



Additions to the Genus Arthrinium (Apiosporaceae) From Bamboos in China

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Feng Y, Liu J-K, Lin C-G, Chen Y-Y, Xiang M-M and Liu Z-Y (2021) Additions to the Genus Arthrinium (Apiosporaceae) From Bamboos in China. Front. Microbiol. 12:661281. doi: 10.3389/fmicb.2021.661281 *Arthrinium* has a widespread distribution occurring in various substrates (e.g., air, soil debris, plants, lichens, marine algae and even human tissues). It is characterized by the basauxic conidiogenesis in the asexual morph, with apiospores in the sexual morph. In this study, seventeen isolates of *Arthrinium* were collected in China. Based on their morphology and phylogenetic characterization, four new species (*A. biseriale, A. cyclobalanopsidis, A. gelatinosum, and A. septatum*) are described and seven known species (*A. arundinis, A. garethjonesii, A. guizhouense, A. hydei, A. neosubglobosa, A. phyllostachium* and *A. psedoparenchymaticum*) are identified, of which the sexual morph of three species (*A. guizhouense, A. phyllostachium* and *A. psedoparenchymaticum*) and asexual morph of *A. garethjonesii* are reported for the first time. The detailed descriptions, illustrations and comparisons with related taxa of these new collections are provided. Phylogenetic analyses of combined ITS, LSU, TUB2, and TEF sequence data support their placements in the genus *Arthrinium* and justify the new species establishments and identifications of known species.

Keywords: 4 new taxa, asexual-sexual morphs, multi-genes, phylogeny, taxonomy

INTRODUCTION

The genus *Arthrinium* Kunze belongs to the family Apiosporaceae, which was introduced by Hyde et al. (1998) and typified by the genus *Apiospora* (Ellis, 1971; Seifert et al., 2011; Hyde et al., 2020). *Arthrinium* is the largest genus within Apiosporaceae and it has a widespread distribution on variety of hosts, 77 species have been recorded by Species Fungrom (March, 2021). Classification of *Arthrinium* was primarily based on conidial shape, conidiophores, sterile cells, and the presence of setae. Some morphologically different taxa (genera) grouped with *Arthrinium* when use the molecular data (Crous and Groenewald, 2013). Thus, those characteristics could be not fully inferred about the phylogenetic relationships for *Arthrinium* (Crous and Groenewald, 2013; Wang et al., 2018; Pintos et al., 2019). Except for being reported as saprobes (Agut and Calvo, 2004; Crous and Groenewald, 2013; Dai et al., 2016, 2017; Jiang et al., 2018; Wang et al., 2018; Jiang et al., 2019, 2020; Luo et al., 2019; Pintos et al., 2019; Yan et al., 2019; Mapook et al., 2020; Senanayake et al., 2020a; Tang et al., 2020), the species of *Arthrinium* also includes phytopathogenic fungi, for instance, *A. arundinis* causing brown culm

streak of *Phyllostachys praecox, A. phaeospermum* causing culm rot on *Phyllostachys viridis* and cutaneous infections of humans (Rai, 1989; Zhao et al., 1990; Martínez-Cano et al., 1992; Mavragani et al., 2007; Crous and Groenewald, 2013; Li et al., 2016; Wang et al., 2018).

we are carrying out the survey of fungal diversity in Karst formations of the Asian region, and many new taxa are described in last few years (Li et al., 2016; Chen et al., 2017, 2020; Zhang et al., 2017, 2019; Feng et al., 2019; Liu et al., 2019; Dissanayake et al., 2020b). In this study, seventeen Arthrinium-like were collected in Guizhou and Guangdong province, China and can be recognized as eleven Arthrinium species based on morphological characters and phylogeny inferred from the multi-gene sequences data (ITS, LSU, TUB2, and TEF) analyses, which four new species (A. biseriale, A. cyclobalanopsidis, A. gelatinosum, and A. septatum) and seven known species (A. arundinis, A. garethjonesii, A. guizhouense, A. hydei, A. neosubglobosa, A. phyllostachium and A. psedoparenchymaticum) are introduced and identified, respectively. The aim of this study is to describe these new taxa with detailed descriptions and illustrations, and also provide their phylogenetic relationships within Arthrinium based on multi-gene analysis.

MATERIALS AND METHODS

Sample Collection, Morphological Studies and Isolation

Samples were collected from Guizhou and Guangdong Province in China. Fungal fruiting bodies were examined by using stereomicroscope (Motic SMZ 168). Free hand sections of fungal structures were mounted in water for microscopic studies and photomicrography. Images were taken by using a Nikon ECLIPSE Ni compound microscope fitted with a Canon EOS 70D digital camera. All measurements were taken by using Tarosoft Image Frame Work software (IFW) (Liu et al., 2010), and photo plates were processed with Adobe Photoshop CS6 software (Adobe Systems, United States).

The single spore isolation followed the method described in Senanayake et al. (2020b). Parts of morphological descriptions were based on sporulated cultures on WA (Water Agar) at room temperature (ca. 25°C). Type specimens were deposited in the herbarium of Cryptogams Kunming Institute of Botany Academia Sinica (HKAS), Kunming, China and Guizhou Academy of Agriculture sciences Herbarium (GZAAS). Pure cultures were deposited in China General Microbiological Culture Collection Center (CGMCC) and Guizhou Culture Collection (GZCC). Faces of Fungi¹ number is obtained as described in the paper by Jayasiri et al. (2015), and the new taxa are registered in Index Fungorum (2021).

DNA Extraction, PCR Amplification and Sequencing

Fungal mycelia were scraped from the pure culture which were growing on PDA (Potato Dextrose Agar) for one week at 25°C in

¹http://www.facesoffungi.org/

dark. DNA was extracted by using Ezup Column Fungi Genomic DNA Purification Kit (Sangon Biotech, China) from fresh fungal mycelia, but some were extracted directly from fruiting bodies by using Forensic DNA Kit (Omega Bio-Tek, China). Four gene regions, large subunit rDNA (LSU), internal transcribed spacer (ITS), beta-tubulin (TUB2) and the translation elongation factor 1-alpha (TEF) gene were amplified by the primer pairs LR0R and LR5 (Vilgalys and Hester, 1990), ITS5 and ITS4, T1 (O'Donnell and Cigelnik, 1997) and Bt2b (Glass and Donaldson, 1995), EF1-728F and EF-2 (O'Donnell et al., 1998; Carbone and Kohn, 1999), respectively. Polymerase chain reaction (PCR) was carried out in 25 μ L reaction volume containing 12.5 μ L 2 \times PCR Master Mix (Sangon Biotech, China), 9.5 µL ddH₂O, 1 µL of each primer and 1 µL DNA template. The annealing temperatures were adjusted to 56°C for ITS, LSU and TUB2, and 55°C for TEF. PCR products were sent to sequence at Sangon Biotech Co., Ltd., China. The PCR products were examined using 1.2% agarose electrophoresis gel stained with ethidium bromide. PCR products were purified and sequenced by Sangon Biotech (Shanghai) Co., Ltd., China. New generated nucleotide sequences were submitted in GenBank (Table 1).

Phylogenetic Analyses

Phylogenetic analyses were performed based on ITS, LSU, TUB2 and TEF sequence data. The related strains of Apiosporaceae (Table 1) used for analysis were referred to BLAST² results and relevant publications (Wang et al., 2018; Jiang et al., 2019, 2020; Pintos et al., 2019; Tang et al., 2020). Sequences were obtained from GenBank and aligned using MAFFT v. 7 (Katoh and Standley, 2013). Manual adjustment was also performed when it is necessary by using BioEdit v. 7.0 (Hall, 1999). The alignment of sequences data used in analyses is deposited in TreeBASE under the accession number S27728. The analyses of maximum parsimony (MP), maximum likelihood (ML) and Bayesian inference (BI) were carried out as detailed in Dissanayake et al. (2020a) and programs used including PAUP v.4.0b 10 (Hillis and Bull, 1993; Swofford, 2002), raxmlGUI v. 1.3 (Silvestro and Michalak, 2012), and MrBayes v3.1.2 (Rannala and Yang, 1996; Huelsenbeck and Ronquist, 2001; Zhaxybayeva and Gogarten, 2002; Nylander, 2004). Trees were visualized with FigTree v1.4.2 (Rambaut, 2012) and the layout was edited using Adobe Illustrator CS6.

RESULTS

Phylogeny

To determine the phylogenetic placement of the new collections in this study, the combined ITS, LSU, TUB2 and TEF data set comprised 119 taxa with *Seiridium phylicae* (CPC 19962 and CPC 19965) as the outgroup taxa. The concatenated alignment comprises 2,770 characters (ITS: 1–635; LSU: 636–1,454; TUB2: 1,455–2,300; TEF: 2,301–2,770) including gaps, of which 1,361 characters were constant, and 1,279 characters are parsimony informative and 130 are parsimony uninformative. Maximum

²https://blast.ncbi.nlm.nih.gov/Blast.cgi

TABLE 1 | GenBank accession numbers of species included in the phylogenetic study.

