idiopathic obstruction to the bile flow. Without prompt

diagnosis, Kasai portoenterostomy (KP), and deliberate follow-

ups, the resulting cholestasis leads to progressive conjugated

hyperbilirubinemia, cirrhosis, and hepatic failure. The BA

# Perioperative reactive oxygen species in infants with biliary atresia

# A retrospective observational study

Naoki Hashizume, MD, PhD<sup>a,\*</sup>, Yoshiaki Tanaka, MD, PhD<sup>a,b</sup>, Kimio Asagiri, MD, PhD<sup>a,c</sup>, Suguru Fukahori, MD, PhD<sup>a</sup>, Shinji Ishii, MD, PhD<sup>a</sup>, Nobuyuki Saikusa, MD, PhD<sup>a</sup>, Motomu Yoshida, MD, PhD<sup>a</sup>, Ken Tanikawa, MD, PhD<sup>d</sup>, Takahiro Asakawa, MD, PhD<sup>a,c</sup>, Minoru Yagi, MD, PhD<sup>a</sup>

# Abstract

Biliary atresia (BA) is a devastating cholestatic disorder of infants that presents during the first several months after birth due to an idiopathic obstruction to the bile flow. Without prompt diagnosis, Kasai portoenterostomy, and deliberate follow-ups, the resulting cholestasis leads to progressive hepatic failure. Oxidative stress is an abnormal phenomenon inside cells or tissues caused by a disturbance in the reactive oxygen species (ROS). We aimed to measure perioperative ROS in BA patients.

Data are presented as median (25th, 75th percentiles). We evaluated 15 BA patients (age 55 [48, 69] days) and measured ROS; serum superoxide dismutase (SOD), urinary 8-iso prostaglandin  $F_{2\alpha}$  (8-iso-PGF<sub>2\alpha</sub>) and 8-hydroxy-2'-deoxyguanosine (8-OHdG) preoperatively and 30 days later to compare values with serum liver function tests and histologic grades of liver cholestasis. For compared BA patients, 4 normal subjects as control group (age 55 [27, 75] days) measured ROS and serum liver function tests.

In BA patients, the preoperative serum SOD was 6.1 IU/mL (4.7, 7.2), urinary 8-iso-PGF<sub>2 $\alpha$ </sub> was 1969 pg/mg Cre (1697, 2374), and urinary 8-OHdG was 37.1 ng/mg Cre (33.1, 53.7). At the postoperative day 30, the serum SOD was 5.2 IU/mL (4.2, 6.7), urinary 8-iso-PGF<sub>2 $\alpha$ </sub> was 1761 pg/mg Cre (1256, 3036), and urinary 8-OHdG was 42.1 ng/mg Cre (29.65, 72.64). In ROS, there were no significant differences between the 2 periods. In control group, urinary 8-iso-PGF<sub>2 $\alpha$ </sub> was significantly lower than that in preoperative BA patient group. However, other ROS were not significant differences between control group and BA patient group. The concentration of urinary 8-iso-PGF<sub>2 $\alpha$ </sub> was positively correlated with total bilirubin and direct bilirubin levels (preoperatively: r = 0.6921, P = .0042 and r = 0.6639, P = .007, postoperatively: r = 0.6036, P = .0172 and r = 0.6464, P = .0092, respectively). The preoperative ROS were not correlated with histologic grades of liver cholestasis. Various factors such as liver inflammation, lipid malabsorption, and tissue disorders due to jaundice might affect the antioxidant activity and elevated urinary 8-iso-PGF<sub>2 $\alpha$ </sub>. However, at least until 30 days later, urinary 8-OHdG as oxidative DNA damage might persist after the operation whether the cholestasis improved or not.

**Abbreviations:** Alb = albumin, ALP = alkaline phosphatase, ALT = alanine aminotransferase, AST = aspartate transaminase, BA = biliary atresia, D-Bil = direct bilirubin, GGT = gamma-glutamyltranspeptidase, 8-iso-PGF<sub>2</sub> $_{\alpha}$  = 8-iso prostagrandin F<sub>2</sub> $_{\alpha}$ , 8-OHdG = 8-hydroxy-2'-deoxyguanosine, ROS = reactive oxygen species, SOD = serum superoxide dismutase, T-Bil = total bilirubin, TP = total protein.

**Keywords:** biliary atresia, 8-hydroxy-2'-deoxyguanosine, 8-iso prostagrandin $F_{2\alpha}$ , oxidative stress, superoxide dismutase

# 1. Introduction

Biliary atresia (BA) is a devastating cholestatic disorder of infants that presents during the first several months after birth due to an

Editor: Somchai Amornyotin.

