peritoneal lavage fluid and log CFU/g for tissues.

mice, further supporting the notion that $LXR\alpha$ may have a regulatory role on the pro-inflammatory pathways. The NF- κ B is a nuclear transcription factor that plays an important role in the coordination of innate and adaptive immune responses in sepsis by modulating the expression of many pro-inflammatory genes.¹³ There is evidence to suggest that at least some of the anti-inflammatory effects of LXRa are mediated through inhibition of the NF-KB pathway.31,32 Our laboratory has also previously demonstrated that LXRa activation inhibits the LPS-induced inflammatory response in murine macrophages by inhibiting the NF-kB pathway.⁹ Moreover, we have shown that activation of LXRa inhibits NF-kB binding to DNA in the lung in a rodent model of hemorrhagic shock and resuscitation.12

Our CLP procedure without fluid resuscitation or antibiotic treatment resulted in a severe septic shock, with 90–100% mortality at 3d after surgery. Nevertheless, despite the rapid demise, the presence of a functional LXR α in young WT LXR $\alpha^{+/+}$ mice was associated with delayed onset of mortality when compared with young LXR $\alpha^{-/-}$ mice. Furthermore, even in this severe model of sepsis, treatment of young septic C57BL/6 mice with the LXR α ligand T0901317 significantly improved survival compared with vehicle treatment. Thus, our data suggest that an adequate LXR α



Figure 6. Plasma levels of (a) IL-6, (b) TNF- α , (c) resistin and (d) leptin WT LXR $\alpha^{+/+}$ and LXR $\alpha^{-/-}$ mice at 0, 6 and 18 h after CLP. Each data point represents the mean \pm SEM of 4–6 animals for each group. *P < 0.05 vs. –WT mice at the same time point.



Figure 7. Sepsis induces up-regulation of NF- κ B activity and decrease of LXR α content in the lung. (a) Activity of the p65 subunit of NF- κ B in lung nuclear extracts at 0, 3, 6 and 18 h after CLP. Data are mean \pm SEM of 4–6 animals for each group and are expressed as optical density units. *P < 0.05 vs. LXR $\alpha^{+/+}$ mice. (b) Western blot analysis of LXR α and β -actin (used as loading control protein) in lung nuclear extracts. (c) Image analyses of LXR α content as determined by densitometry. Data are mean \pm SEM of 4–6 animals for each group and are expressed as relative intensity units. *P < 0.05 vs. time 0.

receptor reserve offers a therapeutic target for the use of specific ligands in young mice.

Age itself is an independent risk factor for death in patients with severe sepsis.^{1,33} In support of the clinical observation, several experimental studies have demonstrated that aged animals experience increased severity of the inflammatory response and mortality when compared with younger animals.^{34–37} In our pharmacological studies, in order to investigate the therapeutic efficacy of LXR α activation, we calibrated injury severity in the CLP procedure to produce similar mortality in young and mature adult mice, thus avoiding the potential mortality disadvantage of aged animals. We demonstrated that post-treatment with the LXR α ligand T0901317 reduced lung neutrophil infiltration and improved survival after sepsis in young mice when compared with vehicle treatment; however, it failed to provide any protective effects in mature adult mice. At molecular analysis, mature adult mice had a significant lower content of LXR α in the lung when compared with young animals. Therefore, it is plausible that the lack of therapeutic effect of the ligand T0901317 in mature adult mice may be secondary to the poor availability of total receptor for ligand engagement. The age-dependent reduced content of the receptor may also explain the fact that mature adult WT LXR $\alpha^{+/+}$ mice had similar outcomes as the mature adult LXR $\alpha^{-/-}$ mice after sepsis.

In conclusion, our study shows that a reduction of LXR α protein is associated with lung injury after sepsis. Young, but not mature adult mice, have an adequate receptor reserve, which may be a valid therapeutic target for sepsis treatment. It is clear, therefore, that the decrease of LXR α levels with advanced age limits the therapeutic potentials of ligands for this receptor. The molecular mechanisms that mediate the age-dependent decrease of LXR α remain to be elucidated.

