# Surveillance of OXA-244-producing Escherichia coli and epidemiologic investigation of cases, Denmark, January 2016 to August 2019

Anette M Hammerum<sup>1</sup>, Lone Jannok Porsbo<sup>2</sup>, Frank Hansen<sup>1</sup>, Louise Roer<sup>1</sup>, Hülya Kaya<sup>1</sup>, Anna Henius<sup>1</sup>, Karina Lauenborg Møller<sup>3</sup>, Ulrik S Justesen<sup>4</sup>, Lillian Søes<sup>5</sup>, Bent L Røder<sup>6</sup>, Philip K Thomsen<sup>7</sup>, Mikala Wang<sup>8</sup>, Turid Snekloth Søndergaard<sup>9</sup>, Barbara Juliane Holzknecht<sup>10</sup>, Claus Østergaard<sup>11</sup>, Anne Kjerulf<sup>2</sup>, Brian Kristensen<sup>2</sup>, Henrik Hasman<sup>1</sup> 1. Department of Microbiology and Infection Control, Statens Serum Institut, Copenhagen, Denmark

- Infectious Disease Epidemiology & Prevention, Statens Serum Institut, Copenhagen, Denmark
   Data Integration and Analysis, Statens Serum Institut, Copenhagen, Denmark

- Department of Clinical Microbiology, Journal Microbiology, Journal Microbiology, Journal Microbiology, Heidovre University Hospital, Odense, Denmark
   Department of Clinical Microbiology, Hvidovre University Hospital, Hvidovre, Denmark
   Department of Clinical Microbiology, Zealand University Hospital, Slagelse, Denmark
   Department of Clinical Microbiology, Aalborg University Hospital, Aalborg, Denmark
   Department of Clinical Microbiology, Aarhus University Hospital, Aalborg, Denmark
   Department of Clinical Microbiology, Hospital Sønderjylland, Sønderborg, Denmark
   Department of Clinical Microbiology, Hospital Sønderjylland, Sønderborg, Denmark

- Department of Clinical Microbiology, Herlev and Gentofte University Hospital, Herlev, Denmark
   Department of Clinical Microbiology, Lillebaelt Hospital, Vejle, Denmark

#### Correspondence: Anette M. Hammerum (ama@ssi.dk)

#### Citation style for this article:

Hammerum Anette M, Porsbo Lone Jannok, Hansen Frank, Roer Louise, Kaya Hülya, Henius Anna, Møller Karina Lauenborg, Justesen Ulrik S, Søes Lillian, Røder Bent L, Thomsen Philip K, Wang Mikala, Søndergaard Turid Snekloth, Holzknecht Barbara Juliane, Østergaard Claus, Kjerulf Anne, Kristensen Brian, Hasman Henrik. Surveillance of OXA-244-producing Escherichia coli and epidemiologic investigation of cases, Denmark, January 2016 to August 2019. Euro Surveill. 2020;25(18):pii=1900742. https://doi.org/10.2807/1560-7917.ES.2020.25.18.1900742

Article submitted on 10 Dec 2019 / accepted on 23 Mar 2020 / published on 07 May 2020

Background: Carbapenemase-producing Escherichia coli are increasing worldwide. In recent years, an increase in OXA-244-producing E. coli isolates has been seen in the national surveillance of carbapenemase-producing organisms in Denmark. Aim: Molecular characterisation and epidemiological investigation of OXA-244-producing E. coli isolates from January 2016 to August 2019. Methods: For the epidemiological investigation, data from the Danish National Patient Registry and the Danish register of civil registration were used together with data from phone interviews with patients. Isolates were characterised by analysing whole genome sequences for resistance genes, MLST and core genome MLST (cgMLST). Results: In total, 24 OXA-244-producing E. coli isolates were obtained from 23 patients. Among the 23 patients, 13 reported travelling before detection of the E. coli isolates, with seven having visited countries in Northern Africa. Fifteen isolates also carried an extended-spectrum beta-lactamase gene and one had a plasmid-encoded AmpC gene. The most common detected sequence type (ST) was ST<sub>3</sub>8, followed by ST69, ST167, ST10, ST361 and ST3268. Three clonal clusters were detected by cgMLST, but none of these clusters seemed to reflect nosocomial transmission in Denmark. Conclusion: Import of OXA-244 E. coli isolates from travelling abroad seems likely for the majority of cases. Community sources were also possible, as many of the patients had no history of hospitalisation and many of the E. coli isolates belonged to STs

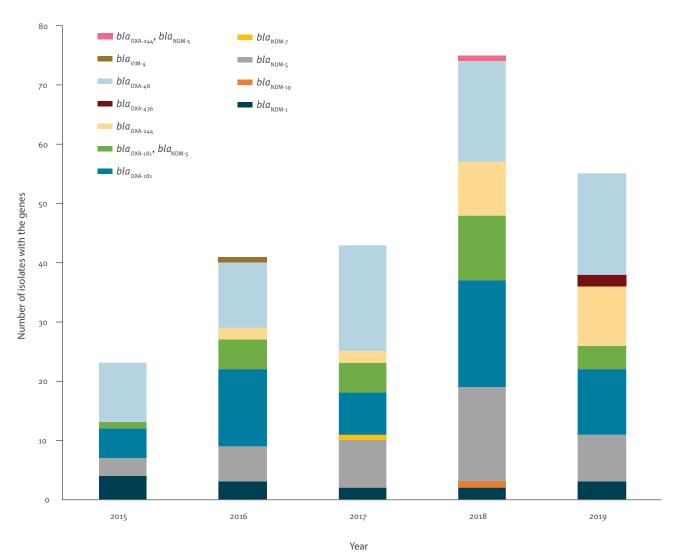
that are present in the community. It was not possible to point at a single country or a community source as risk factor for acquiring OXA-244-producing E. coli.

