



REVIEW

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Serious Non-AIDS events: Immunopathogenesis and interventional strategies

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Abstract

Despite the major advances in the management of HIV infection, HIV-infected patients still have greater morbidity and mortality than the general population. Serious non-AIDS events (SNAEs), including non-AIDS malignancies, cardiovascular events, renal and hepatic disease, bone disorders and neurocognitive impairment, have become the major causes of morbidity and mortality in the antiretroviral therapy (ART) era. SNAEs occur at the rate of 1 to 2 per 100 person-years of follow-up. The pathogenesis of SNAEs is multifactorial and includes the direct effect of HIV and associated immunodeficiency, underlying co-infections and co-morbidities, immune activation with associated inflammation and coagulopathy as well as ART toxicities. A number of novel strategies such as ART intensification, treatment of co-infection, the use of anti-inflammatory drugs and agents that reduce microbial translocation are currently being examined for their potential effects in reducing immune activation and SNAEs. However, currently, initiation of ART before advanced immunodeficiency, smoking cessation, optimisation of cardiovascular risk factors and treatment of HCV infection are most strongly linked with reduced risk of SNAEs or mortality. Clinicians should therefore focus their attention on addressing these issues prior to the availability of further data.

Keywords: Serious non-AIDS events, Immune activation, HIV infection

Introduction

Since the first description of AIDS in 1981, there have been tremendous advances in understanding the biology of the virus, the host's immune response and the clinical management of HIV infection. The introduction of combination antiretroviral therapy (ART) in 1996 has revolutionized HIV treatment, increasing the average life expectancy after HIV diagnosis from 10.5 to 22.5 years from 1996 to 2005 [1]. The estimated life expectancy for a 30 year old male infected with a drug-sensitive virus in 2010 and starting ART at about 6 years post infection can be as high as 75 years in some predictive models [2].

Despite the success of ART, life expectancy in HIV-infected patients is still lower than uninfected persons [2-4] and mortality in HIV-infected patients can be up to 15 times higher when compared with the general population, matched for sex and age [3]. In the pre-ART era, AIDS was the primary cause of death in HIV-infected patients [5-7]. With the use of ART, mortality due to serious

non-AIDS events (SNAEs) has become more prominent especially in resource-rich settings [6,8-13] and in patients with higher CD4 T cell counts [7,14].

Definition of serious non-AIDS events

Non-AIDS events (NAEs) are clinical events that do not meet the definition of AIDS-defining events based on the 1993 US Centers for Disease Control and Prevention (CDC) AIDS indicator conditions [15]. They encompass multiple diseases involving different organ systems, including cardiovascular, liver and renal disease, non-AIDS-defining malignancies, diabetes, neuropsychiatric disorders and bone-related abnormalities [16].

SNAEs are NAEs that result in death, are life-threatening, cause prolonged hospitalization and persistent incapacity or are associated with significant morbidity [12,14,17]. Most studies include cardiovascular, liver and end stage renal disease, as well as non-AIDS-defining cancers [11,14,18,19]. Other studies include an even broader range of conditions such as non-AIDS-related infections and psychiatric events [7,12,16,17,20].

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Incidence of SNAEs

The incidence of SNAEs in ART-treated patients is around 1 to 2 per 100 person-years of follow-up (PYFU) [11,14,17-19,21], (Table 1), but can be up to 60 per 100 PYFU in a cohort of treatment-experienced patients with multidrug resistant virus [12]. The relative contribution of non-AIDS malignancy, cardiovascular, liver and end stage renal disease to SNAEs vary across studies due to inconsistencies in the definition of SNAEs and differences in the rates of underlying co-morbidities e.g. Hepatitis B virus (HBV) and Hepatitis C virus (HCV) co-infection. However, non-AIDS malignancy, cardiovascular disease (CVD) and liver disease combined seem to account for >80% of SNAEs according to several published studies [9,11,14,17,18]. The incidence of non-AIDS malignancy and cardiovascular disease is about 2-fold higher in HIV-infected patients in the ART era when compared to the general population [22-26].

SNAEs are associated with worse outcome than AIDS events in the ART era. Compared with ART-treated patients without events, the risk of death is increased by 7 to 11-fold in those with SNAEs and by 4 to 5-fold in those with AIDS events [11,14]. Amongst SNAEs, liver-related events are associated with the highest mortality [11,14], followed by renal events, malignancies and cardiovascular events, with estimated 12-month mortality of 39.7, 32.7, 29.5 and 6.1% respectively [11].

SNAEs pathogenesis

Prior to the Strategies for Management of Antiretroviral Therapy (SMART) Study, ART toxicities were thought to be a major contributor to SNAEs. In the SMART Study, over 5000 HIV-infected patients with CD4 T cell count >350 cells/ μ L were randomized to either episodic ART (when CD4 T cell count fell below 250 cells/ μ L) or continuous ART. Patients on episodic ART had 1.8-fold increase in mortality and 1.7-fold increase in SNAEs (defined as major cardiovascular, renal or hepatic disease) when compared to those on continuous ART [27], thereby highlighting the role of HIV viraemia and immunodeficiency in the pathogenesis of SNAEs [28].

The pathogenesis of SNAEs is in fact multifactorial and complex (Figure 1). The direct effect of HIV and associated immunodeficiency, underlying co-morbidities and co-infections, immune activation with associated inflammation and coagulopathy as well as ART toxicities can all contribute.

The direct effect of HIV

Uncontrolled HIV replication causes immune activation and progressive decline in CD4 T cell count [29]. In addition, HIV can also contribute directly to organ dysfunction and SNAEs. HIV can infect human hepatic stellate cells and induce collagen expression and pro-

inflammatory cytokines secretion *in vitro* [30,31]. HIV can also mediate dysregulation of glomerular podocytes in HIV-transgenic mouse models [32], as well as apoptosis of human renal tubular cell lines [33]. Therefore, direct effect of HIV may contribute to decline in renal function and increase risk of chronic kidney disease [34]. HIV may contribute directly to non-AIDS malignancy [35], as it can be oncogenic by activating proto-oncogenes [36] or by blocking tumour suppressor genes [37] in cell lines. Increased rates of microsatellite instability in tumours of HIV-infected patients have also been found [38].

Immunodeficiency

Lower nadir or pre-ART CD4 T cell count is associated with increased risk of SNAEs [11,14,16,39]. In addition, the degree of CD4 T cell recovery after ART initiation also influences the incidence of SNAEs [14,18,21,39]. A 100 cells/ μ L lower latest CD4 T cell count in ART-treated patients is associated with a 30% increase in the risk of SNAEs, even after adjusting for smoking status, diabetes mellitus, hyperlipidaemia, HCV and HBV co-infection and alcohol abuse [18]. Lower latest CD4 T cell count in ART-treated patients is also associated with an increase in mortality [40,41].

Suboptimal restoration of CD4 T cells may be secondary to a number of factors including decreased thymic function [42,43] and impaired homeostatic responses and survival of T cells [44]. HIV replication and immune activation stimulate the secretion of transforming growth factor (TGF- β), mainly by regulatory T cells, with macrophages also contributing. TGF- β triggers collagen production by fibroblasts [45-47]. The resultant structural damage and fibrosis of the lymphoid tissues restricts T cell access to interleukin-7 (IL-7) on the fibroblastic reticular cell (FRC) network [45] thus limiting naïve CD4 T cell survival [48,49]. Furthermore, ongoing immune activation leads to rapid CD4 T cell turnover, overwhelming the already impaired renewal mechanisms [44], resulting in suboptimal CD4 recovery [50,51].

Co-infections

Due to common routes of transmission, HIV-infected patients also have increased risk of exposure to other infections that can cause organ dysfunction.

About 6-14% of HIV-infected patients have HBV and about 25-30% have HCV infection, with the rates varying depending on endemicity of HBV in the population and the prevalence of injecting drug use [52]. HBV and/or HCV co-infection greatly increases the risk of SNAEs despite ART [9,11,14,18,39,53]. Co-infection with HCV is associated with increased risk of renal disease (1.5-fold) [54,55], cardiovascular disease (1.5-fold) [56], cirrhosis (19-fold) and hepatocellular carcinoma (5-fold) [57]

Table 1 Summary of studies describing the incidence of SNAEs in various patient populations

Study	Study population	N	Median follow-up (yrs)	Male (%)	Median age (yrs)	Median nadir CD4 count (cells/ μ L)	Median baseline CD4 count (cells/ μ L)	HBV + (%)	HCV + (%)	Rate of SNAEs per 100 PYFU	Ref
EuroSIDA	A prospective observational cohort of HIV-infected patients in Europe, Israel and Argentina followed from 2001-09.	12844		73	39	178	403	6	24	1.8	[14]
SMART (S) ESPRIT(E)	S: HIV-infected patients with CD4 count >350 cells/ μ L were randomized to either CD4 count guided episodic use of ART or to continuous use of ART. E: HIV-infected patients with CD4 count >300 cells/ μ L were randomized to interleukin-2 plus ART or to ART alone.	S: 5472 E: 4111	S: 2.4 E: 6.8	S: 73 E: 81	S: 43 E: 40	S: 250 E: 197	S: 597 E: 457	S: 2	S: 15	1	[11,27,127]
	An observational cohort of HIV-infected patients with CD4 count >500 cells/ μ L in Spain from 1996-2011.	547	10	80	43	348	630	5	28	1.4	[17]
CoRIS	A prospective multicenter observational cohort of HIV-infected patients in Spain followed from 2004-2010.	5185	2.1	79	36		342	4	12	2.9	[16]
	A retrospective study of HIV-infected patients receiving ART in Botswana (B) and Nashville, USA (US); from 2002 (B) and 2003 (US)-2007.	B: 650 US: 1129	B: 3 US: 1.5	B: 31 US: 74	B: 33 US: 40		B: 199 US: 243			B: 1 US: 1.2	[19]
LATINA	A retrospective study of HIV-infected patients in Latin America from 1997-2007.	6007	2.5	70						0.9	[18]
APROCO/ COPILOTE	A prospective observational cohort of HIV-infected patients in France followed from 1997-2006.	1231	7.3	77	36		279	5	23	10.5	[20]
OPTIMA	HIV-infected patients with resistance to at least 2 different multidrug regimens were randomized to (a) re-treatment with either standard (≤ 4) or intensive (≥ 5) antiretroviral drugs and (b) either treatment starting immediately or after a 12-week monitored ART interruption.	368	4	98	48		107	11	22	61.0	[12]
ATHENA	An observational cohort of ART naive HIV-infected patients starting ART in the Netherlands, 1996-2010.	6440	3.9	75	39		200	7	6	1.2	[21]

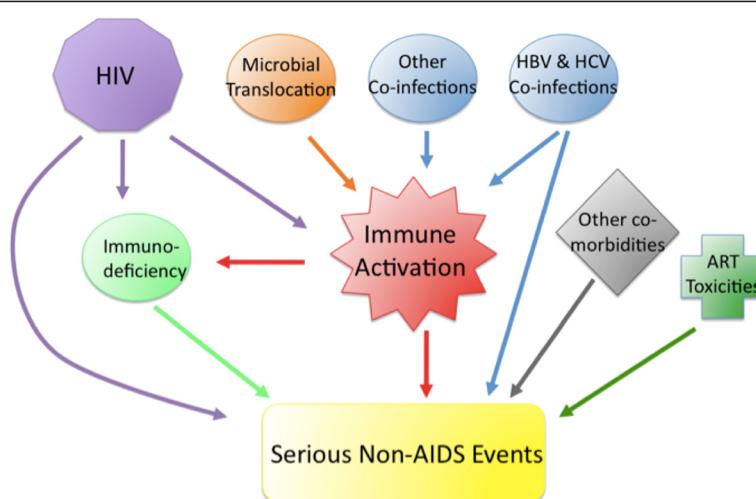


Figure 1 Pathogenesis of serious non-AIDS events. HIV infection causes progressive decline in CD4 T cells through direct cytopathic effects and immune mediated killing of infected cells, as well as indirectly via immune activation. Other drivers of immune activation include co-infections and microbial translocation. HIV can contribute to organ dysfunction through detrimental effects on hepatic stellate cells and renal tubular cells. HIV may also be oncogenic. Co-infection with HBV and HCV is especially important in liver related events. In addition, patients' underlying co-morbidities e.g. smoking, cardiovascular risk factors, and ART related toxicities also contribute to SNAEs.

when compared with HIV mono-infected patients in the ART era.

HIV-infected patients also have increased risk of exposure to Human papillomavirus (HPV) [58], that is implicated in the pathogenesis of cervical and anal cancer [59,60].

Other co-morbidities

HIV-infected patients also have higher prevalence of traditional cardiovascular risk factors such as smoking [61-63], elevated total cholesterol/HDL ratio [61-64] and substance abuse [65]. Some studies also found higher rates of hypertension and diabetes [64,66], but these findings have not been confirmed by others [62,63,67].