Funding

This work was supported by the National Institutes of Health to B.Z. (grants R01 GM-067202 and R01 AG-027990) and J.A.C. (R01 GM-027673). The content of this article is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Conflict of interest

The authors do not have any potential conflicts of interest to declare.

Acknowledgements

We would like to thank Dr David Mangelsdorf (University of Texas Southwestern Medical Center, Dallas, Texas) for having generously provided us with the wild type $(LXR\alpha^{+/+})$ and $LXR\alpha$ -deficient $(LXR\alpha^{-/-})$ breeding mice.

References

- 1. Girard TD, Opal SM and Ely EW. Insights into severe sepsis in older patients: from epidemiology to evidence-based management. *Clin Infect Dis* 2005; 40: 719–727.
- Dellinger R, Levy MM, Rhodes A, et al. Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock: 2012. *Crit Care Med* 2013; 41: 580–637.
- Russell DW. Nuclear orphan receptors control cholesterol catabolism. *Cell* 1999; 97: 539–542.
- Lehmann JM, Kliewer SA, Moore LB, et al. Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. J Biol Chem 1997; 272: 3137–3140.
- Bełtowski J. Liver X receptors (LXR) as therapeutic targets in dyslipidemia. *Cardiovasc Ther* 2008; 26: 297–316.
- Song C, Kokontis JM, Hiipakka RA, et al. Ubiquitous receptor: a receptor that modulates gene activation by retinoic acid and thyroid hormone receptors. *Proc Natl Acad Sci USA* 1994; 91: 10809–10813.

- Willy PJ, Umesono K, Ong ES, et al. LXR, a nuclear receptor that defines a distinct retinoid response pathway. *Genes Dev* 1995; 9: 1033–1045.
- Joseph SB, Castrillo A, Laffitte BA, et al. Reciprocal regulation of inflammation and lipid metabolism by liver X receptors. *Nat Med* 2003; 9: 213–219.
- Piraino G, Cook JA, O'Connor M, et al. Synergistic effect of peroxisome proliferator activated receptor-gamma and liver X receptor-α in the regulation of inflammation in macrophages. *Shock* 2006; 26: 146–153.
- Wang D, Liu M, Wang Y, et al. Synthetic LXR agonist T0901317 attenuates lipopolysaccharide-induced acute lung injury in rats. *Int Immunopharmacol* 2011; 11: 2098–2103.
- Crisafulli C, Mazzon E, Paterniti I, et al. Effects of Liver x receptor agonist treatment on signal transduction pathways in acute lung inflammation. *Respir Res* 2010; 11: 19.
- 12. Solan PD, Piraino G, Hake PW, et al. Liver X receptor α activation with the synthetic ligand T0901317 reduces lung injury and inflammation after hemorrhage and resuscitation via inhibition of the nuclear factor κ B pathway. *Shock* 2011; 35: 367–374.
- Zingarelli B. Nuclear factor-κB. Crit Care Med 2005; 33: S414–S416.
- Rittirsch D, Huber-Lang MS, Flierl MA, et al. Immunodesign of experimental sepsis by cecal ligation and puncture. *Nat Protoc* 2009; 4: 31–36.
- Smoak K, Madenspacher J, Jeyaseelan S, et al. Effects of liver X receptor agonist treatment on pulmonary inflammation and host defense. *J Immunol* 2008; 180: 3305–3312.
- Delvecchio CJ, Bilan P, Radford K, et al. Liver X receptor stimulates cholesterol efflux and inhibits expression of proinflammatory mediators in human airway smooth muscle cells. *Mol Endocrinol* 2007; 21: 1324–1334.
- Gong H, He J, Lee JH, et al. Activation of the liver X receptor prevents lipopolysaccharide-induced lung injury. J Biol Chem 2009; 284: 30113–30121.
- Ware LB and Matthay MA. The acute respiratory distress syndrome. N Engl J Med 2000; 342: 1334–1349.
- Zemans RL, Colgan SP and Downey GP. Transepithelial migration of neutrophils: mechanisms and implications for acute lung injury. *Am J Respir Cell Mol Biol* 2009; 40: 519–535.
- Kumar A, Roberts D, Wood KE, et al. Duration of hypotension prior to initiation of effective antimicrobial therapy is the critical determinant of survival in human septic shock. *Crit Care Med* 2006; 34: 1589–1596.
- Korf H, Vander Beken S, Romano M, et al. Liver X receptors contribute to the protective immune response against *Mycobacterium tuberculosis* in mice. J Clin Invest 2009; 119: 1626–1637.
- 22. Beyer C, Huang J, Beer J, et al. Activation of liver X receptors inhibits experimental fibrosis by interfering with interleukin-6

