### MITOGENOME ANNOUNCEMENT

Taylor & Francis Taylor & Francis Group

OPEN ACCESS Check for updates

# The mitochondrial genome of the black-tailed dasyure (Murexia melanurus)

Ran Tian<sup>a</sup>\* (**b**), Yuepan Geng<sup>a</sup>\*, Patrick B. Thomas<sup>b,c,d</sup> (**b**), Penny L. Jeffery<sup>b,c</sup> (**b**), Thomas Y. Mutton<sup>e</sup>, Lisa K. Chopin<sup>b,c</sup>, Andrew M. Baker<sup>e,f</sup> (**b**) and Inge Seim<sup>a,b,c,g</sup> (**b**)

<sup>a</sup>Integrative Biology Laboratory, College of Life Sciences, Nanjing Normal University, Nanjing, Jiangsu, China; <sup>b</sup>Ghrelin Research Group, Translational Research Institute-Institute of Health and Biomedical Innovation, School of Biomedical Sciences, Queensland University of Technology, Brisbane, Queensland, Australia; <sup>c</sup>Australian Prostate Cancer Research Centre-Queensland, Translational Research Institute – Institute of Health and Biomedical Innovation, Queensland University of Technology, Brisbane, Queensland, Australia; <sup>d</sup>Queensland Bladder Cancer Initiative, Translational Research Institute-Institute of Health and Biomedical Innovation, School of Biomedical Sciences, Queensland University of Technology, Woolloongabba, Queensland, Australia; <sup>e</sup>School of Earth, Environmental and Biological Sciences, Queensland University of Technology, Brisbane, Queensland, Australia; <sup>f</sup>Natural Environments Program, Queensland Museum, South Brisbane, Queensland, Australia; <sup>g</sup>Comparative and Endocrine Biology Laboratory, Translational Research Institute-Institute of Health and Biomedical Innovation, School of Biomedical Sciences, Queensland University of Technology, Woolloongabba, Queensland, Australia;

#### ABSTRACT

In this study, we report the mitochondrial genome of the black-tailed dasyure (*Murexia melanurus*) of New Guinea. The circular genome is 17,736 bp in length and has an AT content of 60.5%. Its gene content – 13 protein-coding genes (PCGs), 2 ribosomal (rRNA) genes, 21 transfer RNA (tRNA) genes, a tRNA pseudogene (*tRNA<sup>Lys</sup>*), and a non-coding control region (CR) – and gene arrangement are consistent with previous marsupial mitogenome assemblies.

#### **ARTICLE HISTORY**

Received 2 September 2019 Accepted 23 September 2019

#### **KEYWORDS**

Mitochondrial genome; marsupial; dasyure; Dasyuridae; New Guinea; *Murexia* 

The marsupial family Dasyuridae (or carnivorous marsupials) includes  $\sim$ 75 species native to mainland Australia and Tasmania, New Guinea, and other adjacent islands (Baker and Dickman 2018). The dasyurid subfamily Phascogalinae comprises three genera: *Antechinus* and *Phascogale* of Australia, and *Murexia* of New Guinea. Although morphological similarities exist, the current evidence suggests these genera derived from a common ancestor some 12.5 million years ago (Mutton et al. 2019). *Murexia* spp. have not received as much attention in evolutionary studies as their Australian counterparts within Phascogalinae, and genetic information on the various constituent species within *Murexia* is poorly represented. Here, we describe the complete mitochondrial genome of the black-tailed dasyure (*Murexia melanurus*).

*Murexia melanurus* genomic DNA was extracted from ear tissue (voucher specimen ABTC46020) collected from Tibi, Papua New Guinea. Paired-end short-insert (200 bp) DNA libraries were sequenced by BGI (Hong Kong, China), to generate  $\sim 30 \times$  genome coverage. Raw data were filtered using Flexbar v3.4.0 (Roehr et al. 2017). To remove microbial contaminants, we used bowtie2 v2.3.4.1 (Langmead and Salzberg 2012) to map reads to all bacterial and fungal sequences in NCBI Genomes, retaining 99.68% of reads. Two to 95 million reads were assembled using NOVOPlasty v2.7.2 (Dierckxsens et al. 2017), with the *ND1* coding sequence from a partial *M. melanurus* mitogenome (GenBank: KJ868127) (Mitchell et al. 2014) as a seed sequence and the parameters 'Type = mito, K-mer = 39, Genome range = 16,000–22,000'. The subset with the lowest number of reads which yielded a circular genome was retained (here: 12 M reads; 0.26% were assembled into a 17,736 bp contig at 179× coverage). Geneious Prime v2019.1.3 (Biomatters Ltd., Auckland, New Zealand) was used to align 95 M reads against the assembled contig and generate a consensus genome sequence, with a 75% masking threshold. The genome was annotated using GenBank features of the northern quoll (*Dasyurus hallucatus;* accession no. NC\_007630). Various genome features were compared to the Virginia opossum (*Didelphis virginiana*), a seminal species in the marsupial mitochondrial genetics literature [e.g. see Janke et al. (1994) and Nilsson (2009)].