Species	Strain no.	GenBank Accession Numbers				
		LSU	ITS	TEF	TUB2	
Arthrinium acutiapicum	KUMCC 20-0209	MT946338	MT946342	MT947359	MT947365	
A. acutiapicum	KUMCC 20-0210	MT946339	MT946343	MT947360	MT947366	
A. aquaticum	MLFU 18-1628	MK835806	MK828608	N/A	N/A	
A. arundinis	CBS 106.12	KF144927	KF144883	KF145015	KF144973	
A. arundinis	CBS 114316	KF144928	KF144884	KF145016	KF144974	
A. arundinis	GZCC 20-0116	MW478899	MW481720	MW522952	MW522968	
A. aureum	CBS 244.83	KF144935	AB220251	KF145023	KF144981	
A. balearicum	CBS 145129	MK014836	MK014869	MK017946	MK017975	
A. bambusae	LC7107	N/A	KY494719	KY705117	KY705187	
A. bambusae	LC7106	KY494794	KY494718	KY806204	KY705186	
A. bambusicola	MFLUCC 20-0144	MW173087	MW173030	MW183262	N/A	
A. biseriale	CGMCC 3.20135	MW478885	MW481708	MW522938	MW522955	
A. biseriale	GZCC 20-0099	MW478886	MW481709	MW522939	MW522956	
A. biseriale	GZCC 20-0100	MW478887	MW481710	MW522940	MW522957	
A. camellia-sinensis	LC8181	KY494837	KY494761	KY705157	KY705229	
A. camellia-sinensis	LC5007	KY494780	KY494704	KY705103	KY705173	
A. caricicola	CBS 145127	MK014838	MK014871	MK017948	MK017977	
A. chinense	CFCC53036	N/A	MK819291	MK818545	MK818547	
A. chinense	CFCC53037	N/A	MK819292	MK818546	MK818548	
A. chromolaenae	MFLUCC 17-1505	MT214436	MT214342	N/A	N/A	
A. cyclobalanopsidis	CGMCC 3.20136	MW478892	MW481713	MW522945	MW522962	
A. cyclobalanopsidis	GZCC 20-0103	MW478893	MW481714	MW522946	MW522963	
A. descalsii	CBS 145130	MK014837	MK014870	MK017947	MK017976	
A. dichotomanthi	LC8175	KY494831	KY494755	KY705151	kY705223	
A. dichotomanthi	LC4950	KY494773	KY494697	KY705096	KY705167	
A. esporlense	CBS 145136	MK014845	MK014878	MK017954	MK017983	
A. euphorbiae	IMI 285638b	AB220335	AB220241	N/A	AB220288	
A. gaoyouense	CFCC 52301	N/A	MH197124	MH236793	MH236789	
A. gaoyouense	CFCC 52302	N/A	MH197125	MH236794	MH236790	
A. garethjonesii	KUMCC16-0202	KY356091	KY356086	N/A	N/A	
A. garethjonesii	GZCC 20-0115	MW478894	MW481715	MW522947	N/A	
A. gelatinosum	KHAS 11962	MW478888	MW481706	MW522941	MW522958	
A. gelatinosum	GZAAS 20-0107	MW478889	MW481707	MW522942	MW522959	
A. guizhouense	LC5318	KY494784	KY494708	KY705107	KY705177	
A. guizhouense	LC5322	KY494785	KY494709	KY705108	KY705178	
A. guizhouense	GZCC 20-0114	MW478895	MW481716	MW522948	MW522964	
A. gutiae	CBS 135835	KR149063	KR011352	KR011351	KR011350	
A. hispanicum	IMI 326877	AB220336	AB220242	N/A	AB220289	
A. hydei	CBS 114990	KF144936	KF144890	KF145024	KF144982	
A. hydei	KUMCC 16-0204	KY356092	KY356087	N/A	N/A	
A. hydei	GZCC 20-0113	MW478900	MW481721	MW522953	N/A	
A. hyphopodii	KUMCC 16-0201	KY356093	KY356088	N/A	N/A	
A. hyphopodii	MFLUCC15-0003	N/A	KR069110	N/A	N/A	
A. hysterinum	CBS 145134	MK014843	MK014876	N/A	N/A	
A. hysterinum	ICMP 6889	MK014841	MK014874	MK017951	MK017980	
A. ibericum	CBS 145137	MK014846	MK014879	MK017955	MK017984	
A. italicum	CBS 145138	MK014847	MK014880	MK017956	MK017985	
A. japonicum	IFO 30500	AB220356	AB220262	N/A	AB220309	
A. japonicum	IFO 31098	AB220358	AB220264	N/A	AB220311	
A. jatrophae	MMI 00052	N/A	JQ246355	N/A	N/A	
A. jatrophae	AMH-9557	N/A	JQ246355	N/A	N/A	

(Continued)

TABLE 1 | Continued

Species	Strain no.	GenBank Accession Numbers			
		LSU	ITS	TEF	TUB2
A. jiangxiense	LC4577	KY494769	KY494693	KY705092	KY705163
A. jiangxiense	LC4578	KY494770	KY494694	KY705093	KY705164
A. kogelbergense	CBS 113332	KF144937	KF144891	KF145025	KF144983
A. kogelbergense	CBS 113333	KF144938	KF144892	KF145026	KF144984
A. locuta-pollinis	LC11683	N/A	MF939595	MF939616	MF939622
A. longistromum	MFLUCC 11-0481	KU863129	KU940141	N/A	N/A
A. longistromum	MFLUCC 11-0479	KU863130	KU940142	N/A	N/A
A. malaysianum	CBS 102053	KF144942	KF144896	KF145030	KF144988
A. malaysianum	CBS 251.29	KF144943	KF144897	KF145031	KF144989
A. marii	CBS 497.90	KF144947	AB220252	KF145035	KF144993
A. mediterranei	IMI 326875	AB220337	AB220243	N/A	AB220290
A. minus	CBS 145131	MK014839	MK014872	MK017949	MK017978
A. mytilomorphum	DAOM 214595	N/A	KY494685	N/A	N/A
A. garethjonesii	KUMCC 18-0192	MK070898	MK070897	N/A	N/A
A. neosubglobosa	KUMCC 16-0203	KY356095	KY356090	N/A	N/A
A. neosubglobosa	JHB006	KY356094	KY356089	N/A	N/A
A. neosubglobosa	GZAAS 20-0099	MW478901	MW481705	MW522954	MW522969
A. obovatum	LC8177	KY494833	KY494757	KY705153	KY705225
A. obovatum	LC4940	KY494772	KY494696	KY705095	KY705166
A. ovatum	CBS 115042	KF144950	KF144903	KF145037	KF144995
A, paraphaeospermum	MFLUCC 13-0644	KX822124	KX822128	N/A	N/A
A. phaeospermum	CBS 114314	KF144951	KF144904	KF145038	KF144996
A. phaeospermum	CBS 114315	KF144952	KF144905	KF145039	KF144997
A. phragmites	CPC 18900	KF144956	KF144909	KF145043	KF145001
A. phyllostachium	MFLUCC 18-1101	MH368077	MK351842	MK340918	MK291949
A. phyllostachium	GZCC 20-0111	MW478896	MW481717	MW522949	MW522965
A. phyllostachium	GZCC 20-0112	MW478897	MW481718	MW522950	MW522966
A. piptatheri	CBS 145149	MK014860	MK014893	MK017969	N/A
A. pseudoparenchvmaticum	LC7234	KY494819	KY494743	KY705139	KY705211
A. pseudoparenchymaticum	LC8173	KY494829	KY494753	KY705149	KY705221
A. pseudoparenchymaticum	GZCC 20-0117	MW478898	MW481719	MW522951	MW522967
A. pseudorasikravindrae	KUMCC 20-0208	N/A	MT946344	MT947361	MT947367
A. pseudorasikravindrae	KUMCC 20-0211	N/A	MT946345	MT947362	MT947368
A. pseudosinense	CPC 21546	KF144957	KF144910	KF145044	N/A
A. pseudosinense	CBS 135459	N/A	KF144910	KF145044	N/A
A. pseudospegazzinii	CBS 102052	KF144958	KF144911	KF145045	KF145002
A. pterospermum	CBS 123185	KF144959	KF144912	N/A	KF145003
A. pterospermum	CPC 20193	KF144960	KF144913	KF145046	KF145004
A. puccinioides	CBS 145150	MK014861	MK014894	MK017970	MK017998
A. puccinioides	CBS 549.86	AB220347	AB220253	N/A	AB220300
A, ainlinaense	CFCC 52303	N/A	MH197120	MH236795	MH236791
A. ginlingense	CECC 52304	N/A	MH197121	MH236796	MH236792
A. rasikravindrae	CBS 145152	MK014863	MK014896	MK017971	MK017999
A. rasikravindrae	LC7115	KY494797	KY494721	KY705118	KY705189
A rasikravindrae	NECCI 2144	N/A	JE326454	N/A	N/A
A sacchari	CBS 212 30	KF144963	KF144917	KF145048	KF145006
A. sacchari	CBS 301 49	KF144962	KF144916	KF145047	KF145005
A. saccharicola	CBS 191 73	KF144966	KF144920	KF145051	KF145009
A. saccharicola	CBS 463 83	KF144968	KF144921	KF145053	KF145011
A. septatum	CGMCC 3.20134	MW478890	MW481711	MW522943	MW522960
A. septatum	GZCC 20-0109	MW478891	MW481712	MW522944	MW522961