release from macrophages. Ann Rheum Dis 2014; doi: 10.1136/ annrheumdis-2013-204401 [Epub ahead of print].

- Wang YY, Ryg U, Dahle MK, et al. Liver X receptor protects against liver injury in sepsis caused by rodent cecal ligation and puncture. *Surg Infect* 2011; 12: 283–289.
- 24. Zingarelli B, Sheehan M, Hake PW, et al. Peroxisome proliferator activator receptor-γ ligands, 15-deoxy-Δ^{12,14}-prostaglandin J₂ and ciglitazone, reduce systemic inflammation in polymicrobial sepsis by modulation of signal transduction pathways. *J Immunol* 2003; 171: 6827–6837.
- Vish M, Mangeshkar P, Pirano G, et al. Proinsulin c-peptide exerts effects in endotoxic shock in mice. *Crit Care Med* 2007; 35: 1348–1355.
- Zingarelli B, Piraino G, Hake PW, et al. Peroxisome proliferatoractivated receptor δ regulates inflammation via NF-κB signaling in polymicrobial sepsis. *Am J Pathol* 2010; 177: 1834–1847.
- 27. Kim MS, Sweeney TR, Shigenaga JK, et al. Tumor necrosis factor and interleukin 1 decrease RXRα, PPARα, PPARγ, LXRα, and the coactivators SRC-1, PGC-1α, and PGC-1β in liver cells. *Metabolism* 2007; 256: 267–279.
- Sanguino E, Roglans N, Rodríguez-Calvo R, et al. Ageing introduces a complex pattern of changes in several rat brain transcription factors depending on gender and anatomical localization. *Exp Gerontol* 2006; 41: 372–379.
- Li X, Zhang S, Blander G, et al. SIRT1 deacetylates and positively regulates the nuclear receptor LXR. *Mol Cell* 2007; 28: 91–106.
- Kim KH, Yoon JM, Choi AH, et al. Liver X receptor ligands suppress ubiquitination and degradation of LXRα by displacing BARD1/BRCA1. *Mol Endocrinol* 2009; 23: 466–474.
- Zelcer N and Tontonoz P. Liver X receptors as integrators of metabolic and inflammatory signaling. J Clin Invest 2006; 116: 607–614.
- Joseph SB, Bradley MN, Castrillo A, et al. LXR-dependent gene expression is important for macrophage survival and the innate immune response. *Cell* 2004; 119: 299–309.
- Martin GS, Mannino DM and Moss M. The effect of age on the development and outcome of adult sepsis. *Crit Care Med* 2006; 34: 15–21.
- Turnbull IR, Wlzorek JJ, Osborne D, et al. Effects of age on mortality and antibiotic efficacy in cecal ligation and puncture. *Shock* 2003; 19: 310–313.
- Hyde SR, Stith RD and McCallum RE. Mortality and bacteriology of sepsis following cecal ligation and puncture in aged mice. *Infect Immun* 1990; 58: 619–624.
- Saito H, Sherwood ER, Varma TK, et al. Effects of aging on mortality, hypothermia, and cytokine induction in mice with endotoxemia or sepsis. *Mech Ageing Dev* 2003; 124: 1047–1058.
- Turnbull IR, Clark AT, Stromberg PE, et al. Effects of aging on the immunopathologic response to sepsis. *Crit Care Med* 2009; 37: 1018–1023.