

Research Article

MPTP Neurotoxicity and Testosterone Induce Dendritic Remodeling of Striatal Medium Spiny Neurons in the C57Bl/6 Mouse

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Nigrostriatal damage is increased in males relative to females. While estrogen is neuroprotective in females, less is known about potential protective effects of testosterone in males. We determined if castration enhances neuronal injury to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). Castrated or sham-castrated mice were sacrificed 1 week following injection of MPTP (4×20 mg/kg) or saline ($n = 11-12$ /group). The right striatum was immunostained for tyrosine hydroxylase (TH). The left hemisphere was stained by Golgi Cox to quantify neuronal morphology in medium spiny neurons (MSNs) of the dorsolateral striatum. MPTP reduced TH, but there was no effect of castration and no interaction. For MSN dendritic morphology, MPTP decreased the highest branch order and increased spine density on 2nd-order dendrites. Castrated males had shorter 5th-order dendrites. However, there was no interaction between gonadal status and MPTP. Thus, castration and MPTP exert nonoverlapping effects on MSN morphology with castration acting on distal dendrites and MPTP acting proximally.

1. Introduction

Gonadal steroid hormones are potent modulators of neuronal survival and neuronal morphology [1]. In the adult, steroid hormones exert activational effects in steroid-responsive brain regions which include protective effects against neurodegeneration [2, 3]. However, sex differences exist in many neurodegenerative disorders, suggesting that the male and female brains are not equally responsive to gonadal steroids.

In this regard, Parkinson's disease (PD) is a common neurological disorder that demonstrates a substantial sex difference, with a one- to twofold higher incidence in men [4]. PD results from the progressive loss of dopaminergic neurons in the substantia nigra pars compacta (SNc; [5]). Dopaminergic efferents from SNc project rostrally as part of the nigrostriatal pathway to the dorsolateral striatum, where

they synapse onto medium spiny neurons (MSNs). These efferents are reduced in PD, leading to a depletion of striatal dopamine. Gonadal steroids modulate the function of the nigrostriatal system and are thought to contribute, in part, to this sex difference.

In females, estrogen promotes the function of the nigrostriatal system by enhancing striatal dopamine release, increasing dopamine metabolism and altering both dopamine receptors and uptake sites [6, 7]. In response to neurotoxic insult, estrogen is also neuroprotective in the nigrostriatal system [7]. This has been well demonstrated using 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and 6-hydroxydopamine (6-OHDA), neurotoxins which selectively deplete dopaminergic SNc neurons [8]. In both models, estrogen attenuates the loss of striatal dopamine and reduces the loss of SNc dopaminergic neurons [9–15].

In parallel to its effects in the female, estrogen also modulates nigrostriatal function in males [16, 17]. Estrogen in males is derived by the local aromatization of testosterone, although testosterone can also act as an androgen in the male brain. Whether testosterone has neuroprotective effects in the male nigrostriatal system, similar to the effects of estrogen in the female, is not well-established. After MPTP in castrated mice, striatal dopamine loss is attenuated by estrogen but not by testosterone [18, 19].

Previous studies investigating the effects of testosterone on nigrostriatal function in male rats and mice after 6-OHDA or MPTP have measured dopamine content and release [18–21], dopaminergic striatal input [21], and dopamine transporter binding [19, 21]. It is unknown whether testosterone reduces the loss of dopaminergic innervation to the striatum after MPTP in mice. Ultimately, the effects of MPTP lesion may also extend beyond the SNC dopamine neurons themselves. Loss of dopaminergic input may remodel MSN morphology, and testosterone has potential to attenuate this MPTP-induced deafferentation. The current study used castrated and gonad-intact adult male mice to determine whether testosterone reduces MPTP-induced deficits in striatal dopamine neurochemistry and MSN neuronal architecture.

2. Materials and Methods

2.1. Animals. Forty-five C57BL/6 adult male mice (8–10 weeks old) were obtained from Charles River Laboratories (Wilmington, MA). Mice were group-housed on a 12:12 LD photoperiod with access to food and water *ad libitum*. Experimental procedures were approved by USC's Institutional Animal Care and Use Committee and conducted in accordance with the NIH Guide for the Care and Use of Laboratory Animals (DHEW Publication 80-23, revised 1985, Office of Science and Health reports, DRR/NIH, Bethesda, MD).

Initially, half of the mice ($n = 23$) were castrated (OrchX) via a midline scrotal incision. The other half ($n = 22$) received sham castrations. As measured by androgen-sensitive seminal vesicle weight, castration was effective (187.5 ± 14.2 mg in sham males versus 5.6 ± 1.4 mg in castrated males, $P < .05$). Two weeks later, half of the mice in each group received MPTP and half were given saline. MPTP (Sigma, St. Louis, MO) was dissolved in 0.9% saline and was administered in 4 injections of 20 mg/kg (free-base).ip. with an interinjection interval of 2 hours. Control mice received equivalent injections of 0.1 mL saline. This lesioning paradigm is a well-established method that leads to ca. 67% loss of nigrostriatal neurons and 90–95% depletion of striatal DA, as reported in previous studies from our laboratories [22, 23].

