MITOGENOME ANNOUNCEMENTS

Taylor & Francis Taylor & Francis Group

OPEN ACCESS

The complete mitochondrial genome of the smallest known free-living insect *Scydosella musawasensis*

Artem Nedoluzhko^a, Fedor Sharko^b, Eugenia Boulygina^a, Svetlana Tsygankova^a, Alexey Sokolov^b, Alexander Mazur^b, Alexey Polilov^c, Egor Prokhortchouk^{b,c} and Konstantin Skryabin^{a,b,c}

^aGenome Analysis Laboratory, National Research Centre "Kurchatov Institute", Moscow, Russia; ^bInstitute of Bioengineering, Research Center of Biotechnology of the Russian Academy of Sciences, Moscow, Russia; ^cFaculty of Biology, Lomonosov Moscow State University, Moscow, Russia

ABSTRACT

The mitochondrial genome of the smallest known free-living insect *Scydosella musawasensis* (Polilov, 2015) is published in this paper. The mitochondrial DNA (mtDNA) is 14 719 base pairs (bp) in length and contained 13 protein-coding genes, 2 rRNA genes and 21 tRNA genes. The overall base composition of the genome in descending order was 40.59% – A, 13.85% – C, 36.82% – T and 8.73% – G, with a significant AT bias of 77.41%.

ARTICLE HISTORY

Received 10 January 2016 Revised 18 January 2016 Accepted 31 January 2016

KEYWORDS

Coleoptera; mitochondrial genome; next-generation sequencing; Ptiliidae; Scydosella musawasensis

Scydosella musawasensis individuals were collected in Chicaque National Park, Colombia, 10 km west of Bogotá, 2200 m above sea level, on the fungus *Steccherinum* sp. (Meruliaceae). Specimens were deposited in the entomological collection of the Zoological Museum of Lomonosov Moscow State University (ZMMU I-P-00844-854). DNA was extracted using NucleoSpin Tissue XS kit (Macherey-Nagel, Germany). DNA-libraries were constructed using Ovation Ultralow Systems V2 kit (NuGEN, San Carlos, CA). Mitochondrial genome was sequenced used Illumina Hiseq 1500 (Illumina, USA) with 100 bp pared-end reads.

Totally 226 571 628 Illumina paired-end reads were generated. Reads were merged (up to 60%) using Pear software (Zhang et al. 2014). After PCR-duplicate trimming, we used MITObim software (Hahn et al. 2013) and mitogenome of the American carrion beetle *Necrophila americana* (Coleoptera: Silphidae) as reference sequence for reconstructing the mitochondrial genome of *S. musawasensis*.

Surprisingly, we did not find *trnl* gene in our assembly. SPAdes software was used for *de novo* assembly (Bankevich et al. 2012) as proof that *trnl* is absent in *S. musawasensis* mitogenome. Totally 249 988 contigs were assembled (N50 = 1314 bp), but contigs mapping on the MITObim assembly did not yield a *trnl* gene in mitogenome.

As a result, the mitogenome of *S. musawasensis* consists of 14 719 bp (GenBank accession number: KU302777), including 13 protein-coding genes (PCGs), 2 rRNA genes and 21 tRNA genes. Despite that arthropod mitochondrial DNA typically

contain the 22 tRNAs (Boore 1999), our data showed that *S. musawasensis* have lost *trn1* gene.

Six of the 13 PCGs (COX1, NAD5, NAD4L, NAD6, COB, NAD1) used ATT as a start codon, and other three (NAD2, COX2, NAD4) used ATA, ATP6 started with ATG codon, ATP8 and NAD3 with ATC, and COX3 with CAC. Three genes (ATP8, COX2, NAD3) ended with a TGA stop codon, NAD1, NAD5 and NAD4L ended with ATT, NAD2 and NAD4 ended with AAA, COX1 gene ended with a TTA codon, ATP6 with TCT, COX3 with AGT, NAD6 with CGA, and COB with ATT. The *rrnL* and *rrnS* genes were located between *trnL1*(tag) and *trnQ*(ttg) genes, with 631 and 739 bp in length, respectively.