## Introduction

Carbapenems are used for treatment of infections with multi-resistant Gram-negative bacteria, e.g. extendedspectrum beta-lactamase (ESBL)-producing Escherichia *coli*. Carbapenemase production can be caused by the presence of various carbapenemases. One of the frequently detected carbapenemases in Europe is OXA-48. Besides OXA-48, 17 other variants belong to the OXA-48 carbapenemase group) [1] including OXA-244. In comparison to OXA-48, OXA-244 has a single amino acid substitution (Arg-222-Gly) and has reduced carbapenemase activity [2]. It was first described in Spain from a *Klebsiella pneumoniae* isolate in 2013 [2]. Subsequently, OXA-244-producing *E. coli* isolates were reported from patients in the United Kingdom (UK), France and Egypt, from a healthy person in Germany, from a Dutch traveller and her spouse visiting Indonesia, from river water in Algeria and from an estuary in Lebanon [3-8]. OXA-244-producing E. *coli* can be a challenge for clinical laboratories as they may not grow on selective media used for detection of carbapenemase producers or may not be detected by carbapenemase-specific tests [4]. Furthermore, OXA-244-producing E. coli isolates can have low minimal inhibitory concentrations (MIC) for temocillin and meropenem [4,9].

## FIGURE 1

Carbapenemase-producing Escherichia coli, Denmark, January 2015-August 2019 (n = 237)



On 18 February 2020, the European Centre for Disease Prevention and Control (ECDC) published a Rapid Risk Assessment (RRA) on the increase in OXA-244-producing *E. coli*, including in Denmark [10]. Before the invitation to contribute data to the RRA, the national reference laboratory at Statens Serum Institute (SSI), Copenhagen had already detected an increase in OXA-244-producing *E. coli* in Denmark, whereas *bla* <sub>OXA-244</sub> was not detected from other carbapenemase-producing Enterobacterales.

In this study, we characterised by whole genome sequencing (WGS), all OXA-244-producing *E. coli* isolates submitted to SSI during 2016 through July 2019. Data from the Danish National Patient Registry were used for epidemiological investigation together with data from phone interviews with patients infected/ colonised with OXA-244-producing *E. coli* isolates from January 2018 through July 2019. Our aim was to investigate if the increase in OXA-244-producing *E. coli* was related to hospital outbreaks or was related to travel from abroad.

# Methods

### Setting

Before 2018, Danish departments of clinical microbiology (DCM) submitted on voluntary basis, carbapenemase-producing organisms (CPO) for verification and genotyping at the national reference laboratory at SSI [11]. On 5 September 2018, the Danish Health Authority made CPO notifiable and published a national guideline to prevent the spread of CPO, including criteria for screening for CPO colonisation after hospitalisation abroad [12]. The CPO isolates are both from screening, e.g. patients travelling abroad or during outbreak investigations, and clinical samples from hospitals and primary care facilities detected during routine susceptibility testing.

Screening isolates are obtained using faecal material from a faecal swabs added to a selective Brain heart infusion (BHI) broth with 0.25 mg/L meropenem. After overnight incubation (18-24 hours) at 35°C, 10  $\mu$ l from the BHI broth are plated in a selective agar e.g. chro-mID CARBA SMART (bioMérieux, Ballerup, Denmark)

#### TABLE 1

Sample information, demographic data and travel history for patients with OXA-244-producing *Escherichia coli*, Denmark, January 2016–August 2019 (n = 23)

Patient number	Sample date	Sample	Sample site	Region	Age group (years)ª	Sex	Travel⁵
1	May 2016	Urine	General practitioner	Capital	31-50	Female	No travel
2	Nov 2016	Urine	Emergency department	Zealand	51-65	Male	No information
3	Jun 2017	Urine	Hospital	Southern Denmark	31-50	Female	No travel
4	Dec 2017	Urine	General practitioner	Southern Denmark	51-65	Female	No information
5	Feb 2018	Blood	Hospital	Southern Denmark	51-65	Female	Greece
6	Feb 2018	Urine	General practitioner	Zealand	> 65	Female	No
7	May 2018	Faeces	Hospital	Capital	> 65	Female	Egypt
8	Jun 2018	Urine	General practitioner	Capital	3-10	Female	No information
9	Jun 2018	Urine	Hospital	Central Denmark	> 65	Female	No travel
10	Aug 2018	Urine	General practitioner	Southern Denmark	> 65	Female	Tunisia
11	Sep 2018	Blood	Hospital	Zealand	> 65	Male	Poland, Austria
12	Sep 2018	Faeces	Emergency department	Capital	31-50	Female	Egypt, Turkey
13	Oct 2018	Urine	General practitioner	North Denmark	31-50	Female	No travel
14	Feb 2019	Urine	Hospital	Capital	> 65	Female	Egypt
15	Mar 2019	Faeces	Emergency department	Capital	> 65	Male	India
16	Apr 2019	Urine	General practitioner	Central Denmark	> 65	Female	Egypt, Turkey, Italy
17	May 2019	Urine	Hospital	Capital	3-10	Female	Sweden, Portugal
18	May 2019	Urine	General practitioner	Southern Denmark	60	Female	Germany
19	Jun 2019	Urine	General practitioner	Southern Denmark	31-50	Female	Egypt
20	May 2019	Urine	General practitioner	Central Denmark	> 65	Female	No
21	Jun 2019	Urine	General practitioner	Southern Denmark	> 65	Female	Egypt
22	Jun 2019	Urine	Hospital	Central Denmark	51-65	Female	No information
23	Jul 2019	Urine	Hospital	Southern Denmark	> 65	Female	Spain

<sup>a</sup> The age of the patients ranged from 5 to 79 years, with a median of 58 years.