One week following MPTP, animals were sacrificed via intracardiac perfusion. This duration is sufficient for MPTP-induced cell death to occur [22]. Mice were deeply anesthetized with sodium pentobarbital (150 mg/kg BW) and perfused intracardially with 150 mL of 0.1 M sodium phosphate buffer (PB, pH = 7.4) containing 0.9% NaCl and

0.1% NaNO_3 . The brains were removed and hemisected. To allow us to obtain TH and neuronal morphologic measures in the same animals, the right hemisphere for each brain was processed for TH immunocytochemistry and the left hemisphere was processed for Golgi-Cox staining. We are unaware of any evidence of laterality in striatal damage after ip. MPTP injections in the mouse.

2.2. Tyrosine Hydroxylase Immunocytochemistry. The right hemispheres from each brain were postfixed in 4% paraformaldehyde in PB overnight at 4°C, then cryoprotected for 5 days at 4°C with 20% sucrose in PB. Hemispheres were rapidly frozen and sectioned coronally at 25 μm thickness through the rostrocaudal extent of the striatum. Sections were stored in PB with 0.01% sodium azide at 4°C until processed for TH immunocytochemistry.

Sections through the striatum at or rostral to the anterior commissure corresponding to Plates 18–28 of Paxinos and Franklin [24] were stained for TH. Tissue from mice in different groups was stained at the same time. Sections were incubated overnight at room temperature (RT) in polyclonal rabbit anti-TH antibody (1:5000; Chemicon, Temecula, CA) with 4% normal donkey serum and 0.3% Triton X-100 in PB. The following day, sections were incubated in biotinylated donkey antirabbit secondary antibody (1:200; Jackson Immunoresearch, West Grove, PA) and the avidin-biotin-horseradish peroxidase complex (Vector Elite Kit; Vector Laboratories, Burlingame, CA), each for 1 hour at RT with extensive washes in between. TH-labeled cells were visualized using NiCl-enhanced 3',3'-diaminobenzidine tetrahydrochloride with 0.25% hydrogen peroxide. Sections were mounted onto gelatin-coated slides, dehydrated in alcohols, cleared in xylenes, and coverslipped with Permount.

The relative expression of TH immunoreactivity was measured in dorsolateral striatum on coded slides by an observer blind to the treatment group. To ensure that differences in staining intensity were due to differences in antigen expression, multiple sections from each of the different treatment groups were handled concurrently in identical staining conditions. Control experiments excluding either primary or secondary antibody were also carried out to verify staining specificity. Three striatal sections rostral to the anterior commissure (Bregma 0.25–1.25 in [24]) were sampled per animal ($n = 8$ –9 animals/group) using methods previously described by our laboratory [25, 26]. Briefly, striatal sections were digitally photographed at low magnification. The dorsolateral quadrant of each striatal section was outlined, and TH immunostaining was measured in a 1.6 mm² circular region of interest at the dorsolateral boundary of this quadrant (Figure 1). In previous studies [25, 26], this region shows the largest decrease in TH immunostaining after MPTP. The relative optical density (expressed as arbitrary units within the linear range of detection) was determined by subtracting the relative optical density of the corpus callosum as background. This measurement reflects both the area and intensity of TH immunostaining within the striatum. To ensure that the gray values represented an optical density within the nonsaturated