The authors have no funding and conflicts of interest to disclose.

1

Received: 27 March 2020 / Received in final form: 29 May 2020 / Accepted: 14 June 2020

http://dx.doi.org/10.1097/MD.00000000021332

Hematic, urinary, and hepatic biopsies were obtained after satisfactory informed consent was obtained from these parents. This study was approved by the ethics committee for clinical research of Kurume University School of Medicine (approval no: 2007-008).

All data generated or analyzed during this study are included in this published article [and its supplementary information files].

<sup>&</sup>lt;sup>a</sup> Department of Pediatric Surgery, Kurume University School of Medicine, <sup>b</sup> Division of Medical Safety Management, Kurume University Hospital, <sup>c</sup> Department of Pediatric Surgery, St Mary's Hospital, <sup>d</sup> Departments of Pathology, Kurume University School of Medicine, Kurume, Fukuoka, Japan.

<sup>\*</sup> Correspondence: Naoki Hashizume, Department of Pediatric Surgery, Kurume University School of Medicine, 67 Asahi-machi, Kurume, Fukuoka 830-0011, Japan (e-mail: naoki\_hashidume@med.kurume-u.ac.jp).

Copyright © 2020 the Author(s). Published by Wolters Kluwer Health, Inc.

This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Hashizume N, Tanaka Y, Asagiri K, Fukahori S, Ishii S, Saikusa N, Yoshida M, Tanikawa K, Asakawa T, Yagi M. Perioperative reactive oxygen species in infants with biliary atresia: a retrospective observational study. Medicine 2020;99:31(e21332).



**Figure 1.** Cellular liver damage associated with reactive oxygen species (ROS) production. Overproduction of reactive oxygen species results in oxidative stress. Antioxidant enzymes catalyze the decomposition of ROS. One antioxidant enzyme is superoxide dismutase (SOD), which is a metalloenzyme capable of scavenging superoxides. 8-lso-prostaglandine  $F_{2\alpha}$  (8-iso-PGF<sub>2\alpha</sub>) is generated by the nonenzymic free radical-initiated peroxidation of arachidonic acid which is present in phospholipid membranes. 8-lso-PGF<sub>2\alpha</sub> has been proposed as a valuable biomarker for the assessment of oxidative stress. 8-Hydroxy-2'-deoxyguanosine (8-OHdG) is the product of the oxidation of guanine, an intracellular compound and a component of DNA. 8-OHdG antibody has been widely used to evaluate oxidative DNA damage in animal and human tissues.

patients and end-stage liver disease have no other option but to undergo liver transplantation.

Oxidative stress is an abnormal phenomenon inside cells or tissues caused by a disturbance in the balance between the production of reactive oxygen species (ROS) and both enzymatic and nonenzymatic antioxidants.<sup>[1]</sup> Excess free radicals can damage fatty tissues and DNA leading to abnormal gene expression, with receptor activity and proliferation disturbances or cell death.<sup>[2]</sup>

Antioxidant enzymes in the cells include superoxide dismutase (SOD), glutathione peroxidase, and catalase (Fig. 1). SOD is a metalloenzyme capable of scavenging superoxide radicals by catalyzing their dismutation to hydrogen peroxide and oxygen.<sup>[1,3]</sup> This function appears to protect cells from the toxic effects of the endogenously generated superoxide radicals. SOD is a measure of antioxidant activity.

Arachidonic acid is a ubiquitous polyunsaturated fatty acid present in phospholipid membranes in humans. F<sub>2</sub> isoprostanes are generated by nonenzymatic free radical-initiated peroxidation of arachidonic acid and are frequently used as an accurate measure of oxidative stress during lipid damage.<sup>[4–8]</sup> One isomer of the F<sub>2</sub> isoprostane family, 8-iso-prostaglandin F<sub>2α</sub> (8-iso $PGF_{2\alpha}$ ) has been proposed as a valuable biomarker of phospholipid membranes for oxidative stress assessments.<sup>[1,5,7]</sup> 8-Iso-PGF<sub>2 $\alpha$ </sub> can be detected in plasma, urine, and other biologic fluids.<sup>[4,5,7,8]</sup>

8-Hydroxy-2'-deoxyguanosine (8-OHdG) is a product of the oxidation of guanine, an intracellular component of DNA. 8-OHdG is a relatively stable substance outside the cells produced during DNA repair processes and gets excreted through the blood into urine.<sup>[1]</sup> Urinary 8-iso-PGF2a and urinary 8-OHdG are measures of ROS (Fig. 1).

