Shoot Induction in White Eggplant (*Solanum melongena* L. Cv. Bulat Putih) using 6-Benzylaminopurine and Kinetin

Pei Ching Foo, Ze Hong Lee, Chee Keong Chin, Sreeramanan Subramaniam and Bee Lynn Chew*

School of Biological Sciences, Universiti Sains Malaysia, 11800 USM Pulau Pinang, Malaysia

Published online: 6 July 2018

To cite this article: Pei Ching Foo, Ze Hong Lee, Chee Keong Chin, Sreeramanan Subramaniam and Bee Lynn Chew. (2018). Shoot induction in white eggplant (*Solanum melongena* L. cv. bulat putih) using 6-benzylaminopurine and kinetin. *Tropical Life Sciences Research* 29(2): 119–129. https://doi.org/10.21315/tlsr2018.29.2.9 **To link to this article:** https://doi.org/10.21315/tlsr2018.29.2.9

Abstrak: Solanum melongena L. biasanya dikenali sebagai terung adalah dari famili Solanaceae, yang sama asal usulnya dengan pokok tomato dan pokok ubi kentang. Ia adalah tanaman yang penting secara ekonomi di seluruh dunia dan dikaji untuk sifat perubatannya. nilai pemakanan dan peranannya sebagai model alternatif bagi tumbuhan. Buah terung telah digunakan sejak dahulu untuk rawatan pelbagai penyakit seperti bronkitis, asma, arthritis dan kencing manis serta sifat khasiatnya yang bermanfaat kepada diet manusia. Kajian transformasi tumbuhan pada terung telah dikaji secara meluas untuk penghasilan terung transgenik yang membawa gen yang bermanfaat untuk pertumbuhan tanaman yang optimum dan pengeluaran buah yang bermutu. Penginduksian pucuk adalah langkah penting yang diperlukan untuk penjanaan semula tisu tumbuhan yang berjaya kerana ianya merupakan prasyarat penting dalam transformasi menggunakan Agrobakterium sebagai pengantara. Terung tempatan cv. Bulat Putih adalah kultivar terung tempatan di Malaysia dengan buah putih dan bulat menjadikannya satu model potensi untuk kajian pengumpulan pigmen warna tumbuhan untuk tanaman buah-buahan. Kajian ini bertujuan untuk menyiasat potensi penginduksian pucuk menggunakan 6-benzylaminopurine (BAP) dan Kinetin dari eksplan kotiledon terung cv. Bulat Putih. Keputusan menunjukkan bahawa kedua-dua BAP dan kinetin boleh menginduksikan regenerasi kalus dari eksplan kotiledon. Selain itu, kinetin pada kepekatan 2.0 mg/L berjaya menginduksikan pucuk pada nilai 1.50 ± 0.22 pucuk untuk setiap eksplan manakala BAP sahaja tidak dapat menginduksikan pucuk. Kajian ini menunjukkan bahawa Kinetin sahaja sudah mencukupi untuk menginduksi pucuk dalam terung cv. Bulat Putih tanpa kehadiran BAP.

Kata Kunci: 6-Benzylaminopurine, Kinetin, Solanum melongena L., Induksi Pucuk

Abstract: Solanum melongena L. commonly known as the eggplant or brinjal comes from the family of Solanaceae, sharing the same ancestor with the tomato and potato. It is an economically important crop worldwide, being well studied for its medicinal properties, nutritional values and its role as an alternative model plant. The eggplant fruit has been

^{*}Corresponding author: beelynnchew@usm.my

[©] Penerbit Universiti Sains Malaysia, 2018. This work is licensed under the terms of the Creative Commons Attribution (CC BY) (http://creativecommons.org/licenses/by/4.0/).