The *M. melanurus* mitochondrial genome (GenBank: MK977600) is 17,736 bp and has a base composition of 32% A, 28.5% T, 14.1% G, and 25.4% C. As in other marsupials, the genome has 13 protein-coding genes (PSGs), 2 ribosomal (rRNA) genes, and 21 transfer RNA (tRNA) genes. The genome shares unique features with all marsupial mitogenomes reported to date. These include: an 'ACWNY' tRNA gene rearrangement (Paabo et al. 1991); a *tRNA<sup>Lys</sup>* pseudogene (Janke et al. 1994; Dorner et al. 2001); and lack of an anticodon for aspartic acid, which is likely rescued by

**CONTACT** Inge Seim inge@seimlab.org integrative Biology Laboratory, College of Life Sciences, Nanjing Normal University, Nanjing 210046, Jiangsu, China \*These authors contributed equally to this work.

© 2019 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Figure 1. Phylogenetic tree of black-tailed dasyure (*Murexia melanurus*; indicated in bold blue font), nine other species in the marsupial family Dasyuridae, and the outgroup species *Thylacinus cynocephalus*. Because there are no complete mitogenomes from the genera *Antechinus* and *Murexia* in GenBank, phylogenetic reconstruction was performed with coding sequences of 12 protein-coding genes (excluding *ND6*). The number at each node is ML/BI bootstrap support value.

RNA-editing (Janke and Paabo 1993). In agreement with a recent molecular appraisal (Mitchell et al. 2014; Westerman et al. 2016), phylogenetic analysis revealed that *M. melanurus* and *Murexia habbema* are sisters, to the exclusion of the closely-related genera *Phascogale* and *Antechinus* (Figure 1). Maximum-likelihood (ML; estimated using IQ-TREE (Nguyen et al. 2015)) and Bayesian Interference (BI; implemented in MrBayes v3.2.7 (Ronquist and Huelsenbeck 2003)) gave the same tree topology.

# **Geolocation information**

Geospatial coordinates for the black-tailed dasyure (Murexia melanurus) ear tissue collection:  $6^{\circ}11'S$ ,  $143^{\circ}9'E$ 

# **Acknowledgements**

We thank Leanne Wheaton (South Australian Museum) for providing the *M. melanurus* ear tissue sample.

### **Disclosure statement**

The authors declare that they have no competing interests.

# Funding

This work was supported by the Queensland University of Technology Faculty of Health/School of Biomedical Sciences Industry Collaboration Preparedness Pilot Research Support Scheme (to LKC, AB, and IS), the National Natural Science Foundation of China [31950410545 to IS; 31900310 to RT], a Young Foreign Experts in Economic and Technological Sector Grant (to IS), the China Postdoctoral Science Foundation [2018M642278 to RT], and the Priority Academic Programme Development of Jiangsu Higher Education Institutions (PAPD; to IS).

## ORCID

Ran Tian (b) http://orcid.org/0000-0002-4564-1886 Patrick B. Thomas (b) http://orcid.org/0000-0002-3649-7923 Penny L. Jeffery (b) http://orcid.org/0000-0003-1229-8724 Andrew M. Baker (b) http://orcid.org/0000-0001-8825-1522 Inge Seim (b) http://orcid.org/0000-0001-8594-7217

## References

- Baker A, Dickman C. 2018. Secret lives of carnivorous marsupials. Clayton (Australia): CSIRO Publishing.
- Dierckxsens N, Mardulyn P, Smits G. 2017. NOVOPlasty: de novo assembly of organelle genomes from whole genome data. Nucl Acids Res. 45(4): e18.
- Dorner M, Altmann M, Paabo S, Morl M. 2001. Evidence for import of a lysyl-tRNA into marsupial mitochondria. MBoC. 12(9):2688–2698.
- Janke A, Feldmaier-Fuchs G, Thomas WK, von Haeseler A, Paabo S. 1994. The marsupial mitochondrial genome and the evolution of placental mammals. Genetics. 137(1):243–256.
- Janke A, Paabo S. 1993. Editing of a tRNA anticodon in marsupial mitochondria changes its codon recognition. Nucl Acids Res. 21(7): 1523–1525.
- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. Nat Methods. 9(4):357–359.
- Mitchell KJ, Pratt RC, Watson LN, Gibb GC, Llamas B, Kasper M, Edson J, Hopwood B, Male D, Armstrong KN, et al. 2014. Molecular phylogeny, biogeography, and habitat preference evolution of marsupials. Mol Biol Evol. 31(9):2322–2330.

- Mutton TY, Phillips MJ, Fuller SJ, Bryant LM, Baker AM. 2019. Systematics, biogeography and ancestral state of the Australian marsupial genus *Antechinus* (Dasyuromorphia: Dasyuridae). Zool J Linn Soc. 186(2): 553–568.
- Nguyen LT, Schmidt HA, von Haeseler A, Minh BQ. 2015. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. Mol Biol Evol. 32(1):268–274.
- Nilsson MA. 2009. The structure of the Australian and South American marsupial mitochondrial control region. Mitochondrial DNA. 20(5–6): 126–138.
- Paabo S, Thomas WK, Whitfield KM, Kumazawa Y, Wilson AC. 1991. Rearrangements of mitochondrial transfer RNA genes in marsupials. J Mol Evol. 33(5):426–430.
- Roehr JT, Dieterich C, Reinert K. 2017. Flexbar 3.0 SIMD and multicore parallelization. Bioinformatics. 33(18):2941–2942.
- Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics. 19(12):1572–1574.
- Westerman M, Krajewski C, Kear BP, Meehan L, Meredith RW, Emerling CA, Springer MS. 2016. Phylogenetic relationships of Dasyuromorphian marsupials revisited. Zool J Linn Soc. 176(3): 686–701.