In another study, we found that postoperative BA patients and a median age of 5.2 years had significantly increased production of ROS, including higher serum SOD, urine 8-iso-PGF<sub>2 $\alpha$ </sub>, and 8-OHdG, than individuals without BA.<sup>[9]</sup> However, studies on serum and urinary ROS in the pediatric population with liver disease are few,<sup>[9–11]</sup> and those measuring serum SOD, urinary 8iso-PGF<sub>2 $\alpha$ </sub>, and urinary 8-OHdG levels are even more scarce. Moreover, we know of no reports of the assessment of ROS during the perioperative BA period.

This study aimed to assess the serum and urinary OSAs during the perioperative BA period focusing on serum SOD, urinary 8-iso-PGF<sub>2 $\alpha$ </sub>, and urinary 8-OHdG levels.

## 2. Materials and methods

## 2.1. Patients

We enrolled 15 patients (5 boys and 10 girls) with BA and 4 subjects (3 boys and 1 girl) with an umbilical hernia as the control group who were patients from 2006 to 2017 at the Kurume University School of Medicine. In this period, we excluded four patients, 1 with BA operated during day 4, 1 with BA and intracranial hemorrhage due to vitamin K malabsorption, and 2 with sepsis after the operations.

#### 2.2. Study design

In 15 BA patients, preoperative sample collections were performed at days 1 or 0 and the postoperative serum sample collections on day 30, defining the day of the operation as day 0. We measured aspartate transaminase (AST), alanine aminotransferase (ALT), total protein (TP), albumin (Alb), alkaline phosphatase (ALP), gamma-glutamyltranspeptidase (GGT), total bilirubin (T-Bil), direct bilirubin (D-Bil), cholinesterase, and total biliary acid levels. In addition, we obtained type IV collagen levels during the preoperative period.

We also measured the following ROS: serum SOD, urinary 8iso-PGF<sub>2 $\alpha$ </sub>, and urinary 8-OHdG. Serum SOD was measured by nitrous transformation (Roche Diagnostics, Tokyo, Japan), urinary 8-iso-PGF<sub>2 $\alpha$ </sub> by an enzymatic immune-assay (8-Isoprostane EIA kit; Cayman Chemical Co, Ann Arbor, MI), and urinary 8-OHdG by an enzymatic immunoassay (ELISA System kit; Institute of Senescence Control of Japan, Shizuoka, Japan). Liver biopsies were performed during the KP procedures, and a pathologist (KT) examined the specimens after fixing them in formalin and staining them with hematoxylin/eosin. Preoperatively, all the BA patients fed breast milk and milk included medium chain triglyceride.

In 4 patients as control group, we measured AST, ALT, TP, Alb, ALP, GGT, T-Bil, D-Bil, serum SOD, urinary 8-iso-PGF<sub>2 $\alpha$ </sub>, and urinary 8-OHdG. All patients in control group fed breast milk.

To correlate histologic findings with liver cholestasis disease, we used a published grading system,<sup>[12,13]</sup> and our preliminary experiment results were based on 4 pathologic parameters including the presence of a portal area bile plug, cholestasis, hepatocyte degeneration, and portal area inflammatory cell infiltration. We used a 12-point histologic scoring system to score the severity of liver cholestasis disease. We classified portal ductal bile plugs into 3 grades: grade 0, bile plug absent; grade 1, bile plugs present without bile lake formation; and grade 2, bile plugs present with bile lake formations. We also classified cholestasis into 3 grades based on the scope of bile pigment deposition and distribution: grade 1, mild cholestasis (pigment deposition within the bile capillary); grade 2, moderate cholestasis (pigment deposition in the small bile duct or hepatocytes); and grade 3, severe cholestasis (diffuse bile pigment deposition in the lobular bile capillary and small bile ductules). We divided hepatocyte degeneration into four grades: grade 1, swollen; grade 2, ballooning degeneration; grade 3, multinuclear hepatocyte formation; and grade 4, necrosis. Additionally, we subclassified portal area inflammatory cell infiltration into 3 categories: stage 1, portal area inflammatory cell infiltration <20%; stage 2, portal area inflammatory cell infiltration <50% without lymphocyte accumulation; and stage 3, portal area inflammatory cell infiltration (including lymphocytes) >50%.

We divided patients into 2 groups: group A (n=8) comprised patients with T-bil  $\geq 2.0$  mg/dL on postoperative day 30 and group B (n=7) comprised patients with T-bil <2.0 mg/dL on postoperative day 30.