Foo Pei Ching et al.

previously used for treatments of various diseases such as bronchitis, asthma, arthritis and diabetes as well as its nutritive properties that are beneficial to the human diet. Plant transformation studies on the eggplant have been widely done for the production of transgenic eggplants harbouring genes that are beneficial for optimal plant growth and fruit production. Shoot induction is an essential step required for the successful regeneration of transformed plant tissues and therefore is an essential pre-requisite in *Agrobacterium*-mediated transformation. The local eggplant cv. Bulat Putih is a local cultivar of eggplant in Malaysia with white and round fruits making it a potential model plant colour pigment accumulation studies in fruit crops. The current work aims to investigate the shoot induction potential of 6-benzylaminopurine (BAP) and Kinetin from cotyledon explants of eggplant cv. Bulat Putih. Results indicated that both BAP and Kinetin were able to induce the regeneration of callus from cotyledon explants. On the other hand, Kinetin at the concentration of 2.0 mg/L successfully induced shoots at the value of 1.50 ± 0.22 shoots per explant, whereas BAP alone did not trigger any formation of shoots. This study indicated that kinetin alone is sufficient to induce shoots in eggplant cv. Bulat Putih without the presence of BAP.

Keywords: 6-Benzylaminopurine, Kinetin, Solanum melongena L., Shoot Induction

INTRODUCTION

Solanum melongena L. with the common name brinjal or eggplant is an economically well-known crop grown in many countries particularly at regions with tropical and temperate conditions (Collonnier et al. 2001). It belongs to the Solanaceae (Nightshade) family and shares the common ancestor with other wellknown members in this family such as the tomato, potato, peppers and tobacco (Fukuoka et al. 2010). Generally, the eggplant is divided into three main categories according to the shape of their fruits. They are the egg-shaped (S. melongena var. esculentum), dwarf (S. melongena var. depressum) and long slender shaped (S. melongena var. serpentium) (Kashyap et al. 2003). Eggplant can grow well in regions with high rainfall as well as high temperature whereby conditions as such induces higher yield (Bhatti et al. 2013). The flowers of the eggplant are purplish, reddish or white in colour depending on the cultivar producing fruits either globular or long in shape, ranging from purple, white, green, brown or yellow in colour (Daunay et al. 2008). The eggplant is commonly known for its nutritive values as well as its medicinal properties to the human diet. Eggplant are used in cuisines worldwide especially in Asia, and also a popular ingredient in vegetarian dishes. The eggplant is packed with high soluble fiber and mineral contents as calcium, iron, potassium and phosphorus. Vitamins such vitamin C, vitamin B-6, vitamin K, folate and choline are also considerably high in the fruits making it beneficial to the human health (Bhatti et al. 2013). The low calorie and fat content of the eggplant fruit also contributes to weight loss and lowering risks of cardiovascular diseases (Robinson & Saranya 2013). The consumption of the eggplant fruit has also been linked to prevention of several diseases such as bronchitis, asthma, diabetes and arthritis, whereby this is mainly due to the presence of phenolic compounds such as chlorogenic acid in its fruit (Pratap et al. 2011; Scalzo et al. 2016). Previous

investigations have indicated the importance of chlorogenic acid in increasing glucose tolerance in human body and subsequently reducing the risk of diabetes and obesity in human (Rodriguez & Hadley 2002; Niggeweg *et al.* 2004; Van Dijk *et al.* 2009).

Several approaches including conventional plant breeding and biotechnological methods have been employed to increase the resistance of eggplant against pest and pathogen attack. However, the introduction of resistance gene into the plant either via traditional cross breeding methods or protoplast fusion brings alongside the issues of plant sterility in progenies (Fári et al. 1995). Previous attempts such as in-vitro selection, embryo rescue, somatic hybridization and genetic engineering have greatly improved the varieties of eggplant (Swamynathan et al. 2010; Rattan et al. 2015). Intensive studies on invitro regeneration of eggplant have been successfully performed on cell suspension culture (Wang et al. 2013), anther culture (Salas et al. 2012; Rotino 2016) and invitro shoot organogenesis (Bhat et al. 2013; Bhatti et al. 2014). As reviewed by Magioli and Mansur (2005), the efficiency of in-vitro organogenesis of eggplant is greatly dependent on the type of explants used and the combinations of plant growth regulators supplemented.