<sup>b</sup> Travel 6 months before the OXA-244-producing *Escherichia coli* was detected.

and incubated for 18–24 h at 35°C [13]. Isolates (from clinical and screenings samples) suspected to be carbapenemase-producing according to the European Committee on Antimicrobial Susceptibility Testing (EUCAST) guidelines [14,15] are submitted to SSI from the Danish DCM [11].

#### Samples

From January 2016 to August 2019, 24 OXA-244producing *E. coli* isolates from 23 patients were detected in Denmark as part of the national surveillance of CPO. For patients from whom more than one OXA-244-producing *E. coli* isolate was reported within a rolling 12-month period, only the first isolate was included in the study. From a single patient, two isolates were included as the second isolate carried an additional carbapenemase gene. The numbers of OXA-244 producing *E. coli* isolates were compared with the total numbers of carbapenemase-producing *E. coli* during the same period.

## Epidemiological data and sources

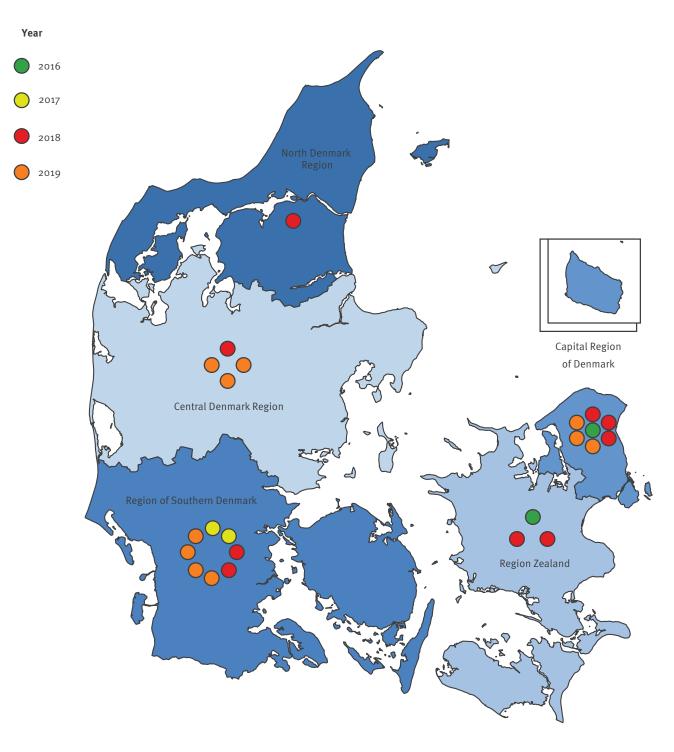
Available data on travel history abroad 6 months before detection of the OXA-244-producing *E. coli* isolates

were, as part of the national surveillance, reported by the DCM or by the responsible clinician/physician. Information on age, sex and postal code for residence was collected from the Danish Civil Registration System. Furthermore, patients found positive with OXA-244-producing *E. coli* during January 2018 to August 2019 (or their parents if children under 15 years of age) were interviewed by phone if possible during the summer of 2019, using a questionnaire created for this study containing questions on travelling within 6 months before date of sampling, food consumption and animal contact.

For investigation of possible nosocomial transmission in Denmark, hospitalisation data were retrieved from the DNPR for all patients during the period November 2015 to February 2019. The DNPR data included administration data, diagnoses, examinations and treatment procedures. Administration data included among others; hospital and department codes, dates of admission and discharge on patient level [16]. On 1 February 2019, the coding procedure for DNPR changed. Therefore, DNPR hospital data were only available until 1 February 2019 for epidemiological analysis in the present study.

#### FIGURE 2

OXA-244-producing *Escherichia coli* by date of detection and region, Denmark, January 2016–August 2019 (n = 24 isolates)



# Whole genome sequencing and in silico analysis

The genomic DNA was extracted (DNeasy Blood and Tissue Kit, Qiagen, Copenhagen, Denmark), with subsequent library construction (Nextera Kit, Illumina, Little Chesterford, UK) and finally sequenced (MiSeq or Nextseq, Illumina) according to the manufacturer's instructions to obtain paired-end reads of 2x250 or 2X150 bp in length. Quality control was performed on the raw reads, using the Bifrost pipeline at SSI (https://github.com/ssi-dk/bifrost) with accepted average coverage of 30.00x or above. The WGS data were either used as raw data or de novo assembled using the assemblies generated with SKESA in Bifrost. The raw reads of all 24 isolates were assembled into draft genomes using SKESA version 2.2 in the Bifrost pipeline.

Sequences are available from GenBank under accession number PRJEB36710.

