### MITOGENOME ANNOUNCEMENT

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# Complete mitochondrial genome and phylogenetic analysis of Huangshan Black chicken (*Gallus gallus*)

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#### ABSTRACT

In this study, we sequenced the complete mitochondrial genome (mitogenome) of the Huangshan Black chicken (HBC). Results showed that the complete HBC mitogenome was 16,785 bp in size, comprising 22 transfer RNA genes, 2 ribosomal RNA genes, 13 protein-coding genes, and 1 non-coding control region (D-loop). The overall nucleotide composition was 32.5% for C, 30.3% for A, 23.7% for T, and 13.5% for G. Phylogenetic analysis showed that the HBC mitogenome was clustered with Xianju chicken, which belonged to the haplogroup D2. Our results therefore demonstrate that the origin of HBC corresponds to haplogroup D2 distribution and might have at least one maternal lineage originated from Southeast Asia.

ARTICLE HISTORY

Received 31 July 2020 Accepted 2 December 2020

**KEYWORDS** Huangshan Black chicken; mitochondrial genome; phylogenetic analysis

The domestic chicken is an important source of protein for humans, providing large quantities of meat and eggs. The Huangshan Black chicken (HBC, *Gallus gallus*) is one of the most important domestic chicken breeds in Anhui province, which was documented as a local poultry genetic resource in 2009. It is very well-known for its strong disease resistance, adaptability, superior meat quality, and high polyunsaturated fatty acids percentage (Yang et al. 2018). However, the genetic resources of HBC should be preserved because increased use of highly productive commercial breeds has reduced genetic diversity in chickens (Gao et al. 2017). In this study, we sequenced the complete mitochondrial genome (mitogenome) of HBC for the first time and used these data to perform phylogenetic analyses.

Blood samples of HBC were collected from Yi County N29°47′-30°11′), (E117°38′–118°6′, Huangshan, China. Genomic DNA was isolated from the specimens (voucher no. HBC190916) and stored at -70 °C in the laboratory of Poultry Genetics and Breeding, College of Animal Science and Technology, Anhui Agricultural University. DNA integrity was evaluated using 1.0% agarose gel electrophoresis. We then performed whole-genome shotgun sequencing using pairedend Illumina NovaSeg, with a read length of 150 bp (TSINGKE Biological Technology, Beijing, China). The genome was assembled by SPAdes (Bankevich et al. 2012), annotated by GeSeg (Tillich et al. 2017), and tRNA Scan-SE 1.21 (http://lowelab.ucsc.edu/tRNA Scan-SE/) was used to search for tRNA genes in the genomic sequences.

The complete mitogenome was 16,785 bp in size and typical in structure, containing 13 protein-coding genes, 22 transfer RNA genes, 2 ribosomal RNA genes, and one noncoding control region (D-loop) (GenBank accession No. MT762346). Base composition was 32.5% for C, 30.3% for A, 23.7% for T, and 13.5% for G. The start codon for all proteincoding genes was ATG, except *COX1*, which had GTG as the start codon. These genes had 15 spaces and 7 overlaps, both 1–9 bp in length. Eight tRNA genes ( $tRNA^{Gln}$ ,  $tRNA^{Ala}$ ,  $tRNA^{Asn}$ ,  $tRNA^{Cys}$ ,  $tRNA^{Tyr}$ ,  $tRNA^{Ser}$ ,  $tRNA^{Pro}$ , and  $tRNA^{Glu}$ ) and one protein-coding gene (*ND6*) were encoded on the L strand; the rest of the genes were encoded on the H strand. We identified three different complete stop codons: TAG for *ND2*, AGG for *COX1*, and TAA for *ND1*, *COX2*, *ATPase8*, *ATPase6*, *ND3*, *ND4L*, *ND5*, *Cytb*, and *ND6*. For *COX3* and *ND4*, the stop codon was incomplete (T––) and was located in the 5' terminal of the adjacent gene (Anderson et al. 1981). The D-loop region was 1232 bp in length and was located between  $tRNA^{Glu}$  and  $tRNA^{Phe}$ .