Cytokinin and auxin are the commonly known plant growth regulators being widely used in explant regeneration as different combination ratios of these two would bring significant difference in regeneration processes. Cytokinins are commonly used in plant tissue culture to initiate cell growth and also to induce the formation of shoots whereby the are actively take part in various plant physiological processes (Davies 1995). This includes shoot multiplication (Kaviani *et al.* 2013; Bhat *et al.* 2013; Abu-Romman *et al.* 2015), tuberous root production (Deshwal & Trivedi 2011), callus induction (Borjian & Arak 2013), leaf senescence (Riefler *et al.* 2006) and shoot apical meristem activity regulators (Tucker & Laux 2007). Cytokinin such as kinetin is known to initiate cell division in the presence of auxin and is widely used together with auxins for callus formation or to induce shoots where low levels of auxin is available. Cytokinin like 6-Benzylaminopurine (BAP) or benzyl adenine on the other hand is used for growth acceleration.

Genetic transformation of crops through *Agrobacterium* mediated transformation have been utilised to introduce pest resistant genes in eggplant to increase plant survival rate and product yield (Rotino & Gleddie 1990). The establishment of *in-vitro* regeneration protocol for transformed tissues is an essential stage required of the production of transgenic plants (Billings *et al.* 1997; Franklin *et al.* 2004; Rahman *et al.* 2006). Previous transformation attempts of transformation in eggplant include introducing the genes linked to resistance to the Colorado Potato Beetle and tolerance to abiotic stresses (Arpaia *et al.* 1997; Prabhavathi *et al.* 2002; Sidhu *et al.* 2014).

Up to the current knowledge, there are no reports on the regeneration protocol for the local Malaysian white eggplant cv. Bulat Putih, particularly in *invitro* shoot regeneration. This white cultivar is a suitable model for transformation and genetic studies linked pigment accumulation. The current study functions as a preliminary study to assess the regeneration potential of cotyledons using cytokinins

such BAP and kinetin. The current findings aim to aid future transformation studies for this cultivar particularly in genetic investigations of pigment accumulation in eggplant.

MATERIALS AND METHODS

Seed Surface Sterilization and *in-vitro* Germination

The seeds of round white *Solanum melongena* L. cv. Bulat Putih were obtained from Soon Huat Seeds Co. Sdn. Bhd. Penang, Malaysia. The seeds were washed with 70% ethanol for one minute, followed by rinsing with sterile distilled water for three times. Next, the seeds were washed with 50% Clorox® (commercial bleach solution) for 20 min and rinsed with sterile distilled water. Sterilised seeds were dried on sterile filter paper and placed in solid half-strength MS (Murashige & Skoog 1962) media. The sterilised seeds left to germinate in the culture room with 16 h light/8 h dark photoperiod at 24°±1°C.

In-vitro Shoot Regeneration

Cotyledons obtained from 3-week-old seedlings were used as explants for shoot regeneration. The cotyledons were first excised from the seedlings and cut at both ends. The cotyledons were then cultured on solid full-strength MS medium supplemented with 6-benzylaminopurine (BAP) and Kinetin. Kinetin (0.5, 1.0, 1.5, 2.0 mg/L) and BAP (1.0, 2.0, 3.0, 4.0 mg/L) were used singly or in combinations and maintained in the culture room under 16 hours light/8 hours dark photoperiod at $24^{\circ}\pm1^{\circ}$ C. The explants were sub-cultured every three weeks. The percentage of callus, shoot induction and the average number of shoots induced per explant were observed and recorded after six weeks of culture.

Statistical analysis

The experiment was performed according to the Complete Randomized Design and data were analysed using one-way analysis of variance (ANOVA) followed by comparison of means using Duncan's multiple range test at p < 0.05.