(Continued)

TABLE 1 | Continued

Species	Strain no.	GenBank Accession Numbers				
		LSU	ITS	TEF	TUB2	
A. serenense	IMI 326869	AB220344	AB220250	N/A	AB220297	
A. setostromum	KUMCC 19-0217	MN528011	MN528012	MN527357	N/A	
A. sporophleum	CBS 145154	MK014865	MK014898	MK017973	MK018001	
A. subglobosum	MFLUCC 11-0397	KR069113	KR069112	N/A	N/A	
A. subroseum	LC7292	KY494828	KY494752	KY705148	KY705220	
A. thailandicum	MFLUCC 15-0199	KX986111	KU940146	N/A	N/A	
A. thailandicum	MFLUCC 15-0202	KU863133	KU940145	N/A	N/A	
A. trachycarpum	CFCC 53038	N/A	MK301098	MK303396	MK303394	
A. trachycarpum	CFCC 53039	N/A	MK301099	MK303397	MK303395	
A. urticae	IMI 326344	AB220339	AB220245	N/A	N/A	
A. vietnamese	IMI 99670	KX986111	KX986096	N/A	KY019466	
A. xenocordella	CBS478.86	KF144970	KF144925	KF145055	KF145013	
A. xenocordella	CBS 595.66	KF144971	KF144926	N/A	N/A	
A. yunnanum	DDQ 00281	KU863136	KU940148	N/A	N/A	
A. yunnanum	MFLU 15-0002	N/A	KU940147	N/A	N/A	
Seiridium phylicae	CPC 19962	NG 042759	LT853092	LT853189	LT853239	
Seiridium phylicae	CPC 19965	KC005809	LT853093	LT853190	LT853240	

The newly generated sequence is shown in bold. AMH: Ajrekar Mycological herbarium, Pune, Maharashtra, India; CBS: Westerdijk Fungal Biodiversity Institute, Utrecht, Netherlands; CFCC: China Forestry Culture Collection Center, Beijing, China; CGMCC: China General Microbiological Culture Collection Center, Beijing, China; CPC: Culture collection of Pedro Crous, housed at the Westerdijk Fungal Biodiversity Institute; DAOM: Canadian Collection of Fungal Cultures, Ottawa, Canada; DDQ: D.Q. Dai; GZAAS: Guizhou Academy of Agricultural Sciences herbarium, China; GZCC: Guizhou Culture Collection, China; HKAS: Herbarium of Cryptogams Kunming Institute of Botany Academia Sinica; ICMP: International Collection of Microorganisms from Plants, New Zealand; IFO: Institute for Fermentation, Osaka, Japan; IMI: Culture collection of CABI Europe UK Centre, Egham, UK; JHB: H.B. Jiang; KUMCC: Culture Collection of Kunming Institute of Botany, Yunnan, China; LC: Working collection of Lei Cai, housed at CAS, China; MFLUCC: Mae Fah Luang University Culture Collection, Chinag Rai, Thailand; NFCCI: National Fungal Culture Collection of India.

likelihood, maximum parsimony and Bayesian analyses were performed, respectively, and presented consistent topologies. The best scoring RAxML tree (**Figure 1**) is obtained with a final likelihood value of -27544.434044. Estimated base frequencies were as follows: A = 0.236829, C = 0.253015, G = 0.251523, T = 0.258634; substitution rates AC = 1.154254, AG = 2.738844, AT = 1.073582, CG = 0.896658, CT = 4.134885, GT = 1.000000; The gamma distribution shape parameter alpha is equal to 0.332641 and the Tree-Length equal to 3.795226.

Phylogenetic analyses showed that our newly collected seventeen taxa clustered into eleven clades and can be recognized as seven known species (*Arthrinium arundinis*, *A. garethjonesii*, *A. guizhouense*, *A. hydei*, *A. neosubglobosa*, *A. phyllostachium*, *A. psedoparenchymaticum*) and four new species (*A. biseriale*, *A. cyclobalanopsidis*, *A. gelatinosum*, *A. septatum*) (Figure 1).

Taxonomy

Arthrinium biseriale Y. Feng and Z.Y. Liu, sp. nov. Figure 2

Index Fungorum number: IF558136

Facesoffungi number: FoF 09569

Etymology: The epithet refers to the ascospores are arranged in two rows in the ascus.

Holotype: HKAS 111961

Saprobic on dead bamboo culms, forming black, lenticular spots on the host surface, with stromata breaking through raised cracks with black center, and merge with each other with age, forming an erumpent black mass visible at the naked

eye. Sexual morph: Stromata scattered to gregarious, immersed to erumpent, later becoming superficial, dark brown to black, fusiform, forming a slit-like opening at the apex, multi-loculate, membranous, with a periphysate ostiole. Ascomata 122-153 µm high, 138–207 μ m diam, arranged in rows, dark brown to black. Peridium 11-19 µm wide, composed of several layers of dark brown to hyaline cells of textura angularis. Hamathecium 3-4 µm wide, comprising dense, hyaline, septa paraphyses. Asci 84–116 μ m × 18–25 μ m (\bar{x} = 97 μ m × 21 μ m, n = 20), 8spored, unitunicate, clavate, apically rounded, with an indistinct pedicel. Ascospores 22–28 μ m \times 7–11 μ m (\bar{x} = 25 μ m \times 9 μ m, n = 30), biseriate, fusiform, curved at the bottom, obtuse at both ends, slightly wider in the middle, hyaline, 1-septate, constricted at the septum, mostly curved at the lower cell, rarely straight, with a large upper cell and a small lower cell, smoothwalled. The lower cell has 1-guttulate, the upper cell has 1-3 big guttulate in the middle surrounded by multiple small guttulate with a shallow 4–7 μ m thick gelatinous sheath in the early. Growing to a later stage, the guttulate filled the entire spore, and the gelatinous sheath dissolves easily. Asexual morph: On WA, Hyphae 2.5-6.0 µm diam, hyaline, branched, septate, some curled in a ring structure. Conidiophores 12.0-44.0 μ m × 2.5–5.0 μ m (\bar{x} = 20.0 μ m × 3.5 μ m, n = 20), straight or flexuous, smooth, thin-walled, unbranched, hyaline to pale brown, cylindrical, cyathiform, having transverse septa, often reduced to conidiogenous cells. Conidiogenous cells 5.0-22.0 μ m × 2.5–5.0 μ m (\bar{x} = 10.0 μ m × 3.5 μ m, n = 20), integrated, hyaline to pale brown, doliiform to ampulliform, or lageniform.

Conidia 7–9 μ m long (n = 30), brown, smooth in surface view, and 7–11 μ m long (n = 30), lenticular, with a paler equatorial slit in side view, globose to ellipsoid with many guttules.

Culture characters

Ascospores germinated on WA within 24 h and germ tubes produced from middle and lower end. Colonies fast





FIGURE 1 | RAXML tree of representatives based on a combined dataset of ITS, LSU, TUB2 and TEF sequences. Bootstrap support values for ML, MP (\geq 75%) and Bayesian (\geq 0.95) are given at the nodes (ML/MP/BI). Branches with ML, MP and BI equal 100, 100 and 1 are in bold. The tree is rooted with *Seiridium phylicae* (CPC 19962 and CPC 19965). New strains are shown in red.



FIGURE 2 | *Arthrinium biseriale* (HKAS 111961, holotype). (A) Appearance of stromata on bamboo host. (B) Vertical section of stroma. (C) Peridium. (D) Paraphyses. (E) Germinating ascospore. (F–H) Asci. (I–K) Ascospore. (L) Culture. (M) Hyphae. (N,O) Conidiophore and conidiogenous cells. (P–T) Conidia. Scale bars: (B) = 50 μm. (C) = 20 μm. (D) = 10μm. (E–H) = 20 μm. (I–K) = 10 μm. (M) = 20 μm. (N) = 10 μm. (O) = 20 μm. (P–T) = 5 μm.

grown on PDA at 25° C, reached 7 cm in 7 days at 25° C. Colonies evenly tiled, with a large number of aerial hyphae, white, velvety, thin, gray-white on the reverse side and dirty white in the center.

Materials examined

China, Guizhou Province, Chishui City, Zhuhai National Forest Park, on dead culms of bamboo, 10 July 2019, Yao Feng, CS19-17 (HKAS 111961, holotype; GZAAS 20–0102, isotype), ex-type living cultures, CGMCC 3.20135 = GZCC 20–0101. *Ibid.*, on dead base of the bamboo stem, 10 July 2019, Yao Feng, CS 19-25 (GZAAS 20–0101), living culture, GZCC 20–0100. *Ibid.*, on dead branch of bamboo, 11 July 2019, Ya-Ya Chen, CS 013 (GZAAS 20–0100), living culture, GZCC 20–099.