A maximum-likelihood (ML) tree was constructed using MEGA 7.0 (Kimura 1980; Kumar et al. 2016) with 1000 bootstrap replicates, based on the complete HBC mitogenome sequence and 14 chicken breeds sequences from the National Center for Biotechnology Information (NCBI). Phylogenetic analysis (Figure 1) showed that the HBC mitogenome was clustered with Xianju chicken. Previous studies have shown that the mitogenome of Xianju chicken (GU261677) belonged to haplogroup D2, which most likely originated from Southeast Asia (Liu et al. 2006; Miao et al. 2013). We therefore conclude that the origin of HBC corresponds to haplogroup D2 distribution and might have at least one maternal lineage originated from Southeast Asia. This study will provide fundamental data for conservation, breeding, and further genetic studies of HBC and other indigenous chicken breeds.

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(a)



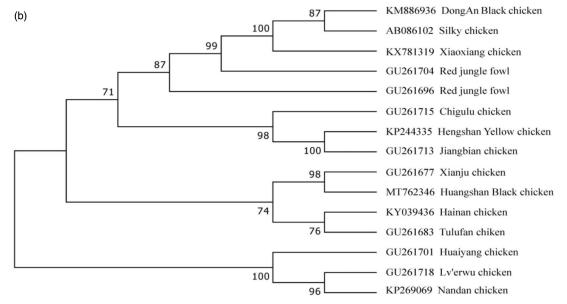


Figure 1. Characteristic of Huangshan Black chicken (HBC) and phylogenetic analysis based on the complete mitochondrial genome sequences of HBC. (a) Phenotypes of HBC. (b) The Phylogenetic tree. A maximum-likelihood (ML) tree was constructed from the complete HBC mitogenome and 14 chicken breed sequences. The 14 sequences were obtained from GenBank (accession numbers on the figure).

### **Disclosure statement**

No potential conflict of interest was reported by the author(s).

### Funding

This work was supported by Natural Science Foundation of Anhui Province [1808085QC61] and Key Science and Technology Program of Anhui Province [1704a07020091].

### Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at (https://www.ncbi.nlm.nih.gov/) under the accession no. MT762346. The associated BioProject, SRA, and BioSample numbers are PRJNA678580, SRP292644, and SAMN16807199, respectively.

## References

Anderson S, Bankier AT, Barrell BG, de Bruijn MH, Coulson AR, Drouin J, Eperon IC, Nierlich DP, Roe BA, Sanger F, et al. 1981. Sequence and organization of the human mitochondrial genome. Nature. 290(5806): 457–465.

- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol. 19(5):455–477.
- Gao YS, Jia XX, Tang XJ, Fan YF, Lu JX, Huang SH, Tang MJ. 2017. The genetic diversity of chicken breeds from Jiangxi, assessed with BCDO2 and the complete mitochondrial DNA D-loop region. PLOS One. 12(3): e0173192.
- Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J Mol Evol. 16(2):111–120.
- Kumar S, Stecher G, Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol. 33(7): 1870–1874.
- Liu YP, Wu GS, Yao YG, Miao YW, Luikart G, Baig M, Beja-Pereira A, Ding ZL, Palanichamy MG, Zhang YP. 2006. Multiple maternal origins of chickens: out of the Asian jungles. Mol Phylogenet Evol. 38(1):12–19.
- Miao YW, Peng MS, Wu GS, Ouyang YN, Yang ZY, Yu N, Liang JP, Pianchou G, Beja-Pereira A, Mitra B, et al. 2013. Chicken domestication: an updated perspective based on mitochondrial genomes. Heredity. 110(3):277–282.
- Tillich M, Lehwark P, Pellizzer T, Ulbricht-Jones ES, Fischer A, Bock R, Greiner S. 2017. GeSeq – versatile and accurate annotation of organelle genomes. Nucleic Acids Res. 45(W1):W6–W11.
- Yang S, Wang Y, Wang L, Shi Z, Ou X, Wu D, Zhang X, Hu H, Yuan J, Wang W, et al. 2018. RNA-Seq reveals differentially expressed genes affecting polyunsaturated fatty acids percentage in the Huangshan Black chicken population. PLOS One. 13(4):e0195132.