RESULTS AND DISCUSSION

The present study describes the regeneration potential of cotyledon explants of eggplant cv. Bulat Putih. The *in-vitro* seeds germinated two weeks after surface sterilisation. In the current study, all treatments of 6-Benzylaminopurine (BAP), Kinetin and the combinations of both induced white, friable callus (Fig. 1). The calluses were able to proliferate on the same treatment medium after the first

subculture. Treatments with BAP alone induced only callus but no shoot formation (Figs. 1C and 1D), indicating that media supplemented with BAP is more suitable for callus induction in the cultivar Bulat Putih. As reported by Sarker et al. (2006), cotyledon explants were identified to be the most suitable explant for shoot regeneration for eggplant as compared to the shoot tip, hypocotyl and roots. Previous studies on Solanum melongena cv. Larga Negra and Black Beauty reported the induction of callus in the treatment MS media supplemented with 0.5 mg/mL BAP + 2.0 mg/mL NAA whereby this combination favours the highest callus induction in cotyledon (90.0%) and hypocotyls (63.3%) (Zayova et al. 2008). On the other hand, Huda et al. (2007) also reported the combination of NAA and BAP initiated the formation of callus of Solanum melongena cv. Loda. In their research, MS media fortified with 0.05 mg/L BAP and 2.0 mg/L NAA had the highest response of callus formation from cotyledonary explants (100%). The same observation was reported by Ashrafuzzaman et al. (2009), whereby callus induction was the highest (95%) in MS medium supplied with 5.0 mg/mL BAP in combination with 0.1 mg/mL of NAA for hypocotyl of Capsicum annuum as compared to cotyledons (80%). In this study, hypocotyl explants treated with 4.0 mg/mL BAP were producing callus after 18 days of culture indicating BAP was able to induce callus for different types of explant of different Solanaceae species. Shah et al. (2015) also reported the formation of callus from hypocotyl explants of Solanum lycopersicum Mill. cv. Rio Grande in MS media supplemented with 2.0 mg/L Indole-3-acetic acid (IAA) and 2.5 mg/mL BAP (67.48 ± 0.7%).

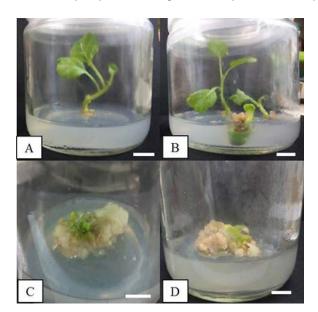


Figure 1: Shoot and callus formation in cotyledon explants of Solanum melongena L. cv. Bulat Putih. (A) MS medium + 1.5 mg/L kinetin. (B) MS medium + 2.0 mg/L kinetin. (C) MS medium + 3.0 mg/L BAP. (D) MS + 4.0 mg/L BAP after six weeks of culture. Scale bars represent 1 cm.

Foo Pei Ching et al.

Shoot formation was induced only in Kinetin supplemented media either alone or in combination with BAP. Kinetin at the concentration of 2.0 mg/L induced the highest percentage of shoot formation (65%) from the cotyledon explants with an average number of shoots of 1.50 ± 0.22 shoots per explant (Figs. 2 and 3). Shoot formation was also observed in treatments with lower concentrations of kinetin (1.5 mg/L) or in combination with BAP such as, 2.0 mg/L kinetin + 2.0 mg/L BAP and 1.0 mg/L kinetin + 3.0 mg/L BAP with average number of 0.80 ± 0.25 , $0.80 \pm$ 0.36 and 0.60 ± 0.40 shoots per explant, respectively (Fig. 3). In the current study, it was evident that the use of kinetin alone was sufficient to induce shoots from the cotyledon explants. There were no shoot and callus formation observed in full-strength MS basal medium without any plant growth regulators whereby only the formation of roots was noted. Root formation has been successfully induced on MS basal medium without plant growth regulators in other eggplant cultivars such as cv. Pusa Purple Long, cv. Black Jack and cv. Loda (Chen et al. 1995; Rahman et al. 2006; Pratap et al. 2011) suggesting that MS basal medium without plant growth regulators can be used for root regeneration. These results also indicated that this cultivar of Solanum melongena L. does not require auxins for root production.