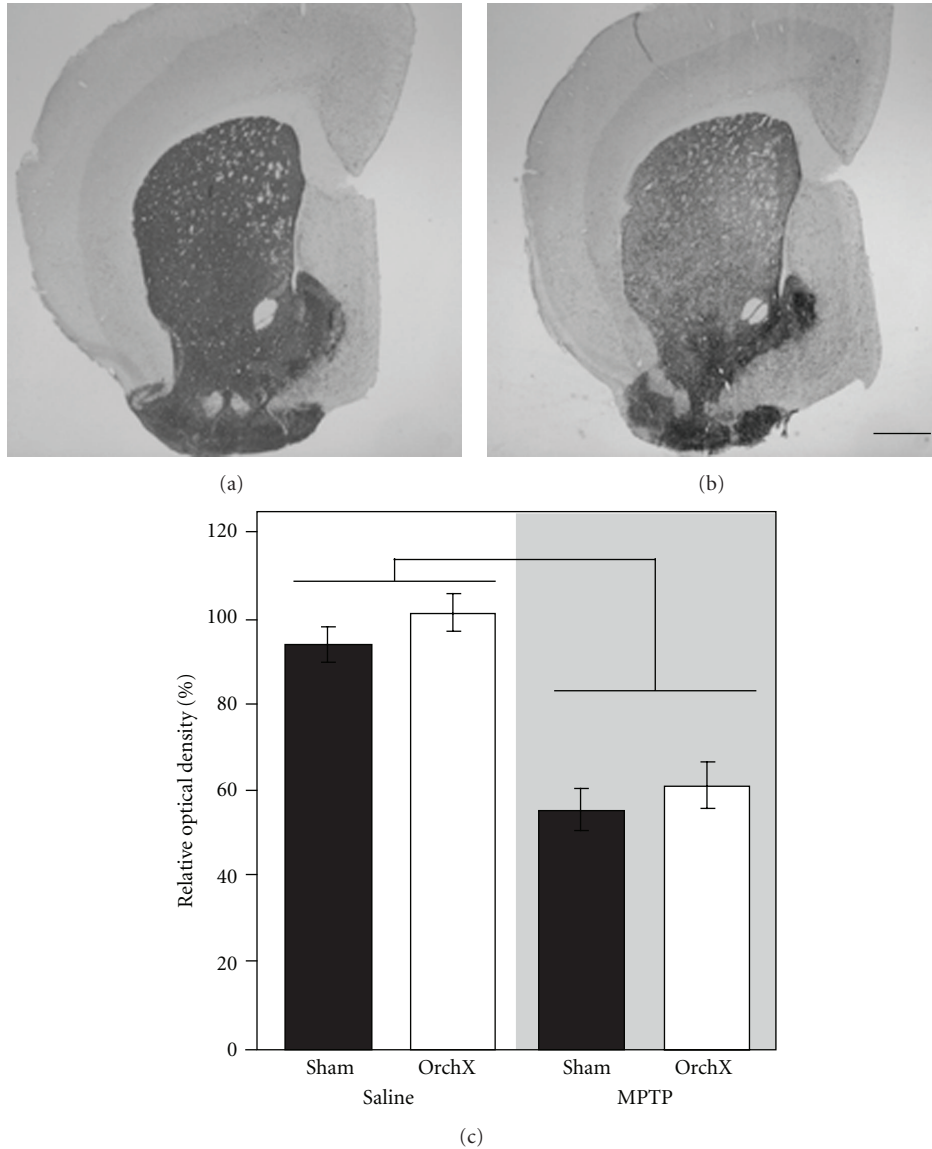


FIGURE 1: TH staining in the dorsolateral striatum. Photomicrographs of TH staining in gonad-intact saline-injected (a) and MPTP-treated (b) mice. Scale bar = 500 μm . (c) The density of TH staining in the dorsolateral striatum of saline-injected (clear) and MPTP-treated (shaded) gonad-intact (black bars) and orchidectomized (white bars) male mice ($n = 8-10$ mice/group). The bar represents an effect of MPTP ($P < .05$). OrchX: orchidectomized; Sham: sham-orchidectomized.

range of the image analysis, a Kodak photographic step tablet (density range to 255 OD units) captured by the CCD camera was used. Maximal tissue immunostaining relative OD units did not exceed the relative OD units of the tablet.

2.3. Golgi-Cox Staining. Golgi-Cox staining was performed on the left hemisphere of each brain using the FD Rapid GolgiStain Kit (FD NeuroTechnologies, Ellicott City, MD). Pilot studies using Golgi staining (according to Gomez and Newman [27]), Golgi-cox staining (according to Gibb and Kolb [28]), and the rapid Golgi kit (FD NeuroTechnologies) were conducted to determine an optimal way to visualize neuronal morphology in our striatal tissue. The rapid Golgi

kit provided the most complete staining of medium spiny neurons.

The hemispheres were placed in Golgi-Cox solution containing mercuric chloride, potassium dichromate, and potassium chromate for 2 weeks, and the solution was replaced after the first 24 hours. The brains were moved to a cryoprotection solution (GolgiStain Kit) for 48 hours and then sectioned coronally at 200 μm on a vibratome (Vibratome Series 1000). Sections through the rostral-caudal extent of the striatum were mounted on gelatin-coated slides. Slides were stored in a humidity chamber overnight and developed the following day according to the Rapid GolgiStain Kit protocol. Briefly, slides were rinsed in distilled water and placed in a developing solution for 10 minutes.

Immediately afterwards, the slides were rinsed, dehydrated in alcohols, cleared in xylenes, and coverslipped with Cytoseal-60 mounting medium (Richard-Allan Scientific). Slides were stored in the dark at RT until morphological analysis.