Notes

Three strains representing *Arthrinium biseriale* clustered in a well-supported clade which are closely related to *A. gelatinosum*, but phylogenetically distinct and can be recognized as two different species (99% sequence similarity in ITS; 99% in TEF; 98% in TUB2). Morphologically, *Arthrinium biseriale* has smaller stromata (122–153 μ m × 138–207 μ m vs. 144–199





 $\mu m \times$ 184–214 $\mu m)$ and the spores of A. biseriale are more curved than those of A. gelatinosum.

Arthrinium gelatinosum Y. Feng and Z.Y. Liu, sp. nov. Figure 3

Index Fungorum number: IF558137

Facesoffungi number: FoF 09570

Etymology: The epithet refers to the ascospore surrounded by gelatinous sheath.

Holotype: HKAS 111962

Saprobic on dead bamboo culms, forming black, lenticular spots on the host surface, with stromata breaking through raised cracks with black center. Sexual morph: Stromata solitary to gregarious, immersed to erumpent, fusiform, with long axis broken at the top by one cracks. Ascomata 144–199 μ m high μ m × 184–214 μ m wide, uniseriate or irregularly arranged beneath stromata, pseudothecial, black, globose to subglobose with a flattened base. Peridium composed of 5 or 6 layers of

brown cells arranged in *textura angularis*, with a conspicuous perfusate ostiole. *Hamathecium paraphyses* hyphae-like. *Asci* 85–121 μ m × 15–24 μ m ($\bar{x} = 100 \mu$ m × 17 μ m, n = 20), 8-spored, unitunicate, clavate, apically rounded, broadly cylindrical, with an indistinct pedicel. *Ascospores* (27–) 28-31 (–32) μ m × 6–8 μ m ($\bar{x} = 30 \mu$ m × 7 μ m, n = 30), apiosporic, clavate to fusiform with narrowly rounded ends, composed of a large guttate and small guttate, hyaline, smooth-walled, surrounded by a gelatinous sheath. Growing to a later stage, the guttate filled the entire spore, and the gelatinous sheath dissolves easily. **Asexual morph:** undetermined.

Materials examined

China, Guizhou Province, Chishui City, Zhuhai National Forest Park, on dead culms of bamboo, 10 July 2019, Yao Feng, CS19-32 (HKAS 111962, holotype; GZAAS20–0108, isotype). *Ibid.*, Chishui National Scenic Area, on dead branch of bamboo, 10 July 2019, Yao Feng, CS 19–29 (GZAAS 20–0107).

Notes

Two taxa representing *Arthrinium gelatinosum* cluster in a wellsupported lineage (ML/MP/BI = 93/98/1, **Figure 1**), which is a sister to *A. biseriale*, and they are phylogenetically distinct species.

Arthrinium septatum Y. Feng and Jian K. Liu, sp. nov. Figure 4

Index Fungorum number: IF558138

Facesoffungi number: FoF 09571

Etymology: The epithet refers to the septate conidiophore.

Holotype: HKAS 111960

Saprobic on dead bamboo culms. Sexual morph: Stromata scattered to gregarious, immersed to erumpent, initially breaks through a black spot on the host, later visible as black, raised, lenticular or dome-shaped, and will grow into a linear shape at the later stage of growth. Ostiolate, with the long axis broken at the top revealing the ostioles of pseudothecia. Ascomata 88-195 µm high × 160–185 µm wide ($\bar{x} = 140 \text{ µm} \times 173 \text{ µm}$, n = 10), arranged in rows, brown to dark brown, subglobose with a flattened base. Peridium with several layers of cells arranged in textura angularis, with a conspicuous ostiole 50-90 µm in diameter, periphysate. Hamathecium paraphyses hyphae-like, septate, hyaline. Asci 75–104 μ m \times 17–26 μ m (\bar{x} = 91 μ m \times 20 μ m, *n* = 20), 8-spored, clavate, cylindrical, apically rounded, with distinct pedicel. Ascospores (24–) 25–30 (–32) μ m × (6–) 8– 10 (-11) μ m (\bar{x} = 29 μ m × 9 μ m, n = 30), biseriate, broad fusiform to cylindrical, with a large upper cell and a small lower cell, hyaline, 1-septate, constricted at septum, slightly curved, smooth-walled, with many guttules, with a large guttule at the center of large upper cell, with a distinct gelatinous sheath. Asexual morph: On PDA, *Hyphae* 2–4 µm in diameter, hyaline, branched, septate. Conidiophores 12.0-63.0 \times 2-5 (\bar{x} = 313 μ m × 3 μ m, *n* = 20), straight or flexuous, smooth, thin-walled, septate, hyaline to light brown, cylindrical, sometimes reduced to conidiogenous cells. Conidiogenous cells 4.0–18.0 \times 1.5–4.0 $(\bar{x} = 10.0 \ \mu m \times 2.5 \ \mu m, n = 20)$, solitary on hyphae, integrated, branched, ampuliform, cylindrical, hyaline to brown. Conidia 8-11 (-13) μ m long (*n* = 30), brown, smooth, guttulate, globose to ellipsoid in surface view. and (8-) 9-13 (-14) $\mu m \log$ (n = 30), lenticular with a paler equatorial slit in side view. Sterile cells (13–) 14–21 (–27) μ m × 5–7 (–9) μ m elongated, mixed among conidia.

Culture characteristics

Ascospores germinated on WA within 24 h. Colonies on PDA reached 8 cm in 7 days at 25 $^{\circ}$ C, flat, aerial mycelium white. The hyphae in the center are cottony, dense, and there is a thin circle of hyphae at the edge. Reverse grayish white with a dirty white patch.

Materials examined

China, Guizhou Province, Chishui City, Zhuhai National Forest Park, on dead culms of bamboo, 10 July 2019, Yao Feng, CS19-8 (HKAS 111960, holotype; GZAAS 20–0109, isotype), ex-type living culture CGMCC 3.20134 = GZCC20–0108. *Ibid.*, on dead branch of bamboo, 11 July 2019, Ya-Ya Chen, CS 025 (GZAAS 20–0111), living culture GZCC 20–0109.

Notes

Two isolates, representing *Arthrinium septatum*, grouped in a well-supported clade and appear to be distinct from other *Arthrinium* species phylogenetically (**Figure 1**). *Arthrinium septatum* resembles to *A. biseriale* in having biseriate, broad fusiform to cylindrical ascospores and cylindrical, clavate asci. However, *Arthrinium septatum* differs from *A. biseriale* by having smaller stromata (160–185 µm diam vs. 138–207 µm diam) and asci (75–104 × 17–26 µm vs. 84–116 µm × 18–25 µm).

Arthrinium cyclobalanopsidis Y. Feng and Jian K. Liu, sp. nov. Figure 5

Index Fungorum number: IF558139

Facesoffungi number: FoF 09572

Etymology: The epithet "cyclobalanopsidis" refers to the host plant, *Cyclobalanopsidis glauca* (Thunb.) Oerst.

Holotype: HKAS 111963

Saprobic on Cyclobalanopsidis glauca (Thunb.). Sexual morph: Undetermined. Asexual morph: On PDA, Hyphae 2.5–5.5 μ m in diameter, hyaline, septate, branched with chain structure. Conidiophores reduced to the conidiogenous cells. Conidiogenous cells 6.0–19.0 μ m × 2.5–7.0 μ m ($\bar{x} = 11.0 \ \mu$ m × 4.5 μ m, n = 20), aggregated in clusters on hypha, pale brown, ampuliform or cylindrical. Conidia 8–12 μ m long (n = 30), brown, smooth, globose to ellipsoid in surface view, and 10–14 μ m long (n = 30), lenticular, with a paler equatorial slit in side view. Sterile cells elongated, rolled up, sometimes mixed among conidia.

Culture characteristics

Conidia germinated on WA within 12 h. Sporulated on PDA, Colonies flat, margin circular, fluffy, sparse, white, with dirty white patches in center, reverse white, with sparse aerial mycelium, reached 8 cm in 7 days at 25°C.

Material examined

China, Guizhou Province, Qianxinan Buyi and Miao Autonomous Prefecture, Ceheng County, on Leaf of *cyclobalanopsidis glauca* (Thunb.) Oerst., 13 May 2018, Yao Feng, G81 (HKAS 111963, holotype; GZAAS 20–0096, isotype),



FIGURE 4 | *Arthrinium septatum* (HKAS 111960, holotype). (A) Appearance of stromata on bamboo host. (B) Vertical section of stroma. (C) Peridium. (D) Germinating ascospore. (E) Paraphyses. (F,G) Culture. (H–J) Asci. (K–P) Ascospore. (Q) Colony on PDA producing conidia masses. (R–T) Conidiophore and conidiogenous cells. (U–X) Conidia. (Y,Z) Sterile cell. Scale bars: (B) = 50 μm. (C,D) = 25 μm. (E) = 10 μm. (H–J) = 20 μm. (K–P) = 10 μm. (U–Z) = 5 μm.

ex-type living cultures, CGMCC 3.20136 = GZCC 20–0102. *Ibid.*, 20 Oct. 2019, Yao Feng, G82 (GZAAS 20–0097), living culture GZCC 20–0103.