Smoking is a major cause of increased mortality in ART-treated patients, accounting for a loss of >12 life years, and is associated with >4-fold increase in all-cause mortality, >5-fold increase in non-AIDS mortality, >4-fold increase in cardiovascular-related mortality and >3-fold increase in cancer-related mortality [68].

Though uncontrolled HIV replication, immunodeficiency, co-infection and co-morbid conditions are important contributors to the pathogenesis of SNAEs, these factors only partially account for the increased risk of SNAEs in HIV-infected patients. HIV elite controllers have increased coronary plaques and carotid intima media thickness when compared to uninfected controls even after adjustment for traditional cardiovascular risk factors [69,70], demonstrating that factors other than uncontrolled viral replication, immunodeficiency, traditional risk factors, and ART toxicities contribute to increased cardiovascular risks in HIV-infected patients.

Immune activation

Inflammation is central to the process of atherosclerosis [71,72], tumour progression [73,74] and liver fibrosis [75,76] in the general population. High levels of biomarkers associated with inflammation (C-reactive protein, CRP, and IL-6) and coagulopathy (D-dimer and fibrinogen) are associated with increased risk of cardiovascular disease [77-80], increased cancer risks [81-83] and mortality [80,84-86] in the general population.

In HIV-infected patients, CRP, IL-6 and D-dimer levels [61] as well as markers of T cell activation [50,87] remain higher than uninfected controls despite suppressive ART. Furthermore, higher CRP, IL-6 and D-dimer [88-90], soluble CD14 (sCD14, a marker of LPS-induced monocyte activation) [91] and lymphocyte activation markers [92] are associated with higher mortality.

In ART-treated patients, CRP, IL-6 and D-dimer levels are also associated with increased risk of CVD, independent of other CVD risk factors [93,94]. These markers are also associated with increased risks of both infection-related and infection unrelated cancers even after adjusting for demographics and CD4 T cell count [95]. Recently, higher levels of tumour necrosis factor (TNF) were also found to be significantly associated with increased risk of SNAEs [96]. Therefore these data suggest that immune activation plays a very important role in SNAEs pathogenesis.

Intermittent viraemia

The drivers of immune activation are diverse. Intermittent HIV viraemia can occur in 20-30% of ART-treated patients [97,98]. The presence of viraemia whilst on

ART is associated with higher IL-6, D-dimer and sCD14 levels [62] and SNAEs [11,39]. In addition, viraemia below the limit of detection of conventional assays also correlates with persistent T cell activation [99]. Therefore, residual viraemia may partially be responsible for persistent immune activation despite ART.

Co-infections

The presence of co-infections also contributes to continual stimulation and activation of the immune system. Asymptomatic CMV infection has been associated with CD8 T cell activation in ART-treated patients [100]. CMV specific CD4 T cells can cause a systemic inflammatory response that is sustained even during latent infection [101] and is associated with atherosclerosis [102,103]. Hepatitis C co-infection is also associated with increased CD8 T cell activation when compared with HIV mono-infected patients despite ART [104,105].

Microbial translocation

Microbial translocation due to impaired mucosal barrier integrity may be another cause of immune activation in HIV infection. Rhesus macaques with chronic SIV infection have disruptions of the epithelial barrier of the colon and increased lipopolysaccharide (LPS) staining. The levels of LPS staining in the colon also correlated with the levels of LPS in the draining lymph nodes and remote peripheral lymph nodes [106]. African green monkeys are natural hosts of SIV. In chronic SIV infection, they display little immune activation, maintain mucosal barrier integrity and do not progress to AIDS [107]. However, the injection of LPS into SIV-infected African green monkeys was associated with increases in T cell activation, sCD14 and SIV viraemia [108]. In HIV-infected patients, plasma LPS also correlated with plasma interferon (IFN)- α and T cell activation levels [109]. These data suggest that impaired mucosal barrier integrity and microbial translocation may induce immune activation.

Markers of microbial translocation e.g. LPS, sCD14 [110] and bacterial 16 s rDNA [111] do not always normalise with ART. LPS can induce tissue factor expression on monocytes [112]. Tissue factor is the initiator of the coagulation cascade [113] and its expression on monocytes is correlated with D-dimer levels [112]; suggesting that microbial translocation may contribute to atherosclerosis and increased CVD [112].

Antiretroviral therapy

Finally, adverse effects of ART may also contribute to SNAEs. A detailed review of ART-related toxicities is beyond the scope of this review. Protease inhibitors as a class, as well as indinavir, lopinavir and abacavir may be associated with increased risk of CVD [25,114,115].

Some studies found that the rate of liver-related deaths is increased per year of ART [116,117]. The cohort described by Weber et al have high rates of HBV and HCV infection, 17% and 66%, respectively [116]. Though patients with HBV or HCV infection have about 5-fold greater risk of hepatotoxicity after ART initiation [118-120], ART is essential as it slows fibrosis progression [121] and reduces liver-related mortality by about 10-fold [122]. In patients without chronic viral hepatitis, ART toxicity rarely causes liver-related mortality, at a rate of 0.04 per 1000 person years [123].

ART has been associated with improved renal function and a slower rate of eGFR decline in HIV-infected patients [124,125]. Though tenofovir use is associated with higher risk of acute renal injury and greater loss of renal function than other ART regimen, the overall risk of serious renal event is not high, in 0.5% of patients [126].

Interventions to reduce SNAEs

Strategies to reduce SNAEs include preventing and reversing immunodeficiency, the modification of traditional risk factors, treatment of co-infections and addressing drivers of immune activation. A wide variety of agents are currently being examined for their potential effects in reducing immune activation and SNAEs (Table 2). However, the majority of studies that have been performed are small, are heterogeneous in terms of ART status and show conflicting findings. The majority of studies used markers of immune activation, in particular CD8 T cell activation as outcome measures. Some recent studies have also included sCD14 and D-dimer. However, randomized placebo-controlled trials that use clinical outcome measures are rare [127].

Preventing and reversing immunodeficiency

ART initiation prior to advanced immunodeficiency

Data from randomized controlled trials suggest that deferral of ART initiation until CD4 T cell count <250 cells/ μ L was associated with increased SNAEs, AIDS-related events and mortality [128-130]. Observational studies suggest that ART initiation at CD4 T cell count >350 cells/ μ L is associated with lower risk of SNAEs [14], AIDS-defining illness or death when compared to deferring ART [131-134]. The benefit of ART initiation at CD4 T cell count >350 cells/ μ L when compared to deferring until CD4 T cell count ~350 cells/ μ L is insignificant in some studies when the analysis is restricted to mortality alone [131,134]. Results from the START study (NCT00867048), a multicenter international trial designed to assess the risks and benefits of initiating ART at CD4 T cell count at >500 or <350 cells/ μ L will likely shed further light on this issue. Nonetheless, given that the majority of HIV-infected patients start ART with CD4 T cell count <250 cells/ μ L [135,136], earlier HIV diagnosis and initiation

Table 2 Potential strategies to reduce SNAEs

Potential strategies to reduce SNAEs	Interventions investigated or under evaluation	References
Preventing immunodeficiency	Initiate ART prior to advanced immunodeficiency	[14,128-134]
Increasing CD4 T cell recovery		
Cytokine therapy	Subcutaneous IL-2 Subcutaneous IL-7	[127] [137,138]
Modulating lymphoid tissue fibrosis	Pirfenidone Angiotensin receptor antagonist ACE inhibitor	Human data pending Human data pending Human data pending
Managing co-morbidities	Smoking cessation Optimise blood pressure, lipids and diabetic control ART switch	[68,145] [146,147] [152-160]
Reducing chronic antigen stimulation		
Residual viraemia	Raltegravir intensification Maraviroc intensification	[161-170] [171-174]
HBV and HCV co-infection	Hepatitis B and C treatment	[105,175,176]
CMV co-infection	Valganciclovir	[178]
HSV co-infection	Valacyclovir	[179]
Reducing inflammation	Statins COX-2 inhibitors Aspirin Hydroxychloroquine & Chloroquine Leflunomide Prednisone	[182-187] [195,196] [199] [191-193] [200] [201-204]
Reducing microbial translocation		
Balancing microbiota	Prebiotic, probiotic and symbiotic	[210-213]
Reducing bacterial/endotoxin load	Rifaximin Bovine colostrum Sevelamer	Human data pending [167] Human data pending
Improving mucosal integrity	Lubiprostone	Human data pending
Reducing inflammation in the gut	Mesalamine	Human data pending

of ART before advanced immunodeficiency will likely reduce SNAEs.

Improving CD4 T cell recovery

A number of studies have investigated the use of cytokines critical for T cell homeostasis e.g. IL-2 and IL-7 to enhance CD4 T cell recovery. Though subcutaneous IL-2 administration in concert with ART resulted in sustained increase in CD4 T cell count, this did not translate into clinical benefit [127]. Subcutaneous IL-7 administration also leads to increase in CD4 T cells in phase I and II studies [137,138] but clinical outcomes have not yet been assessed. The restoration of TH17 cells and improvement in TH17/T regulatory cell ratio may be especially important given their roles in mucosal immunity [139,140].

Modulating the effects of lymphoid tissue fibrosis

Lymphoid tissue fibrosis is associated with poor CD4 T cell restoration after ART initiation [49]. TGF- β is key in the process of lymphoid tissue fibrosis. Pirfenidone can reduce TGF- β production and has anti-fibrotic effects [45,141]. Pirfenidone administered to rhesus macaques prior to SIV infection was associated with reduced lymph node fibrosis and preservation of lymph node CD4 T cells [142]. TNF blockade with adalimumab was also associated with attenuated TGF- β expression, reduced lymph node fibrosis and preserved lymph node architecture in a recent rhesus macaque study [47]. The effect of pirfenidone or adalimumab on lymph node fibrosis in HIV infection has not been studied to date.

The renin-angiotensin pathway is involved in cardiac, renal and liver fibrosis. Binding of angiotensin II to the angiotensin 1 receptor on cardiac fibroblast, hepatic stellate cells or mesangial cells leads to proliferation as well as collagen and TGF- β synthesis [143,144]. Trials on the effect of angiotensin converting enzyme (ACE) inhibitor e.g. lisinopril (ClinicalTrials.gov identifier: NCT01535235), angiotensin II receptor antagonists e.g. losartan (NCT01852942, NCT01529749) and telmisartan (NCT01928927) in modulating lymphoid tissue fibrosis are currently underway.

Optimizing cardiovascular risk factors

In the D.A.D study, patients who stopped smoking had about 30% reduction in the risk of CVD [145]. Surprisingly, a reduction in mortality was not seen. This may be because patients ceased smoking after the diagnosis of a serious illness and succumb before the benefit of smoking cessation on mortality can be seen [145]. In the Danish HIV cohort, previous smokers had a >1.5-fold reduction in mortality when compared with current smokers. In addition, though previous smokers have higher rates of AIDS-related deaths when compared with never smokers, the incidence of non-AIDS-related death was not different between previous and never smokers [68]. These data suggest that smoking cessation alone would result in significant benefits and should be encouraged.

Modification of other cardiovascular risk factors e.g. treatment of hypertension, dyslipidaemia and optimal glycaemic control in diabetic patients is critical. Suggested target levels have been published [146,147]. Each 10 mmHg reduction in systolic blood pressure and each 38 mg/dL reduction in total cholesterol is associated with a 5 and 20% reduction in risk of CVD, respectively [148]. However, in patients with known hypertension, diabetes or dyslipidaemia meeting indication for treatment, over 40% were not on treatment [149]. Given that a significant proportion of SNAEs are cardiovascular events, more aggressive detection and management of cardiovascular risk factors will likely reduce SNAEs.

ART modification is a potential strategy to reduce cardiovascular risk [150]. A recent review of switch studies have been published [151]. Switching from stavudine to tenofovir was associated with a reduction in total cholesterol and triglycerides and an increase in limb fat [152]. Switching from protease inhibitors to efavirenz or nevirapine was associated with reduction in total cholesterol [153]. However, this switch is not possible in patients with non-nucleoside reverse transcriptase inhibitor resistance. Switching from lopinavir/ritonavir to atazanavir (both boosted with ritonavir 100 mg or unboosted) was associated with reduction of total cholesterol and triglycerides, though greater reductions were seen with unboosted atazanavir [154-156]. Switching to

atazanavir was also associated with a reduction in cardiovascular risk score [157]. Though switching from lopinavir/ritonavir to raltegravir was associated with improvement in lipid profile [158,159], no change in endothelial function was detected [160]. The importance of having fully active backbone antiretrovirals was highlighted in the SWITCHMRK study where patients switched to raltegravir had higher rates of virologic failure [158].

