

in the occurrence of cardiovascular disease, cerebrovascular disease and tumor angiogenesis.

Chemokine receptor-4 (CXCR4) belongs to the G-protein coupled receptor (GPCR) family (7). The interactions between chemokines and chemokine receptors can induce the chemotactic migration of target cells, rearrangement of cell cytoskeleton, and intensive adhesive capacity between target cells and endothelial cells, which are extensively involved in biological functions such as cell growth, differentiation and apoptosis (8). Presently, it is known that one of the most important natural ligands of CXCR4 is stromal cell-derived factor-1 (SDF-1/CXCL12) (7). The combination of CXCR4 and SDF-1 can activate downstream signaling pathways including Ca^{2+} influx, phosphatidylinositol 3-kinase (PI3K), Akt, mitogen-activated protein (MAP) kinase, nuclear factor- κ B (NF- κ B), proline-rich tyrosine kinase 2 and the CT10 regulator of kinase (Crk) pathway (9). The significant effects of SDF-1/CXCR4 interaction have been highlighted in the regulation of cellular functions including cell adhesion, cell proliferation, cell migration, cell survival and angiogenesis (10,11). However, the functions and mechanisms of CXCR4 in BM-derived EPCs (BM-EPCs) have not previously been investigated.

The aim of the present study was to investigate the effects of CXCR4 overexpression on the proliferation, apoptosis and migration of hypoxia-treated BM-EPCs, and to explore the involvement of the downstream PI3K signaling pathway in these behaviors. BM-EPCs were pre-treated with hypoxia, in order to identify the regulatory mechanism of CXCR4 overexpression in EPCs under hypoxic conditions.

Materials and methods

Experimental animals. A total of 20 female Sprague Dawley (SD) rats (aged 4-6 weeks, weighing 200-240 g) were provided by Shrek (Shanghai, China). Animals were allowed free access to food and water, maintained in a 12-h light/dark cycle at room temperature (21-25°C) in 50-60% relative humidity. Routine cleaning and disinfection were performed daily. The duration of the experiment was 10 months. Animal health was monitored every 3 months. The animals used in the present study were cared for in accordance with the Guide for the Care and Use of Laboratory Animals published by the National Institutes of Health, and methods were approved by the Animal Research Committee of Hunan Provincial People's Hospital [Changsha, China; approval no. (2019)-041].

Isolation and culture of EPCs. SD rats were anesthetized by intraperitoneal injection of 3% pentobarbital sodium (30 mg/kg body weight) and then euthanized by means of cervical dislocation. Respiration and heartbeat were stopped to confirm the death of the animal, and the bodies were subsequently soaked in 75% ethanol solution for 15 min. The femur and tibia were then separated under aseptic conditions, with the bone surface of the femur and tibia carefully removed and sheared accordingly. The BM cavity was repeatedly washed with low-glucose Dulbecco's modified Eagle's medium (L-DMEM; Beyotime Institute of Biotechnology), after which the flushing fluid was collected. The collected flushing fluid was incubated with the equivalent volume of lymphocyte separation medium (Wisent,

Inc.). The solution was centrifuged at 2,000 x g for 15 min at room temperature to collect the cells, which were seeded into a culture dish with 200 ng/ml fibrinolyisin at a concentration of $1 \times 10^6/cm^2$. The cells were then incubated in L-DMEM with 10% fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc.) at 37°C with 5% CO_2 .

Phenotype identification of BM-EPCs. The BM-EPCs were characterized by immunofluorescent staining and flow cytometry analysis. For immunofluorescent staining, the BM-EPCs were harvested and washed with phosphate buffered saline (PBS) and incubated with 200 μ l Acetylated DiI lipoprotein from human plasma (DiI-Ac-LDL; 10 μ g/ml; Molecular Probes; Thermo Fisher Scientific, Inc.) for 4 h at 37°C with 5% CO_2 . Samples were then fixed with 4% paraformaldehyde for 30 min at 37°C. Cells were stained with 200 μ l fluorescein isothiocyanate labeled ulex europaeus lectin-1 (FITC-UEA-1; 10 μ g/ml; Sigma Aldrich; Merck KGaA) for 1 h at 37°C with 5% CO_2 , followed by observation under a fluorescence microscope (magnification, x120; Olympus Corporation).

For flow cytometry analysis, the BM-EPCs were lysed in PBS (Invitrogen; Thermo Fisher Scientific, Inc.) with 0.5% paraformaldehyde (Sigma Aldrich; Merck KGaA) and 0.5% saponin (Sigma Aldrich; Merck KGaA) for 5 min at 4°C. The cells were labeled with rabbit anti-mouse FITC-CD14 (BD Biosciences), rabbit anti-mouse PE-CD34 (Bio Legend, Inc.), and rabbit anti-mouse PerCP/Cy5.5-KDR (BioLegend, Inc.) for 1 h at room temperature. The same fluorescein-labeled isotype IgG was used as a control to define the negative population for each stain. After washing, cells were analyzed with the FACS™ Canto II flow cytometer. CD14, CD34 and KDR were examined using the FACSDiva software (v.6; BD Biosciences) for fluorescence-activated cell sorting, based on the manufacturer's instructions.