#### 2.3. Ethical approval and studies and informed consent

We obtained serum, urinary, and hepatic biopsies after obtaining written informed consents from the patients' guardians. The ethics committee for clinical research of Kurume University School of Medicine approved this study (approval number 2007-008).

#### 2.4. Statistical analysis

Data are presented as median (25th, 75th percentiles). First, the comparison between preoperative BA patients group and control group were performed. Second, the comparison between preoperative and postoperative periods in BA patients was performed. Finally, the comparisons between the 2 subgroups; group A and group B were performed. All comparisons were using the Mann–Whitney U and Chi-squared tests. The Spearman rank correlation analysis to assess the association between liver function tests and ROS and to evaluate the correlation between the labeling index and the grades was used. All of the statistical analyses were performed using the JMP Pro 14 software package (SAS, Cary, NC), and *P* values of <.05 were considered statistically significant.

#### 3. Results

Table 1 shows the characteristics of BA patients and normal subjects. During the preoperative period, all BA patients showed jaundice and cholestasis liver injury. At the postoperative day 30, AST, T-bil, and D-bil levels were significantly lower than those in the preoperative period in all cases, respectively. There were no significant differences between the other preoperative and postoperative liver function test. There were no significant differences between the preoperative ROS.

In age and gender, there was no difference between BA patient and control groups. At the control group, AST, ALT, Alb, GGT, T-bil, and D-bil levels were significantly lower than BA patient group in the preoperative period, respectively. At the control group, urinary 8-iso-PGF<sub>2 $\alpha$ </sub> was significantly lower than at BA patient group. The serum SOD and urinary 8-OHdG values at the control group were marginally lower than those at BA patient group, but there were not significant differences.

Preoperatively, the concentration of urinary 8-iso-PGF<sub>2α</sub> was positively correlated with T-bil and D-bil levels (r=0.6921, P=.0042; and r=0.6639, P=.007, respectively; Fig. 2). No other preoperative liver function tests were correlated with OSA. At postoperative day 30, the concentration of urinary 8-iso-PGF<sub>2α</sub> was positively correlated with T-bil and D-bil levels (r=0.6036, P=.0172 and r=0.6464, P=.0092, respectively; Fig. 3). No other postoperative liver function tests were correlated with ROS.

Table 2 shows the characteristics comparison of BA patients in terms of between group A and group B. There were no significant differences in terms of preoperative liver function test findings between the group A and group B. At postoperative day 30, T-Bil and D-Bil were significantly higher in group A than in the group B. In group A, there were no significant differences between the preoperative and postoperative liver function tests. In group B at

# Table 1

#### The characteristics of BA patients and control groups.

	BA patients group (n = 15)			
	Day 0	Day 30	Control group (n=4)	
Age, d	55 (48, 69)		55 (27, 75)	
Gender, male/female	10/5	3/1		
Liver function test				
AST, IU/L	172 (133, 304)	115 (83.2, 179) <sup>*</sup>	36 (28.3, 45.3) <sup>†</sup>	
ALT, IU/L	115 (66, 174)	118 (106, 152)	20.5 (14, 25.3) <sup>†</sup>	
TP, g/dL	5.49 (5.26, 5.85)	5.94 (5.73, 6.28)	5.82 (5.67, 5.89)	
Alb, g/dL	3.66 (3.41, 4.02)	3.76 (3.62, 3.99)	4.14 (3.95, 4.17)*	
GGT, IU/L	380 (217, 943)	775 (530, 1106)	46 (32.8, 88.5) <sup>†</sup>	
T-Bil, mg/dL	7.69 (7.06, 10.33)	2.16 (1.12, 5.84) <sup>†</sup>	1.08 (0.42, 1.78) <sup>†</sup>	
D-Bil, mg/dL	5.57 (4.83, 7.54)	1.20 (0.47, 4.10) <sup>†</sup>	0.13 (0.06, 0.19) <sup>†</sup>	
ALP, IU/L	1315 (1076, 1923)	1235 (919, 1528)	1170 (989, 1684)	
TBA, mg/dL	114.9 (95.9, 141.6)	98.8 (52.2, 142)	_	
CHE, IU/L	284 (179, 328)	342 (240, 393)	_	
Reactive oxygen species				
Serum SOD, IU/mL	6.1 (4.7, 7.2)	5.2 (4.2, 6.7)	3.3 (2.9, 6.5)	
Urinary 8-iso-PDF <sub>2a</sub> , pg/mg Cre	1969 (1697, 2374)	1761 (1256, 3036)	1025 (580, 1496)*	
Urinary 8-OHdG, ng/mg Cre	37.1 (33.1, 53.7)	42.1 (29.65, 72.64)	27.0 (19.4, 93.0)	

Data are median (1st and 3rd quartiles).