Suppressing chronic antigen stimulation

Reducing residual viraemia

Intensification studies have been performed to assess the impact of adding antiretroviral agents to a suppressive ART regimen (as measured by conventional assays). None of the raltegravir intensification studies were able to demonstrate reduction in ultra-sensitive plasma HIV-RNA levels [161-167]. In addition, the majority of studies also found no reduction in markers of T cell [161-164,167] or monocyte activation [167]. However, some studies have noted a reduction in D-dimer levels [168], T cell activation [165,166,169] as well as an early transient increase in 2-LTR circles post raltegravir intensification [168,170], suggesting that residual viraemia was occurring prior to raltegravir intensification and was contributing to immune activation in some patients.

Maraviroc intensification studies have also been performed and yielded conflicting data. Some found reduction in T cell activation [171-173] whilst others found increase in CD4 and CD8 T cell activation both in the peripheral blood and in the rectal mucosa after maraviroc intensification [174]. Hunt et al postulated that the binding of maraviroc to CCR5 prevents the interaction between CCR5 and its natural ligands. Excess CCR5 ligands may then bind to other chemokine receptor such as CCR3 and CCR4 on T cells, leading to T cell activation [174]. Therefore, the beneficial effect of adding antiretroviral agents to an already suppressive ART regimen is uncertain based on currently available data.

Treating other co-infections

HCV treatment and suppression of HCV viraemia is associated with reductions in CD4 and CD8 T cell activation [105]. Sustained virologic response is associated with reduced liver-related complications as well as both liver-related and non liver-related mortality in co-infected patients [175,176]. Unfortunately, HCV treatment may be limited by contraindications, adverse events, high costs, and drug interactions. Next generation agents with higher efficacy and better side effect profiles may revolutionise the management of HIV/HCV co-infected patients [177].

Treatment of other persistent viral infection has also been investigated. Eight weeks of valganciclovir in ART-treated, CMV seropositive patients led to a significant reduction of CMV viraemia as well as a reduction in CD8

T cell activation [178]. In a study targeting HSV co-infection using 12 weeks of valacyclovir in ART-treated, HSV-1 and HSV-2 seropositive patients, no change in T cell activation, CRP or IL-6 levels was demonstrated [179].

Anti-inflammatory agents

Statins are 3-hydroxy-3-methyl-glutaryl-coenzyme A reductase inhibitors. Not only do statins reduce serum cholesterol [180], they may also have anti-inflammatory properties [181]. Statin use is associated with reduced monocyte activation (unpublished data McComsey et al), decline in CRP levels [182] as well as reduced T cell activation [183] in ART-treated and in untreated, HIV-infected patients [184]. A retrospective observational study of ART-treated patients showed that statin use is associated with a 3-fold reduction in mortality [185]. Though not statistically significant, a trend for reduction in SNAEs [186] and mortality [187] has also been seen in other retrospective observational studies.

Hydroxychloroquine (HCQ) and its analogue chloroquine (CQ) have immunomodulatory, anti-inflammatory and anti-HIV properties [188-190]. In patients with uncontrolled viral replication, the use of CQ was associated with reduced CD8 T cell activation [191] whereas the same effect was not seen with HCQ [192]. However a non-randomized study of HCQ in 20 ART-treated patients showed decline in plasma LPS, IL-6 and reduced T cell and monocyte activation [193]. Thus findings are inconclusive.

COX-2 inhibitors inhibit cyclooxygenase type 2, reducing Prostaglandin E2 production, thereby reducing activation of T cells through the cyclic adenosine monophosphate (cAMP) pathway [194]. Studies on COX-2 inhibitors have been small and reduction in T cell activation tended to occur in viraemic patients [195,196]. However, it is important to bear in mind that COX-2 inhibitors are associated with increased cardiovascular risk, via a direct pharmacologic consequence of inhibition of COX-2 [197]. Therefore assessing the utility of COX-2 inhibitors without using clinical outcome measures may be insufficient.

Aspirin is a cornerstone in the secondary prevention of vascular disease [198]. In a pilot study, aspirin use was associated with reduced platelet activation, a decrease in sCD14 in monocytes and reductions in CD38 and HLA-DR on CD4 and CD8 T cells. However, there was no change in IL-6, D-dimer and CRP [199]. An aspirin study with larger number of participants is in development with the AIDS Clinical Trials Group.

Leflunomide is an immunomodulatory agent that is used in the treatment of rheumatoid arthritis. The administration of leflunomide in untreated, HIV-infected patients for 28 days was associated with a decrease in CD8 T cell activation [200].

The use of prednisone in patients with untreated chronic HIV infection was associated with less CD4 T cell depletion, a decline in CD4 T cell activation and stable HIV viral load [201,202]. Prednisone at 0.5 mg/kg/day in ART-treated patients was also associated with a reduction in CD8 T cell activation and TNF levels as well as a transient decrease in IL-6 [203]. However, in another study using prednisone at 40 mg/day, no reduction in CD4 or CD8 T cell activation, plasma IL-6 or TNF levels was found [204]. Furthermore, long-term prednisone use, especially at doses >7.5 mg/day is associated with significant adverse effects such as osteoporosis, impaired glucose tolerance, dyslipidaemia, weight gain, cataract formation and increased risk of infections [205]. Even short courses have been associated with an increased risk of osteonecrosis in HIV-infected patients [206].

Targeting microbial translocation

Given that HIV infection has been associated with depressed levels of beneficial gut microbiota and elevated levels of pathogenic microbiota [207], a range of prebiotics (selectively fermented ingredients that changes the growth and/or activity of certain gut microflora, resulting in health benefits [208]), probiotics (live microorganisms that when consumed, confer a health benefit [209]) and synbiotics (combinations of pre and probiotics) are under investigation.

A prebiotic oligosaccharide mixture has been associated with improvement in microbiota composition and reduction in sCD14 in untreated HIV-infected patients [210]. A retrospective cohort study on both ART-treated and untreated HIV-infected patients found that probiotic yogurt consumption was associated with a greater increase in CD4 T cell count even after adjustment for ART [211]. A double-blind randomized placebo-controlled trial in 20 untreated HIV-infected patients found reductions in plasma bacterial DNA and IL-6 levels in patients receiving synbiotics [212]. However, a symbiotic agent in ART-treated women found no change in microbial translocation nor immune activation status despite improvement in the levels of probiotic species [213]. Therefore, more randomized controlled clinical trial data are needed to clarify the effects of pre and probiotics in reducing immune activation.

Bovine colostrum contains oligosaccharides, growth factors, immunoglobulins and antimicrobial peptides and has some activity in alleviating HIV-associated diarrhoea in single arm studies [214-216]. However a randomized controlled trial on the addition of bovine colostrum to suppressive ART found no change in CD4 T cell count, markers of microbial translocation nor T cell activation [167].

A number of new agents that target microbial translocation are under evaluation. Rifaximin is a minimally

absorbed oral rifamycin antibiotic that has activity against both gram-positive and gram-negative enteric bacteria [217]. It is effective in the treatment of hepatic encephalopathy, by reducing ammonia-producing enteric bacteria [218,219]. There are currently 3 clinical trials of rifaximin in HIV-infected patients (ClinicalTrials.gov identifier: NCT01654939, NCT01866826 and NCT01466595). Lubiprostone is a chloride channel activator that is used in the treatment of constipation [220]. It has been found to enhance recovery of mucosal barrier function in ischaemic porcine colon [221]. A pilot study of lubiprostone in ART treated, virologically suppressed patients with CD4 T cell count <350 cells/ μ L (NCT01839734) is currently recruiting. Sevelamer is a phosphate binder that is used in patients with end-stage renal failure [222]. It can also bind to endotoxins and reduce CRP, IL-6 and sCD14 in patients on haemodialysis [223,224]. The trial in untreated, HIV-infected patients (NCT01543958) was completed in June 2013 and results are pending. Mesalamine (5-aminosalicylic acid) is an anti-inflammatory agent used in the management of inflammatory bowel disease [225]. A trial using mesalamine in ART treated, virologically suppressed patients with CD4 T cell count <350 cells/ μ L (NCT01090102) is currently enrolling.

Conclusions

Despite the use of ART, HIV-infected patients still have higher mortality and morbidity when compared to the general population. SNAEs occur at the rate of about 1-2 per 100 PYFU and are the predominant causes of morbidity and mortality in HIV-infected patients in the ART era. Many factors contribute to the pathogenesis of SNAEs including the direct effect of HIV and associated immunodeficiency, underlying co-morbidities, immune activation and ART toxicities. Though multiple interventions have been investigated or are ongoing, most of the studies are small, of short duration and clinical outcome measures have not been ascertained. The cost required to investigate the effectiveness of an intervention to reduce SNAEs may be prohibitively high as it will require thousands of participants with possibly several years of follow up.

Currently, the interventions with evidence to suggest an association with reduced risk of SNAEs or mortality are starting ART before advanced immunodeficiency, smoking cessation, optimisation of cardiovascular risk factors and treatment of HCV infection. Clinicians should focus their attention on addressing these issues prior to the availability of further data.

Abbreviations

ACE: Angiotensin converting enzyme; ART: Antiretroviral therapy; cAMP: Cyclic adenosine monophosphate; CDC: Centers for disease control and prevention; CQ: Chloroquine; CRP: C-reactive protein; CVD: Cardiovascular disease; EBV: Epstein-Barr virus; FRC: Fibroblastic reticular cell; HBV: Hepatitis B

virus; HCV: Hepatitis C virus; HCQ: Hydroxychloroquine; HPV: Human papillomavirus; IL-7: Interleukin-7; IFN: Interferon; LPS: Lipopolysaccharide; NAEs: Non-AIDS events; PYFU: Person-years of follow-up; sCD14: Soluble CD14; SIR: Standardized incidence ratio; SMART: Strategies for management of antiretroviral therapy; SNAEs: Serious non-AIDS events; START: Strategic timing of antiretroviral treatment; TGF: Transforming growth factor; TNF: Tumour necrosis factor.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

DH, JA, IS contributed to the writing of the manuscript. All authors read and approved the final manuscript.

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References

1. Harrison KM, Song R, Zhang X: Life expectancy after HIV diagnosis based on national HIV surveillance data from 25 states, United States. *J Acquir Immune Defic Syndr* 2010, 53:124–130.
2. Nakagawa F, Lodwick RK, Smith CJ, Smith R, Cambiano V, Lundgren JD, Delpech V, Phillips AN: Projected life expectancy of people with HIV according to timing of diagnosis. *AIDS* 2012, 26:335–343.
3. Lohse N, Hansen AB, Pedersen G, Kronborg G, Gerstoft J, Sorensen HT, Vaeth M, Obel N: Survival of persons with and without HIV infection in Denmark, 1995–2005. *Ann Intern Med* 2007, 146:87–95.
4. Life expectancy of individuals on combination antiretroviral therapy in high-income countries: a collaborative analysis of 14 cohort studies. *Lancet* 2008, 372:293–299.
5. Krentz HB, Kliewer G, Gill MJ: Changing mortality rates and causes of death for HIV-infected individuals living in Southern Alberta, Canada from 1984 to 2003. *HIV Med* 2005, 6:99–106.
6. Palella FJ Jr, Baker RK, Moorman AC, Chmiel JS, Wood KC, Brooks JT, Holmberg SD: Mortality in the highly active antiretroviral therapy era: changing causes of death and disease in the HIV outpatient study. *J Acquir Immune Defic Syndr* 2006, 43:27–34.
7. Lau B, Gange SJ, Moore RD: Risk of non-AIDS-related mortality may exceed risk of AIDS-related mortality among individuals enrolling into care with CD4+ counts greater than 200 cells/mm³. *J Acquir Immune Defic Syndr* 2007, 44:179–187.
8. Reisler RB, Han C, Burman WJ, Tedaldi EM, Neaton JD: Grade 4 events are as important as AIDS events in the era of HAART. *J Acquir Immune Defic Syndr* 2003, 34:379–386.
9. Causes of death in HIV-1-infected patients treated with antiretroviral therapy, 1996–2006: collaborative analysis of 13 HIV cohort studies. *Clin Infect Dis* 2010, 50:1387–1396.
10. Lewden C, May T, Rosenthal E, Burty C, Bonnet F, Costagliola D, Jouglard E, Semaille C, Morlat P, Salmon D, et al: Changes in causes of death among adults infected by HIV between 2000 and 2005: the "Mortalite 2000 and 2005" surveys (ANRS EN19 and Mortavic). *J Acquir Immune Defic Syndr* 2005, 2008(48):590–598.
11. Neuhaus J, Angus B, Kowalska JD, La Rosa A, Sampson J, Wentworth D, Mocroft A: Risk of all-cause mortality associated with nonfatal AIDS and serious non-AIDS events among adults infected with HIV. *AIDS* 2010, 24:697–706.
12. Anis AH, Nosy B, Sun H, Guh DP, Bansback N, Li X, Barnett PG, Joyce V, Swanson KM, Kyriakides TC, et al: Quality of life of patients with advanced HIV/AIDS: measuring the impact of both AIDS-defining events and non-AIDS serious adverse events. *J Acquir Immune Defic Syndr* 2009, 51:631–639.
13. Kowalska JD, Reekie J, Mocroft A, Reiss P, Ledergerber B, Gatell J, d'Arminio Monforte A, Phillips A, Lundgren JD, Kirk O: Long-term exposure to