Adenovirus transfection. The BM-EPCs at passage number 2 were used for transduction. Viral vector, packaging plasmid, adenovirus related vectors (rAAV6 vector and pHelper1.0 vector) and 293T cells were purchased from Cyagen Biosciences Inc. 293T cells were transfected by the calcium phosphate method. The recombinant plasmids pAAV2.1-CMV-GFP (0.5 μ g/ml) or pAAV2.1-CMV-CXCR4 (5 μ g/ml) and the packaging plasmid pRC6 (2 μ g/ml) and pHelper (10 μ g/ml) were added to $CaCl_2$, then mixed with 2X HBS to make Calcium-DNA. 293T cells were then co-transfected with Calcium-DNA at 37°C with 5% CO_2 . After 48 h, the viral supernatant was harvested with centrifugation at 2,000 x g for 20 min at 4°C. The concentration of recombinant adeno-associated virus was determined by reverse transcription-quantitative polymerase chain reaction (RT-qPCR). BM-EPCs ($1 \times 10^5/well$) were seeded in 24-well cell culture plates and cultured overnight. Then, BM-EPCs were subsequently transfected with rAAV6-GFP and rAAV6-CXCR4 at the indicated multiplicity of infection (moi). After incubation at 37°C for 48 h, the transfection efficiency of the lentivirus vectors in transduced-EPCs was identified using fluorescence microscope. The level of CXCR4 expression was detected by western blotting, RT-qPCR and flow cytometry.

Cell treatment. After the BM-EPCs were transfected with rAAV₆-GFP and rAAV₆-CXCR4, they were cultured at 37°C

with 5% CO₂ in a humidified atmosphere with Endothelial Cell Growth Medium-2 (Lonza Group, Inc.), supplemented with 10% FBS, 100 U/ml penicillin and streptomycin sulfate (Thermo Fisher Scientific, Inc.), and SDF-1 at 20 µg/l for conventional culture. Initially, the BM-EPCs in the normoxia group were cultured under normoxic conditions (5% CO₂, 95% air atmosphere). BM-EPCs in the hypoxia group were cultured under anaerobic conditions (5% CO₂, 95% N₂) free of FBS for 4 h, and then cultured under normoxic conditions with PBS for 6 h. The BM-EPCs were subsequently divided into four groups: Normoxia + Sham (Empty vector) group, Normoxia + CXCR4 group, Hypoxia + Sham group, and Hypoxia + CXCR4 group. In additional experiments, the BM-EPCs were divided into these four groups: Hypoxia + Sham group, Hypoxia + CXCR4 group, Hypoxia + CXCR4 + PI3K inhibitor (LY294002, 20 µmol/l for 1 h) group, and Hypoxia + CXCR4 + CXCR4 inhibitor (AMD3100) group.

Cell counting kit-8 (CCK-8) assay. The treated BM-EPCs (2x10³ cells/well) were seeded into 96-well plates at 37°C. CCK-8 reagent (cat. no. CK04; Dojindo Molecular Technologies, Inc.) was added at the 24, 48, 72 and 96 h time points. After an additional 3 h, the absorbance value was determined by a microplate reader (Multiscan MK3; Thermo Fisher Scientific, Inc.) at 450 nm. The mean optical density (OD) of the three wells in each group was used to reflect the percentage of cell proliferation.

Transwell assay. The treated BM-EPCs (5x10⁴ cells/well) were added to the top of chambers (BD Biosciences) with serum-free medium. The complete medium was then added to the lower chambers. After 24 h, the non-migrated cells were removed, while the migrated cells were fixed using 4% paraformaldehyde for 20 min, and stained with 0.1% crystal violet solution for 5-10 min at 37°C (Sigma Aldrich; Merck KGaA). Lastly, images were captured under a light microscope (magnification, x200). The experiment was performed three times.

Annexin V-FITC/PI double staining assay. The treated BM-EPCs (1x10⁶ cells/ml) were stained with Annexin V-FITC/PI (Multisciences) under dark conditions for 15 min at room temperature. Cell apoptosis was detected using a FACS Calibur flow cytometry (BD Biosciences) with CellQuest software (version FACS101; BD Biosciences). The experiment was performed three times.