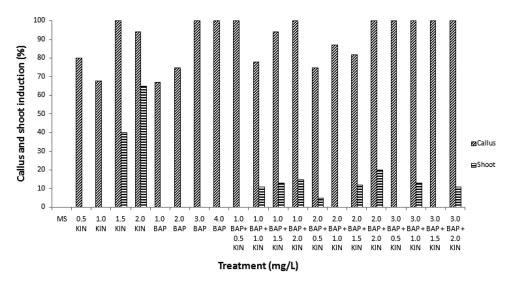


Figure 2: Percentage of callus and shoots induced on MS medium supplemented with different concentrations and combinations of cytokinins.

Previous regeneration studies in *Solanum melongena* L. reported the use of kinetin in combination with other plant growth regulators to induce shoots. Sarker *et al.* (2006) used a combination of kinetin and BAP to induce a high number of shoots, ranging from 0.75 to 4.4 shoots per explant in the cultivar Singhnath and Kazla. Jamil *et al.* (2013) reported that kinetin is necessary to be included together with other plant growth regulators in inducing shoots for eggplant cultivar NS-797. They

reported in their investigation the use of MS medium added with 25% of coconut milk, 1.5 mg/L of kinetin and 0.5 mg/L IAA induced shoots, whereby 70% of the total embryogenic calli produced turned green. Robinson and Saranya (2013) also reported a high number of shoot induction $(5.9 \pm 2.5 \text{ shoots per explant})$ with a percentage of 90% of shoot formation was achieved by supplementing 1.5 mg/L BAP in the culture media indicated the efficiency of BAP in shoot induction for a different cultivar. Other cytokinin such as Thidiazuron (TDZ) has also been used for shoot organogenesis of eggplant from other cultivars producing as high as 20 shoots per explant (Sharma & Rajam 1995; Magioli *et al.* 1998; 2000).

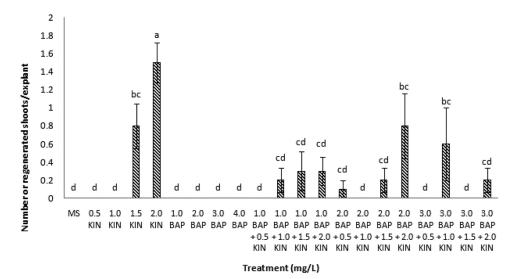


Figure 3: Average number of shoots regenerated on medium supplemented with different concentrations and combinations of cytokinins. Means marked by the same letters were not significantly different (Duncan Test, p < 0.05). Data represent mean ± standard error, n = 10.

Eggplant has been used as the model plant in various fruiting plant developmental studies and the establishment of biotechnological new approaches such as embryo rescue, somatic hybridisation and *in-vitro* selection, indicated the importance of this plant species (Magioli & Mansur 2005). Up to now, various studies on eggplant has been reported mainly on the common purple variety and not much being conducted on the white cultivar. The white eggplant cultivar is an important cultivar for gene expression studies especially the expression of genes associated to the accumulation of colour pigments or phytochemicals such as carotenoids and anthocyanins. This study aids the establishment of regeneration protocol for the Malaysian local cultivar of white eggplant.

Foo Pei Ching et al.

CONCLUSION

MS basal medium supplemented with 2.0 mg/L of kinetin was found to induce the highest number of shoots in comparison to other treatments applied. BAP was found to be suitable for callus induction of this cultivar of eggplant instead for shoot induction. The current study is a preliminary assessment to investigate the induction shoots from the local white eggplant cultivar in Malaysia. Future studies will involve gene expression studies in this cultivar particularly colour pigment genes for quality enhancement.

ACKNOWLEDGEMENTS

The authors would like to acknowledge Universiti Sains Malaysia for funding the project under the USM Research University Grant (1001/PBIOLOGY/811258).