MSN morphology was analyzed on coded slides by an observer blinded to the treatment groups using a Nikon Eclipse 80i microscope (Nikon Instruments, Inc., Melville, NY) with motorized stage and MicroFire camera (Olympus America, Inc., Center Valley, PA). To compare dendritic morphology after castration and MPTP lesion, the entire dendritic tree from one primary dendrite was traced under a 100x oil immersion lens using the Neuron Tracing function in NeuroLucida (MicroBrightField, Inc., Williston, VT). Brains with well-differentiated Golgi-Cox labeling from 5 mice in each experimental group were selected for morphologic analysis; 5 neurons from each mouse were analyzed. Morphologic data from the 5 neurons/mouse were averaged to provide a single data point for each animal used in statistical comparison ($n = 5/\text{group}$). MSNs selected for analysis were located in the dorsolateral quadrant of the striatum at or rostral to the level of the anterior commissure (Plates 18–28 of [24]). Selected MSNs were fully impregnated with Golgi stain and had clearly visible spines with minimal or absent obstruction by neighboring Golgi-stained cells or blood vessels. Morphometric analysis was conducted using NeuroExplorer software (MicroBrightField, Inc.). Briefly, each dendritic segment was assigned a branch order with the dendritic segment proximal to the soma identified as the first branch order. Dendritic lengths, number of spines, and spine density were computed for each branch order. All dendrites subject to morphologic analysis had at least 3 branch orders. However, because not all dendrites had 4th- and 5th-order branches, the variability in dendritic length increased at higher branch orders. In addition, total spine density and total dendrite length were calculated for the entire dendritic tree. Due to the relative lack of spines on primary dendrites (typically, 1 or 2 spines/primary dendrite), branch order analysis was not performed on first-order dendrites.

2.4. Statistics. For comparison of both TH and Golgi-Cox labeling, morphologic data from each mouse were averaged to provide a single data point used in statistical comparison. Group differences for the 5 animals in each group were analyzed by two-factor (gonadal status and lesion) analysis of variance (ANOVA). Post hoc comparisons using the Fisher's LSD test were conducted when statistically significant differences ($P < .05$) were found.

3. Results

3.1. Striatal TH. Striatal TH immunocytochemistry was used here as a measure of lesion damage in gonad-intact and castrated males. Damage to midbrain DA neurons causes a loss of striatal DA terminals, which produces equivalent changes in striatal levels of presynaptic dopamine transporter and TH [23, 29, 30]. Similar to previous reports by our lab and others [25, 26, 31, 32], MPTP decreased TH immunoreactivity in the striatum (Figures 1(a) and 1(b), $n =$

8–10/group). Striatal TH was reduced by 40% after MPTP (mean gray level 59.4 ± 3.7 versus 98.5 ± 3.1 in unlesioned mice, $F(1, 31) = 64.883$, $P < .05$, Figure 1(c)). However, there was no effect of castration on TH immunoreactivity and no interaction between MPTP and castration ($P > .05$, Figure 1(c)). This finding parallels previous studies which have found no effect of castration on the number of TH-positive neurons in SNC [33].

3.2. Medium Spiny Neuron Morphology

3.2.1. Spine Density. MSNs have elaborate dendritic arbors with a high density of dendritic spines (Figure 2). In saline-treated sham-castrate controls, spines were largely absent from primary dendrites (1.2 ± 0.2 spines/ $10 \mu\text{m}$), but increased on more distal dendrites (5th-order dendrites 6.8 ± 0.4 spines/ $10 \mu\text{m}$). The density of dendritic spines in the present study ($n = 5/\text{group}$) is comparable to that reported previously in mice [34–36]. When combining castrated and gonad-intact mice, MPTP produced a modest but significant increase in total spine density (spines/ $10 \mu\text{m}$) on MSNs (6.9 ± 0.1 versus 6.3 ± 0.1 in saline-injected castrate and intact mice, $F(1, 16) = 10.22$, $P < .05$, Figure 3(a)). When analyzed according to branch order, the increase in spine density was restricted to proximal dendrites (Figure 4(a)). Specifically, MPTP-treated mice had a higher spine density on 2nd-order (5.3 ± 0.3) and 3rd-order (7.1 ± 0.2) dendrites, compared with 4.2 ± 0.2 and 6.5 ± 0.2 in saline-injected mice, respectively ($F(1, 16) = 9.659$ and 5.700 , $P < .05$, Figure 4(a)). However, there was no effect of castration on spine density and no interaction ($P > .05$, Figures 3(a) and 4(a)).

3.2.2. Branch Order. MPTP significantly decreased the average highest branch order (3.9 ± 0.1) compared to saline-injected males (4.3 ± 0.1 , $F(1, 16) = 4.595$, $P < .05$, Figure 3(b)). As with other measures of overall neuronal morphology, castration was without effect and there was no interaction (Figure 3(b)).