RT-qPCR assay. The total RNA of the treated BM-EPCs was extracted using TRIzol reagent (cat. no. 9109; Takara Biotechnology Co. Ltd.). Then, 3 µg RNA was reverse transcribed into cDNA using a cDNA synthesis kit (cat. no. 6130; Takara Biotechnology Ltd.) following the manufacturer's instructions. The expression levels of CXCR4 were detected using SYBR Green PCR Master Mix kit (Takara Biotechnology Co. Ltd.) on an ABI-Prism 7300 system (Applied Biosystems; Thermo Fisher Scientific, Inc.). β-actin was used as an endogenous control. Results were calculated using the 2^{-ΔΔC_t} method (12). The following primers were used: β-actin (product length, 150 bp) forward, 5'-GGAGATTAC TGCCCTGGCTCCTA-3'; β-actin reverse, 5'-GACTCATCG

TACTCCTGCTTGCTG-3'; CXCR4 (product length 113 bp) forward, 5'-CCATCCTCTACGCCTTCC-3'; CXCR4 reverse, 5'-TCCACCCCGTTTCCCTT-3'. The qPCR reaction mixture consisted of the following: 1 µl cDNA, 0.5 µl forward primers and 0.5 µl reverse primers, 10 µl SYBR Green PCR Master mix and 8 µl ddH₂O. The PCR reaction was as follow: (94°C, 2 min; 94°C, 20 sec; 58°C, 20 sec; 72°C, 20 sec; 40 cycles in total) followed by a dissolution curve.

Western blotting. Total protein from the treated BM-EPCs was extracted using a cell lysis buffer for Western and IP with proteinase inhibitors (Beyotime Institute of Biotechnology, Inc.). The concentration of total protein was determined using a BCA kit (cat. no. 23227; Thermo Fisher Scientific, Inc.). Then, 40 µg protein was separated using 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to polyvinylidene fluoride membranes (EMD Millipore). The membranes were blocked with 5% milk for 2 h at room temperature, then blotted with primary antibodies at 4°C overnight, followed by blotting with specific HRP-conjugated AffiniPure goat anti-rabbit secondary antibodies (cat. no. BA1054; Boster Biological Technology) at 37°C for 1 h after TBST (TBS with 0.05% Tween-20) washing. Finally, protein bands were visualized using enhanced chemiluminescence reagents (Pierce; Thermo Fisher Scientific, Inc.) on a Gel Imaging System (cat. no. 1708370; Edwards Life Sciences). The primary antibodies used were: Anti-GAPDH (1:10,000; RC-5G5, Kangcheng), anti-CXCR4 (1:1,000; cat. no. PA1237; Boster Biological Technology), anti-PI3K (1:1,000; cat. no. A01091-1; Boster Biological Technology) and anti-Akt (1:1,000; cat. no. A00024; Boster Biological Technology).

Statistical analysis. All data are presented as mean ± standard deviation (SD). Statistical analysis was performed using SPSS software (version 13.0; SPSS, Inc.). One-way analysis of variance (ANOVA) was used for data with n < 50. The least significant difference method was employed to compare samples when the variance was homogeneous. The Dunnett T3 test was used to compare samples when variance was not homogeneous. P < 0.05 was considered to indicate a statistically significant difference.

Results

Phenotype identification of BM-EPCs. The phenotype of BM-EPCs was identified through the application of immunofluorescent staining with Dil-ac-LDL and FITC-UEA-1. As depicted in Fig. 1A, the cell nuclei of BM-EPCs were blue after staining with DAPI. Endocytic vesicles were fluorescent red around the nucleus within the cytoplasm because of Dil-ac-LDL. The cell membrane was green after staining with FITC-UEA-1. The aforementioned results indicate that the expression of the EPC-specific surface marker CD14 was negative, while that of CD34 and KDR was positive (Fig. 1B).

CXCR4 is overexpressed in BM-EPCs. In order to investigate the mechanism as well as the functions associated with CXCR4 in BM-EPCs, adenoviruses were constructed (rAAV6-GFP and rAAV6-CXCR4), and the empty vector or adenovirus-CXCR4 were transfected into BM-EPCs. CXCR4