REFERENCES

- Abu-Romman S M, Al-Hadid K A and Arabiyyat A R. (2015). Kinetin is the most effective cytokinin on shoot multiplication from cucumber. *Journal of Agricultural Science* 7(10): 159–165.
- Arpaia S, Mennella G, Onofaro V, Perri E, Sunseri F and Rotino G L. (1997). Production of transgenic eggplant (*Solanum melongena* L.) resistant to Colorado potato beetle (*Leptinotarsa decemlineata* Say). *Theoretical and Applied Genetics* 95(3): 329– 334. https://doi.org/10.1007/s001220050567
- Ashrafuzzaman M, Hossain M M, Ismail M R, Haque M S, Shahidullah S M and Uz-zaman S. (2009). Regeneration potential of seedling explants of chilli (*Capsicum annuum*). *African Journal of Biotechnology* 8(4): 591–596.
- Bhat S V, Jadhav A S, Pawar B D, Kale A A, Chimote V P and Pawar S V. (2013). In vitro shoot organogenesis and plantlet regeneration in brinjal (Solanum melongena L.). The Bioscan 8(3): 821–824.
- Bhatti K H, Jamil M D and Tufail M. (2014). Direct organogenesis (shoot and root) of eggplant (Solanum melongena L.) through tissue culture. World Applied Sciences Journal 30(3): 317–321.
- Bhatti K H, Kausar N, Rashid U, Hussain K, Nawaz K and Siddiqi E H. (2013). Effects of biotic stresses on eggplant (*Solanum melongena* L.). *World Applied Sciences Journal* 26(3): 302–311.
- Billings S, Jelenkovic G, Chin C K and Eberhardt J. (1997). The effect of growth regulators and antibiotics on eggplant transformation. *Journal of the American Society for Horticultural Science* 122(2): 158–162.
- Borjian L and Arak H. (2013). A study on the effect of different concentration of plant hormones (BAP, NAA, 2, 4-D, and Kinetin) on callus induction in *Brassica napus*. *International Research Journal of Applied and Basic Sciences* 5: 519–521.

- Chen Q, Jelenkovic G, Chin C K, Billings S, Eherhardt J, Goffreda J C and Day P. (1995). Transfer and transcriptional expression of Coleopteran *CryIIIB* endotoxin gene of *Bacillus thuringiensis* in eggplant. *Journal of the American Society for Horticultural Science* 120(6): 921–927.
- Collonnier C, Fock I, Kashyap V, Rotino G L, Daunay M C, Lian Y, Mariska I K, Rajam M V, Servaes A, Ducreux G and Sihachakr D. (2001). Applications of biotechnology in eggplant. *Plant Cell, Tissue and Organ Culture* 65(2): 91–107. https://doi.org/10.1023/A:1010674425536
- Daunay M C, Laterrot H and Janick J. (2008). Iconography and history of Solanaceae: Antiquity to the 17th century. In J Janick (ed.), *Horticultural Reviews*, Volume 34. Hoboken: John Wiley & Sons, Inc., pp. https://doi.org/10.1002/9780470380147. ch1
- Davies P J. (1995). The plant hormones: Their nature, occurrence, and functions. In: P J Davies (ed.), *Plant hormones: Physiology, biochemistry and molecular biology*. Dordrecht: Springer Netherlands, 1–12. https://doi.org/10.1007/978-94-011-0473-9_1
- Deshwal R K and Trivedi P. (2011). Effect of kinetin on enhancement of tuberous root production of *Chlorophytum borivilianum*. *International Journal of Innovations in Biological and Chemical Sciences* 1: 28–31.
- Fári M, Nagy I, Csányi M, Mitykó J and Andrásfalvy A. (1995). Agrobacterium mediated genetic transformation and plant regeneration via organogenesis and somatic embryogenesis from cotyledon leaves in eggplant (Solanum melongena L. cv. 'Kecskemetilila'). Plant Cell Reports 15(1–2): 82–86. https://doi.org/10.1007/ BF01690259
- Franklin G, Sheeba C J and Sita G L. (2004). Regeneration of eggplant (Solanum melongena L.) from root explants. In Vitro Cellular and Developmental Biology–Plant 40(2): 188–191. https://doi.org/10.1079/IVP2003491
- Fukuoka H, Yamaguchi H, Nunome T, Negoro S, Miyatake K and Ohyama A. (2010). Accumulation, functional annotation, and comparative analysis of expressed sequence tags in eggplant (*Solanum melongena* L.), The third pole of the genus *Solanum* species after tomato and potato. *Gene* 450(1): 76–84. https://doi. org/10.1016/j.gene.2009.10.006
- Huda A, Bari M A, Rahman M and Nahar N. (2007). Somatic embryogenesis in two varieties of eggplant (*Solanum melongena* L.). *Research Journal of Botany* 2: 195–201. https://doi.org/10.3923/rjb.2007.195.201
- Jamil M D, Parvaiz M, Tufail M, Arshad J, Hussain S and Imtiaz S. (2013). Callogenesis, regeneration of shoot and root of brinjal (*Solanum melongena* L.). *World Applied Sciences Journal* 26: 1039–1045.
- Kashyap V, Kumar S V, Collonnier C, Fusari F, Haicour R, Rotino G L and Rajam M V. (2003). Biotechnology of eggplant. *Scientia Horticulturae* 97(1): 1–25. https://doi. org/10.1016/S0304-4238(02)00140-1
- Kaviani B, Hesar A A, Tarang A, Zanjani S B, Hashemabadi D and Ansari M H. (2013). Effect of kinetin (Kn) and naphthalene acetic acid (NAA) on the micropropagation of *Matthiola incana* using shoot tips, and callus induction and root formation on the leaf explants. *African Journal of Agricultural Research* 8(30): 4134–4139.
- Magioli C and Mansur E. (2005). Eggplant (*Solanum melongena L*.): Tissue culture, genetic transformation and use as an alternative model plant. *Acta Botonica Brasilica* 19(1): 139–148. https://doi.org/10.1590/S0102-33062005000100013