- combination antiretroviral therapy and risk of death from specific causes: no evidence for any previously unidentified increased risk due to antiretroviral therapy. *AIDS* 2012, 26:315–323.
- 14. Mocroft A, Reiss P, Gasiorowski J, Ledergerber B, Kowalska J, Chiesi A, Gatell J, Rakhamanova A, Johnson M, Kirk O, Lundgren J: **Serious fatal and nonfatal non-AIDS-defining illnesses in Europe.** *J Acquir Immune Defic Syndr* 2010, 55:262–270.
 - 15. CDC: 1993 revised classification system for HIV infection and expanded surveillance case definition for AIDS among adolescents and adults. *MMWR Recomm Rep* 1992, 41:1–19.
 - 16. Masia M, Padilla S, Alvarez D, Lopez JC, Santos I, Soriano V, Hernandez-Quero J, Santos J, Tural C, del Amo J, Gutierrez F: **Risk, predictors, and mortality associated with non-AIDS events in newly diagnosed HIV-infected patients: role of antiretroviral therapy.** *AIDS* 2013, 27:181–189.
 - 17. Lucero C, Torres B, Leon A, Calvo M, Leal L, Perez I, Plana M, Arnedo M, Mallolas J, Gatell JM, Garcia F: **Rate and Predictors of Non-AIDS Events in a Cohort of HIV-Infected Patients with a CD4 T Cell Count Above 500 Cells/mm³.** *AIDS Res Hum Retroviruses* 2013, 29:1161–1167.
 - 18. Beloso WH, Orellana LC, Grinsztejn B, Madero JS, La Rosa A, Veloso VG, Sanchez J, Ismerio Moreira R, Crabtree-Ramirez B, Garcia Messina O, et al: **Analysis of serious non-AIDS events among HIV-infected adults at Latin American sites.** *HIV Med* 2010, 11:554–564.
 - 19. Wester CW, Koethe JR, Shepherd BE, Stinnette SE, Rebeiro PF, Kipp AM, Hong H, Bussmann H, Gaolathate T, McGowan CC, et al: **Non-AIDS-defining events among HIV-1-infected adults receiving combination antiretroviral therapy in resource-replete versus resource-limited urban setting.** *AIDS* 2011, 25:1471–1479.
 - 20. Ferry T, Raffi F, Collin-Filleul F, Dupon M, Dellamonica P, Waldner A, Strady C, Chene G, Leport C, Moing VL: **Uncontrolled viral replication as a risk factor for non-AIDS severe clinical events in HIV-infected patients on long-term antiretroviral therapy: APROCO/COPILOTE (ANRS CO8) cohort study.** *J Acquir Immune Defic Syndr* 2009, 51:407–415.
 - 21. Zhang S, van Sighem A, Kesselring A, Gras L, Smit C, Prins JM, Kauffmann R, Richter C, de Wolf F, Reiss P: **Episodes of HIV viremia and the risk of non-AIDS diseases in patients on suppressive antiretroviral therapy.** *J Acquir Immune Defic Syndr* 2012, 60:265–272.
 - 22. Powles T, Robinson D, Stebbing J, Shamash J, Nelson M, Gazzard B, Mandelis S, Moller H, Bower M: **Highly active antiretroviral therapy and the incidence of non-AIDS-defining cancers in people with HIV infection.** *J Clin Oncol* 2009, 27:884–890.
 - 23. Bedimo RJ, McGinnis KA, Dunlap M, Rodriguez-Barradas MC, Justice AC: **Incidence of non-AIDS-defining malignancies in HIV-infected versus noninfected patients in the HAART era: impact of immunosuppression.** *J Acquir Immune Defic Syndr* 2009, 52:203–208.
 - 24. Shieh MS, Cole SR, Kirk GD, Poole C: **A meta-analysis of the incidence of non-AIDS cancers in HIV-infected individuals.** *J Acquir Immune Defic Syndr* 2009, 52:611–622.
 - 25. Islam FM, Wu J, Jansson J, Wilson DP: **Relative risk of cardiovascular disease among people living with HIV: a systematic review and meta-analysis.** *HIV Med* 2012, 13:453–468.
 - 26. Martinez E, Milinkovic A, Buira E, de Lazzari E, Leon A, Larrousse M, Lonca M, Laguno M, Blanco JL, Mallolas J, et al: **Incidence and causes of death in HIV-infected persons receiving highly active antiretroviral therapy compared with estimates for the general population of similar age and from the same geographical area.** *HIV Med* 2007, 8:251–258.
 - 27. El-Sadr WM, Lundgren J, Neaton JD, Gordin F, Abrams D, Arduino RC, Babiker A, Burman W, Clumeck N, Cohen CJ, et al: **CD4+ count-guided interruption of antiretroviral treatment.** *N Engl J Med* 2006, 355:2283–2296.
 - 28. Strategies for Management of Antiretroviral Therapy Study Group, Lundgren JD, Babiker A, El-Sadr W, Emery S, Grund B, Neaton JD, Neuhaus J, Phillips AN: **Inferior clinical outcome of the CD4+ cell count-guided antiretroviral treatment interruption strategy in the SMART study: role of CD4+ Cell counts and HIV RNA levels during follow-up.** *J Infect Dis* 2008, 197:1145–1155.
 - 29. Lawn SD, Butera ST, Folks TM: **Contribution of immune activation to the pathogenesis and transmission of human immunodeficiency virus type 1 infection.** *Clin Microbiol Rev* 2001, 14:753–777.
 - 30. Tuyama AC, Hong F, Saiman Y, Wang C, Ozkok D, Mosoian A, Chen P, Chen BK, Klotman ME, Bansal MB: **Human immunodeficiency virus (HIV)-1 infects human hepatic stellate cells and promotes collagen I and monocyte chemoattractant protein-1 expression: implications for the pathogenesis of HIV/hepatitis C virus-induced liver fibrosis.** *Hepatology* 2010, 52:612–622.
 - 31. Hong F, Saiman Y, Si C, Mosoian A, Bansal MB: **X4 Human immunodeficiency virus type 1 gp120 promotes human hepatic stellate cell activation and collagen I expression through interactions with CXCR4.** *PLoS One* 2012, 7:e33659.
 - 32. Zuo Y, Matsusaka T, Zhong J, Ma J, Ma LJ, Hanna Z, Jolicœur P, Foggo AB, Ichikawa I: **HIV-1 genes vpr and nef synergistically damage podocytes, leading to glomerulosclerosis.** *J Am Soc Nephrol* 2006, 17:2832–2843.
 - 33. Kapasi AA, Fan S, Singhal PC: **p300 modulates HIV-1 gp120-induced apoptosis in human proximal tubular cells: associated with alteration of TGF-beta and Smad signaling.** *Nephron Exp Nephrol* 2006, 102:e30–e38.
 - 34. Wyatt CM, Meliambro K, Klotman PE: **Recent progress in HIV-associated nephropathy.** *Annual review of medicine* 2012, 63:147–159.
 - 35. Deeken JF, Tjen ALA, Rudek MA, Okulair C, Young M, Little RF, Dezube BJ: **The rising challenge of non-AIDS-defining cancers in HIV-infected patients.** *Clin Infect Dis* 2012, 55:1228–1235.
 - 36. Wright S, Lu X, Peterlin BM: **Human immunodeficiency virus type 1 tat directs transcription through attenuation sites within the mouse c-myc gene.** *J Mol Biol* 1994, 243:568–573.
 - 37. Harrod R, Nacsá J, Van Lint C, Hansen J, Karpova T, McNally J, Franchini G: **Human immunodeficiency virus type-1 Tat/co-activator acetyltransferase interactions inhibit p53Lys-320 acetylation and p53-responsive transcription.** *J Biol Chem* 2003, 278:12310–12318.
 - 38. Bedi GC, Westra WH, Farzadegan H, Pitha PM, Sidransky D: **Microsatellite instability in primary neoplasms from HIV + patients.** *Nat Med* 1995, 1:65–68.
 - 39. Baker JV, Peng G, Rapkin J, Abrams DJ, Silverberg MJ, MacArthur RD, Cavert WP, Henry WK, Neaton JD: **CD4+ count and risk of non-AIDS diseases following initial treatment for HIV infection.** *AIDS* 2008, 22:841–848.
 - 40. Marin B, Thiebaut R, Bucher HC, Rondeau V, Costagliola D, Dorrucci M, Hamouda O, Prins M, Walker S, Porter K, et al: **Non-AIDS-defining deaths and immunodeficiency in the era of combination antiretroviral therapy.** *AIDS* 2009, 23:1743–1753.
 - 41. Smith C, Sabin CA, Lundgren JD, Thiebaut R, Weber R, Law M, Monforte A, Kirk O, Friis-Møller N, Phillips A, et al: **Factors associated with specific causes of death amongst HIV-positive individuals in the D:A:D Study.** *AIDS* 2010, 24:1537–1548.
 - 42. Haynes BF, Markert ML, Sempowski GD, Patel DD, Hale LP: **The role of the thymus in immune reconstitution in aging, bone marrow transplantation, and HIV-1 infection.** *Annu Rev Immunol* 2000, 18:529–560.
 - 43. Douek DC, McFarland RD, Keiser PH, Gage EA, Massey JM, Haynes BF, Polis MA, Haase AT, Feinberg MB, Sullivan JL, et al: **Changes in thymic function with age and during the treatment of HIV infection.** *Nature* 1998, 396:690–695.
 - 44. Zeng M, Haase AT, Schacker TW: **Lymphoid tissue structure and HIV-1 infection: life or death for T cells.** *Trends Immunol* 2012, 33:306–314.
 - 45. Zeng M, Smith AJ, Wietgrefe SW, Southern PJ, Schacker TW, Reilly CS, Estes JD, Burton GF, Silvestri G, Lifson JD, et al: **Cumulative mechanisms of lymphoid tissue fibrosis and T cell depletion in HIV-1 and SIV infections.** *J Clin Invest* 2011, 121:998–1008.
 - 46. Estes JD, Wietgrefe S, Schacker T, Southern P, Beilman G, Reilly C, Milush JM, Lifson JD, Sodora DL, Carlis JV, Haase AT: **Simian immunodeficiency virus-induced lymphatic tissue fibrosis is mediated by transforming growth factor beta 1-positive regulatory T cells and begins in early infection.** *J Infect Dis* 2007, 195:551–561.
 - 47. Tabb B, Morcock DR, Trubey CM, Quinones OA, Hao XP, Smedley J, Macallister R, Piatak M Jr, Harris LD, Piaiardini M, et al: **Reduced inflammation and lymphoid tissue immunopathology in rhesus macaques receiving anti-tumor necrosis factor treatment during primary simian immunodeficiency virus infection.** *J Infect Dis* 2013, 207:880–892.
 - 48. Schacker TW, Brenchley JM, Beilman GJ, Reilly C, Pambuccian SE, Taylor J, Skarda D, Larson M, Douek DC, Haase AT: **Lymphatic tissue fibrosis is associated with reduced numbers of naive CD4+ T cells in human immunodeficiency virus type 1 infection.** *Clin Vaccine Immunol* 2006, 13:556–560.
 - 49. Zeng M, Southern PJ, Reilly CS, Beilman GJ, Chipman JG, Schacker TW, Haase AT: **Lymphoid tissue damage in HIV-1 infection depletes naive T**