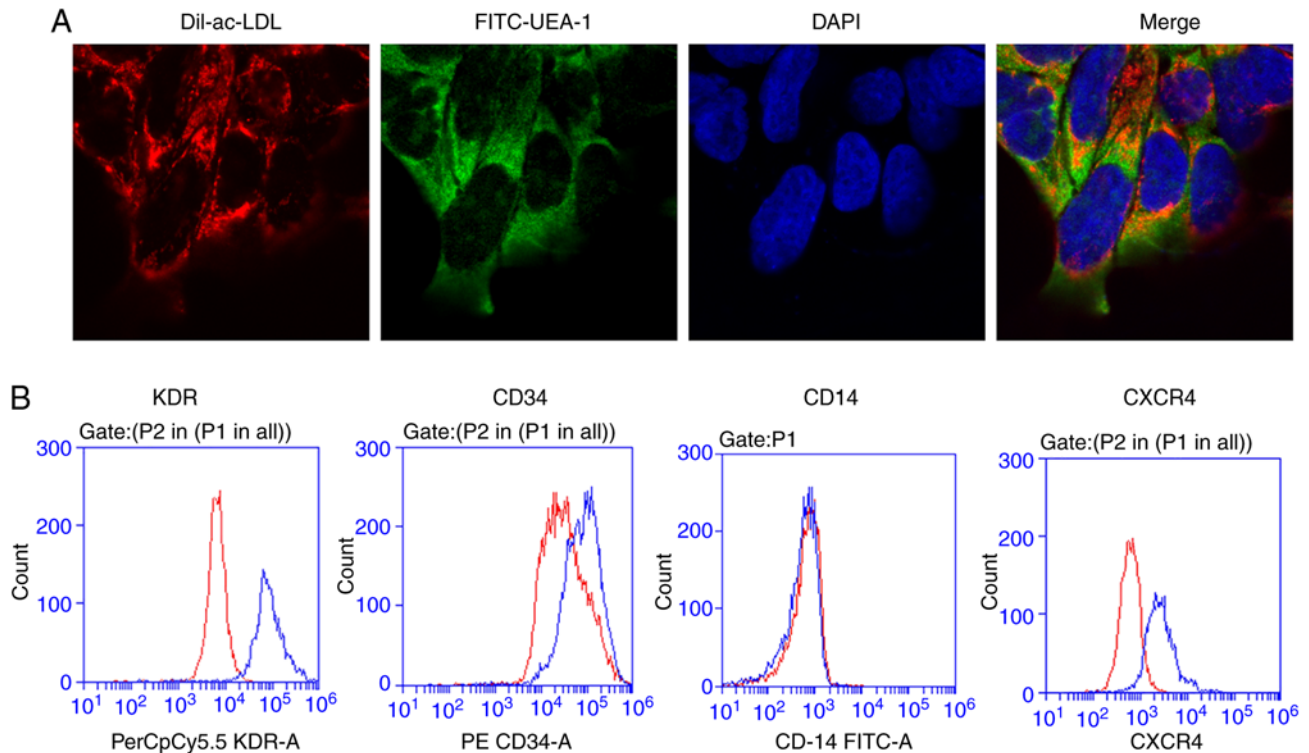


Figure 1. Phenotype identification of BM-EPCs observed following IF staining. (A) Dil-ac-LDL- and FITC-UEA-1-positive levels measured using an IF assay on BM-EPCs under a fluorescence microscope (magnification, x120). (B) CD14, CD34 and KDR detected by flow cytometry, and the levels of CXCR4 expression in BM-EPCs that were transfected with empty vector or adenovirus-CXCR4 detected by flow cytometry. BM-EPCs, bone marrow-derived endothelial progenitor cells; Dil-ac-LDL, dil-labelled acetylated low-density lipoprotein; FITC-UEA-1, fluorescein isothiocyanate labeled ulex europaeus lectin-1; DAPI, 4',6-diamidino-2-phenylindole. Immunofluorescence; CXCR4, chemokine receptor-4.

was stably overexpressed in BM-EPCs, as measured through flow cytometry assays (Fig. 1B). The mRNA and protein expression of CXCR4 was determined by RT-qPCR and western blot assays, respectively. The RT-qPCR results showed that CXCR4 mRNA was highly expressed in the CXCR4 group compared with the vector group ($P < 0.01$; Fig. 2A). Western blotting results revealed that CXCR4 protein was also highly expressed in the CXCR4 group compared with the vector group (Fig. 2B). The aforementioned results confirmed efficient transfection, and subsequent high expression of adenovirus-CXCR4 in BM-EPCs.

CXCR4 promotes proliferation and migration, while inhibiting apoptosis of hypoxia-induced BM-EPCs. In order to demonstrate the functional mechanism of CXCR4 in BM-EPCs under hypoxic conditions, BM-EPCs were treated with Hypoxia + CXCR4, Normoxia + CXCR4, Hypoxia + Sham (Empty vector), or Normoxia + Sham. Initially, western blotting revealed that the expression level of CXCR4, PI3K and Akt were all significantly elevated in the Normoxia + CXCR4 group compared with the Normoxia + Sham group ($P < 0.01$). The expression level of CXCR4, PI3K and Akt were also significantly increased in the Hypoxia + CXCR4 group vs. the Hypoxia + Sham group ($P < 0.01$; Fig. 3A). Thus, it was concluded that CXCR4 upregulated the expression level of PI3K and Akt in BM-EPCs under hypoxic conditions. Moreover, the present results indicated that cell proliferation ability was significantly strengthened in the Normoxia + CXCR4 group compared with the Normoxia + Sham group ($P < 0.01$), while a dramatic increase was observed

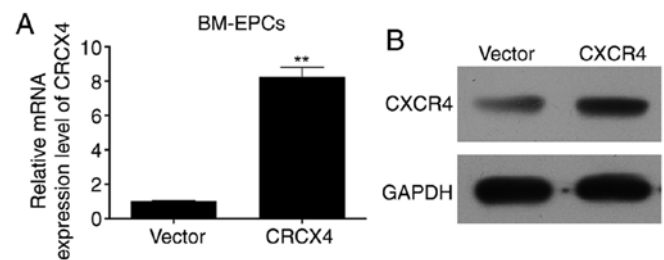


Figure 2. CXCR4 is overexpressed in BM-EPCs. BM-EPCs were transfected with empty vector or adenovirus-CXCR4. (A) The mRNA expression level of CXCR4 was measured by reverse transcription-quantitative PCR assay in BM-EPCs ($^{**}P < 0.01$ vs. vector group). (B) The protein expression level of CXCR4 was examined by western blotting assay in BM-EPCs. BM-EPCs, bone marrow-derived endothelial progenitor cells; CXCR4, chemokine receptor-4.

in the Hypoxia + CXCR4 group compared with the Hypoxia + Sham group ($P < 0.05$). In addition, cell proliferation ability was significantly inhibited in the Hypoxia + Sham group compared with the Normoxia + Sham group, and was also markedly suppressed in the Hypoxia + CXCR4 group compared with the Normoxia + CXCR4 group ($P < 0.01$; Fig. 3B).