Resistance genes were identified with ResFinder version 2.1 [17] (included in the Bifrost pipeline), using a threshold of 100% identities for identifying genes encoding beta-lactamases and carbapenemases, and

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1         1         1 $bula_{max}$ <th>TIC JIM</th> <th></th> <th>number</th> <th>date</th> <th>number</th> <th>Beta-lactam</th> <th>Aminoglycoside</th> <th>Fluoroquinolone</th> <th>Phenicol</th> <th>Sulphonamide</th> <th>Tetracycline</th> <th>Trimethoprim</th>	TIC JIM		number	date	number	Beta-lactam	Aminoglycoside	Fluoroquinolone	Phenicol	Sulphonamide	Tetracycline	Trimethoprim
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S18         id         Feb 200         i $bb_{(cue,u)}$ $abb_{(cue,u)}$	ST38	ST8	5	Feb 2018	1	bla <sub>CTX-M-27</sub>	aph (6)-Id, aph(3")-Ib, aadA5	1	I	sul1, sul2	tet(A)	dfrA17
158         16         Aproxo         1 $bb_{0,m_{M,M}}$ $aph(0), d_a, aph(3), d_B$ $aph(0), d_A, aph(3), d_A, aph(3), d_B$ $aph(0), d_A, d_B, d_B, d_B, d_B, d_B, d_B, d_B, d_B$	ST38	ST8	14	Feb 2019	1	bla <sub>cTX-M-27</sub>	aph (6)-1d, aph(3")-1b	I	I	sul2	tet(A)	I
518 $7.7$ $Way$ $1.7$ $$ $ $ $$	ST38	ST8	16	Apr 2019	1	bla <sub>CTX-M-27</sub>	aph (6)-1d, aph(3")-1b	I	I	sul2	tet(A)	I
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5 T69 Ju 2019 $bla_{(TriALA)}$ $aph (6) \cdot (1, aph (3') \cdot b)$ $  -$	ST38	ST8	∞	Jun 2018	2	bla <sub>CTX-M-14b</sub> , bla <sub>TEM-1B</sub>	aadA1,aph(3')-la, strA, strB	1	I	sul2	1	dfrA1
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ST8130t ctools $bla_{Crystardy} bla_{Tutkina}$ $aph(6).id, aph(3).ib,$ su/2su/2-ST818 $2039$ - $bula_{Crystardy} bla_{Tutkina}$ $aph(6).id, aph(3).ib,$ $$ $cadAi$ $$ $cuty$ $ter(0)$ ST823 $1u1$ corols- $bula_{Crystardy} bla_{Tutkina}$ $aph(6).id, ikk, aph(3).ib,$ $m/2$ $$ $ter(0)$ ST321 $1u1$ corols3 $$ $aph(6).id, ikk, aph(3).ib, ikk,$ $m/2$ $$ $urols$ $ter(0)$ ST322 $1un$ corols3 $$ $$ $$ $$ $$ $$ $$ $$ ST322 $1un$ corols3 $$ $$ $$ $$ $$ $$ $$ $$ $$ ST322 $1un$ corols $$ <	ST38	ST8	10	Aug 2018	I	bla <sub>cMY-2</sub> , bla <sub>TEM-1B</sub>	aph (6)-1d, aph(3")-1b	I	I	sul2	1	dfrA8
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ST2         12         Sep 2018         - $bla_{\text{NDM-1}}, bla_{\text{CXM-13}}, bla_{\text{DMM-1}}$ $aadA2, aac(6')lb-cr$ $aac(6')lb-cr$ $catB4$ $sult$ -         -	ST69	ST <sub>3</sub>	2	Nov 2016	I	I	1	QnrB19	I	I	tet(A), tet(U)	I
ST43712Sep 2018- $bla_{TX,M+15}$ , $bla_{TEM+16}$ , $bla_{0XM-1}$ $aph(6) \cdot  d_{1}$ , $aph(3) \cdot  a_{1}$ , $adA5$ , $aac(6) / b \cdot cr$ , $QrS1$ $catB4$ , $sul2$ , $sul1$ $sul2$ , $sul1$ ST6507May- $bla_{CX,M+14}$ , $bla_{TEM+16}$ $aph(3) \cdot  a_{1}$ , $adA5$ , $aac(6) / b \cdot cr$ , $QrS1$ $catB4$ , $sul2$ , $sul1$ $sul2$ , $sul1$ ST6507May- $bla_{CX,M+14}$ , $bla_{TEM+18}$ $aph(3) \cdot  a_{1}$ , $aph(3) \cdot  b_{1}$ , $aph(6) \cdot  d_{1}$ $-$ - $sul2$ $sul2$ ST53520May- $bla_{CX,M+15}$ , $bla_{TEM+18}$ $aph(6) \cdot  d_{1}$ , $aph(3) \cdot  b_{1}$ $qnrS1$ - $sul2$ $sul2$	ST167	ST2	12	Sep 2018	I	bla <sub>NDM-5</sub> , bla <sub>CTX-M-15</sub> , bla <sub>OXA-1</sub>	aadA2, aac(6')Ib-cr	aac(6')Ib-cr	catB4	sulı	I	dfrA12, dfrA14
ST650       7       May 2018       - $bla_{CTXM:4b}$ , $bla_{TEM:1b}$ $aph(3')-la, aph(6)-ld$ -       -       -       sul2         8       ST535       20       May 2019       - $bla_{CTXM:45}$ , $bla_{TEM:1b}$ $aph(6)-ld, aph(3'')-lb, aph(6)-ld$ -       sul2       sul2	ST167	ST437	12	Sep 2018	I	bla <sub>cTX-M-15</sub> , bla <sub>TEM-176</sub> , bla <sub>oXA-1</sub>	aph (6)-Id, aph(3')-Ia, aph(3")-Ib, aadA5, aac(6')Ib-cr	aac(6')Ib-cr, QnrS1	catB4, floR	sul2, sul1	tet(A)	dfrA17, dfrA14
ST535         20         May         -         bla <sub>crxw15</sub> , bla <sub>TEW18</sub> aph (6)-ld, aph(3")-lb         qnrS1         -         sul2	ST361	ST650	7	May 2018	I	bla <sub>CTX:M-14</sub> b, bla <sub>TEM-1B</sub>	aph(3')-Ia,aph(3")-Ib, aph (6)-Id	I	I	sul2	I	dfrA14
	ST3268	ST535	20	May 2019	I	bla <sub>CTX-M-15</sub> , bla <sub>TEM-1B</sub>	aph (6)-Id, aph(3")-Ib	qnrS1	I	sul2	tet(A)	dfrA14

MLST: multilocus sequence typing. <sup>a</sup> Samples taken from 23 patients. <sup>b</sup> MLST 1: Achtman scheme [18]. <sup>c</sup> MLST 2: Pasteur scheme [19]. 98.00% ID for all other genes encoding transferable antimicrobial resistance. Two different MLST schemes were used, i.e. the Achtman scheme (MLST 1) [18] and the Pasteur scheme (MLST 2) [19].