3.2.3. Dendritic Length. There was no effect of MPTP on total dendrite length ($402.0 \pm 17.4 \mu\text{m}$ versus $415.5 \pm 20.2 \mu\text{m}$ in saline-injected mice, $P > .05$, Figure 3(c)). However, in parallel to the increase in spine density, we observed a selective increase in dendritic length after MPTP on 2nd-order dendrites ($80.2 \pm 6.8 \mu\text{m}$ versus $61.6 \pm 4.6 \mu\text{m}$ in saline-injected mice, $F(1, 16) = 4.796$, $P < .05$, Figure 4(b)). Castration had no effect on total dendritic length. However, castrated mice had a prominent reduction in dendritic length of distal dendrites. Specifically, 5th-order dendritic length was significantly reduced in castrated mice ($78.9 \pm 14.8 \mu\text{m}$) compared with gonad-intact males ($138.3 \pm 13.2 \mu\text{m}$, $F(1, 16) = 6.659$, $P < .05$, Figure 4(b)). There was no interaction between MPTP treatment and gonadectomy (Figures 3(c) and 4(b)).

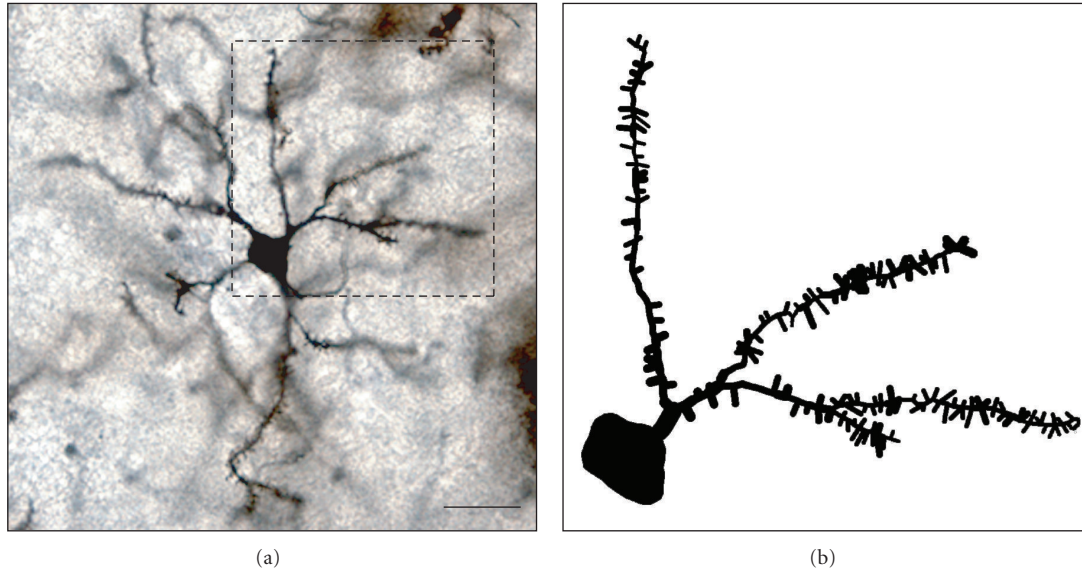


FIGURE 2: Golgi stained MSNs. Photomicrograph of a representative Golgi-impregnated medium spiny neuron with high-magnification inserts of a primary dendritic branch and fifth-order dendritic branch (a) and corresponding neuroLucida tracing (b). Scale bar = 10 μm .

4. Discussion

The current study used the MPTP mouse model of PD to investigate the effects of castration on dopamine-depleting lesions of the nigrostriatal system. MPTP decreased striatal TH immunoreactivity, reduced the average highest branch order on MSNs, and increased proximal spine density. Separately, castration reduced dendritic length of distal dendrites. We predicted that testosterone would act as a neuroprotectant to attenuate the effects of MPTP and that castrated mice would have increased nigrostriatal damage after MPTP compared with gonad-intact mice. However, there was no interaction between gonadal hormone status and MPTP, suggesting that testosterone does not attenuate the neurotoxic effects of MPTP in the nigrostriatal system of males.

The current study used a well-established lesioning protocol [22, 23] that produced a moderate lesion, as measured by TH immunostaining. This is relevant to the study of how gonadal steroid hormones act on the nigrostriatal system because the neuroprotective effects of gonadal hormones are likely to be evident earlier in PD. For example, PD symptom severity is sexually dimorphic in early stages of the disease, with women experiencing less severe motor impairments [4, 37, 38]. This has been attributed, in part, to the neuroprotective effects of estrogen on the nigrostriatal system. In later stages of PD, sex differences are not reported, presumably because severe nigrostriatal degeneration obscures the effects of neuroprotective factors, including estrogen. This is paralleled in animal studies, where moderate lesions do not overwhelm the potential for gonadal hormones to attenuate the nigrostriatal response to MPTP. In 6-OHDA-lesioned rats, Gillies et al. [21] have demonstrated sex differences with small doses of 6-OHDA that disappear

with larger doses. Using MPTP, our laboratory has demonstrated sex differences in motor impairments after relatively small lesions [39]. Even so, in the current study, castration did not alter the morphologic response to MPTP. Larger lesions should produce a more dramatic depletion of TH, but seem unlikely to reveal an interaction with castration.