Notes

Two isolates, representing *Arthrinium cyclobalanopsidis*, cluster together with *A. camelliae-sinensis* which was introduced by Wang et al. (2018) from *Camellia sinensis* (Figure 1). *Arthrinium cyclobalanopsidis* can be distinguished from *A. camelliae-sinensis* (567/572 in ITS; 390/414 in TEF; 715/748 in TUB2). Morphologically, *Arthrinium cyclobalanopsidis* resembles to

A. camelliae-sinensis in having similar conidia (8–12 μ m × 10–14 μ m vs. 9.0–13.5 μ m × 7.0–12.0 μ m), but can be distinguished by its relatively longer conidiogenous cells (6.0–19.0 μ m vs. 4.0–9.5 μ m).

Arthrinium garethjonesii D.Q. Dai and H.B. Jiang, Mycosphere 7 (9): 1337 (2017). Figure 6

Saprobic on dead bamboo branch. **Sexual morph**: See Dai et al. (2016). **Asexual morph**: *Sporodochia* on host with hair-like setae, also grow in the gaps of the perithecia and scatter on the surface of the perithecia, black. *Conidiophores* reduced to conidiogenous



FIGURE 5 | Arthrinium cyclobalanopsidis (HKAS 111963, holotype) (A) Conidia masses on host. (B) Culture. (C) Sterile cell mixed among conidia. (D) Hyphae of chain structure. (E–H) Conidiogenous cells and conidia. (I–L) Conidia. Scale bars: (C–H) = 10 µm. (I–L) = 5 µm.

cells. *Conidiogenous cells* (5–) 6–19 (–20) μ m × (2–) 3–5 (–7) μ m (\bar{x} = 11 μ m × 4 μ m, *n* = 20), aggregated in black sporodochia, hyaline to pale brown, smooth, ampulliform. *Conidia* (14–) 16–19 (–20) μ m diam, brown, smooth, granular, globose to subglobose in surface view, and (16–) 17–22 (–23) μ m diam, with pale equatorial slit in side view.

Culture characteristics

Conidia germinated on WA within 12 h, colonies fast growing on PDA, reached 8 cm in 7days at 25°C, fluffy, circular, dense, raised at center, white, reverse reddish. Hyphae 2–4 μ m diam, hyaline to pale brown, branched, septate.

Material examined

China, Guizhou Province, Chishui City, Zhuhai National Forest Park, on dead branches of bamboo, 10 July 2019, Yao Feng, CS19-9 (GZAAS 20–0117); living culture GZCC 20–0115.

Notes

Arthrinium garethjonesii was originally described by Dai et al. (2016) based on the sexual morph from dead bamboo culms (HKAS 96289) collected from Yunnan Province, China. Our phylogenetic result (**Figure 1**) indicates that our collection is identical to Arthrinium garethjonesii and we report its asexual morph for the first time in this study.



Arthrinium guizhouense M. Wang and L. Cai, Mycokeys 34: 13 (2018). Figure 7

Saprobic on dead bamboo culms, forming black, lenticular spots on the host surface, stromata breaking through raised cracks with black center. **Sexual morph:** Stromata solitary to gregarious, immersed to erumpent, fusiform, with long axis broken at the top by one cracks. Ascomata 188–220 μ m high \times 170–200 μ m wide, uniseriate or irregularly

arranged beneath stromata, pseudothecial, black, globose to subglobose. *Peridium* composed of 5 or 6 layers of brown cells arranged in *textura angularis*, *Hamathecium paraphyses* $3-5 \ \mu\text{m}$, hyaline, hyphae-like, septate. *Asci* (80–) 94–106 (–107) $\ \mu\text{m} \times (20-) \ 21-23 \ (-24) \ \mu\text{m}$, 8-spored, clavate, apically rounded, broadly cylindrical, with an indistinct pedicel. *Ascospores* (24–) 25–32 (–33) $\ \mu\text{m} \times (6-) \ 7-9 \ (-10) \ \mu\text{m}$ ($\bar{x} = 31 \ \mu\text{m} \times 8 \ \mu\text{m}$, n = 30), apiosporic, clavate to fusiform



Conidia. (W) Sterile cell. Scale bars: (B,C) = 50 μm. (D-F) = 30 μm. (G) = 50 μm. (J-O) = 20 μm. (Q,R) = 10 μm. (S-W) = 5 μm.

with narrowly rounded ends, composed of a large upper cell and small lower cell, hyaline, smooth-walled, surrounded an inconspicuous gelatinous sheath. **Asexual morph:** *Hyphae* 2.5– 7.5 μ m diam, hyaline, branched, septate. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* 4–12 μ m \times 2– 5 μ m ($\bar{x} = 8 \ \mu$ m \times 3 μ m, n = 20), erect, aggregated in clusters on hyphae, pale brown, smooth, subglobose, ampulliform or doliiform. *Conidia* 5–8 μ m long (n = 30), dark brown to black, smooth, globose or subglobose, and 6-8 μ m long (n = 30), lenticular, with a paler equatorial slit in side view. *Sterile cells* elongated, rolled up, sometimes mixed among conidia.

Culture characteristics

On PDA, colonies very fast, reached 8 cm in 8 days at 25° C, velvety, circular, with regular edge, middle densely and raised white, dense at above the from margin, aerial mycelia, surface initially white, became grayish and reverse white.

Material examined

China, Guangdong Province, Guangzhou City, on decaying bamboo culms, 03 Sep. 2019, Yao Feng, GZ 23 (GZAAS 20–0114); living culture GZCC 20–0114.

Notes

Arthrinium guizhouense was introduced from air in a karst cave (asexual morph was provided from the culture) in Guizhou province, China by Wang et al. (2018). In this study, one collection was found as saprobe on bamboo in Guangzhou, China and it is identified as *A. guizhouense* based on the phylogeny and morphology evidences. In addition, our new collection provides the sexual morph which only the asexual morph was illustrated by Wang et al. (2018) and Senanayake et al. (2020a).

Arthrinium phyllostachium C.L. Yang, X.L. Xu and K.D. Hyde, Phytotaxa 406 (2): 102 (2019). Figure 8

Saprobic on dead bamboo culms, Sexual morph: Stromata scattered to gregarious, immersed to erumpent, later becoming superficial, dark brown to black, fusiform, forming a slit-like opening at the apex, with stromata breaking through raised cracks with black center. Ascomata 135-185 µm high, 157-215 µm diam, multi-loculate, with a periphysate ostiole, arranged in rows, clustered, gregarious, ampulliform, dark brown to black. Peridium 20-25 µm wide, composed of several layers of dark brown to brown cells of textura angularis. Hamathecium 3-5 µm wide, comprising dense, hyaline, septate paraphyses, hyphaelike. Asci 66–98 × 17–27 ($\bar{x} = 85 \ \mu m \times 21 \ \mu m$, n = 20), 8-spored, unitunicate, clavate, apedicellate, apically rounded. Ascospores 29–34 μ m \times 7–10 μ m (\bar{x} = 32 μ m \times 9 μ m, n = 30), biseriate, elliptical, hyaline, 1-septate, constricted at the septum, mostly curved at the lower cell, rarely straight, with a large upper cell and a small lower cell, smoothwalled, with a shallow gelatinous sheath. Asexual morph: On WA, Hyphae 1.5-4.0 µm in diameter, hyaline, septate, branched. Conidiophores reduced to the conidiogenous cells. Conidiogenous cells (8–) 9–28 (–31.5) $\mu m \times (1.5-)$ 2–4 (-6) μm ($\bar{x} = 20 \ \mu m \times 3 \ \mu m$, n = 20), aggregated in clusters on hypha or solitary, erect, ampuliform or cylindrical, arising holoblastically from vegetative hyphae, monoblastic, or polyblastic, sympodial, terminal, cylindrical to clavate, ampulliform, hyaline, smooth, thin-walled. Conidia 5-8 µm long (n = 30), brown, smooth, globose to ellipsoid in surface view, and 7–9 μ m long (n = 30), lenticular, with a paler equatorial slit in side view. Sterile cells light brown, elongated, and occasionally irregularly angled.

Culture characters

Ascospores germinated on WA within 24 h and germ tubes produced from Upper, middle and lower end. Colonies fast growed on PDA at 25°C, flat, spreading, the middle is white, smooth, the edge surface tapetum, gray-white, indistinct aerial hyphae, the reverse side is yellowish in the middle, and the edge is grayish white.

Materials examined

China, Guizhou Province, Chishui City, Zhuhai National Forest Park, on dead culms of bamboo, 10 July 2019, Yao Feng, CS 19-23 (GZAAS 20–0112), living culture GZCC 20–0111; *Ibid.*, Chishui National Scenic Area, on dead culms of bamboo, 10 July 2019, Ya-Ya Chen, CS004 (GZAAS 20–0113), living culture GZCC 20–0112.

Notes

Arthrinium phyllostachium was introduced by Yang et al. (2019) based on the asexual morph and phylogeny analyses. It was collected from culms of *Phyllostachys* heteroclada (Poaceae) in China (Yang et al., 2019). The phylogenetic results showed that our new collections are identical to *A. phyllostachium*, Yang et al. (2019) only provided the asexual morph and the sexual morph is given in this study.