# **Phylogenetic analyses**

The contigs of the 24 OXA-244-producing *E. coli* in this study were uploaded to SeqSphere+version 5.1.0 (Ridom GmbH, Münster, Germany (http://www.ridom. de/seqsphere/)) using the Enterobase *E. coli* cgMLST scheme based on 2,513 genes with a cluster distance threshold of  $\leq$  10 allele differences.

# **Ethical statement**

Data on cases were collected as part of the national CPO surveillance according to guidelines for the prevention of spread of CPO by the Danish Health Authority according to which, both patients and treating medical doctor may be contacted for investigation [12]. All data were anonymised and cannot be inferred directly or indirectly to a person.

# Results

# Patients with OXA-244-producing Escherichia coli

From January 2016 to August 2019, 24 OXA-244producing *E. coli* isolates were detected in 23 patients in Denmark (Figure 1). From one of the patients, an OXA-244-producing *E. coli* and an OXA-244/NDM-5producing *E. coli* were obtained from the same faecal sample; both isolates were included in the study (Table 1). The remaining 22 OXA-244-producing *E. coli* isolates were obtained from blood samples (n = 2), urine samples (n = 18) and faecal samples (n = 2).

The 24 isolates were collected from hospitals and primary healthcare facilities across all five Danish regions (Figure 2, Table 1).

The age of the patients ranged from 5 to 79 years, with a median of 58 years, 20 were female and three were male. In comparison, during the study period, 63.5%(136/214) of patients with carbapenemase-producing *E*. *coli* were female in the Danish CPO surveillance, while the majority of the patients with OXA-244-producing *E*. *coli* in our study were female (p<0.05).

Of the 23 patients, 13 had been travelling 6 months before detection of the OXA-244-producing *E. coli* isolates, six had no reported history of recent travel and for the remaining four patients, travel information was unavailable (Table 1). Of the 13 patients reporting travel abroad within 6 months before detection of the OXA-244-producing *E. coli* isolate, seven had been to countries in Northern Africa (six to Egypt, one to Tunisia).

During the study period, Carbapenemase-producing *E. coli* isolates were detected from 214 patients; of these, 21 patient samples were taken at a general practitioner, 11 of these patients had OXA-244-producing *E.* 

*coli* isolates. Furthermore, OXA-244-producing *E. coli* isolates were detected from the samples from three patients visiting an emergency department and from nine patients hospitalised at different hospital wards.

Data from DNPR were available for the 13 patients with positive samples before February 2019. None of the 13 patients had been hospitalised at the same time at the same hospitals. Among these 13 patients, seven had not been hospitalised in Denmark, from January 2016 to February 2019, and their samples were either from a visit at the general practitioner or at the emergency department.

# Questionnaire of food consumption and animal contact

Fifteen of 19 patients with OXA-244-producing *E. coli* isolates during January 2018 to August 2019 were interviewed on the phone, while four patients were unavailable.

All 15 patients were eating meat and eggs. Four patients had been in contact with cats, 12 with dogs and two with wild birds. None of the patients had been in contact with livestock, hunting animals or reptiles.

# **Phylogenetic analysis**

The 24 OXA-244-producing *E. coli* isolates belonged to six different STs; ST10 (n=1), ST38 (n=13), ST69 (n=6), ST167 (n=2), ST361 (n=1) and ST3268 (n=1) (Table 2), and 15 had an ESBL gene (*bla*  $_{CTX-M-14b}$ , *bla*  $_{CTX-M-27}$ ) and one had a plasmid-encoded AmpC gene (*bla*  $_{CMY-2}$ ). Genes encoding colistin resistance, i.e. *mcr* genes, were not detected in any of the isolates. In four isolates, *bla*  $_{OXA-244}$  was the only detected resistance gene (Table 2).

One patient had both an OXA-244-producing ST167 *E. coli* (CPO20180141) and an OXA-244/NDM-5-producing ST167 *E. coli* (CPO20180142), but the two isolates did not cluster together in the cgMLST analysis and they had different resistance gene profiles (Table 2).

Three phylogenetic clusters were detected by cgMLST for the 24 isolates from this study (Table 2).

### Core-genome multilocus sequence typing clusters

The five isolates in Cluster 1 belonged to ST<sub>3</sub>8 (Table 2). The samples were obtained from December 2017 to May 2019 from three different regions in Denmark. Data from DNPR until February 2019 did not detect any overlapping hospitalisations between the first three patients. Travel information was missing for the first patient in this cluster. Six months prior to detection of the OXA-244- producing *E. coli* isolates, the second patient had been to Greece, the third patient had visited Egypt, the fourth patient had travelled to Egypt, Turkey as well as Italy, and the last patient had been to Sweden and Portugal. There were no geographical associations for place of residence among the patients in Cluster 1.