The aforementioned results suggest that hypoxia inhibits cell proliferation of BM-EPCs, while CXCR4 promotes the proliferation ability of BM-EPCs with or without hypoxia treatment. Transwell assays further indicated that hypoxia resulted in the inhibition of BM-EPC cell migration, in addition to demonstrating that CXCR4 significantly enhanced the migration ability of BM-EPCs with or without hypoxia

treatment ($P < 0.01$; Fig. 3C and E). Furthermore, hypoxia markedly accelerated cell apoptosis of BM-EPCs, and it was found that CXCR4 markedly suppressed the apoptosis ability of BM-EPCs with or without hypoxia treatment ($P < 0.01$; Fig. 3D and F).

PI3K inhibitor and AMD3100 suppresses proliferation and migration, and accelerates apoptosis of CXCR4-mediated BM-EPCs under the hypoxic condition. Based on the observation that CXCR4 could upregulate the expression levels of PI3K and Akt in BM-EPCs, it was subsequently hypothesized that CXCR4 is implicated in the biological process of BM-EPCs by regulating the PI3K/Akt signaling pathway. In order to test the hypothesis, BM-EPCs were treated with Hypoxia + Sham, Hypoxia + CXCR4, Hypoxia + CXCR4 + PI3K inhibitor (LY294002), or Hypoxia + CXCR4 + CXCR4 inhibitor (AMD3100). The CCK-8 assay indicated that cell proliferation ability was significantly increased in the Hypoxia + CXCR4 group compared with the Hypoxia + Sham group ($P < 0.01$). A significant decrease in cell proliferation ability was identified in the Hypoxia + CXCR4 + LY294002 group compared with the Hypoxia + CXCR4 group ($P < 0.05$), and similar results were observed in the Hypoxia + CXCR4 + AMD3100 group compared with the Hypoxia + CXCR4 group ($P < 0.01$; Fig. 4A).

The aforementioned findings suggest that CXCR4 functioning on the EPC surface promoted BM-EPC proliferation under hypoxic conditions, and that the PI3K inhibitor acts to attenuate the stimulation mediated by CXCR4 in BM-EPCs under hypoxic conditions. Transwell assays further verified that CXCR4 significantly accelerated the migration of BM-EPCs under hypoxic conditions, while illustrating that PI3K inhibitor and CXCR4 inhibitor weakened the acceleration mediated by CXCR4 in BM-EPCs under hypoxic conditions ($P < 0.01$; Fig. 4B and D). More importantly, CXCR4 significantly repressed the apoptosis of BM-EPCs under hypoxic conditions, but PI3K inhibitor or CXCR4 inhibitor blocked the inhibition mediated by CXCR4 in BM-EPCs under hypoxic conditions ($P < 0.01$; Fig. 4C and E). These findings have significant implications on the understanding of how CXCR4 promotes the proliferation and migration of BM-EPCs, and suppresses their apoptosis through the PI3K signaling pathway.

Discussion

The pathogenesis of the majority of CVDs involves vascular endothelial damage (13), and hypoxia represents a chief factor implicated in vascular endothelial damage (14). Therefore, strategies to prevent vascular endothelial cell injury could potentially decrease the occurrence and severity of CVDs. The present study focused on the role of CXCR4 in protecting BM-derived EPCs in hypoxia, and found that the overexpression of CXCR4 promoted the proliferation and migration abilities of hypoxia-induced BM-EPCs and simultaneously inhibited the apoptosis ability of hypoxia-induced BM-EPCs through the PI3K/Akt signaling pathway.

Firstly, mononuclear cells were successfully isolated from the bone marrow of rats under a hypoxic environment and EPCs were characterized by immunofluorescence assay with

DIL-ac-LDL and FITC-UEA-1 labeling, resulting in cells that were CD14-negative, CD34- and KDR-positive, suggesting that EPCs have similar characteristics to endothelial cells. EPCs are defined as precursors of endothelial cells with positive expression of CD34, CD133 and VEGFR2 (15). EPCs are also BM mononuclear cells with partial endothelial function and differentiation ability, which can bind to ulex europaeus agglutinin-1 (UEA-1) and acetylated low-density lipoprotein (AcLDL) (16). EPCs derived from BM possess the ability to proliferate, migrate and differentiate into mature endothelial cells, as well as participate in angiogenesis, bind to blood vessels, and stimulate the proliferation of neighboring endothelial cells (17). Studies have highlighted the key role of EPCs in CVDs (18). Therefore, CXCR4 may be a potential therapeutic target for vascular damage and regeneration.