- Magioli C, Pinheiro M M and Mansur E. (2000). Establishment of an efficient *Agrobacterium*mediated transformation system for eggplant and study of a potential biotechnologically useful promoter. *Journal of Plant Biotechnology* 2(1): 43–49.
- Magioli C, Rocha A P M, De Oliveira D E and Mansur E. (1998). Efficient shoot organogenesis of eggplant (*Solanum melongena* L.) induced by thidiazuron. *Plant Cell Reports* 17(8): 661–663. https://doi.org/10.1007/s002990050461
- Murashige T and Skoog F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum* 15(3): 473–497. https://doi. org/10.1111/j.1399-3054.1962.tb08052.x
- Niggeweg R, Michael A J and Martin C. (2004). Engineering plants with increased levels of the antioxidant chlorogenic acid. *Nature Biotechnology* 22(6): 746–754. https://doi.org/10.1038/nbt966
- Prabhavathi V, Yadav J S, Kumar P A and Rajam M V. (2002). Abiotic stress tolerance in transgenic eggplant (*Solanum melongena* L.) by introduction of bacterial mannitol phosphodehydrogenase gene. *Molecular Breeding* 9(2): 137–147. https://doi.org/10.1023/A:1026765026493
- Pratap D, Kumar S, Raj S K and Sharma A K. (2011). *Agrobacterium*-mediated transformation of eggplant (*Solanum melongena* L.) using cotyledon explants and coat protein gene of cucumber mosaic virus. *Indian Journal of Biotechnology* 10(1): 19–24.
- Rahman M, Asaduzzaman M, Nahar N and Bari M A. (2006). Efficient plant regeneration from cotyledon and midrib derived callus in eggplant (*Solanum melongena* L.). *Journal of Biosciences* 14: 31–38.
- Rattan P, Kumar S, Salgotra R K, Samnotra R K and Sharma F. (2015). Development of interspecific F1 hybrids (*Solanum melongena × Solanum khasianum*) in eggplant through embryo rescue technique. *Plant Cell, Tissue and Organ Culture* 120(1): 379–386. https://doi.org/10.1007/s11240-014-0591-4
- Riefler M, Novak O, Strnad M and Schmülling T. (2006). Arabidopsis cytokinin receptor mutants reveal functions in shoot growth, leaf senescence, seed size, germination, root development, and cytokinin metabolism. The Plant Cell 18: 40–54. https://doi.org/10.1105/tpc.105.037796
- Robinson J P and Saranya S. (2013). An improved method for the *In Vitro* propagation of Solanum melongena L. International Journal of Current Microbiology and Applied Sciences 2(6): 299–306.
- Rodriguez de S D V and Hadley M. (2002). Chlorogenic acid modifies plasma and liver concentrations of: cholesterol, triacylglycerol, and minerals in (fa/fa) Zucker rats. *Journal of Nutritional Biochemistry* 13(12): 717–726. https://doi.org/10.1016/ S0955-2863(02)00231-0
- Rotino G L and Gleddie S. (1990).Transformation of eggplant (Solanum melongena L.) using a binary Agrobacterium tumefaciens vector. Plant Cell Reports 9(1): 26–29. https://doi.org/10.1007/BF00232129
- Rotino G L. (2016). Anther culture in eggplant (*Solanum melongena* L.). *Methods in Molecular Biology* 1359: 453–66. https://doi.org/10.1007/978-1-4939-3061-6_25
- Salas P, Rivas-Sendra A, Prohens J and Seguí-Simarro J M. (2012). Influence of the stage for anther excision and heterostyly in embryogenesis induction from eggplant anther cultures. *Euphytica* 184(2): 235–250. https://doi.org/10.1007/s10681-011-0569-9
- Sarker R H, Yesmin S and Hoque M I. (2006). Multiple shoot formation in eggplant (*Solanum melongena* L.). *Plant Tissue Culture and Biotechnology* 16(1): 53–61.