Both isolates in Cluster 2 belonged to ST<sub>3</sub>8 (Table 2). The isolates were sampled in May 2016 and June 2018 in the same region. The first patient had not been travelling 6 months before detection of the ST<sub>3</sub>8 OXA-244-producing *E. coli* isolate, whereas travel information was missing for the other patient. No epidemiologic links regarding hospitalisation or place of residence were detected between the patients.

The three isolates in Cluster 3 belonged to ST69 (Table 2). The samples were from three different regions and the patients also lived in different parts of the country. Unfortunately, DNPR data were not available for these patients as they all were detected after February 2019. The first patient had been to Poland and Austria 6 months before detection and the second patient had been to Egypt, while travel information was unavailable for the third patient.

# Discussion

In recent years, several hospital-related outbreaks with CPO have been observed in Denmark [11]. The increase in OXA-244-producing E. coli in Denmark described here did not appear to be related to Danish hospital outbreaks. Unfortunately, travel information was not available for all patients. Despite this, repeated import from abroad of the OXA-244-producing E. coli isolates appears likely. Many of the patients had been travelling 6 months before detection of the OXA-244-producing E. coli, predominantly to Northern Africa destinations. Intestinal persistence of CPO for more than 6 months has been reported, so a limitation in our study could by the lack of information about travel prior to 6 months [20]. It could be speculated that the OXA-244-producing E. coli could have been acquired during travel abroad longer than 6 months ago (e.g. a single patient reported travel to Egypt 5 years ago, data not shown).

Among the 15 patients interviewed by phone, 12 reported contacts to dogs and four to cats. To our knowledge, OXA-244-producing *E. coli* isolates have neither been reported from dogs nor cats, but a recent study reported on OXA-181-producing *E. coli* isolates obtained from dogs and cats after hospitalisation in a veterinary clinic in Switzerland [21].

All 15 patients interviewed by phone were eating meat, but none of them had been in direct contact with livestock. OXA-244-producing *E. coli* have neither been reported from meat nor from livestock. More generally, CPO are very rarely reported from livestock and meat in Europe. In contrast to Europe, dissemination of CPO among livestock are more common in China, India and Northern Africa [22].

Community sources for the OXA-244-producing *E. coli* isolates seem possible since many of the patients had not been hospitalised and the *E. coli* isolates belonged to STs that are present in the community

[23-32]. The ECDC RRA identified one main geographically dispersed ST<sub>3</sub>8 OXA-244-producing *E. coli* cluster with chromosomal-encoded *bla*  $_{0XA-244}$  present in all 10 countries, including Denmark, that submitted data for the RRA [10]. The exact location (plasmid or chromosomal) of *bla*  $_{0XA-244}$  was not investigated in our study as is would require utilisation of long read sequencing techniques (e.g. MinION and PacBio). The *bla*  $_{0XA-244}$ gene can be part of the Tn*51098* transposon [33]. This transposon has been reported at the same location of the chromosomal position among different STs [34].

In conclusion, an increase in OXA-244-producing *E. coli* has been detected in Denmark and other countries in Europe. Import of OXA-244 *E. coli* isolates from travelling abroad seems most likely for the majority of cases. Community sources of infections are also possible, as many of the patients had no history of hospitalisation. Furthermore, many of the *E. coli* isolates belonged to STs that are present in the community. It was not possible to point at a single country or a community source as risk factor for acquiring OXA-244-producing *E. coli* isolates.

## Acknowledgements

Karin Sixhøj Pedersen, Pia Thurø Hansen, Alexandra Medina, Lone Ryste Hansen Kildevang Mette Bar Ilan, Luise Müller and the Interview Center at Infectious Disease Preparedness, SSI are thanked for excellent assistance.

# **Conflict of interest**

None declared.

### Authors' contributions

Lone Jannok Porsbo, Frank Hansen, Louise Roer, Hülya Kaya, Anna Henius, Karina Lauenborg Møller, Ulrik S Justesen, Lillian Søes, Bent Røder, Philip K Thomsen, Mikala Wang, Turid Snekloth Søndergaard, Barbara Juliane Holzknecht, Claus Østergaard, Anne Kjerulf, Brian Kristensen and Henrik Hasman, contributed to the revision of the manuscript and approved the final version. Louise Roer, Frank Hansen, Hülya Kaya, Anette M Hammerum and Henrik Hasman did the molecular analysis. Ulrik S Justesen, Lillian Søes, Bent L Røder, Philip K Thomsen, Mikala Wang, Turid Snekloth Søndergaard, Barbara Juliane Holzknecht, Claus Østergaard detected CPOs at the DCMs in Denmark. Lone Jannok Porsbo and Anette M Hammerum drafted the questionnaire. Lone Jannok Porsbo, Anette M Hammerum, Henrik Hasman, Anna Henius, Karina Lauenborg Møller, Anne Kjerulf and Brian Kristensen were involved in the epidemiology analysis. Anette M Hammerum drafted the manuscript, incorporated comments, additions and feedback throughout the revision.