- cells and limits T cell reconstitution after antiretroviral therapy. *PLoS Pathog* 2012, 8:e1002437.
50. Hunt PW, Martin JN, Sinclair E, Bredt B, Hagos E, Lampiris H, Deeks SG: **T cell activation is associated with lower CD4+ T cell gains in human immunodeficiency virus-infected patients with sustained viral suppression during antiretroviral therapy.** *J Infect Dis* 2003, 187:1534–1543.
 51. Lederman MM, Calabrese L, Funderburg NT, Clagett B, Medvik K, Bonilla H, Grishovker B, Salata RA, Taegel A, Ligaris M, et al: **Immunologic failure despite suppressive antiretroviral therapy is related to activation and turnover of memory CD4 cells.** *J Infect Dis* 2011, 204:1217–1226.
 52. Alter MJ: **Epidemiology of viral hepatitis and HIV co-infection.** *J Hepatol* 2006, 44:S6–S9.
 53. Thio CL, Seaberg EC, Skolasky R Jr, Phair J, Visscher B, Munoz A, Thomas DL: **HIV-1, hepatitis B virus, and risk of liver-related mortality in the Multicenter Cohort Study (MACS).** *Lancet* 2002, 360:1921–1926.
 54. Fischer MJ, Wyatt CM, Gordon K, Gibert CL, Brown ST, Rimland D, Rodriguez-Barradas MC, Justice AC, Parikh CR: **Hepatitis C and the risk of kidney disease and mortality in veterans with HIV.** *J Acquir Immune Defic Syndr* 2010, 53:222–226.
 55. Wyatt CM, Malvestutto C, Coca SG, Klotman PE, Parikh CR: **The impact of hepatitis C virus coinfection on HIV-related kidney disease: a systematic review and meta-analysis.** *AIDS* 2008, 22:1799–1807.
 56. Freiberg MS, Chang CC, Skanderson M, McGinnis K, Kuller LH, Kraemer KL, Rimland D, Goetz MB, Butt AA, Rodriguez Barradas MC, et al: **The risk of incident coronary heart disease among veterans with and without HIV and hepatitis C.** *Circ Cardiovasc Qual Outcomes* 2011, 4:425–432.
 57. Giordano TP, Kramer JR, Soucek J, Richardson P, El-Serag HB: **Cirrhosis and hepatocellular carcinoma in HIV-infected veterans with and without the hepatitis C virus: a cohort study, 1992–2001.** *Arch Intern Med* 2004, 164:2349–2354.
 58. Mooij SH, van der Klis FR, van der Sande MA, Scheppe RM, Speksnijder AG, Bogaards JA, de Melker HE, de Vries HJ, Snijders PJ, van der Looff MF: **Seroepidemiology of High-Risk HPV in HIV-Negative and HIV-Infected MSM: The H2M Study.** *Cancer Epidemiol Biomarkers Prev* 2013, 22:1698–1708.
 59. Palmer JG, Scholefield JH, Coates PJ, Shepherd NA, Jass JR, Crawford LV, Northover JM: **Anal cancer and human papillomaviruses.** *Dis Colon Rectum* 1989, 32:1016–1022.
 60. Munoz N, Castellsague X, de Gonzalez AB, Gissmann L: **Chapter 1: HPV in the etiology of human cancer.** *Vaccine* 2006, 24(Suppl 3):S3/1–10.
 61. Neuhaus J, Jacobs DR Jr, Baker JV, Calmy A, Duprez D, La Rosa A, Kuller LH, Pett SL, Ristola M, Ross MJ, et al: **Markers of inflammation, coagulation, and renal function are elevated in adults with HIV infection.** *J Infect Dis* 2010, 201:1788–1795.
 62. Armah KA, McGinnis K, Baker J, Gibert C, Butt AA, Bryant KJ, Goetz M, Tracy R, Oursler KK, Rimland D, et al: **HIV status, burden of comorbid disease, and biomarkers of inflammation, altered coagulation, and monocyte activation.** *Clin Infect Dis* 2012, 55:126–136.
 63. Saves M, Chene G, Ducimetiere P, Leport C, Le Moal G, Amouyel P, Arveiler D, Ruidavets JB, Reynes J, Bingham A, Raffi F: **Risk factors for coronary heart disease in patients treated for human immunodeficiency virus infection compared with the general population.** *Clin Infect Dis* 2003, 37:292–298.
 64. Triant VA, Lee H, Hadigan C, Grinspoon SK: **Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease.** *J Clin Endocrinol Metab* 2007, 92:2506–2512.
 65. Goulet JL, Fultz SL, Rimland D, Butt A, Gibert C, Rodriguez-Barradas M, Bryant K, Justice AC: **Aging and infectious diseases: do patterns of comorbidity vary by HIV status, age, and HIV severity?** *Clin Infect Dis* 2007, 45:1593–1601.
 66. Cockerham L, Scherzer R, Zolopa A, Rimland D, Lewis CE, Bacchetti P, Grunfeld C, Shlipak M, Tien PC: **Association of HIV infection, demographic and cardiovascular risk factors with all-cause mortality in the recent HAART era.** *J Acquir Immune Defic Syndr* 2010, 53:102–106.
 67. Bergersen BM, Sandvik L, Dunlop O, Birkeland K, Bruun JN: **Prevalence of hypertension in HIV-positive patients on highly active retroviral therapy (HAART) compared with HAART-naïve and HIV-negative controls: results from a Norwegian study of 721 patients.** *Eur J Clin Microbiol Infect Dis* 2003, 22:731–736.
 68. Helleberg M, Afzal S, Kronborg G, Larsen CS, Pedersen G, Pedersen C, Gerstoft J, Nordestgaard BG, Obel N: **Mortality attributable to smoking among HIV-1-infected individuals: a nationwide, population-based cohort study.** *Clin Infect Dis* 2013, 56:727–734.
 69. Pereyra F, Lo J, Triant VA, Wei J, Buzon MJ, Fitch KV, Hwang J, Campbell JH, Burdo TH, Williams KC, et al: **Increased coronary atherosclerosis and immune activation in HIV-1 elite controllers.** *AIDS* 2012, 26:2409–2412.
 70. Hsue PY, Hunt PW, Schnell A, Kalapus SC, Hoh R, Ganz P, Martin JN, Deeks SG: **Role of viral replication, antiretroviral therapy, and immunodeficiency in HIV-associated atherosclerosis.** *AIDS* 2009, 23:1059–1067.
 71. Libby P, Ridker PM, Maseri A: **Inflammation and atherosclerosis.** *Circulation* 2002, 105:1135–1143.
 72. Ross R: **Atherosclerosis—an inflammatory disease.** *N Engl J Med* 1999, 340:115–126.
 73. Coussens LM, Werb Z: **Inflammation and cancer.** *Nature* 2002, 420:860–867.
 74. Grivennikov SI, Greten FR, Karin M: **Immunity, inflammation, and cancer.** *Cell* 2010, 140:883–899.
 75. Bataller R, Brenner DA: **Liver fibrosis.** *J Clin Invest* 2005, 115:209–218.
 76. Hernandez-Gea V, Friedman SL: **Pathogenesis of liver fibrosis.** *Annu Rev Pathol* 2011, 6:425–456.
 77. Kaptoge S, Di Angelantonio E, Lowe G, Pepys MB, Thompson SG, Collins R, Danesh J: **C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: an individual participant meta-analysis.** *Lancet* 2010, 375:132–140.
 78. Danesh J, Kaptoge S, Mann AG, Sarwar N, Wood A, Angleman SB, Wensley F, Higgins JP, Lennon L, Eiriksdottir G, et al: **Long-term interleukin-6 levels and subsequent risk of coronary heart disease: two new prospective studies and a systematic review.** *PLoS Med* 2008, 5:e78.
 79. Danesh J, Whincup P, Walker M, Lennon L, Thomson A, Appleby P, Rumley A, Lowe GD: **Fibrin D-dimer and coronary heart disease: prospective study and meta-analysis.** *Circulation* 2001, 103:2323–2327.
 80. Danesh J, Lewington S, Thompson SG, Lowe GD, Collins R, Kostis JB, Wilson AC, Folsom AR, Wu K, Benderly M, et al: **Plasma fibrinogen level and the risk of major cardiovascular diseases and nonvascular mortality: an individual participant meta-analysis.** *JAMA* 2005, 294:1799–1809.
 81. Il'yasova D, Colbert LH, Harris TB, Newman AB, Bauer DC, Satterfield S, Kritchevsky SB: **Circulating levels of inflammatory markers and cancer risk in the health aging and body composition cohort.** *Cancer Epidemiol Biomarkers Prev* 2005, 14:2413–2418.
 82. Siemers C, Visser LE, Coebergh JW, Splinter TA, Witteman JC, Uitterlinden AG, Hofman A, Pols HA, Stricker BH: **C-reactive protein levels, variation in the C-reactive protein gene, and cancer risk: the Rotterdam Study.** *J Clin Oncol* 2006, 24:5216–5222.
 83. Heikkila K, Harris R, Lowe G, Rumley A, Yarnell J, Gallacher J, Ben-Shlomo Y, Ebrahim S, Lawlor DA: **Associations of circulating C-reactive protein and interleukin-6 with cancer risk: findings from two prospective cohorts and a meta-analysis.** *Cancer Causes Control* 2009, 20:15–26.
 84. Harris TB, Ferrucci L, Tracy RP, Corti MC, Wacholder S, Ettinger WH Jr, Heimovitz H, Cohen HJ, Wallace R: **Associations of elevated interleukin-6 and C-reactive protein levels with mortality in the elderly.** *Am J Med* 1999, 106:506–512.
 85. Cohen HJ, Harris T, Pieper CF: **Coagulation and activation of inflammatory pathways in the development of functional decline and mortality in the elderly.** *Am J Med* 2003, 114:180–187.
 86. Shorr AF, Thomas SJ, Alkins SA, Fitzpatrick TM, Ling GS: **D-dimer correlates with proinflammatory cytokine levels and outcomes in critically ill patients.** *Chest* 2002, 121:1262–1268.
 87. Valdez H, Connick E, Smith KY, Lederman MM, Bosch RJ, Kim RS, St Clair M, Kuritzkes DR, Kessler H, Fox L, et al: **Limited immune restoration after 3 years' suppression of HIV-1 replication in patients with moderately advanced disease.** *AIDS* 2002, 16:1859–1866.
 88. Kuller LH, Tracy R, Belloso W, De Wit S, Drummond F, Lane HC, Ledergerber B, Lundgren J, Neuhaus J, Nixon D, et al: **Inflammatory and coagulation biomarkers and mortality in patients with HIV infection.** *PLoS Med* 2008, 5:e203.
 89. Tien PC, Choi AI, Zolopa AR, Benson C, Tracy R, Scherzer R, Bacchetti P, Shlipak M, Grunfeld C: **Inflammation and mortality in HIV-infected adults: analysis of the FRAM study cohort.** *J Acquir Immune Defic Syndr* 2010, 55:316–322.