The phylogenetic analysis was performed for the Staphylinoidea superfamily and other Coleoptera species (Phacomorphus fratyi (KT780668.1); Sciodrepoides watsoni (KT780675.1); Tetartopeus terminatus (NC_028613.1); Sepedophilus bipunctatus (NC_028611.1); Scaphidium quadrimaculatum (NC_028609.1); Pselaphinae sp. 5 EF-2015 (KT7 80684.1); Pselaphinae sp. 4 EF-2015 (KT780682.1); Aleocharinae sp. 6 EF-2015 (KT780687.1); Aleocharinae sp. 5 EF-2015 (KT780685.1); Oxypoda acuminata (NC_028606.1); Rugilus geniculatus (NC 028608.1); Olophrum piceum (NC 028605.1); Myrmecocephalus concinnus (NC_028604.1); Liogluta microptera (NC 028602.1); Gabronthus thermarum (NC 028601.1); Euryusa optabilis (NC 028600.1); Dacrila fallax (NC 028599.1); Callicerus obscurus (NC_028598.1); Atrecus affinis (NC_028597.1); Thinonoma atra (KT780699.1); Necrophila americana (NC_018352.1); Euspilotus scissus (NC_018353.1); Margarinotus

CONTACT Artem Nedoluzhko 🐼 nedoluzhko@gmail.com; 🗈 Genome Analysis Laboratory, National Research Centre "Kurchatov Institute", Moscow, Russia; Egor Prokhortchouk 🖾 prokhortchouk@gmail.com 🗈 Genome Analysis Laboratory, National Research Centre "Kurchatov Institute", Moscow, Russia

© 2016 The Author(s). Published by Taylor & Francis. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/ licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

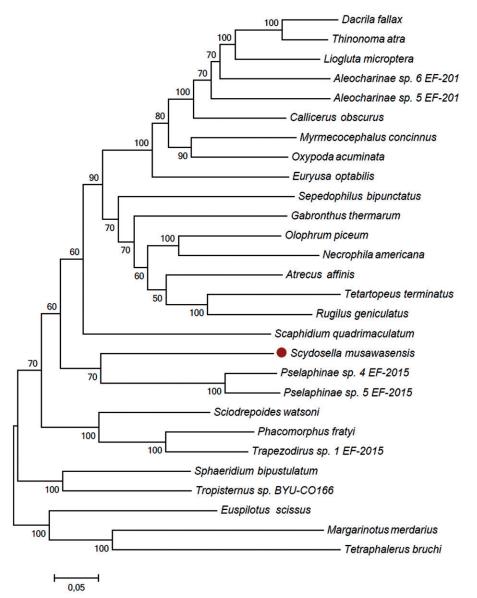


Figure. 1. Neighbor-Joining tree for Scydosella musawasensis, Staphylinoidea superfamily and other Coleoptera species and primitive beetle Tetraphalerus bruchi as outgroup based on sequence of 9 protein-coding genes (COX3, ATP6, COX2, COX1, NAD5, NAD4, NAD4L, NAD6, and COB) and 9 transfer RNAs.

merdarius (NC_028603.1); *Sphaeridium bipustulatum* (NC_028612.1); *Tropisternus sp.* BYU-CO166 (NC_018349.1) and *Tetraphalerus bruchi* (NC_011328.1) (Figure 1). For phylogenetic analysis, 9 protein-coding genes (*COX3, ATP6, COX2, COX1, NAD5, NAD4, NAD4L, NAD6, and COB*) and 9 transfer RNAs were used as marker genes. The phylogenetic tree was constructed by neighbor-Joining method, using the MEGA5.0 (Tamura et al. 2011).

Disclosure statement

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

Funding information

This work is supported by the Russian Science Foundation (Grant No. 14-24-00175).

References

- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single–cell sequencing. J Comput Biol. 19:455–477.
- Boore JL. 1999. Animal mitochondrial genomes. Nucleic Acids Res. 27:1767–1780.
- Hahn C, Bachmann L, Chevreux B. 2013. Reconstructing mitochondrial genomes directly from genomic next-generation sequencing reads a baiting and iterative mapping approach. Nucleic Acids Res. 41:e129.
- Polilov AA. 2015. How small is the smallest? New record and remeasuring of *Scydosella musawasensis* Hall, 1999 (Coleoptera, Ptiliidae), the smallest known free-living insect. Zookeys. 526:61–64.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol. 28: 2731–2739.
- Zhang J, Kobert K, Flouri T, Stamatakis A. 2014. PEAR: a fast and accurate Illumina paired-end reAd mergeR. Bioinformatics. 30:614–620.