Arthrinium pseudoparenchymaticum M. Wang and L. Cai, Mycokeys 34 (1): 17 (2018), Figure 9

Saprobic on dead bamboo culms, forming black, lenticular spots on the host surface, with stromata breaking through raised cracks with black center. Sexual morph: Ascomata 187-242 μ m high \times 242–373 μ m wide, uniseriate, or irregularly arranged beneath stromata, black, globose to subglobose with a flattened base. Peridium composed of 5-7 layers of brown cells arranged in *textura angularis*, with a conspicuous perfusate ostiole. Hamathecium paraphyses 4-8 µm, hyphae-like, septa hyaline. Asci (95-) 107-110 (-133) µm × 23-25 (-27) µm, 8-spored, broadly cylindrical, clavate or subglobose, apically rounded, with an indistinct pedicel. Ascospores (35-) 35-43 (-44) × (10-) 11-13 μ m ($\bar{x} = 41 \times 12$, n = 30), apiosporic, clavate to fusiform with narrowly rounded ends, composed of a large upper cell and small lower cell, hyaline, smooth-walled, surrounded by a gelatinous sheath. Asexual morph: On WA, Hyphae 1.5-4 µm diam, hyaline to pale brown, branched, septate. Conidiophore extends from the vegetative hypha, up to 60 µm long. Conidiogenous cells 10.0-40.0 μ m \times 3.0–6.0 μ m, scattered in clusters on hyphae, smooth, unbranched, hyaline to pale yellow, smooth, erect, subcylindrical. *Conidia* 17–27 μ m × 17–21 μ m (\bar{x} = 23 μ m × 19 μ m, n = 30), pale to light brown, smooth, globose to subglobose, sometimes lobed or dentate, polygonal or irregular in surface view.

Culture characteristics

Ascospores germinating on WA within 24 h and germ tubes produced from upper. Colonies fast growing on PDA at 25°C, under 12 h light/12 h dark, cottony, circular, sparse, raised, with irregular edge, white in center.

Material examined

China, Guangdong Province, Guangzhou City, on decaying bamboo culms, 3 Sep. 2019, Yao Feng, GZ18 (GZAAS 20–0115), living culture GZCC20–0117.

Notes

Arthrinium pseudoparenchymaticum was introduced by Wang et al. (2018) based on the asexual morph characters and phylogeny analyses. It was originally collected from bamboo in China (Wang et al., 2018). In this study, a fresh specimen was collected and it



FIGURE 8 | Arthrinium phyllostachium (GZAAS 20–0112) (A) Appearance of stromata on bamboo host. (B) Vertical section of stroma. (C) Peridium.
(D) Germinating ascospore. (E) Paraphyses. (F–I) Asci. (J–M) Ascospore. (N,O) Culture. (P–S) Conidiogenous cells and conidia. Scale bars: (B) = 50 μm. (C) = 10 μm. (D) = 20 μm. (E) = 20 μm. (F–I) = 30 μm. (J–M) = 10 μm. (P,Q) = 20 μm. (R–S) = 10 μm.

is identical to *A. pseudoparenchymaticum* (Figure 1), both sexual and asexual morphs were described and illustrated (Figure 9).

Arthrinium arundinis (Corda) Dyko and B. Sutton, Mycotaxon 8: 119 (1979). Figure 10.

Saprobic on dead bamboo culms. Asexual morph: On PDA, Hyphae 2–3 μ m diam, consisting of smooth, hyaline, branched, septate. Conidiophores reduced to conidiogenous cells. Conidiogenous cells 3–8 μ m \times 2–5 μ m, aggregated in clusters on hyphae, pale brown,

smooth, ampulliform. *Conidia* (4–) 5–6 (–7) μ m, brown, smooth, globose in surface view, and (4–) 5–7 (–8) μ m diam, lenticular with pale equatorial slit in side view. *Sterile cells* at times intermingled among conidia. **Sexual morph:** Undetermined.

Culture characteristics

The colony is flat, cotton-like, thick and dense, with sparse aerial mycelia. The surface of PDA is white and the reverse side is grayish white.



(D) Paraphyses. (E) Peridium. (F) Germinating ascopere. (G) Culture. (H–J) Asci. (K–O) Ascospore. (P) Ascospore in Indian ink and present clear gelatinous sheath. (Q) Colony on WA producing conidia masses. (R) Dentate conidia. (S–U) Conidiogenous cells giving rise to conidia. Scale bars: (C) = 50 μm. (D–F) = 20 μm. (H–J) = 30 μm. (K–P) = 15 μm. (R–U) = 15 μm.

Materials examined

China, Guizhou Province, Chishui City, Zhuhai National Forest Park, on dead culms of bamboo, 10 July 2019, Yao Feng, CS 19-7 (GZAAS 20–0116), living culture GZCC 20–0116.

Notes

Our collection clusters together with the isolates of *Arthrinium arundinis* (Figure 1) and its morphology lines up with the type species. Therefore we identify it as *Arthrinium arundinis*.

Arthrinium hydei Crous, IMA Fungus 4 (1): 142 (2013). Figure 11

Saprobic on bamboo leaves. Asexual morph: Colonies on the host punctiform, pulvinate, blackish brown. Conidiophores pale brown, smooth, transversely septate, subcylindrical. Conidiogenous cells $5-15 \times 3-6 \mu m$, brown, smooth, subcylindrical to doliiform to lageniform. Conidia (13–) 14–19 (–20) μm diam in surface view, brown, roughened, globose, and 15–20 μm diam,



FIGURE 10 | Arthrinium arundinis (GZAAS 20–0116) (A,B) culture. (C) Colonies on culture. (D) Sterile cell mixed among conidia. (E–J) Conidiogenous cells and conidia. (K–N) Conidia. Scale bars: (D) = 50 µm. (E) = 5 µm. (F) = 10 µm. (G–N) = 5 µm.

lenticular with pale equatorial slit in side view. Sexual morph: Undetermined.

Culture characteristics

Colonies flat, spreading, with sparse aerial mycelium. On PDA surface and reverse pale luteous. Mycelium consisting of smooth, hyaline to pale brown, branched, septate, 2.0–4.5 μ m diam hyphae.

Materials examined

China, Guizhou Province, Guiyang City, Baihua Lake, on bamboo leaves, 20 April 2018, Yao Feng, 67 (GZAAS 20–0098), living culture GZCC20–0113.

Notes

This collection is identified as *Arthrinium hydei* based on both morphological characters and molecular data. Crous and Groenewald (2013) originally described *A. hydei*



Scale bars: (C) = 50 μ m. (D–M) = 5 μ m. (N) = 25 μ m.

based on the asexual morph from a culture (CBS 114990) which was isolated from bamboo culms in Hong Kong, China; we found this species in Guizhou from the substrate in nature with its asexual morph.

Arthrinium neosubglobosa D.Q. Dai and H.B. Jiang, Mycosphere 7 (9): 1337 (2017) Figure 12.

Saprobic on dead bamboo culms. Sexual morph: Stromata scattered to gregarious, superficial to raised, with a slit-like

opening, dark brown to black, naviculate, with black papillate ostiole, multi-loculate. *Ascomata* 205–328 μ m high, 168–345 μ m, perithecial, arranged in a row, immersed in stromata, later becoming erumpent through host surface to superficial, obpyriform to ampulliform, dark brown, membranous. Ostiole raised from center of *Ascomata*, internally lined with periphyses. *Peridium* 4 layers, outer layer composed of dark brown, cells of *textura prismatica*, inner layer thin, with hyaline cells of *textura angularis*. *Hamathecium* 4.0–5.5 μ m wide, comprising dense



paraphyses, indistinctly aseptate, unbranched, not an astomosing, filamentous, clustered embedded in gelatinous matrix. Asci 80–119 µm × 20–37 µm ($\bar{x} = 97$ µm × 28 µm, n = 20), 8-spored, unitunicate, clavate, with a short pedicel, apically rounded. Ascospores 25–36 µm × 11–15 µm ($\bar{x} = 29$ µm × 13 µm, n = 30), 2-seriate, elliptical, hyaline, 1-septate, constricted at the septum, mostly curved at the lower cell, rarely straight, with a large upper cell and a small lower cell, smoothwalled, 1-guttulate, with a shallow 8–12 µm thick gelatinous sheath. Asexual morph: Undetermined.

Materials examined

China, Guizhou Province, Zunyi City, Daozhen County, on dead culms of bamboo, 15 August 2018, Yao Feng, DZ22 (GZAAS 20–0099).

Notes

Arthrinium neosubglobosa was introduced by Dai et al. (2016) based on the sexual morph characters and phylogeny analyses. The pure culture was attempted by single spore isolation and the DNA was extracted directly from the fruiting body, the

new collection is identified as *A. neosubglobosa* based on the phylogeny (**Figure 1**) and morphology evidences.

DISCUSSION

Arthrinium species have been reported from many hosts, includes hive-stored pollen lichens, marine algae, soil debris, gut of insects and nodules of human skin (Sharma et al., 2014; Crous et al., 2015, Senanayake et al., 2015; Wijayawardene et al., 2017, Zhao et al., 2018), it can be concluded that *Arthrinium* is ecologically diverse. Bamboo, as one of the most reported host, is a gramineous plant integrating economy and ornamental value (Gratani et al., 2008; Kelchner and Bamboo Phylogeny Group, 2013; Dai et al., 2016, 2017; Jiang et al., 2018, 2019, 2020; Wang et al., 2018; Yang et al., 2019), there is more than 115 genera with approximately 1,450 species. According to incomplete statistics, more than 1,100 species of fungi on bamboo were reported (Hyde et al., 2002a,b; Dai et al., 2016, 2017; Senanayake et al., 2020a; Tang et al., 2020; Wijesinghe et al., 2020). It is of great significance to excavate and identify the fungi on bamboo.