90. Andrade BB, Hullsiek KH, Boulware DR, Rupert A, French MA, Ruxrungtham K, Montes ML, Price H, Barreiro P, Audsley J, et al: **Biomarkers of inflammation and coagulation are associated with mortality and hepatitis flares in persons coinfecte**d with HIV and hepatitis viruses. *J Infect Dis* 2013, **207**:1379–1388.
91. Sandler NG, Wand H, Roque A, Law M, Nason MC, Nixon DE, Pedersen C, Ruxrungtham K, Lewin SR, Emery S, et al: **Plasma levels of soluble CD14 independently predict mortality in HIV infection**. *J Infect Dis* 2011, **203**:780–790.
92. Hunt PW, Cao HL, Muzoora C, Ssewanyana I, Bennett J, Emenyonu N, Kembabazi A, Neilands TB, Bangsberg DR, Deeks SG, Martin JN: **Impact of CD8+ T-cell activation on CD4+ T-cell recovery and mortality in HIV-infected Ugandans initiating antiretroviral therapy**. *AIDS* 2011, **25**:2123–2131.
93. Duprez DA, Neuhaus J, Kuller LH, Tracy R, Belloso W, De Wit S, Drummond F, Lane HC, Ledergerber B, Lundgren J, et al: **Inflammation, coagulation and cardiovascular disease in HIV-infected individuals**. *PLoS One* 2012, **7**:e44454.
94. Ford ES, Greenwald JH, Richterman AG, Rupert A, Dutcher L, Badralmaa Y, Natarajan V, Rehm C, Hadigan C, Sereti I: **Traditional risk factors and D-dimer predict incident cardiovascular disease events in chronic HIV infection**. *AIDS* 2010, **24**:1509–1517.
95. Borges AH, Silverberg MJ, Wentworth D, Grulich AE, Fatkenheuer G, Mitsuyasu R, Tambussi G, Sabin CA, Neaton JD, Lundgren JD: **Predicting risk of cancer during HIV infection: the role of inflammatory and coagulation biomarkers**. *AIDS* 2013, **27**:1433–1441.
96. McComsey GA, Kitch D, Sax PE, Tierney C, Nahed NC, Melbourne K, Ha B, Brown TT, Bloom A, Fedarko N, Daar ES: **Associations of Inflammatory Markers with AIDS and non-AIDS Clinical Events after Initiation of Antiretroviral Therapy: AIDS Clinical Trials Group A5224s, a substudy of ACTG A5202**. *J Acquir Immune Defic Syndr* 2013;. Epub.
97. Havlir DV, Bassett R, Levitan D, Gilbert P, Tebas P, Collier AC, Hirsch MS, Ignacio C, Condra J, Gunthard HF, et al: **Prevalence and predictive value of intermittent viremia with combination hiv therapy**. *JAMA* 2001, **286**:171–179.
98. Grennan JT, Loutfy MR, Su D, Harrigan PR, Cooper C, Klein M, Machouf N, Montaner JS, Rourke S, Tsoukas C, et al: **Magnitude of virologic blips is associated with a higher risk for virologic rebound in HIV-infected individuals: a recurrent events analysis**. *J Infect Dis* 2012, **205**:1230–1238.
99. Mavigner M, Delobel P, Cazabat M, Dubois M, L'Faqihi-Olive FE, Raymond S, Pasquier C, Marchou B, Massip P, Izopet J: **HIV-1 residual viremia correlates with persistent T-cell activation in poor immunological responders to combination antiretroviral therapy**. *PLoS One* 2009, **4**:e7658.
100. Wittkop L, Bitard J, Lazaro E, Neau D, Bonnet F, Mercie P, Dupon M, Hessamfar M, Ventura M, Malvy D, et al: **Effect of cytomegalovirus-induced immune response, self antigen-induced immune response, and microbial translocation on chronic immune activation in successfully treated HIV type 1-infected patients: the ANRS CO3 Aquitaine Cohort**. *J Infect Dis* 2013, **207**:622–627.
101. van de Berg PJ, Heutink KM, Raabe R, Minnee RC, Young SL, van Donselaar-van der Pant KA, Bemelman FJ, van Lier RA, ten Berge IJ: **Human cytomegalovirus induces systemic immune activation characterized by a type 1 cytokine signature**. *J Infect Dis* 2010, **202**:690–699.
102. Hsue PY, Hunt PW, Sinclair E, Bredt B, Franklin A, Kilian M, Hoh R, Martin JN, McCune JM, Waters DD, Deeks SG: **Increased carotid intima-media thickness in HIV patients is associated with increased cytomegalovirus-specific T-cell responses**. *AIDS* 2006, **20**:2275–2283.
103. Sacre K, Hunt PW, Hsue PY, Maidji E, Martin JN, Deeks SG, Autran B, McCune JM: **A role for cytomegalovirus-specific CD4+CX3CR1+ T cells and cytomegalovirus-induced T-cell immunopathology in HIV-associated atherosclerosis**. *AIDS* 2012, **26**:805–814.
104. Kovacs A, Al-Harthi L, Christensen S, Mack W, Cohen M, Landay A: **CD8+ T cell activation in women coinfecte**d with human immunodeficiency virus type 1 and hepatitis C virus. *J Infect Dis* 2008, **197**:1402–1407.
105. Gonzalez VD, Falconer K, Blom KG, Reichard O, Morn B, Laursen AL, Weis N, Alaeus A, Sandberg JK: **High levels of chronic immune activation in the T-cell compartments of patients coinfecte**d with hepatitis C virus and human immunodeficiency virus type 1 and on highly active antiretroviral therapy are reverted by alpha interferon and ribavirin treatment. *J Virol* 2009, **83**:11407–11411.
106. Estes JD, Harris LD, Klatt NR, Tabb B, Pittaluga S, Paiardini M, Barclay GR, Smedley J, Pung R, Oliveira KM, et al: **Damaged intestinal epithelial integrity linked to microbial translocation in pathogenic simian immunodeficiency virus infections**. *PLoS Pathog* 2010, **6**:e1001052.
107. Pandrea IV, Gautam R, Ribeiro RM, Brenchley JM, Butler LF, Pattison M, Rasmussen T, Marx PA, Silvestri G, Lackner AA, et al: **Acute loss of intestinal CD4+ T cells is not predictive of simian immunodeficiency virus virulence**. *J Immunol* 2007, **179**:3035–3046.
108. Pandrea I, Gaufin T, Brenchley JM, Gautam R, Monjure C, Gautam A, Coleman C, Lackner AA, Ribeiro RM, Douek DC, Apetrei C: **Cutting edge: experimentally induced immune activation in natural hosts of simian immunodeficiency virus induces significant increases in viral replication and CD4+ T cell depletion**. *J Immunol* 2008, **181**:6687–6691.
109. Brenchley JM, Price DA, Schacker TW, Asher TE, Silvestri G, Rao S, Kazzaz Z, Bornstein E, Lambotte O, Altmann D, et al: **Microbial translocation is a cause of systemic immune activation in chronic HIV infection**. *Nat Med* 2006, **12**:1365–1371.
110. Cassol E, Malfeld S, Mahasha P, van der Merwe S, Cassol S, Seebregts C, Alfano M, Poli G, Rossouw T: **Persistent microbial translocation and immune activation in HIV-1-infected South Africans receiving combination antiretroviral therapy**. *J Infect Dis* 2010, **202**:723–733.
111. Jiang W, Lederman MM, Hunt P, Sieg SF, Haley K, Rodriguez B, Landay A, Martin J, Sinclair E, Asher AI, et al: **Plasma levels of bacterial DNA correlate with immune activation and the magnitude of immune restoration in persons with antiretroviral-treated HIV infection**. *J Infect Dis* 2009, **199**:1177–1185.
112. Funderburg NT, Mayne E, Sieg SF, Asaad R, Jiang W, Kalinowska M, Luciano AA, Stevens W, Rodriguez B, Brenchley JM, et al: **Increased tissue factor expression on circulating monocytes in chronic HIV infection: relationship to in vivo coagulation and immune activation**. *Blood* 2010, **115**:161–167.
113. Martin DM, Boys CW, Ruf W: **Tissue factor: molecular recognition and cofactor function**. *FASEB J* 1995, **9**:852–859.
114. Bavinger C, Bendavid E, Niehaus K, Olshen RA, Olkin I, Sundaram V, Wein N, Holodniy M, Hou N, Owens DK, Desai M: **Risk of cardiovascular disease from antiretroviral therapy for HIV: a systematic review**. *PLoS One* 2013, **8**:e59551.
115. Mocroft A, Phillips AN, Gatell J, Horban A, Ledergerber B, Zilmer K, Jevtic D, Maltez F, Podlekareva D, Lundgren JD: **CD4 cell count and viral load-specific rates of AIDS, non-AIDS and deaths according to current antiretroviral use**. *AIDS* 2013, **27**:907–918.
116. Weber R, Sabin CA, Friis-Moller N, Reiss P, El-Sadr WM, Kirk O, Dabis F, Law MG, Pradier C, De Wit S, et al: **Liver-related deaths in persons infected with the human immunodeficiency virus: the D:A:D study**. *Arch Intern Med* 2006, **166**:1632–1641.
117. Mocroft A, Soriano V, Rockstroh J, Reiss P, Kirk O, de Wit S, Gatell J, Clotet B, Phillips AN, Lundgren JD: **Is there evidence for an increase in the death rate from liver-related disease in patients with HIV?** *AIDS* 2005, **19**:2117–2125.
118. Aceti A, Pasquazzi C, Zechini B, De Bac C: **Hepatotoxicity development during antiretroviral therapy containing protease inhibitors in patients with HIV: the role of hepatitis B and C virus infection**. *J Acquir Immune Defic Syndr* 2002, **29**:41–48.
119. Wit FW, Weverling GJ, Weel J, Jurriaans S, Lange JM: **Incidence of and risk factors for severe hepatotoxicity associated with antiretroviral combination therapy**. *J Infect Dis* 2002, **186**:23–31.
120. Sulkowski MS, Thomas DL, Chaisson RE, Moore RD: **Hepatotoxicity associated with antiretroviral therapy in adults infected with human immunodeficiency virus and the role of hepatitis C or B virus infection**. *JAMA* 2000, **283**:74–80.
121. Brau N, Salvatore M, Rios-Bedoya CF, Fernandez-Carbia A, Paronetto F, Rodriguez-Orengo JF, Rodriguez-Torres M: **Slower fibrosis progression in HIV/HCV-coinfected patients with successful HIV suppression using antiretroviral therapy**. *J Hepatol* 2006, **44**:47–55.
122. Qurishi N, Kreuzberg C, Luchters G, Effenberger W, Kupfer B, Sauerbruch T, Rockstroh JK, Spengler U: **Effect of antiretroviral therapy on liver-related mortality in patients with HIV and hepatitis C virus coinfection**. *Lancet* 2003, **362**:1708–1713.
123. Kovari H, Sabin CA, Ledergerber B, Ryom L, Worm SW, Smith C, Phillips A, Reiss P, Fontas E, Petoumenos K, et al: **Antiretroviral drug-related liver mortality among HIV-positive persons in the absence of hepatitis B or C**