 $AST = aspartate transaminase, ALT = alanine aminotransferase, BA = biliary atresia, TP = total protein, Alb = albumin, GGT = gamma-glutamyltranspeptidase, T-Bil = total bilirubin, D-Bil = direct bilirubin, ALP = alkaline phosphatase, TBA = total biliary acid, CHE = cholinesterase, SOD = serum superoxide dismutase, 8-iso-PGF<sub>2</sub><sub>\alpha</sub>=8-iso prostagrandin F<sub>2</sub><sub>\alpha</sub>, 8-OHdG = 8-hydroxy-2'-deoxyguanosine.$ 

\* *P*<.05 vs day 0.

 $^{+}P < .01$  vs day 0.

the postoperative day 30, the AST, T-bil, and D-bil levels were significantly lower than at the preoperative period, respectively.

Table 3 shows the characteristics comparison between group A and group B. There were no significant differences in terms of preoperative ROS between group A and group B. At postoperative day 30, urinary 8-iso-PGF<sub>2α</sub> was significantly higher in group A than in group B. However, the serum SOD urinary 8-OHdG values were similar in group A and group B. In each group, there were no significant differences between the preoperative and postoperative ROS.

At the pathologic parameters of BA patients, according to the histologic grades and scores, the bile plugs in portal ductules was classified as 1 (1, 1). All patients had severe cholestasis with diffused bile pigment deposition in the lobular bile capillaries as well as in the small bile duct and were classified the grade of all patients' cholestasis as grade 3. The hepatocyte degeneration was 3 (2, 4), the portal area inflammatory cell infiltration was 1 (1, 2), and the total scored points were 8 (7, 9.25) respectively. Preoperative OSA did not correlate with the grade or scores of histologic findings.



**Figure 2.** Correlation between preoperative urinary 8-iso-prostaglandine  $F_{2\alpha}$  (8-iso-PGF<sub>2 $\alpha$ </sub>) and T-bil (solid line), and urinary 8-iso-PGF<sub>2 $\alpha$ </sub> and D-bil (dotted line) (Spearman rank correlation; r=0.6921, P=.0042 and r=0.6639, P=.007).



**Figure 3.** Correlation between postoperative urinary 8-iso-prostaglandine  $F_{2\alpha}$  (8-iso-PGF<sub>2\alpha</sub>) and T-bil (solid line), and between urinary 8-iso-PGF<sub>2\alpha</sub> and D-bil (dotted line) (Spearman rank correlation; r=0.6036, P=.0172 and r=0.6464, P=.0092).

# Table 2

Comparison of the liver function tests between biliary atresia (BA) patients with T-bil  $\geq$  2.0 mg/dL (group A) and BA patients with T-bil < 2.0 mg/dL (group B) on postoperative day 30.

	Group A (n=8)	Group B (n=7)	Р
Age at operation, d	55 (49, 67)	55 (47, 74)	1.000
Gender, male/female	4/4	1/6	.143
Liver function test	Day 0		
AST, IU/L	185 (140, 329)	150 (104, 183)	.385
ALT, IU/L	137.5 (68, 230.3)	102 (58, 166)	.685
TP, g/dL	5.75 (5.35, 5.97)	5.31 (5.24, 5.69)	.272
Alb, g/dL	3.91 (3.46, 4.06)	3.59 (3.37, 3.77)	.385
GGT, IU/L	331 (210, 760)	782 (217, 1030)	.224
T-Bil, mg/dL	7.64 (5.95, 9.74)	7.75 (7.06, 12.6)	.603
D-Bil, mg/dL	5.59 (4.27, 7.08)	5.48 (4.83, 8.91)	.685
TBA, mg/dL	118.5 (98, 140.6)	112 (84.4, 158.7)	.954
ALP, IU/L	1357 (1199, 1860)	1315 (1071, 1895)	.524
CHE, IU/L	298 (188, 341)	280 (151, 299)	.443
Type 4 collagen, ng/mL	22 (19.5, 27.3)	17 (17, 26)	.429
	Day 30		
AST, IU/L	125 (95, 226)	99 (78, 121) <sup>*</sup>	.125
ALT, IU/L	117 (98, 159)	117 (97, 148)	.609
TP, g/dL	5.90 (5.42, 6.44)	6.02 (5.79, 6.28)	.602
Alb, g/dL	3.66 (3.43, 3.94)	3.79 (3.75, 4.31)	.224
GGT, IU/L	693 (425, 1031)	869 (536, 1548)	.307
T-Bil, mg/dL	5.66 (4.93, 7.11)	1.26 (0.98, 1.75) <sup>*</sup>	.002
D-Bil, mg/dL	3.64 (3.11, 5.15)	0.51 (0.33, 0.71) <sup>*</sup>	.002
TBA, mg/dL	134 (112, 148)	56.9 (22.3 86.9)	.055
ALP, IU/L	1350 (901, 1537)	1138 (700, 1340)	.307
CHE, IU/L	325 (216, 370)	352 (277, 437)	.443

Data are median (1st and 3rd quartiles).