Several studies have shown (Crous and Groenewald, 2013; Dai et al., 2016, Dai et al., 2017; Wang et al., 2018; Yang et al., 2019) that it is difficult to identify the Arthrinium species solely rely on morphology and the multi-gene phylogenetic analyses are needed in the identification and classification of Arthrinium. The morphology of conidia is variable which can be depending on the period of incubation on different habitats, for example, A. biseriale, A. gelatinosum and A. septatum, are very similar in morphology, but molecular data distinguish them into different species; Our collection A. pseudoparenchymaticum differs from the type specimen (LC 8173) in the morphology of conidiophores, the size of conidiogenous cells is also different, while the molecular data supported them as the same species. These results are in agreement with the previous observations and publications (Crous and Groenewald, 2013; Dai et al., 2016, Dai et al., 2017; Wang et al., 2018, Yang et al., 2019). In addition, as a high diverse group, it is also difficult to distinguish species within Arthrinium by only using ITS and LSU gene regions, and the protein genes (TEF and TUB2) are not available for many species in the genus which bring potential problem once the new or existing taxa are introduced and identified, respectively. For example the absent of the protein genes (TEF and TUB2) of Arthrinium garethjonesii would bring the troubles

REFERENCES

- Agut, M., and Calvo, M. A. (2004). In vitro conidial germination in Arthrinium aureum and Arthrinium phaeospermum. Mycopathologia 157, 363–367. doi: 10.1023/B:MYCO.0000030432.08860.f3
- Carbone, I., and Kohn, L. M. (1999). A method for designing primer sets for speciation studies in filamentous ascomycetes. *Mycologia* 91, 553–556. doi: 10.1080/00275514.1999.12061051
- Chen, Y. Y., Dissanayake, A., Liu, Z. Y., and Liu, J. K. (2020). Additions to Karst Fungi 4: Botryosphaeria spp. associated with woody hosts in Guizhou province, China including B. guttulata sp. nov. Phytotaxa 454, 186–202. doi: 10.11646/ phytotaxa.454.3.2

in identification of *Arthrinium setostromum* as it is hard to confirm whether they are same species or not as they are identical in ITS and LSU gene regions, as well as the close phylogenetic relationship (**Figure 1**). It would be necessary to provide protein genes when new taxa are introduced in these well-study and diverse groups.

DATA AVAILABILITY STATEMENT

The data presented in the study can be found in the Genbank. The accession numbers of the sequences deposited in GenBank are ITS: MW481705—MW481721; LSU: MW478885–MW478901; TEF:MW522938–MW522954; and TUB: MW522955–MW522969.

AUTHOR CONTRIBUTIONS

YF and J-KL: conceptualization. YF and Y-YC: methodology. YF, C-GL, and J-KL: formal analysis. YF, Y-YC, M-MX, and J-KL: resources. YF: writing—original draft preparation. C-GL, Z-YL, and J-KL writing—review and editing. Z-YL and J-KL: supervision. All authors approved to publish the version of final manuscript.

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- Chen, Y. Y., Maharachchikumbura, S. S. N., Liu, J. K., Hyde, K. D., Nanayakkara, R. R., Zhu, G. S., et al. (2017). Fungi from asian karst formations I. *Pestalotiopsis photinicola* sp.nov., causing leaf spots of *Photinia serrulata*. *Mycosphere* 8, 103–110. doi: 10.5943/mycosphere/8/1/9
- Crous, P. W., and Groenewald, J. Z. (2013). A phylogenetic re-evaluation of *Arthrinium. IMA Fungus* 4, 133–154. doi: 10.5598/imafungus.2013.04.01.13
- Crous, P. W., Wingfield, M. J., Le Roux, J. J., Richardson, D. M., Strasberg, D., Shivas, R. G., et al. (2015). Fungal planet description sheets: 371-399. *Persoonia* 35, 264–327. doi: 10.3767/003158515X690269
- Dai, D. Q., Jiang, H. B., Tang, L. Z., and Bhat, D. J. (2016). Two new species of *Arthrinium* (Apiosporaceae, Xylariales) associated with bamboo from Yunnan, China. *Mycosphere* 7, 1332–1345. doi: 10.5943/mycosphere/7/9/7

- Dai, D. Q., Phookamsak, R., Wijayawardene, N. N., Li, W. J., Bhat, D. J., Xu, J. C., et al. (2017). Bambusicolous fungi. Fung. Divers. 82, 1–105.
- Dissanayake, A. J., Bhunjun, C. S., Maharachchikumbura, S. S. N., and Liu, J. K. (2020a). Applied aspects of methods to infer phylogenetic relationships amongst fungi. *Mycosphere* 11, 2652–2676. doi: 10.5943/mycosphere/ 11/1/18
- Dissanayake, A. J., Chen, Y. Y., and Liu, J. K. (2020b). Unravelling diaporthe species associated with woody hosts from karst formations (Guizhou) in China. *J. Fungi* 6:251. doi: 10.3390/jof6040251
- Ellis, M. B. (1971). *Dematiaceous Hyphomycetes*. Kew: Commonwealth Mycological Institute, 608.
- Feng, Y., Zhang, S. N., and Liu, Z. Y. (2019). Tremateia murispora sp. nov. (Didymosphaeriaceae, Pleosporales) from Guizhou, China. Phytotaxa 416, 79– 87. doi: 10.11646/phytotaxa.416.1.10
- Glass, N. L., and Donaldson, G. C. (1995). Development ofprimer sets designed for use with the PCR to amplify conserved genes from filamentous ascomycetes. *Appl. Environ. Microbiol.* 61, 1323–1330. doi: 10.1128/aem.61.4.1323-1330. 1995
- Gratani, L., Crescente, M. F., Varone, L., Fabrini, G., and Digiulio, E. (2008). Growth pattern and photosynthetic activity of different bamboo species growing in the Botanical Garden of Rome. *Flora Morphol. Distrib. Funct. Ecol. Plants* 203, 77–84. doi: 10.1016/j.flora.2007.11.002
- Hall, T. A. (1999). "BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT," in *Nucleic Acids Symposium Series*. 95–98.
- Hillis, D. M., and Bull, J. J. (1993). An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. *Syst. Biol.* 42, 182–192. doi: 10.1093/sysbio/42.2.182
- Huelsenbeck, J. P., and Ronquist, F. (2001). MRBAYES: bayesian inference of phylogenetic trees. *Bioinformatics* 17, 754–755. doi: 10.1093/bioinformatics/17. 8.754
- Hyde, K. D., Fröhlich, J., and Taylor, J. E. (1998). Fungi from palms. XXXVI. Reflections on unitunicate ascomycetes with apiospores. Sydowia 50, 21–80.
- Hyde, K. D., Norphanphoun, C., Maharachchikumbura, S. S. N., Bhat, D. J., Jones, E. B. G., Bundhun, D., et al. (2020). Refined families of Sordariomycetes. *Mycosphere* 11, 305–1059. doi: 10.5943/mycosphere/11/1/7
- Hyde, K. D., Zhou, D., and Dalisayl, T. (2002a). Bambusicolous fungi: a review. *Fung. Divers.* 9, 1–14.
- Hyde, K. D., Zhou, D., McKenzie, E., Ho, W., and Dalisay, T. (2002b). Vertical distribution of saprobic fungi on bamboo culms. *Fungal Divers.* 11, 109–118.
- Jayasiri, C. S., Hyde, K. D., Ariyawansa, H. A., Bhat, J., Buyck, B., Cai, L., et al. (2015). The faces of fungi database: fungal names linked with morphology, phylogeny and human impacts. *Fungal Divers*. 74, 3–18. doi: 10.1007/s13225-015-0351-8
- Jiang, H. B., Hyde, K. D., Doilom, M., Karunarathna, S. C., Xu, J. C., and Phookamsak, R. (2019). Arthrinium setostromum (Apiosporaceae, Xylariales), a novel species associated with dead bamboo from Yunnan, China. Asia. J. Mycol. 2, 254–268. doi: 10.5943/ajom/2/1/16
- Jiang, N., Li, J., and Tian, C. M. (2018). Arthrinium species associated with bamboo and reed plants in China. Fungal Syst. Evol. 2, 1–9. doi: 10.3114/fuse.2018. 02.01
- Jiang, N., Liang, Y. M., and Tian, C. M. (2020). A novel bambusicolous fungus from China, Arthrinium chinense (Xylariales). Sydowia 72, 77–83. doi: 10.12905/ 0380
- Katoh, K., and Standley, D. M. (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol. Biol. Evol.* 30, 772–780. doi: 10.1093/molbev/mst010
- Kelchner, S. A., and Bamboo Phylogeny Group (2013). Higher level phylogenetic relationships within the bamboos (Poaceae: Bambusoideae) based on five plastid markers. *Mol. Phylogenet. Evol.* 67, 404–413. doi: 10.1016/j.ympev.2013. 02.005
- Li, B. J., Liu, P. Q., Jiang, Y., Weng, Q. Y., and Chen, Q. H. (2016). First report of culm rot caused by *Arthrinium phaeospermum* on *Phyllostachys viridis* in China. *Plant Dis.* 100, 1013–1013. doi: 10.1094/PDIS-08-15-0901-PDN
- Liu, J. K., Chomnunti, P., Cai, L., Phookamsak, R., Chukeatirote, E., Jones, E. B. G., et al. (2010). Phylogeny and morphology of *Neodeightonia palmicola* sp. nov. from palms. *Sydowia* 62, 261–276.