Nonetheless, our results do demonstrate a broad impact of MPTP on basal ganglia circuitry. Although MPTP selectively kills dopamine-producing neurons of SNC, its effects are not limited to dopaminergic neurons themselves. In fact, morphological effects of MPTP were observed on efferent targets in the striatum. Specifically, MPTP increased dendritic spine density on proximal dendrites of striatal MSNs. This result was initially surprising because loss of dopaminergic input to MSNs is expected to reduce spine density, at least as demonstrated *in vitro* [34, 40]. Importantly, the increased spine density observed in our study was restricted to proximal dendrites. Proximal MSN dendrites receive synaptic inputs from within the striatum, while distal dendrites receive extrinsic inputs from the cortex and SNC [41]. This suggests that the increase in proximal dendritic spine density after MPTP is driven by intrinsic striatal neurons, rather than dopaminergic neurons of SNC. This may reflect a compensatory reaction to dopamine depletion and demonstrates the dynamics of spine morphology with degree of lesion, model, and time postlesion.

Striatal MSNs receive a variety of afferent inputs which include glutamatergic inputs from cortex as well as dopaminergic projections from SNC. Interestingly, MSN spine density is increased after methamphetamine, which also depletes striatal dopamine [42]. However, the methamphetamine-induced increase in spine density is selective to distal dendrites [43–45], while the effect that we observed occurred proximally. Given the importance of dopaminergic projections to the striatum, the absence of MPTP-induced