- Scalzo R L, Fibiani M, Francese G, D'Alessandro A, Rotino G L, Conte P and Mennell G. (2016). Cooking influence on physico-chemical fruit characteristics of eggplant (Solanum melongena L.). Food Chemistry 194: 835–842. https://doi.org/10.1016/j. foodchem.2015.08.063
- Shah S H, Ali S, Jan S A, Din J and Ali G M. (2015). Callus induction, in-vitro shoot regeneration and hairy root formation by the assessment of various plant growth regulators in tomato (*Solanum lycopersicum* Mill.). *Journal of Animal and Plant Sciences* 25(2): 528–538.
- Sharma P and Rajam M V. (1995). Genotype, explant and position effects on organogenesis and somatic embryogenesis in eggplant (*Solanum melongena* L.). *Journal of Experimental Botany* 46(1): 135–141. https://doi.org/10.1093/jxb/46.1.135
- Sidhu M K, Dhatt A S, Sandhu J S and Gosal S S. (2014). Biolistic transformation of cry 1Ac gene in eggplant (Solanum melongena L.). International Journal of Agriculture, Environment and Biotechnology 7(4): 679.
- Swamynathan B, Nadanakunjidam S, Ramamourti A, Sindhu K and Ramamoorthy D. (2010). *In vitro* plantlet regeneration through somatic embryogenesis in *Solanum melongena* (Thengaithittu variety). *Academic Journal of Plant Sciences* 3(2): 64– 70.
- Tucker M R and Laux T. (2007). Connecting the paths in plant stem cell regulation. *Trends in Cell Biology* 17(8): 403–410. https://doi.org/10.1016/j.tcb.2007.06.002
- Van Dijk A E, Olthof M R, Meeuse J C, Seebus E, Heine R J and Van Dam R M. (2009). Acute effects of decaffeinated coffee and the major coffee components chlorogenic Acid and trigonelline on glucose tolerance. *Diabetes Care* 32(6): 1023–1025. https://doi.org/10.2337/dc09-0207
- Wang F, Li G, Chen S, Jiang Y and Wang S. (2013). Callus induction and cell suspension culture of eggplant (*Solanum melongena* L.). *Journal of Agricultural Science and Technology* 14(9): 1220.
- Zayova E, Nikova V, Ilieva K and Philipov P. (2008). Callusogenesis of eggplant (Solanum melongena L.). Comptes Rendus de l'Academie Bulgare Des Sciences 63(12): 1749–1756.