- Liu, N. G., Bhat, D. J., Hyde, K. D., and Liu, J. K. (2019). Conioscypha tenebrosa sp. nov. (Conioscyphaceae) from China and notes on Conioscypha species. *Phytotaxa* 413, 159–171. doi: 10.11646/phytotaxa.413.2.5
- Luo, Z., Hyde, K. D., Liu, J. K., Maharachchikumbura, S. S. N., Jeewon, R., Bao, D. F., et al. (2019). Freshwater Sordariomycetes. *Fungal Divers*. 99, 451–660. doi: 10.1007/s13225-019-00438-1
- Mapook, A., Hyde, K. D., McKenzie, E. H. C., Gareth, J. E. B., Bhat, D. J., Jeewon, R., et al. (2020). Taxonomic and phylogenetic contributions to fungi associated with the invasive weed *Chromolaena odorata* (Siam weed). *Fungal Divers*. 101, 1–175. doi: 10.1007/s13225-020-00444-8
- Martínez-Cano, C., Grey, W. E., and Sands, D. C. (1992). First report of Arthrinium arundinis causing kernel blight on barley. Plant Dis. 76, 1077. doi: 10.1094/PD-76-1077B
- Mavragani, D. C., Abdellatif, L., McConkey, B., Hamel, C., and Vujanovic, V. (2007). First report of damping-off of durum wheat caused by Arthrinium sacchari in the semi-arid Saskatchewan fields. Plant Dis. 91:469. doi: 10.1094/ pdis-91-4-0469a
- Nylander, J. A. A. (2004). *MrModeltest v2. Program Distributed by the Author*. Uppsala: Evolutionary Biology Centre, Uppsala University.
- O'Donnell, K., and Cigelnik, E. (1997). Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus Fusarium are nonorthologous. *Mol. Phylogenet. Evol.* 7, 103–116. doi: 10.1006/mpev.1996. 0376
- O'Donnell, K., Kistler, H. C., Cigelnik, E., and Ploetz, R. C. (1998). Multiple evolutionary origins of the fungus causing Panama disease of banana: concordant evidence from nuclear and mitochondrial gene genealogies. *Proc. Natl. Acad. Sci. U.S.A.* 95, 2044–2049. doi: 10.1073/pnas.95.5.2044
- Pintos, A., Alvarado, P., Planas, J., and Jarling, R. (2019). Six new species of *Arthrinium* from Europe and notes about *A. caricicola* and other species found in Carex spp. *Hosts. Mycokeys* 49, 15–48. doi: 10.3897/mycokeys.49.32115
- Rai, M. K. (1989). Mycosis in man due to Arthrinium phaeospermum var. indicum. First case report. Mycoses 32, 472–475. doi: 10.1111/j.1439-0507.1989.tb0 2285.x
- Rambaut, A. (2012).). FigTree, Version 1.4. 2. Edinburgh: University of Edinburgh.
- Rannala, B., and Yang, Z. (1996). Probability distribution of molecular evolutionary trees: a new method of phylogenetic inference. J. Mol. Evol. 43, 304–311. doi: 10.1007/BF02338839
- Seifert, K., Morgan-Jones, G., Gams, W., and Kendrick, B. (2011). The Genera of Hyphomycetes.[CBS Biodiversity Series 9]. Utrecht: CBS-KNAW Fungal Biodiversity Centre.
- Senanayake, I. C., Bhat, J. D., Cheewangkoon, R., and Xie, N. (2020a). Bambusicolous Arthrinium species in guangdong province. China. Front. Microbiol. 11:602773. doi: 10.3389/fmicb.2020.602773
- Senanayake, I. C., Maharachchikumbura, S. S., Hyde, K. D., Bhat, J. D., Jones, E. G., McKenzie, E. H., et al. (2015). Towards unraveling relationships in Xylariomycetidae (Sordariomycetes). *Fungal Divers*. 73, 73–144. doi: 10.1007/ s13225-015-0340-y
- Senanayake, I. C., Rathnayake, A. R., Marasinghe, D. S., Calabon, M. S., Gentekaki, E., Lee, H. B., et al. (2020b). Morphological approaches in studying fungi: collection, examination, isolation, sporulation and preservation. *Mycosphere* 11, 2678–2754. doi: 10.5943/mycosphere/11/1/20
- Sharma, R., Kulkarni, G., Sonawane, M. S., and Shouche, Y. S. (2014). A new endophytic species of *Arthrinium* (Apiosporaceae) from Jatropha podagrica. *Mycoscience* 55, 118–123. doi: 10.1016/j.myc.2013.06.004
- Silvestro, D., and Michalak, I. (2012). raxmlGUI: a graphical front-end for RAxML. *Org. Divers. Evol.* 12, 335–337. doi: 10.1007/s13127-011-0056-0
- Swofford, D. L. (2002). PAUP*: Phylogenetic Analysis Using Parsimony (*and Other Methods). Version 4. Sunderland, MA: Sinauer Associates.
- Tang, X., Goonasekara, I. D., Jayawardena, R. S., Jiang, H. B., Li, J. F., Hyde, K. D., et al. (2020). Arthrinium bambusicola (Fungi, Sordariomycetes), a new species from Schizostachyum brachycladum in northern Thailand. Biodivers. Data J. 8:e58755. doi: 10.3897/BDJ.8.e58755
- Vilgalys, R., and Hester, M. (1990). Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J. Bacteriol.* 172, 4238–4246. doi: 10.1128/jb.172.8.4238-4246. 1990
- Wang, M., Tan, X. M., Liu, F., and Cai, L. (2018). Eight new Arthrinium species from China. MycoKeys 34, 1–24. doi: 10.3897/mycokeys.34.24221

- Wijayawardene, N. N., Hyde, K. D., Rajeshkumar, K. C., Hawksworth, D. L., Madrid, H., Kirk, P. M., et al. (2017). Notes for genera: ascomycota. *Fungal Divers*. 86, 1–594. doi: 10.1007/s13225-017-0386-0
- Wijesinghe, S. N., Wang, Y., Camporesi, E., Wanasinghe, D. N., Boonmee, S., and Hyde, K. D. (2020). A new genus of Bambusicolaceae (Pleosporales) on *Corylus avellana* (Fagales) from Italy. *Biodivers. Data J.* 8:e55957. doi: 10.3897/BDJ.8. e55957
- Yan, H., Jiang, N., Liang, L. Y., Yang, Q., and Tian, C. M. (2019). Arthrinium trachycarpum sp. nov. from Trachycarpus fortunei in China. Phytotaxa 400, 203–210. doi: 10.11646/phytotaxa.400.3.7
- Yang, C. L., Xu, X. L., Dong, W., Wanasinghe, D. N., Liu, Y. G., and Hyde, K. D. (2019). Introducing Arthrinium phyllostachium sp. nov. (Apiosporaceae, Xylariales) on Phyllostachys heteroclada from Sichuan Province, China. Phytotaxa 406, 091–110. doi: 10.1127/nova_hedwigia/2019/ 0526
- Zhang, J. F., Liu, J. K., Hyde, K. D., Chen, Y. Y., Liu, Y. X., and Liu, Z. Y. (2017). Two new species of *Dyfrolomyces* (Dyfrolomycetaceae, Dothideomycetes) from karst landforms. *Phytotaxa* 313:267. doi: 10.11646/phytotaxa. 313.3.4
- Zhang, J. F., Liu, J. K., Jeewon, R., Wanasinghe, D. N., and Liu, Z. Y. (2019). Fungi from Asian Karst formations III. Molecular and morphological characterization

reveal new taxa in Phaeosphaeriaceae. *Mycosphere* 10, 202–220. doi: 10.5943/ mycosphere/10/1/3

- Zhao, Y. M., Deng, C. R., and Chen, X. (1990). Arthrinium phaeospermum causing dermatomycosis, a new record of China. Acta Mycol. Sin. 9, 232–235.
- Zhao, Y. Z., Zhang, Z. F., Cai, L., Peng, W. J., and Liu, F. (2018). Four new filamentous fungal species from newly-collected and hivestored bee pollen. *Mycosphere* 9, 1089–1116. doi: 10.5943/mycosphere/9/6/3
- Zhaxybayeva, O., and Gogarten, J. P. (2002). Bootstrap, Bayesian probability and maximum likelihood mapping: exploring new tools for comparative genome analyses. *BMC Genom.* 3:4. doi: 10.1186/1471-2164-3-4

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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