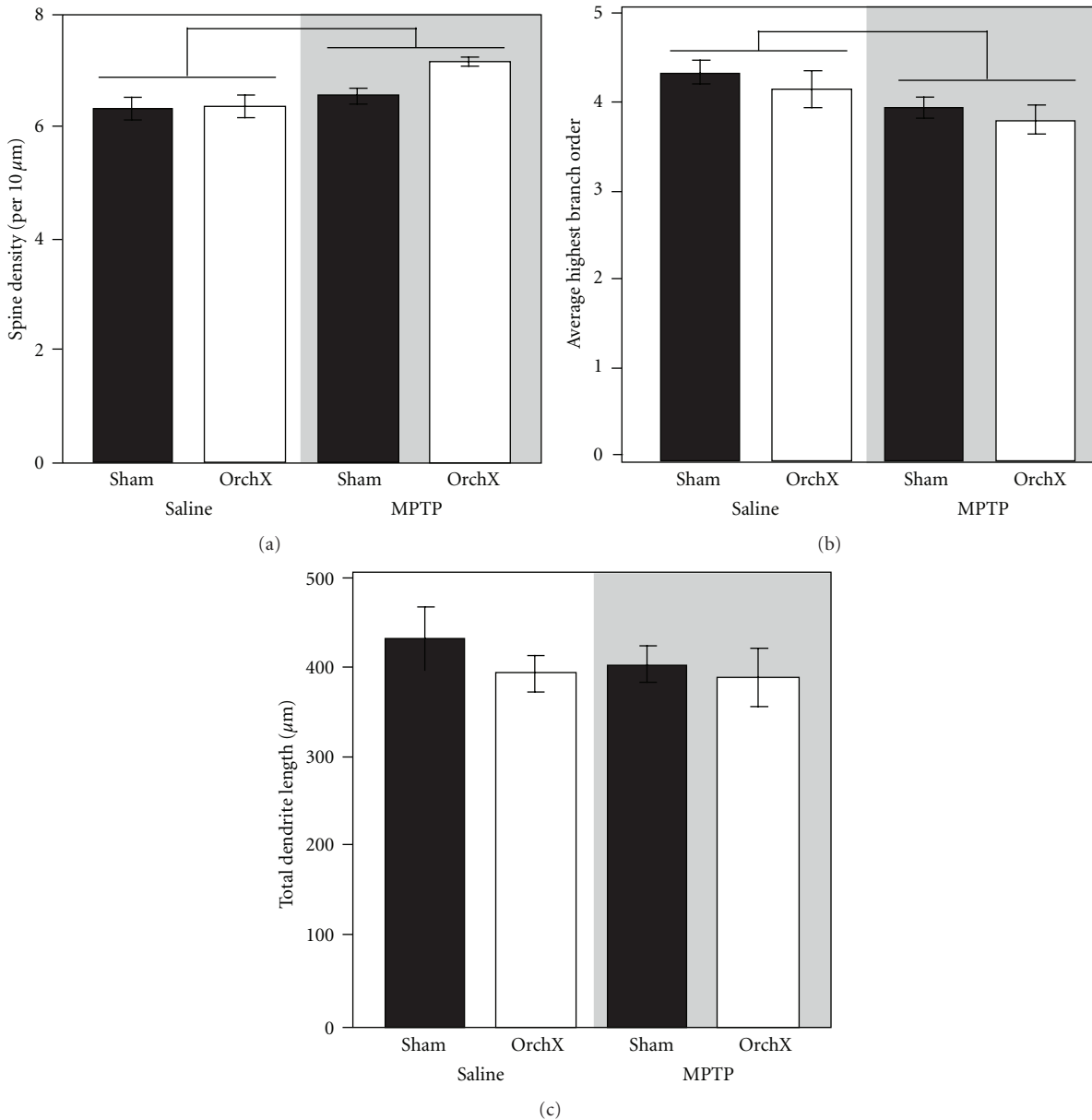


FIGURE 3: Neuronal morphology totals. Total spine density (a), average highest branch order (b), and total dendrite length (c) in saline-injected (unshaded panel) and MPTP-treated (shaded panel) gonad-intact (black bars) and orchidectomized (white bars) male mice ($n = 5$ mice/group). Bars represent an effect of MPTP ($P < .05$). OrchX: orchidectomized; Sham: sham-orchidectomized.

structural changes to MSN distal dendritic spines is indeed unexpected.

The absence of MPTP-induced structural changes to distal dendrites may relate to the distribution of striatal MSNs and the heterogeneity of striatal structure. MSNs represent a heterogeneous population comprised of both D1 receptor-containing neurons of the direct, striatonigral pathway and D2 receptor-containing neurons of the indirect, striatopallidal pathway [41]. The indirect pathway has also been implicated behaviorally, with D2 receptor knockout mice exhibiting PD-like akinesia and bradykinesia [46]. Behavioral deficits in D1 receptor knockout mice are minimal [47] or absent [48]. Recently, Day et al. [34] demonstrated a

selective effect of dopamine-depleting lesions on D2 receptor containing MSNs using 6-OHDA. Therefore, it is possible that MPTP-induced spine changes are also confined to the D2 receptor-containing subpopulation of MSNs.

The other key observation from our study was that castrated and gonad-intact males had the same response to MPTP, suggesting that testosterone fails to protect against MPTP-induced neuronal damage. Initially, we postulated that testosterone would be neuroprotective in males, similar to the effects of estrogen in females. This is because testosterone can be converted to estrogen and because PD occurs most often in older men whose endogenous androgens are in decline. Although our hypothesis was not supported,

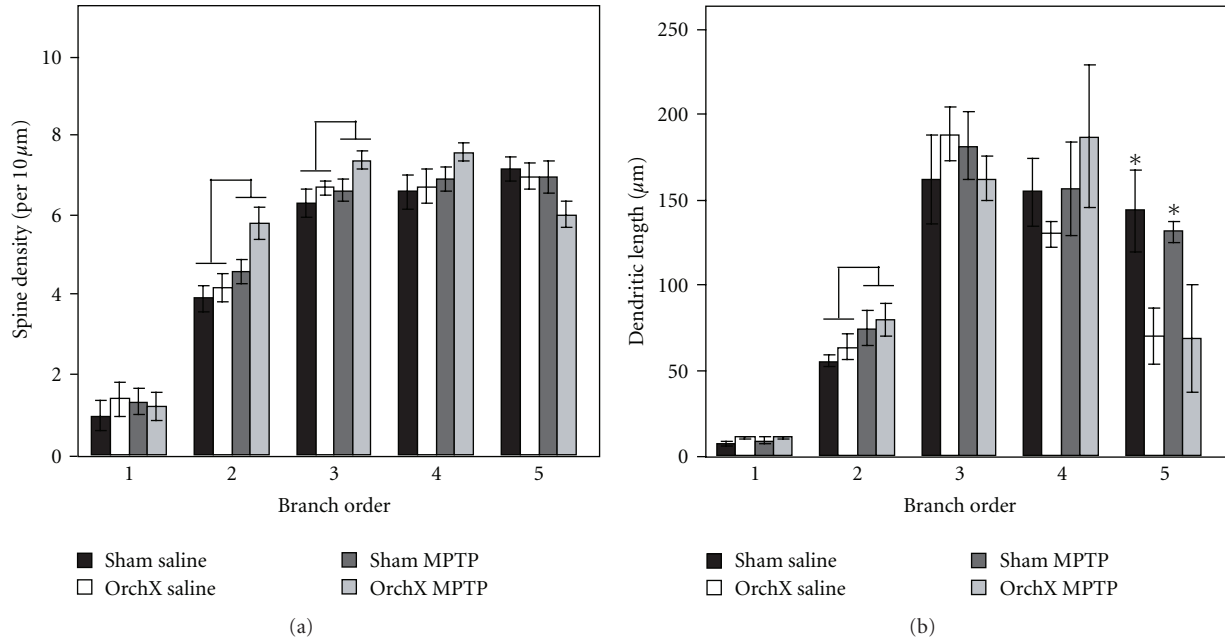


FIGURE 4: Branch order totals. Spine density (a) and dendritic length (b) for first- to fifth-order branches in striatal medium spiny neurons ($n = 5$ mice/group). Saline-injected mice include gonad-intact (black bars) and orchidectomized (white bars) males. MPTP-treated mice include gonad-intact (dark gray bars) and orchidectomized (light gray bars) males. Bars represent an effect of MPTP ($P < .05$), and asterisks represent an effect of gonadectomy ($P < .05$). Abbreviations: OrchX, orchidectomized; Sham, sham-orchidectomized.