Secondly, CXCR4 was found to protect BM-derived EPCs under hypoxia. CXCR4, as a receptor for SDF-1, represents a crucial factor in the regulatory process of EPC homing (19). The blockade of CXCR4 signaling was previously shown to alleviate the decreased homing of EPCs to injured arteries (20), indicating that CXCR4 signaling may be a potential molecular target for gene therapy associated with the dysfunction of EPCs. The SDF-1/CXCR4 signaling pathway plays a key role in cell proliferation, tumor metastasis, angiogenesis, and the reendothelialization capacity of EPCs (21,22). The SDF-1 α /CXCR4 axis depletes EPC apoptosis under serum deprivation conditions through the PI3K/Akt signaling pathway (23), but the functions and mechanisms of CXCR4 in ischemic vascular diseases remain unknown. Adenoviruses were constructed (rAAV6-GFP and rAAV6-CXCR4), and successfully transfected into BM-EPCs. It was found that overexpression of CXCR4 promoted the proliferation and migration abilities of hypoxia-induced BM-EPCs. Simultaneously, the overexpression of CXCR4 inhibited the apoptosis ability of hypoxia-induced BM-EPCs. Therefore, CXCR4 may serve as a potential therapeutic target for the regeneration of impaired blood vessels.

Thirdly, CXCR4 upregulates the expression level of the PI3K and Akt signaling pathway in BM EPCs under hypoxic conditions; PI3K inhibitor (ly294002) inhibits proliferation and migration under hypoxia, and promotes the apoptosis of BM-EPC mediated by CXCR4. The PI3K signaling pathway is activated by various cellular stimuli or toxic insults, and induces the phosphorylation of key downstream effectors such as Akt. The activation of PI3K leads to an increase in phosphatidylinositol-(3,4,5)-triphosphate (PIP3) (24), which can bind to the pleckstrin homology on PDK1. Then, PDK1 can phosphorylate Akt at the activation loop (25). Akt phosphorylates several antiapoptotic proteins, cell-cycle-related proteins and transcription factors (24,25). Akt can regulate intracellular signals, affect cell responses to extrinsic stimuli, and participate in cell proliferation and survival (26). Extensive research has shown that PI3K/Akt are involved in an array of growth factor signal transduction pathways, and exert multiple biological functions (27). The present study found that CXCR4 upregulates PI3K and Akt expression in BM-EPCs under hypoxic conditions. In addition, PI3K inhibitor (LY294002) inhibits proliferation and migration, while promoting the apoptosis of CXCR4-mediated BM-EPCs under hypoxic conditions.

CXCR4 has been found to be expressed in >23 different types of cancer (28). SDF-1/CXCR4 is involved in breast cancer,