 $\begin{array}{l} \text{AST} = \text{aspartate transaminase, ALT} = \text{alanine aminotransferase, TP} = \text{total protein, Alb} = \text{albumin,} \\ \text{GGT} = \text{gamma-glutamyltranspeptidase, T-Bil} = \text{total bilirubin, D-Bil} = \text{direct bilirubin, ALP} = \text{alkaline} \\ \text{phosphatase, TBA} = \text{total biliary acid, CHE} = \text{cholinesterase, SOD} = \text{serum superoxide dismutase, 8-} \\ \text{iso-PGF}_{2\alpha} = 8\text{-}\text{iso prostagrandin F}_{2\alpha}, 8\text{-OHdG} = 8\text{-hydroxy-2'-deoxyguanosine.} \end{array}$ 

<sup>™</sup> P<.05 vs day 0.

Table 4 shows the comparison of pathologic parameters between group A and group B. Hepatocyte degeneration was marginally higher in group A than in group B. Portal area inflammatory cell infiltration and the total score was significantly higher in group A than in group B, respectively.

#### 4. Discussion

Oxidative stress is known to be involved in the pathogenesis of BA.<sup>[10,14,15]</sup> SOD in liver tissue is high in infants with cholestatic liver disease such as BA and Alagille syndrome. Moreover, a significant correlation was found between the liver SOD activity and serum bilirubin in BA patients.<sup>[14]</sup> In patients with cholestatic BA, neonatal hepatitis, and paucity of intrahepatic bile ducts, the levels of SOD, glutathione peroxidase, and catalase in hepatic tissue are significantly increased compared with those in normal liver biopsies.<sup>[10]</sup> Elevated ROS in the liver reflected hepatocyte damage and metabolized in the liver.<sup>[10]</sup> Tiao et al reported that the number of 8-OHdG positive hepatocytes was significantly higher in the preoperative liver biopsy of BA patients when the patients had undergone a KP than when they had undergone liver transplantation and choledochal cyst.<sup>[15]</sup> The early BA stage is associated with a strong inflammatory reaction, with high oxidative DNA included mitochondrial DNA damage (as manifested by higher immunoreactivity, 8-OHdG, and apoptotic

# Table 3

Comparison of the reactive oxygen species between biliary atresia
(BA) patients with T-bil $\geq$ 2.0 mg/dL (group A) and BA patients with
T-bil < 2.0 mg/dL (group B) on postoperative day 30.

	Group A (n=8)	Group B (n=7)	Р
Reactive oxygen species	Day 0		
Serum SOD, IU/mL	6.15 (4.8, 7.2)	6.1 (4.7, 8.4)	.8168
Urinary 8-iso-PDF <sub>2a</sub> , pg/mg Cre	2137 (1827, 2428)	1856 (1245, 2265)	.3854
Urinary 8-OHdG, ng/mg Cre	37.1 (30.8, 53.7)	37.4 (33.5, 70.0)	.7723
	Day	30	
Serum SOD, IU/mL	6.25 (4.48, 8.65)	4.2 (3.4, 5.9)	.1828
Urinary 8-iso-PDF <sub>2a</sub> , pg/mg Cre	2960 (1788, 3628)	1256 (1078, 1761)	.0092
Urinary 8-OHdG, ng/mg Cre	48.45 (37.17, 102.4)	41.95 (28.6, 57.9)	.4519

Data are median (1st and 3rd quartiles).

SOD=serum superoxide dismutase, 8-iso-PGF<sub>2 $\alpha$ </sub>=8-iso prostagrandin F<sub>2 $\alpha$ </sub>, 8-0HdG=8-hydroxy-2'-deoxyguanosine.

activities), and with a decrease in mitochondrial copy numbers. In our study, we found no significant differences between the preoperative and postoperative ROS. The serum SOD and urinary 8-OHdG values at BA patient group were marginally high, but there were not significant differences between control group and BA patient group. Moreover, despite recovery from jaundice, the serum SOD and urinary 8-OHdG levels had not decreased postoperatively at day 30. Our findings suggest that, at least until 30 days later, oxidative DNA included mitochondrial DNA damage persisted after the operation whether the cholestasis improved or not.