our findings are in agreement with previous studies which found no effect of castration on striatal dopamine loss after MPTP [18] or on TH neurons in SNC after 6-OHDA [33]. In addition, studies using methamphetamine, which also depletes striatal dopamine, have shown that dopamine depletion after methamphetamine as well as amphetamine-induced stereotyped behaviors are similar in gonad-intact and castrated animals [49–52]. Interestingly, other studies have reported that testosterone increases neurotoxicity after dopamine-depleting lesions [13, 21, 53]. Due to the tremendous variability among published studies, these results are difficult to interpret. However, testosterone does not appear to have neuroprotective effects in the male nigrostriatal system.

Despite having no effect on MPTP-induced morphologic changes, castration decreased MSN dendritic length, but this effect was restricted to the distal branches of the dendritic tree. These effects of castration suggest that testosterone promotes dendritic growth in striatal MSNs. This raises an important question. How does testosterone promote growth of distal dendrites, but fail to attenuate MPTP-induced structural remodeling? To understand this issue, it is important to understand how hormones modulate neuronal plasticity, where hormones act in relation to the striatum and, in the case of testosterone, whether they act via androgenic or estrogenic mechanisms.

In hormone-sensitive areas of the brain, testosterone-driven changes in structural morphology are driven by classical hormone receptors for androgen or estrogen. For example, castration decreases dendritic branching in the posterior medial amygdala (MeP) and medial preoptic area

(MPOA), and reduces spine density on hippocampal CA1 neurons in adult rodents [31, 54, 55]. The findings in the current study are similar. However, while MeP, MPOA and CA1 each contains an abundance of classical receptors for both androgens and estrogen, the male mouse striatum is largely devoid of classical hormone receptors [56]. Moreover, classical hormone receptors are also sparse in the major dopaminergic input to MSN dendrites from SNC [56]. Although previous studies in rats and mice have found some evidence of AR and ER in SNC [56–58], it appears that few steroid-sensitive neurons are TH-positive and project to the striatum [56, 58]. This relative absence of hormone receptors severely limits the ability of testosterone to exert direct or indirect effects on MSN dendrites via classical mechanisms. It is more likely that testosterone acts through nonclassical mechanisms to drive changes in the striatum, as suggested previously [56, 59]. The present results extend this potential mechanism to include morphologic changes as well.

In the brain, testosterone can act as an androgen, but it can also act via estrogenic mechanisms after aromatization. The ability of gonadal hormones to attenuate nigrostriatal neurotoxicity is largely attributed to estrogen, which is neuroprotective in the female. In animal models, estrogen attenuates striatal dopamine depletion [9, 60, 61] and partially prevents the loss of striatal TH immunoreactivity [14, 62] after MPTP. Indeed, experimental evidence has shown that estrogen also protects against nigrostriatal degeneration in the male striatum. However, while circulating estrogen is abundant in females, striatal estrogen available in males is minimal due to the low levels of aromatase in the striatum [63–66]. Therefore, the inability of testosterone

to act through estrogenic mechanisms effectively prevents hormone-driven neuroprotection in the striatum. However, it does not influence the neurotrophic actions of testosterone on MSN distal dendrites, which likely occur via androgenic mechanisms.

Clinically, the actions of gonadal steroid hormones in the nigrostriatal system are important because estrogen is thought to be neuroprotective in several neurological disorders, including Parkinson's disease. In fact, women are more likely to develop PD after hysterectomy or menopause, when endogenous estrogen is eliminated [67, 68]. One of the potential benefits of hormone replacement therapy on postmenopausal women is the potential to delay the onset and/or decrease the severity of neurodegenerative disease. Men also experience a loss of testosterone with age, albeit less severe than the complete loss of gonadal steroids in the female. Although androgen replacement therapy is available for men with hypoandrogenism, the results of the current study suggest that androgen replacement will not attenuate nigrostriatal neurodegeneration in the male.

Abbreviations

6-OHDA:	6-hydroxydopamine
CA1:	CA1 region of the hippocampus
D1:	Dopamine receptor D1
D2:	Dopamine receptor D2
MeP:	Posterior medial amygdala
MPOA:	Medial preoptic area of the hypothalamus
MPTP:	1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
MSN:	Medium spiny neuron
PD:	Parkinson's disease
SNC:	Substantia nigra pars compacta
TH:	Tyrosine hydroxylase.

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