#### References

Naas T, Oueslati S, Bonnin RA, Dabos ML, Zavala A, Dortet L, et al. Beta-lactamase database (BLDB) - structure and function. J Enzyme Inhib Med Chem. 2017;32(1):917-9. https://doi.org/10. 1080/14756366.2017.1344235 PMID: 28719998

- Oteo J, Hernández JM, Espasa M, Fleites A, Sáez D, Bautista V, et al. Emergence of OXA-48-producing Klebsiella pneumoniae and the novel carbapenemases OXA-244 and OXA-245 in Spain. J Antimicrob Chemother. 2013;68(2):317-21. https://doi. org/10.1093/jac/dks383 PMID: 23034714
- Findlay J, Hopkins KL, Loy R, Doumith M, Meunier D, Hill R, et al. OXA-48-like carbapenemases in the UK: an analysis of isolates and cases from 2007 to 2014. J Antimicrob Chemother. 2017;72(5):1340-9. https://doi.org/10.1093/jac/dkx012 PMID: 28199647
- Hoyos-Mallecot Y, Naas T, Bonnin RA, Patino R, Glaser P, Fortineau N, et al. OXA-244-producing Escherichia coli isolates, a challenge for clinical microbiology laboratories. Antimicrob Agents Chemother. 2017;61(9):e00818-17. https:// doi.org/10.1128/AAC.00818-17 PMID: 28674064
- Valenza G, Nickel S, Pfeifer Y, Eller C, Krupa E, Lehner-Reindl V, et al. Extended-spectrum-β-lactamase-producing Escherichia coli as intestinal colonizers in the German community. Antimicrob Agents Chemother. 2014;58(2):1228-30. https:// doi.org/10.1128/AAC.01993-13 PMID: 24295972
- van Hattem JM, Arcilla MS, Bootsma MC, van Genderen PJ, Goorhuis A, Grobusch MP, et al. Prolonged carriage and potential onward transmission of carbapenemase-producing Enterobacteriaceae in Dutch travelers. Future Microbiol. 2016;11(7):857-64. https://doi.org/10.2217/fmb.16.18 PMID: 27357522
- Tafoukt R, Touati A, Leangapichart T, Bakour S, Rolain J-M. Characterization of OXA-48-like-producing Enterobacteriaceae isolated from river water in Algeria. Water Res. 2017;120:185-9. https://doi.org/10.1016/j.watres.2017.04.073 PMID: 28486169
- Diab M, Hamze M, Bonnet R, Saras E, Madec J-Y, Haenni M. Extended-spectrum beta-lactamase (ESBL)- and carbapenemase-producing Enterobacteriaceae in water sources in Lebanon. Vet Microbiol. 2018;217:97-103. https:// doi.org/10.1016/j.vetmic.2018.03.007 PMID: 29615264
- Potron A, Poirel L, Dortet L, Nordmann P. Characterisation of OXA-244, a chromosomally-encoded OXA-48-like β-lactamase from Escherichia coli. Int J Antimicrob Agents. 2016;47(1):102-3. https://doi.org/10.1016/j.ijantimicag.2015.10.015 PMID: 26655033
- 10. European Centre for Disease Prevention and Control (ECDC). Rapid risk assessment: increase in OXA-244-producing Escherichia coli in the European Union/European Economic Area and the UK since 2013. Stockholm: ECDC; 2020. Available from: https://www.ecdc.europa.eu/en/publications-data/ rapid-risk-assessment-increase-oxa-244-producingescherichia-coli-eu-eea
- 11. National Food Institute, Technical University of Denmark, Statens Serum Institut. DANMAP 2018 - Use of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from food animals, food and humans in Denmark. Lyngby, Copenhagen: National Food Institute, Statens Serum Institut; 2019. Available from: https://www.danmap.org/-/media/ arkiv/projekt-sites/danmap/danmap-reports/danmap-2018/ danmap\_2018.pdf?la=en
- Sundhedsstyrelsen. [Danish Health Authority]. Vejledning om forebyggelse af spredning af CPO. [Guidance on preventing the spread of CPO]. Copenhagen: Sundhedsstyrelsen; 2018. Danish. Available from: https://www.sst.dk/-/media/ Udgivelser/2018/CPO/Vejledning-om-forebyggelse-afspredning-af-CPO.ashx?la=da&hash=o60943943A71EA7E2AA6 B51C229577B87E5937A3
- 13. Wang M, Hansen DS, Littauer P, Schumacher H, Hammerum A. Undersøgelse for Carbapenemase Producerende Organismer (CPO) bærertilstand - en metodevejledning. [Carbapenemase Producing Organism (CPO) Carrier Condition Study - A Method Guide]. Copenhagen: Dansk Selskab for Klinisk Mikrobiologi [Danish Society for Clinical Microbiology]; 2016. Danish. Available from: https://dskm.dk/wp-content/uploads/2016/10/ Unders%c3%b8gelse-for-Carbapenemase-Producerende-Organismer-okt-2016.pdf
- 14. European Committee on Antimicrobial Susceptibility Testing (EUCAST). EUCAST guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance version 1.0. Växjö: EUCAST; 2013. Available from: http://www.eucast.org/ resistance\_mechanisms/
- 15. European Committee on Antimicrobial Susceptibility Testing (EUCAST). EUCAST guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance version 2.0. Växjö: EUCAST; 2017. Available from: http://www.eucast.org/ resistance\_mechanisms/
- 16. Schmidt M, Schmidt SAJ, Sandegaard JL, Ehrenstein V, Pedersen L, Sørensen HT. The Danish National Patient Registry: a review of content, data quality, and research potential. Clin