Preoperatively, the concentration of urinary 8-iso-PGF<sub>2α</sub> was positively correlated with both T-bil and D-bil levels. Moreover, postoperatively, urinary 8-iso-PGF<sub>2α</sub> levels in group A were significantly higher than in group B. Our results suggest that urinary 8-iso-PGF<sub>2α</sub>, reflecting a comprehensive spectrum of lipid peroxidation and antioxidant features, is a significant marker of cholestasis with BA. However, it was suspected that the elevation of urinary 8-iso-PGF<sub>2α</sub> was not specific to BA and present in all forms of cholestasis. Our findings suggest that production of endogenous steroids and fat-soluble vitamins inhibit the action of the arachidonic cascade.<sup>[16,17]</sup> In addition, breast milk has important antioxidant properties,<sup>[18,19]</sup> and improvement of jaundice leads to an increase in milk intake. Improvement in lipid

#### Table 4

Comparison of the pathologic parameters between biliary atresia (BA) patients with T-bil  $\geq$  2.0 mg/dL (group A) and BA patients with T-bil < 2.0 mg/dL (group B) on postoperative day 30.

	Group A (n=8)	Group B (n=7)	Р
Pathologic parameters			
Bile plugs in portal ductules	1.0 (0.75, 1)	1 (1,1)	.9154
Cholestasis	3 (3, 3)	3 (3, 3)	1.0000
Hepatocyte degeneration	3.5 (3, 4)	2 (2, 3.25)	.0753
Inflammatory cell infiltration in portal area	2 (1, 2)	1 (1, 1)	.0268
Scored point	10 (9, 11)	8 (7, 9.25)	.0305

Data are median (1st and 3rd quartiles)

T-Bil=total bilirubin.

absorption and increase in bile emissions may be affected by the decreased lipid peroxidation.

In our study, preoperative production of ROS did not correlate with histologic grades and scores. In the ligated-bile duct rat model, Baron and Muriel found that although antioxidant treatment inhibits lipid peroxidation and glutathione oxidation, it has no effect on conventional biochemical markers of liver injury or liver histology.<sup>[20]</sup> The authors speculated that serum and urinary ROS were not correlated with pathologic findings because oxidative stress does not play a causative role in this model of cholestatic liver disease but is instead a consequence of bile salts inducing solubilization of plasma membranes and cell death.<sup>[20]</sup>

We are aware of the limitations of our study. Our analysis was limited due to the small population and the fact that it was based on a single institution. We measured only total SOD activity. Obtaining measurements of subgroups like Mn-SOD (mitochondrial origin) and the Cu-Zu-containing protein in the cytoplasm may provide further information as to whether abnormalities in oxygen free radical metabolism cause or simply reflect tissue injury. Serum and urinary ROS may still be important and useful for appropriate follow-ups of postoperative patients. There were preliminary studies about standard values of serum and urinary ROS for infants.<sup>[5,9,21,22]</sup> Calculation standard values of serum and urinary ROS for infants is difficult because the standard values change as aging. In terms of the pathologic findings, the liver biopsy specimens were only stained with hematoxylin/eosin and were not evaluated for liver fibrosis. Moreover, extending this study to the include BA outcomes for patients with their native liver in the long term may be a future possibility.

In conclusion, our findings suggest that, in BA patients, urinary 8-iso-PGF<sub>2 $\alpha$ </sub> is high and greatly affected by serum bilirubin. However, serum and urinary ROS were not correlated with pathologic findings. Serum and urinary ROS failed to predict the severity of the preoperative liver injury in BA patients. Various factors such as liver inflammation, lipid malabsorption, and tissue disorders due to jaundice might affect the antioxidant activity and elevated urinary 8-iso-PGF<sub>2 $\alpha$ </sub>. However, at least until 30 days later, urinary 8-OHdG as oxidative DNA damage might persist after the operation whether the cholestasis improved or not.

#### Acknowledgment

The authors thank enago (https://www.enago.jp) for English language editing.

#### **Author contributions**

Conceptualization: Yoshiaki Tanaka.

- Data curation: Ken Tanikawa.
- Investigation: Suguru Fukahori, Shinji Ishii, Nobuyuki Saikusa, Motomu Yoshida, Takahiro Asakawa.
- Supervision: Kimio Asagiri, Takahiro Asakawa.

Writing - original draft: Naoki Hashizume.