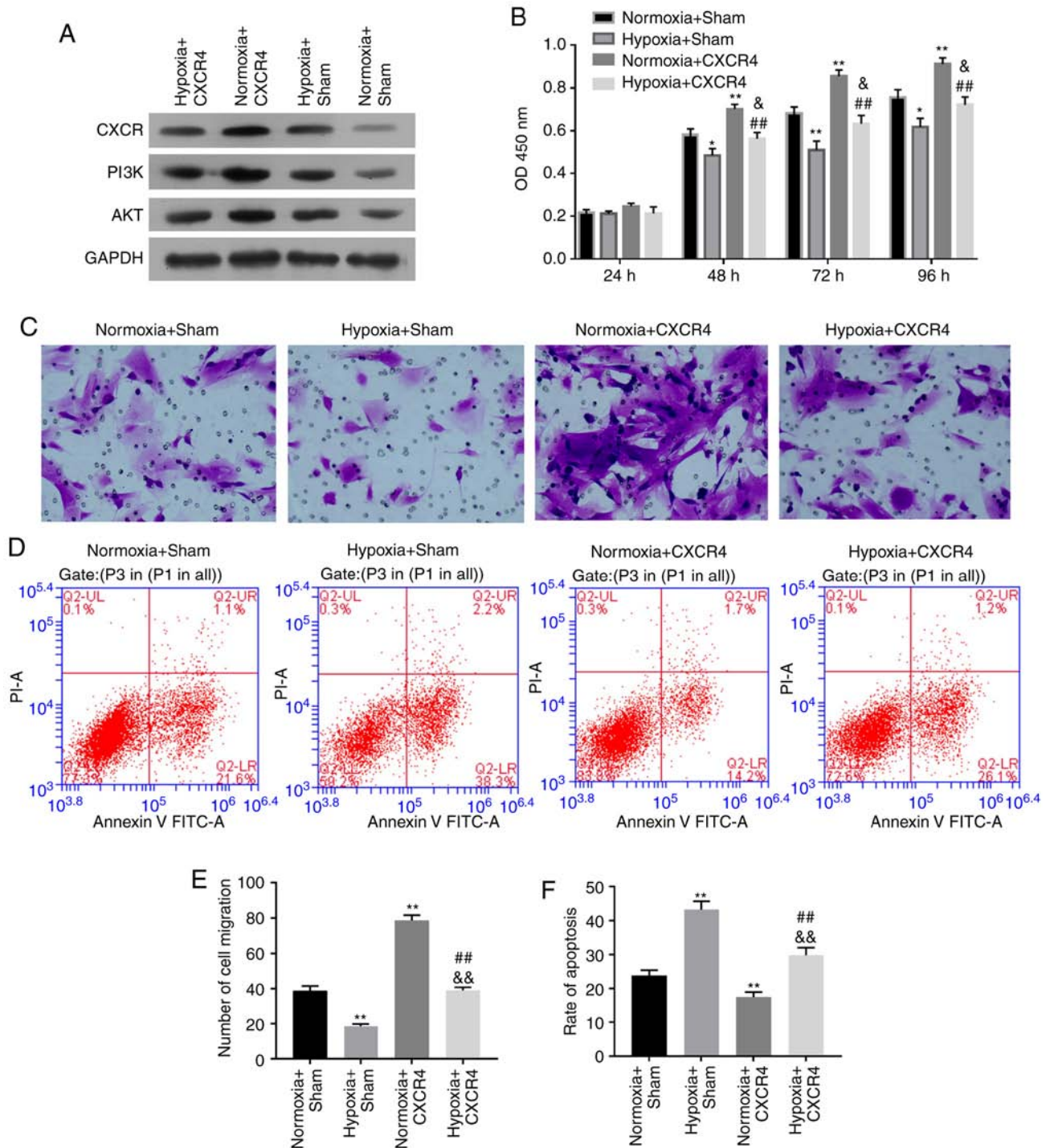


Figure 3. CXCR4 promotes proliferation and migration, and inhibits apoptosis of hypoxia-induced BM-EPCs. BM-EPCs were divided into four groups: Hypoxia + CXCR4, Normoxia + CXCR4, Hypoxia + Sham, and Normoxia + Sham groups. (A) The protein expression levels of CXCR4, PI3K and AKT were measured by western blotting. GAPDH was regarded as the loading control. (B) Cell proliferation ability detected by Cell Counting Kit-8 assay. (C) Migration capacity was determined by Transwell assay (magnification, x200). (D) Apoptosis capacity was detected by flow cytometry. (E) Quantification of CXCR4-promoted migration of hypoxia-induced BM-EPCs. (F) Quantification of CXCR4-inhibited apoptosis in hypoxia-induced BM-EPCs. Experiments were repeated 3 times; data are expressed as mean \pm SD. * $P < 0.05$ and ** $P < 0.01$ vs. Normoxia + Sham; & $P < 0.05$ and && $P < 0.01$ vs. Hypoxia + Sham; ## $P < 0.01$ vs. Normoxia + CXCR4. BM-EPCs, bone marrow-derived endothelial progenitor cells; PI3K, phosphatidylinositol 3-kinase; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; CXCR4, chemokine receptor-4.

glioma and neuroblastoma, which promotes endothelial progenitor cells homing to form neovascularization (29-31). In the present study, CXCR4-overexpressing EPCs showed enhanced cellular proliferation and decreased apoptosis under hypoxia, as well as normoxia. The intrinsic increase in PI3K/Akt is one of the major causes of transformation. Therefore, the potential risk

of tumors and hemangiomas caused by CXCR4-overexpressing EPCs should be carefully considered.

A limitation of the present study was that there were no set gradient values for SDF-1, LY294002 and AMD3100; thus, the most effective relative concentration for the interaction between SDF-1 and CXCR4, and the relative concentration