- virus coinfection: the data collection on adverse events of anti-HIV drugs study. *Clin Infect Dis* 2013, **56**:870–879.
124. Kalayjian RC, Lau B, Mechekano RN, Crane HM, Rodriguez B, Salata RA, Krishnasami Z, Willig JH, Martin JN, Moore RD, et al: Risk factors for chronic kidney disease in a large cohort of HIV-1 infected individuals initiating antiretroviral therapy in routine care. *AIDS* 2012, **26**:1907–1915.
125. Choi AI, Shlipak MG, Hunt PW, Martin JN, Deeks SG: HIV-infected persons continue to lose kidney function despite successful antiretroviral therapy. *AIDS* 2009, **23**:2143–2149.
126. Cooper RD, Wiebe N, Smith N, Keiser P, Naicker S, Tonelli M: Systematic review and meta-analysis: renal safety of tenofovir disoproxil fumarate in HIV-infected patients. *Clin Infect Dis* 2010, **51**:496–505.
127. Abrams D, Levy Y, Losso MH, Babiker A, Collins G, Cooper DA, Darbyshire J, Emery S, Fox L, Gordin F, et al: Interleukin-2 therapy in patients with HIV infection. *N Engl J Med* 2009, **361**:1548–1559.
128. Emery S, Neuhaus JA, Phillips AN, Babiker A, Cohen CJ, Gatell JM, Girard PM, Grund B, Law M, Losso MH, et al: Major clinical outcomes in antiretroviral therapy (ART)-naïve participants and in those not receiving ART at baseline in the SMART study. *J Infect Dis* 2008, **197**:1133–1144.
129. Cohen MS, Chen YQ, McCauley M, Gamble T, Hosseinipour MC, Kumarasamy N, Hakim JG, Kumwenda J, Grinsztejn B, Pilotti JH, et al: Prevention of HIV-1 infection with early antiretroviral therapy. *N Engl J Med* 2011, **365**:493–505.
130. Severe P, Juste MA, Ambroise A, Eliacin L, Marchand C, Apollon S, Edwards A, Bang H, Nicotera J, Godfrey C, et al: Early versus standard antiretroviral therapy for HIV-infected adults in Haiti. *N Engl J Med* 2010, **363**:257–265.
131. Sterne JA, May M, Costagliola D, de Wolf F, Phillips AN, Harris R, Funk MJ, Geskus RB, Gill J, Dabis F, et al: Timing of initiation of antiretroviral therapy in AIDS-free HIV-1-infected patients: a collaborative analysis of 18 HIV cohort studies. *Lancet* 2009, **373**:1352–1363.
132. Kitahata MM, Gange SJ, Abraham AG, Merriman B, Saag MS, Justice AC, Hogg RS, Deeks SG, Eron JJ, Brooks JT, et al: Effect of early versus deferred antiretroviral therapy for HIV on survival. *N Engl J Med* 2009, **360**:1815–1826.
133. Lee L: Timing of HAART initiation and clinical outcomes in human immunodeficiency virus type 1 seroconverters. *Arch Intern Med* 2011, **171**:1560–1569.
134. Cain LE, Logan R, Robins JM, Sterne JA, Sabin C, Bansi L, Justice A, Goulet J, van Sighem A, de Wolf F, et al: When to initiate combined antiretroviral therapy to reduce mortality and AIDS-defining illness in HIV-infected persons in developed countries: an observational study. *Ann Intern Med* 2011, **154**:509–515.
135. Keiser O, Anastos K, Schechter M, Balestre E, Myer L, Boulle A, Bangsberg D, Toure H, Braitstein P, Sprinz E, et al: Antiretroviral therapy in resource-limited settings 1996 to 2006: patient characteristics, treatment regimens and monitoring in sub-Saharan Africa, Asia and Latin America. *Trop Med Int Health* 2008, **13**:870–879.
136. Egger M: Immunodeficiency at the start of combination antiretroviral therapy in low-, middle-and high-income countries. *J Acquir Immune Defic Syndr* 2013. In press.
137. Sereti I, Dunham RM, Spritzler J, Aga E, Proschan MA, Medvik K, Battaglia CA, Landay AL, Pahwa S, Fischl MA, et al: IL-7 administration drives T cell-cycle entry and expansion in HIV-1 infection. *Blood* 2009, **113**:6304–6314.
138. Levy Y, Lacabaratz C, Weiss L, Viard JP, Goujard C, Lelievre JD, Boue F, Molina JM, Rouzioux C, Avettand-Fenoel V, et al: Enhanced T cell recovery in HIV-1-infected adults through IL-7 treatment. *J Clin Invest* 2009, **119**:997–1007.
139. Hunt PW: Th17, gut, and HIV: therapeutic implications. *Curr Opin HIV and AIDS* 2010, **5**:189–193.
140. Kanwar B, Favre D, McCune JM: Th17 and regulatory T cells: implications for AIDS pathogenesis. *Curr Opin HIV and AIDS* 2010, **5**:151–157.
141. Macias-Barragan J, Sandoval-Rodriguez A, Navarro-Partida J, Armendariz-Borunda J: The multifaceted role of pirenidone and its novel targets. *Fibrogenesis Tissue Repair* 2010, **3**:16.
142. Plaeger SF, Collins BS, Musib B, Deeks SG, Read S, Embry A: Immune activation in the pathogenesis of treated chronic HIV disease: a workshop summary. *AIDS Res Hum Retroviruses* 2012, **28**:469–477.
143. Kim S, Iwao H: Molecular and cellular mechanisms of angiotensin II-mediated cardiovascular and renal diseases. *Pharmacol Rev* 2000, **52**:11–34.
144. Abbas G, Silveira MG, Lindor KD: Hepatic fibrosis and the renin-angiotensin system. *Am J Ther* 2011, **18**:e202–e208.
145. Petoumenos K, Worm S, Reiss P, de Wit S, d'Arminio Monforte A, Sabin C, Friis-Møller N, Weber R, Mercie P, Pradier C, et al: Rates of cardiovascular disease following smoking cessation in patients with HIV infection: results from the D:A:D study(*). *HIV Med* 2011, **12**:412–421.
146. EACS Guidelines. http://www.eacsociety.org/Portals/0/Guidelines_Online_131014.pdf.
147. Dube MP, Stein JH, Aberg JA, Fichtenbaum CJ, Gerber JG, Tashima KT, Henry WK, Currier JS, Sprecher D, Glesby MJ: Guidelines for the evaluation and management of dyslipidemia in human immunodeficiency virus (HIV)-infected adults receiving antiretroviral therapy: recommendations of the HIV Medical Association of the Infectious Disease Society of America and the Adult AIDS Clinical Trials Group. *Clin Infect Dis* 2003, **37**:613–627.
148. Friis-Møller N, Thiebaut R, Reiss P, Weber R, Monforte AD, De Wit S, El-Sadr W, Fontas E, Worm S, Kirk O, et al: Predicting the risk of cardiovascular disease in HIV-infected patients: the data collection on adverse effects of anti-HIV drugs study. *Eur J Cardiovasc Prev Rehabil* 2010, **17**:491–501.
149. Reinsch N, Neuhaus K, Esser S, Potthoff A, Hower M, Mostardt S, Neumann A, Brockmeyer NH, Gelbrich G, Erbel R, Neumann T: Are HIV patients undertreated? Cardiovascular risk factors in HIV: results of the HIV-HEART study. *Eur J Prev Cardiol* 2012, **19**:267–274.
150. Lundgren JD, Battegay M, Behrens G, De Wit S, Guaraldi G, Katlama C, Martinez E, Nair D, Powderly WG, Reiss P, et al: European AIDS Clinical Society (EACS) guidelines on the prevention and management of metabolic diseases in HIV. *HIV Med* 2008, **9**:72–81.
151. Melzi S, Carenzi L, Cossu MV, Passerini S, Capetti A, Rizzardini G: Lipid Metabolism and Cardiovascular Risk in HIV-1 Infection and HAART: Present and Future Problems. *Cholesterol* 2010, **2010**:271504.
152. Madruga JR, Cassetti I, Suleiman JM, Etzel A, Zhong L, Holmes CB, Cheng AK, Enejosa J, Study ET: The safety and efficacy of switching stavudine to tenofovir df in combination with lamivudine and efavirenz in hiv-1-infected patients: three-year follow-up after switching therapy. *HIV Clin Trials* 2007, **8**:381–390.
153. Negredo E, Cruz L, Paredes R, Ruiz L, Fumaz CR, Bonjoch A, Gel S, Tuldra A, Balague M, Johnston S, et al: Virological, immunological, and clinical impact of switching from protease inhibitors to nevirapine or to efavirenz in patients with human immunodeficiency virus infection and long-lasting viral suppression. *Clin Infect Dis* 2002, **34**:504–510.
154. Soriano V, Garcia-Gasco P, Vispo E, Ruiz-Sancho A, Blanco F, Martin-Carbonero L, Rodriguez-Novoa S, Morello J, de Mendoza C, Rivas P, et al: Efficacy and safety of replacing lopinavir with atazanavir in HIV-infected patients with undetectable plasma viraemia: final results of the SLOAT trial. *J Antimicrob Chemother* 2008, **61**:200–205.
155. Mallolas J, Podzamczer D, Milinkovic A, Domingo P, Clotet B, Ribera E, Gutierrez F, Knobel H, Cosin J, Ferrer E, et al: Efficacy and safety of switching from boosted lopinavir to boosted atazanavir in patients with virological suppression receiving a LPV/r-containing HAART: the ATAZIP study. *J Acquir Immune Defic Syndr* 2009, **51**:29–36.
156. Gatell J, Salmon-Ceron D, Lazzarin A, Van Wijngaarden E, Antunes F, Leen C, Horban A, Wirtz V, Odesho L, Van den Dungen M, et al: Efficacy and safety of atazanavir-based highly active antiretroviral therapy in patients with virologic suppression switched from a stable, boosted or unboosted protease inhibitor treatment regimen: the SWAN Study (AI424-097) 48-week results. *Clin Infect Dis* 2007, **44**:1484–1492.
157. Colafogli M, Di Giambenedetto S, Bracciale L, Tamburini E, Cauda R, De Luca A: Cardiovascular risk score change in HIV-1-infected patients switched to an atazanavir-based combination antiretroviral regimen. *HIV Med* 2008, **9**:172–177.
158. Eron JJ, Young B, Cooper DA, Youle M, Dejesus E, Andrade-Villanueva J, Workman C, Zajdenverg R, Fatkenheuer G, Berger DS, et al: Switch to a raltegravir-based regimen versus continuation of a lopinavir-ritonavir-based regimen in stable HIV-infected patients with suppressed viraemia (SWITCHMRK 1 and 2): two multicentre, double-blind, randomised controlled trials. *Lancet* 2010, **375**:396–407.
159. Martinez E, Larrousse M, Llibre JM, Gutierrez F, Saumoy M, Antela A, Knobel H, Murillas J, Berenguer J, Pich J, et al: Substitution of raltegravir for ritonavir-boosted protease inhibitors in HIV-infected patients: the SPIRAL study. *AIDS* 2010, **24**:1697–1707.

160. Masia M, Martinez E, Padilla S, Gatell JM, Gutierrez F: Endothelial function in HIV-infected patients switching from a boosted protease inhibitor-based regimen to raltegravir: a substudy of the SPIRAL study. *J Antimicrob Chemother* 2013, **68**:409–413.
161. Gandhi RT, Zheng L, Bosch RJ, Chan ES, Margolis DM, Read S, Kallungal B, Palmer S, Medvik K, Lederman MM, et al: The effect of raltegravir intensification on low-level residual viremia in HIV-infected patients on antiretroviral therapy: a randomized controlled trial. *PLoS Med* 2010, **7**:8.
162. Hatano H, Hayes TL, Dahl V, Sinclair E, Lee TH, Hoh R, Lampiris H, Hunt PW, Palmer S, McCune JM, et al: A randomized, controlled trial of raltegravir intensification in antiretroviral-treated, HIV-infected patients with a suboptimal CD4+ T cell response. *J Infect Dis* 2011, **203**:960–968.
163. Yukl SA, Shergill AK, McQuaid K, Gianella S, Lampiris H, Hare CB, Pandori M, Sinclair E, Gunthard HF, Fischer M, et al: Effect of raltegravir-containing intensification on HIV burden and T-cell activation in multiple gut sites of HIV-positive adults on suppressive antiretroviral therapy. *AIDS* 2010, **24**:2451–2460.
164. Dahl V, Lee E, Peterson J, Spudich SS, Leppla I, Sinclair E, Fuchs D, Palmer S, Price RW: Raltegravir treatment intensification does not alter cerebrospinal fluid HIV-1 infection or immunoactivation in subjects on suppressive therapy. *J Infect Dis* 2011, **204**:1936–1945.
165. Libre JM, Buzon MJ, Massanella M, Esteve A, Dahl V, Puertas MC, Domingo P, Gatell JM, Larrouse M, Gutierrez M, et al: Treatment intensification with raltegravir in subjects with sustained HIV-1 viraemia suppression: a randomized 48-week study. *Antivir Ther* 2012, **17**:355–364.
166. Vallejo A, Gutierrez C, Hernandez-Novoa B, Diaz L, Madrid N, Abad-Fernandez M, Dronda F, Perez-Elias MJ, Zamora J, Munoz E, et al: The effect of intensification with raltegravir on the HIV-1 reservoir of latently infected memory CD4 T cells in suppressed patients. *AIDS* 2012, **26**:1885–1894.
167. Byakwaga H, Kelly M, Purcell DF, French MA, Amin J, Lewin SR, Haskelberg H, Kelleher AD, Garcia R, Boyd MA, et al: Intensification of antiretroviral therapy with raltegravir or addition of hyperimmune bovine colostrum in HIV-infected patients with suboptimal CD4+ T-cell response: a randomized controlled trial. *J Infect Dis* 2011, **204**:1532–1540.
168. Hatano H, Strain MC, Scherzer R, Bacchetti P, Wentworth D, Hoh R, Martin JN, McCune JM, Neaton JD, Tracy RP, et al: Increase in 2-LTR Circles and Decrease in D-dimer After Raltegravir Intensification in Treated HIV-Infected Patients: A Randomized Placebo-Controlled Trial. *J Infect Dis* 2013, **208**(9):1436–1442.
169. Massanella M, Negredo E, Puig J, Puertas MC, Buzon MJ, Perez-Alvarez N, Carrillo J, Clotet B, Martinez-Picado J, Blanco J: Raltegravir intensification shows differing effects on CD8 and CD4 T cells in HIV-infected HAART-suppressed individuals with poor CD4 T-cell recovery. *AIDS* 2012, **26**:2285–2293.
170. Buzon MJ, Massanella M, Llibre JM, Esteve A, Dahl V, Puertas MC, Gatell JM, Domingo P, Paredes R, Sharkey M, et al: HIV-1 replication and immune dynamics are affected by raltegravir intensification of HAART-suppressed subjects. *Nat Med* 2010, **16**:460–465.
171. Cuzin L, Trabelsi S, Delobel P, Barbuat C, Reynes J, Allavena C, Peytavin G, Ghosn J, Lascoix-Combe C, Psomas C, et al: Maraviroc intensification of stable antiviral therapy in HIV-1-infected patients with poor immune restoration: MARIMUNO-ANRS 145 study. *J Acquir Immune Defic Syndr* 2012, **61**:557–564.
172. Wilkin TJ, Lalama CM, McKinnon J, Gandhi RT, Lin N, Landay A, Ribaud H, Fox L, Currier JS, Mellors JW, et al: A pilot trial of adding maraviroc to suppressive antiretroviral therapy for suboptimal CD4+ T-cell recovery despite sustained virologic suppression: ACTG A5256. *J Infect Dis* 2012, **206**:534–542.
173. Gutierrez C, Diaz L, Vallejo A, Hernandez-Novoa B, Abad M, Madrid N, Dahl V, Rubio R, Moreno AM, Dronda F, et al: Intensification of antiretroviral therapy with a CCR5 antagonist in patients with chronic HIV-1 infection: effect on T cells latently infected. *PLoS One* 2011, **6**:e27864.
174. Hunt PW, Shulman NS, Hayes TL, Dahl V, Somsouk M, Funderburg NT, McLaughlin B, Landay AL, Adeyemi O, Gilman LE, et al: The immunologic effects of maraviroc intensification in treated HIV-infected individuals with incomplete CD4+ T-cell recovery: a randomized trial. *Blood* 2013, **121**:4635–4646.
175. Berenguer J, Alvarez-Pellicer J, Martin PM, Lopez-Aldeguer J, Von-Wichmann MA, Quereda C, Mallolas J, Sanz J, Tural C, Bellon JM, Gonzalez-Garcia J: Sustained virological response to interferon plus ribavirin reduces liver-related complications and mortality in patients coinfected with human immunodeficiency virus and hepatitis C virus. *Hepatology* 2009, **50**:407–413.
176. Berenguer J, Rodriguez E, Miralles P, Von Wichmann MA, Lopez-Aldeguer J, Mallolas J, Galindo MJ, Van Den Eynde E, Tellez MJ, Quereda C, et al: Sustained virological response to interferon plus ribavirin reduces non-liver-related mortality in patients coinfected with HIV and Hepatitis C virus. *Clin Infect Dis* 2012, **55**:728–736.
177. Jennings CL, Sherman KE: Hepatitis C and HIV co-infection: new drugs in practice and in the pipeline. *Curr HIV/AIDS Rep* 2012, **9**:231–237.
178. Hunt PW, Martin JN, Sinclair E, Epling L, Teague J, Jacobson MA, Tracy RP, Corey L, Deeks SG: Valganciclovir reduces T cell activation in HIV-infected individuals with incomplete CD4+ T cell recovery on antiretroviral therapy. *J Infect Dis* 2011, **203**:1474–1483.
179. Yi TJ, Walmsley S, Szadkowski L, Raboud J, Rajwans N, Shannon B, Kumar S, Kain KC, Kaul R, Tan DH: A randomized controlled pilot trial of valacyclovir for attenuating inflammation and immune activation in HIV, HSV-2 co-infected adults on suppressive antiretroviral therapy. *Clin Infect Dis* 2013, **57**(9):1331–1338.
180. Baigent C, Keech A, Kearney PM, Blackwell L, Buck G, Pollicino C, Kirby A, Sourjina T, Peto R, Collins R, Simes R: Efficacy and safety of cholesterol-lowering treatment: prospective meta-analysis of data from 90,056 participants in 14 randomised trials of statins. *Lancet* 2005, **366**:1267–1278.
181. Jain MK, Ridker PM: Anti-inflammatory effects of statins: clinical evidence and basic mechanisms. *Nat Rev Drug Discov* 2005, **4**:977–987.
182. Aslangul E, Fellahi S, Assoumou LK, Bastard JP, Capeau J, Costagliola D: High-sensitivity C-reactive protein levels fall during statin therapy in HIV-infected patients receiving ritonavir-boosted protease inhibitors. *AIDS* 2011, **25**:1128–1131.
183. De Wit S, Delforge M, Necsoi CV, Clumeck N: Downregulation of CD38 activation markers by atorvastatin in HIV patients with undetectable viral load. *AIDS* 2011, **25**:1332–1333.
184. Ganesan A, Crum-Cianflone N, Higgins J, Qin J, Rehm C, Metcalf J, Brandt C, Vita J, Decker CF, Sklar P, et al: High dose atorvastatin decreases cellular markers of immune activation without affecting HIV-1 RNA levels: results of a double-blind randomized placebo controlled clinical trial. *J Infect Dis* 2011, **203**:756–764.
185. Moore RD, Bartlett JG, Gallant JE: Association between use of HMG CoA reductase inhibitors and mortality in HIV-infected patients. *PLoS One* 2011, **6**:e21843.
186. Overton ET, Kitch D, Benson CA, Hunt PW, Stein JH, Smurzynski M, Ribaud HJ, Tebas P: Effect of statin therapy in reducing the risk of serious non-AIDS-defining events and nonaccidental death. *Clin Infect Dis* 2013, **56**:1471–1479.
187. Rasmussen LD, Kronborg G, Larsen CS, Pedersen C, Gerstoft J, Obel N: Statin therapy and mortality in HIV-infected individuals; a Danish nationwide population-based cohort study. *PLoS One* 2013, **8**:e52828.
188. Ben-Zvi I, Kivity S, Langevitz P, Shoenfeld Y: Hydroxychloroquine: from malaria to autoimmunity. *Clin Rev Allergy Immunol* 2012, **42**:145–153.
189. Savarino A, Gennaro L, Sperber K, Boelaert JR: The anti-HIV-1 activity of chloroquine. *J Clin Virol* 2001, **20**:131–135.
190. Sperber K, Chiang G, Chen H, Ross W, Chusid E, Gonchar M, Chow R, Liriano O: Comparison of hydroxychloroquine with zidovudine in asymptomatic patients infected with human immunodeficiency virus type 1. *Clin Ther* 1997, **19**:913–923.
191. Murray SM, Down CM, Boulware DR, Stauffer WM, Cavert WP, Schacker TW, Brenchley JM, Douek DC: Reduction of immune activation with chloroquine therapy during chronic HIV infection. *J Virol* 2010, **84**:12082–12086.
192. Paton NI, Goodall RL, Dunn DT, Franzen S, Collaco-Moraes Y, Gazzard BG, Williams IG, Fisher MJ, Winston A, Fox J, et al: Effects of hydroxychloroquine on immune activation and disease progression among HIV-infected patients not receiving antiretroviral therapy: a randomized controlled trial. *JAMA* 2012, **308**:353–361.
193. Piconi S, Parisotto S, Rizzardini G, Passerini S, Terzi R, Argenteri B, Meraviglia P, Capetti A, Biasin M, Trabattoni D, Clerici M: Hydroxychloroquine drastically reduces immune activation in HIV-infected, antiretroviral therapy-treated immunologic nonresponders. *Blood* 2011, **118**:3263–3272.
194. Harris SG, Padilla J, Koumas L, Ray D, Phipps RP: Prostaglandins as modulators of immunity. *Trends Immunol* 2002, **23**:144–150.