Epidemiol. 2015;7:449-90. https://doi.org/10.2147/CLEP. S91125 PMID: 26604824

- 17. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, et al. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother. 2012;67(11):2640-4. https://doi.org/10.1093/jac/dks261 PMID: 22782487
- Wirth T, Falush D, Lan R, Colles F, Mensa P, Wieler LH, et al. Sex and virulence in Escherichia coli: an evolutionary perspective. Mol Microbiol. 2006;60(5):1136-51. https://doi.org/10.1111/ j.1365-2958.2006.05172.x PMID: 16689791
- Jaureguy F, Landraud L, Passet V, Diancourt L, Frapy E, Guigon G, et al. Phylogenetic and genomic diversity of human bacteremic Escherichia coli strains. BMC Genomics. 2008;9(1):560. https://doi.org/10.1186/1471-2164-9-560 PMID: 19036134
- 20. Lázaro-Perona F, Ramos JC, Sotillo A, Mingorance J, García-Rodríguez J, Ruiz-Carrascoso G, et al. Intestinal persistence of a plasmid harbouring the OXA-48 carbapenemase gene after hospital discharge. J Hosp Infect. 2019;101(2):175-8. https:// doi.org/10.1016/j.jhin.2018.07.004 PMID: 30017896
- Nigg A, Brilhante M, Dazio V, Clément M, Collaud A, Gobeli Brawand S, et al. Shedding of OXA-181 carbapenemaseproducing Escherichia coli from companion animals after hospitalisation in Switzerland: an outbreak in 2018. Euro Surveill. 2019;24(39):1900071. https://doi.org/10.2807/1560-7917.ES.2019.24.39.1900071 PMID: 31576806
- 22. Köck R, Daniels-Haardt I, Becker K, Mellmann A, Friedrich AW, Mevius D, et al. Carbapenem-resistant Enterobacteriaceae in wildlife, food-producing, and companion animals: a systematic review. Clin Microbiol Infect. 2018;24(12):1241-50. https://doi. org/10.1016/j.cmi.2018.04.004 PMID: 29654871
- 23. Grönthal T, Österblad M, Eklund M, Jalava J, Nykäsenoja S, Pekkanen K, et al. Sharing more than friendship - transmission of NDM-5 ST167 and CTX-M-9 ST69 Escherichia coli between dogs and humans in a family, Finland, 2015. Euro Surveill. 2018;23(27):1700497. https://doi.org/10.2807/1560-7917. ES.2018.23.27.1700497 PMID: 29991384
- 24. Tarlton NJ, Moritz C, Adams-Sapper S, Riley LW. Genotypic analysis of uropathogenic Escherichia coli to understand factors that impact the prevalence of β-lactam-resistant urinary tract infections in a community. J Glob Antimicrob Resist. 2019;19:173-80. https://doi.org/10.1016/j. jgar.2019.03.002 PMID: 30872040
- 25. Hornsey M, Betts JW, Mehat JW, Wareham DW, van Vliet AHM, Woodward MJ, et al. Characterization of a colistin-resistant Avian Pathogenic Escherichia coli ST69 isolate recovered from a broiler chicken in Germany. J Med Microbiol. 2019;68(1):111-4. https://doi.org/10.1099/jmm.o.000882 PMID: 30475200
- 26. Manges AR, Geum HM, Guo A, Edens TJ, Fibke CD, Pitout JDD. Global Extraintestinal Pathogenic Escherichia coli (ExPEC) Lineages. Clin Microbiol Rev. 2019;32(3):e00135-18. https:// doi.org/10.1128/CMR.00135-18 PMID: 31189557
- 27. Xu L, Wang P, Cheng J, Qin S, Xie W. Characterization of a novel blaNDM-5-harboring IncFII plasmid and an mcr-1-bearing Incl2 plasmid in a single Escherichia coli ST167 clinical isolate. Infect Drug Resist. 2019;12:511-9. https://doi.org/10.2147/IDR. S192998 PMID: 30881056
- 28. Yang P, Xie Y, Feng P, Zong Z. blaNDM-5 carried by an IncX3 plasmid in Escherichia coli sequence type 167. Antimicrob Agents Chemother. 2014;58(12):7548-52. https://doi.org/10.1128/AAC.03911-14 PMID: 25246393
- 29. Zeng X, Chi X, Ho BT, Moon D, Lambert C, Hall RJ, et al. Comparative genome analysis of an extensively drugresistant isolate of avian sequence type 167 Escherichia coli strain Sanji with novel In Silico serotype 089b:H9. mSystems. 2019;4(1):e00242-18. https://doi.org/10.1128/ mSystems.00242-18 PMID: 30834329
- 30. Atterby C, Börjesson S, Ny S, Järhult JD, Byfors S, Bonnedahl J. ESBL-producing Escherichia coli in Swedish gulls-A case of environmental pollution from humans? PLoS One. 2017;12(12):e0190380. https://doi.org/10.1371/journal. pone.0190380 PMID: 29284053
- Freitag C, Michael GB, Kadlec K, Hassel M, Schwarz S. Detection of plasmid-borne extended-spectrum β-lactamase (ESBL) genes in Escherichia coli isolates from bovine mastitis. Vet Microbiol. 2017;200:151-6. https://doi.org/10.1016/j. vetmic.2016.08.010 PMID: 27566885
- 32. Zogg AL, Zurfluh K, Schmitt S, Nüesch-Inderbinen M, Stephan R. Antimicrobial resistance, multilocus sequence types and virulence profiles of ESBL producing and non-ESBL producing uropathogenic Escherichia coli isolated from cats and dogs in Switzerland. Vet Microbiol. 2018;216:79-84. https://doi.org/10.1016/j.vetmic.2018.02.011 PMID: 29519530
- 33. Turton JF, Doumith M, Hopkins KL, Perry C, Meunier D, Woodford N. Clonal expansion of Escherichia coli ST38 carrying a chromosomally integrated OXA-48 carbapenemase gene. J

Med Microbiol. 2016;65(6):538-46. https://doi.org/10.1099/ jmm.o.ooo248 PMID: 26982715

34. Pitout JDD, Peirano G, Kock MM, Strydom KA, Matsumura Y. The global ascendency of OXA-48-type carbapenemases. Clin Microbiol Rev. 2019;33(1):e00102-19. https://doi.org/10.1128/ CMR.00102-19 PMID: 31722889

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