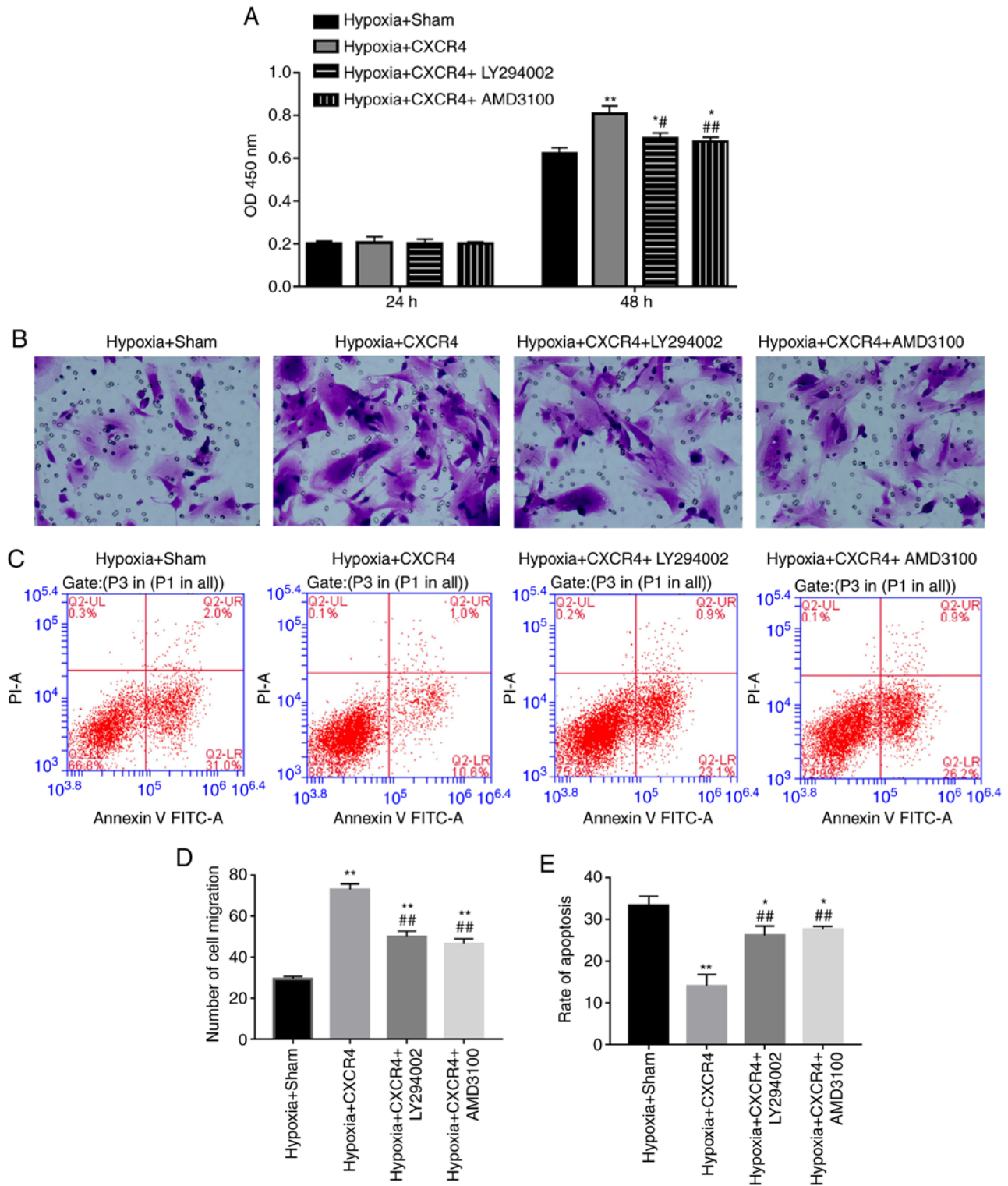


Figure 4. PI3K inhibitor or CXCR4 inhibitor suppresses proliferation and migration, while accelerating apoptosis of CXCR4-mediated BM-EPCs under hypoxic conditions. BM-EPCs were divided into four groups: Hypoxia + Sham, Hypoxia + CXCR4, Hypoxia + CXCR4 + LY294002, and Hypoxia + CXCR4 + AMD3100 groups. (A) Cell Counting Kit-8 assay was used to assess the proliferation abilities of BM-EPCs. (B) Transwell assay to evaluate cell migration ability (magnification, x200). (C) Flow cytometry to analyze cell apoptosis capacity. (D) Quantification of LY294002- and AMD3100-suppressed migration of CXCR4-mediated BM-EPCs under hypoxic conditions. (E) Quantification of LY294002 and AMD3100 increased apoptosis of CXCR4-mediated BM-EPCs under hypoxic conditions. Experiments were repeated 3 times; data are expressed as mean \pm SD. *P<0.05 and **P<0.01 vs. Hypoxia + Sham; #P<0.05 and ##P<0.01 vs. Hypoxia + CXCR4. BM-EPCs, bone marrow-derived endothelial progenitor cells; PI3K, phosphatidylinositol 3-kinase; CXCR4, chemokine receptor-4.

of CXCR4 inhibition by LY294002 and AMD3100 were unknown.

In conclusion, the present study suggests that CXCR4 accelerates proliferation and migration, and suppresses

apoptosis of hypoxia-induced BM-EPCs through the PI3K signaling pathway in rats. These results open new avenues of research to better understand the role of BM-EPCs in vascular repair.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

ZYL, ZFZ, BLZ and QXY conceived and designed the study; YC, JJR, JL, YT, CLW, XP, QCZ, LZ, LBX, FL and ZHY acquired the experimental data; HWP, JH, JZ, JW, YZ, JQP and ZFZ analyzed and interpreted the data; and ZYL, QXY and BLZ drafted the article or revised it critically for intellectual content. ZFZ and BLZ agree to be accountable for all aspects of the work, ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. ZYL, ZFZ, BLZ and QXY were responsible for confirming the authenticity of all the raw data, All authors have read and approved the final version of the manuscript.

Ethics approval and consent to participate

This study was conducted with the approval of the Ethics Committee of the Hunan Provincial People's Hospital (Changsha, China) [approval no. (2019)-041]. The animals used in this study were cared for in accordance with the *Guide for the Care and Use of Laboratory Animals* published by the National Institutes of Health. All efforts were made to minimize the suffering of the animals included in the study.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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