195. Pettersen FO, Torheim EA, Dahm AE, Aaberge IS, Lind A, Holm M, Aandahl EM, Sandset PM, Tasken K, Kvale D: An exploratory trial of cyclooxygenase type 2 inhibitor in HIV-1 infection: downregulated immune activation and improved T cell-dependent vaccine responses. *J Virol* 2011, 85:6557–6566.
196. Kvale D, Ormaasen V, Kran AM, Johansson CC, Aukrust P, Aandahl EM, Froland SS, Tasken K: Immune modulatory effects of cyclooxygenase type 2 inhibitors in HIV patients on combination antiretroviral treatment. *AIDS* 2006, 20:813–820.
197. Cannon CP, Cannon PJ: Physiology. COX-2 inhibitors and cardiovascular risk. *Science* 2012, 336:1386–1387.
198. Baigent C, Blackwell L, Collins R, Emberson J, Godwin J, Peto R, Buring J, Hennekens C, Kearney P, Meade T, et al: Aspirin in the primary and secondary prevention of vascular disease: collaborative meta-analysis of individual participant data from randomised trials. *Lancet* 2009, 373:1849–1860.
199. O'Brien M, Montenont E, Hu L, Nardi MA, Valdes V, Merolla M, Gettenberg G, Cavanagh K, Aberg JA, Bhardwaj N, Berger JS: Aspirin attenuates platelet activation and immune activation in HIV-1-infected subjects on antiretroviral therapy: a pilot study. *J Acquir Immune Defic Syndr* 2013, 63:280–288.
200. Read SW, DeGrezia M, Ciccone EJ, DerSimonian R, Higgins J, Adelsberger JW, Starling JM, Rehm C, Sereti I: The effect of leflunomide on cycling and activation of T-cells in HIV-1-infected participants. *PLoS One* 2010, 5:e11937.
201. Andriu JM, Lu W: Long-term clinical, immunologic and virologic impact of glucocorticoids on the chronic phase of HIV infection. *BMC Med* 2004, 2:17.
202. Ulmer A, Muller M, Bertisch-Mollenhoff B, Frietsch B: Low dose prednisolone reduces CD4+ T cell loss in therapy-naïve HIV-patients without antiretroviral therapy. *Eur J Med Res* 2005, 10:105–109.
203. McComsey GA, Whalen CC, Mawhorter SD, Asaad R, Valdez H, Patki AH, Klaumunzner J, Gopalakrishna KV, Calabrese LH, Lederman MM: Placebo-controlled trial of prednisone in advanced HIV-1 infection. *AIDS* 2001, 15:321–327.
204. Wallis RS, Kalayjian R, Jacobson JM, Fox L, Purdue L, Shikuma CM, Arakaki R, Snyder S, Coombs RW, Bosch RJ, et al: A study of the immunology, virology, and safety of prednisone in HIV-1-infected subjects with CD4 cell counts of 200 to 700 mm³. *J Acquir Immune Defic Syndr* 2003, 32:281–286.
205. Da Silva JA, Jacobs JW, Kirwan JR, Boers M, Saag KG, Ines LB, de Koning EJ, Buttigereit F, Cutolo M, Capell H, et al: Safety of low dose glucocorticoid treatment in rheumatoid arthritis: published evidence and prospective trial data. *Ann Rheum Dis* 2006, 65:285–293.
206. Miller KD, Masur H, Jones EC, Joe GO, Rick ME, Kelly GG, Mican JM, Liu S, Gerber LH, Blackwelder WC, et al: High prevalence of osteonecrosis of the femoral head in HIV-infected adults. *Ann Intern Med* 2002, 137:17–25.
207. Gori A, Tincati C, Rizzardini G, Torti C, Quirino T, Haarman M, Ben Amor K, van Schaik J, Vriesema A, Knol J, et al: Early impairment of gut function and gut flora supporting a role for alteration of gastrointestinal mucosa in human immunodeficiency virus pathogenesis. *J Clin Microbiol* 2008, 46:757–758.
208. Roberfroid M: Prebiotics: the concept revisited. *J Nutr* 2007, 137:830S–837S.
209. Sanders ME: Probiotics: definition, sources, selection, and uses. *Clin Infect Dis* 2008, 46(Suppl 2):S58–S61.
210. Gori A, Rizzardini G, Van't Land B, Amor KB, van Schaik J, Torti C, Quirino T, Tincati C, Bandera A, Knol J, et al: Specific prebiotics modulate gut microbiota and immune activation in HAART-naïve HIV-infected adults: results of the "COPA" pilot randomized trial. *Mucosal Immunol* 2011, 4:554–563.
211. Irvine SL, Hummelen R, Hekmat S, Looman CW, Habbema JD, Reid G: Probiotic yogurt consumption is associated with an increase of CD4 count among people living with HIV/AIDS. *J Clin Gastroenterol* 2010, 44:e201–e205.
212. Gonzalez-Hernandez LA, Jave-Suarez LF, Fafutis-Morris M, Montes-Salcedo KE, Valle-Gutierrez LG, Campos-Loza AE, Enciso-Gomez LF, Andrade-Villanueva JF: Synbiotic therapy decreases microbial translocation and inflammation and improves immunological status in HIV-infected patients: a double-blind randomized controlled pilot trial. *Nutr J* 2012, 11:90.
213. Schunter M, Chu H, Hayes TL, McConnell D, Crawford SS, Luciw PA, Bengmark S, Asmuth DM, Brown J, Bevins CL, et al: Randomized pilot trial of a synbiotic dietary supplement in chronic HIV-1 infection. *BMC Complement Altern Med* 2012, 12:84.
214. Floren CH, Chinenye S, Elfstrand L, Hagman C, Ihse I: ColoPlus, a new product based on bovine colostrum, alleviates HIV-associated diarrhea. *Scand J Gastroenterol* 2006, 41:682–686.
215. Plettenberg A, Stoehr A, Stellbrink HJ, Albrecht H, Meigel W: A preparation from bovine colostrum in the treatment of HIV-positive patients with chronic diarrhea. *Clin Investig* 1993, 71:42–45.
216. Rump JA, Arndt R, Arnold A, Bendick C, Dichtelmuller H, Franke M, Helm EB, Jager H, Kampmann B, Kolb P, et al: Treatment of diarrhea in human immunodeficiency virus-infected patients with immunoglobulins from bovine colostrum. *Clin Investig* 1992, 70:588–594.
217. Jiang ZD, DuPont HL: Rifaximin: in vitro and in vivo antibacterial activity—a review. *Chemotherapy* 2005, 51(Suppl 1):67–72.
218. Festi D, Mazzella G, Orsini M, Sottilli S, Sangermano A, Li Bassi S, Parini P, Ferrieri A, Falcucci M, Grossi L: Rifaximin in the treatment of chronic hepatic encephalopathy; results of a multicenter study of efficacy and safety. *Curr Ther Res* 1993, 54:598–609.
219. Bass NM, Mullen KD, Sanyal A, Poordad F, Neff G, Leevy CB, Sigal S, Sheikh MY, Beavers K, Frederick T, et al: Rifaximin treatment in hepatic encephalopathy. *N Engl J Med* 2010, 362:1071–1081.
220. Ambizas EM, Ginzburg R: Lubiprostone: a chloride channel activator for treatment of chronic constipation. *Ann Pharmacother* 2007, 41:957–964.
221. Moeser AJ, Nighot PK, Engelke KJ, Ueno R, Blikslager AT: Recovery of mucosal barrier function in ischemic porcine ileum and colon is stimulated by a novel agonist of the CIC-2 chloride channel, lubiprostone. *Am J Physiol Gastrointest Liver Physiol* 2007, 292:G647–G656.
222. Manns B, Stevens L, Miskulin D, Owen WF Jr, Winkelmayr WC, Tonelli M: A systematic review of sevelamer in ESRD and an analysis of its potential economic impact in Canada and the United States. *Kidney Int* 2004, 66:1239–1247.
223. Stinghen AE, Goncalves SM, Bucharles S, Branco FS, Gruber B, Hauser AB, Pecóis-Filho R: Sevelamer decreases systemic inflammation in parallel to a reduction in endotoxemia. *Blood Purif* 2010, 29:352–356.
224. Navarro-Gonzalez JF, Mora-Fernandez C, Muros de Fuentes M, Donate-Correia J, Cazana-Perez V, Garcia-Perez J: Effect of phosphate binders on serum inflammatory profile, soluble CD14, and endotoxin levels in hemodialysis patients. *Clin J Am Soc Nephrol* 2011, 6:2272–2279.
225. Burger D, Travis S: Conventional medical management of inflammatory bowel disease. *Gastroenterology* 2011, 140:1827–1837. e1822.

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