Writing - review & editing: Yoshiaki Tanaka, Minoru Yagi.

## References

- Tsukahara H. Biomarkers for oxidative stress: clinical application in pediatric medicine. Curr Med Chem 2007;14:339–51.
- [2] Aboutwerat A, Pemberton PW, Smith A, et al. Oxidant stress is a significant feature of primary biliary cirrhosis. Biochim Biophys Acta 2003;1637:142–50.
- [3] Southorn PA, Powis G. Free radicals in medicine. I. Chemical nature and biologic reactions. Mayo Clin Proc 1988;63:381–9.
- [4] Awad JA, Roberts LJ, Burk RF, et al. Isoprostanes-prostaglandin-like compounds formed in vivo independently of cyclooxygenase: use as clinical indicators of oxidant damage. Gastroenterol Clin North Am 1996;25:409–27.
- [5] Kauffman LD, Sokol RJ, Jones RH, et al. Urinary F2-isoprostanes in young healthy children at risk for type 1 diabetes mellitus. Free Radic Biol Med 2003;35:551–7.
- [6] Milne GL, Morrow JD. Isoprostanes and related compounds: update 2006. Antioxid Redox Signal 2006;8:1379–84.
- [7] Basu S. F2-isoprostanes in human health and diseases: from molecular mechanisms to clinical implications. Antioxid Redox Signal 2008; 10:1405–34.
- [8] Keskin O, Uluca U, Keskin M, et al. The efficacy of single-high dose inhaled corticosteroid versus oral prednisone treatment on exhaled leukotriene and 8-isoprostane levels in mild to moderate asthmatic children with asthma exacerbation. Allergol Immunopathol (Madr) 2016;44:138–48.
- [9] Asakawa T, Tanaka Y, Asagiri K, et al. Oxidative stress profile in the post-operative patients with biliary atresia. Pediatr Surg Int 2009;25: 93–7.
- [10] Ismail NA, Okasha SH, Dhawan A, et al. Antioxidant enzyme activities in hepatic tissue from children with chronic cholestatic liver disease. Saudi J Gastroenterol 2010;16:90–4.
- [11] El-Shabrawi MH, Kamal NM, Halawa FA, et al. Serum superoxide dismutase activity in acute and chronic paediatric liver diseases. Arab J Gastroenterol 2014;15:72–5.
- [12] Chen CJ, Tang KS, Huang YH, et al. The characteristics of antioxidant activity after liver transplantation in biliary atresia patients. Biomed Res Int 2015;2015: 421413.
- [13] Lee WS, Chai PF. Clinical features differentiating biliary atresia from other causes of neonatal cholestasis. Ann Acad Med Singapore 2010;39:648–54.
- [14] Broide E, Klinowski E, Koukoulis G, et al. Superoxide dismutase activity in children with chronic liver diseases. J Hepatol 2000;32:188–92.
- [15] Tiao MM, Lin TK, Kuo FY, et al. Early stage of biliary atresia is associated with significant changes in 8-hydroxydeoxyguanosine and mitochondrial copy number. J Pediatr Gastroenterol Nutr 2007;45: j329–34.
- [16] Kim KR, Kim MJ, Rhee SJ. Effects of vitamin E on arachidonic acid cascade in platelets and aorta of acute cadmium-poisoned rats. Nutr Res 2001;21:657–65.
- [17] Flower RJ, Blackwell GJ. Anti-inflammatory steroids induce biosynthesis of a phospholipase A2 inhibitor which prevents prostaglandin generation. Nature 1979;278:456–9.
- [18] Aycicek A, Erel O, Kocyigit A, et al. Breast milk provides better antioxidant power than does formula. Nutrition 2006;22:616–9.
- [19] Vohr BR, Poindexter BB, Dusick AM, et al. Beneficial effects of breast milk in the neonatal intensive care unit on the developmental outcome of extremely low birth weight infants at 18 months of age. Pediatrics 2006;118:e115–23.
- [20] Baron V, Muriel P. Role of glutathione, lipid peroxidation and antioxidants on acute bile-duct obstruction in the rat. Biochim Biophys Acta 1999;1472:173–80.
- [21] Shoji H, Ikeda N, Hosokawa M, et al. Oxidative stress early in infancy and neurodevelopmental outocome in very low-birthweight infants. Pediatr Int 2014;56:709–13.
- [22] Fukuda M, Yamauchi H, Yamamoto H, et al. The evaluation of oxidative DNA damage in children with brain damage using 8hydroxydeoxyguanosine levels. Brain Dev 2008;30:131–6.