



# Genomes of Bacteriophages Belonging to the Orders *Caudovirales* and *Petitvirales* Identified in Fecal Samples from Pacific Flying Fox (*Pteropus tonganus*) from the Kingdom of Tonga

Jasmine K. M. Lopez,<sup>a</sup> Maketalena Aleamotu'a,<sup>b</sup> Viliami Kami,<sup>c</sup> Daisy Stainton,<sup>d</sup> Michael C. Lund,<sup>a</sup> Simona Kraberger,<sup>a</sup>  
Arvind Varsani<sup>a,e</sup>

<sup>a</sup>Biodesign Center for Fundamental and Applied Microbiomics, School of Life Sciences, Center for Evolution and Medicine, Arizona State University, Tempe, Arizona, USA

<sup>b</sup>School of Environmental and Life Sciences, The University of Newcastle, Callaghan, New South Wales, Australia

<sup>c</sup>Land Resource Division, The Pacific Community, Narere Campus, Suva, Fiji

<sup>d</sup>Department of Entomology and Plant Pathology, Division of Agriculture, University of Arkansas System, Fayetteville, Arkansas, USA

<sup>e</sup>Structural Biology Research Unit, Department of Integrative Biomedical Sciences, University of Cape Town, Cape Town, South Africa

**ABSTRACT** Twenty-nine circular genomes of bacteriophages in the orders *Caudovirales* and *Petitvirales* were identified from fecal samples from Pacific flying foxes that were collected from their roosting sites on the Pacific Island of Tonga in 2014 and 2015. The vast majority are microviruses ( $n = 25$ ), with 2 siphoviruses, 1 myovirus, and 1 podovirus.

Pacific flying foxes (*Pteropus tonganus*) are frugivorous bats that are found throughout the Pacific region (1) and are the sole bat species found on the Tongan archipelago (2), playing an important role in pollination and seed dispersal (1, 3, 4). Our previous work on Pacific flying foxes from Tonga reported various cressdnaviruses (5). Here, we expand on that work, focusing on bacteriophages.

Fecal samples from four roosting sites ('Atele, Ha'avakatolo, Kolovai, and Lapaha) located on Tongatapu island were collected in April 2014 and January 2015 (5). From each, 5- to 10-g samples were pooled based on sample year, resuspended in 45 mL of SM buffer (50 mM Tris HCl, 10 mM MgSO<sub>4</sub>, 0.1 M NaCl [pH 7.5]), and processed for viral nucleic acid extraction as described by Male et al. (5). The High Pure viral nucleic acid kit (Roche Diagnostics, USA) was used to extract viral DNA. The extracted DNA samples were enriched for circular sequences using rolling circle amplification with the TempliPhi 100 kit (GE Healthcare, USA); they were then used by Beijing Genomics Institute (Hong Kong) to prepare 2 × 90-bp libraries using their custom protocol, and the libraries were sequenced using a HiSeq 2000 sequencer (Illumina, USA). The raw reads were trimmed with Trimmomatic v0.39 (6) and then *de novo* assembled using metaSPAdes v3.12.0 (7). In order to identify bacteriophage-like sequences, we used VirSorter (8), and sequences were determined to be circular (based on terminal redundancy). From the pooled samples from April 2014 (Tbat1) and from January 2015 (Tbat2), 29 full bacteriophage genomes were identified. Open reading frames were identified with RASTtk (9), and the annotations were refined with the HMMER web server with the Pfam database (10) and Cenote-Taker 2 (11). The coverage depth and number of mapped reads for each genome were determined with BMAP (12). All bioinformatic software was used with default settings. The genomes have varied read depths of 10.99× to 24,465.65×, with 741 to 1,592,983 reads (Table 1).

Of the 29 bacteriophages, 25 members of the *Microviridae* family (order *Petitvirales*) were identified in Tbat1 and 23 in Tbat2 (Fig. 1). These 25 microviruses have genome lengths of 3,911 to 6,376 nucleotides (nt) and GC contents of 36 to 60% (Fig. 1 and Table 1). All of the identified microvirus genomes encode a homologous major capsid

**Editor** Simon Roux, DOE Joint Genome Institute

**Copyright** © 2022 Lopez et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Arvind Varsani, arvind.varsani@asu.edu.

The authors declare no conflict of interest.

**Received** 19 January 2022

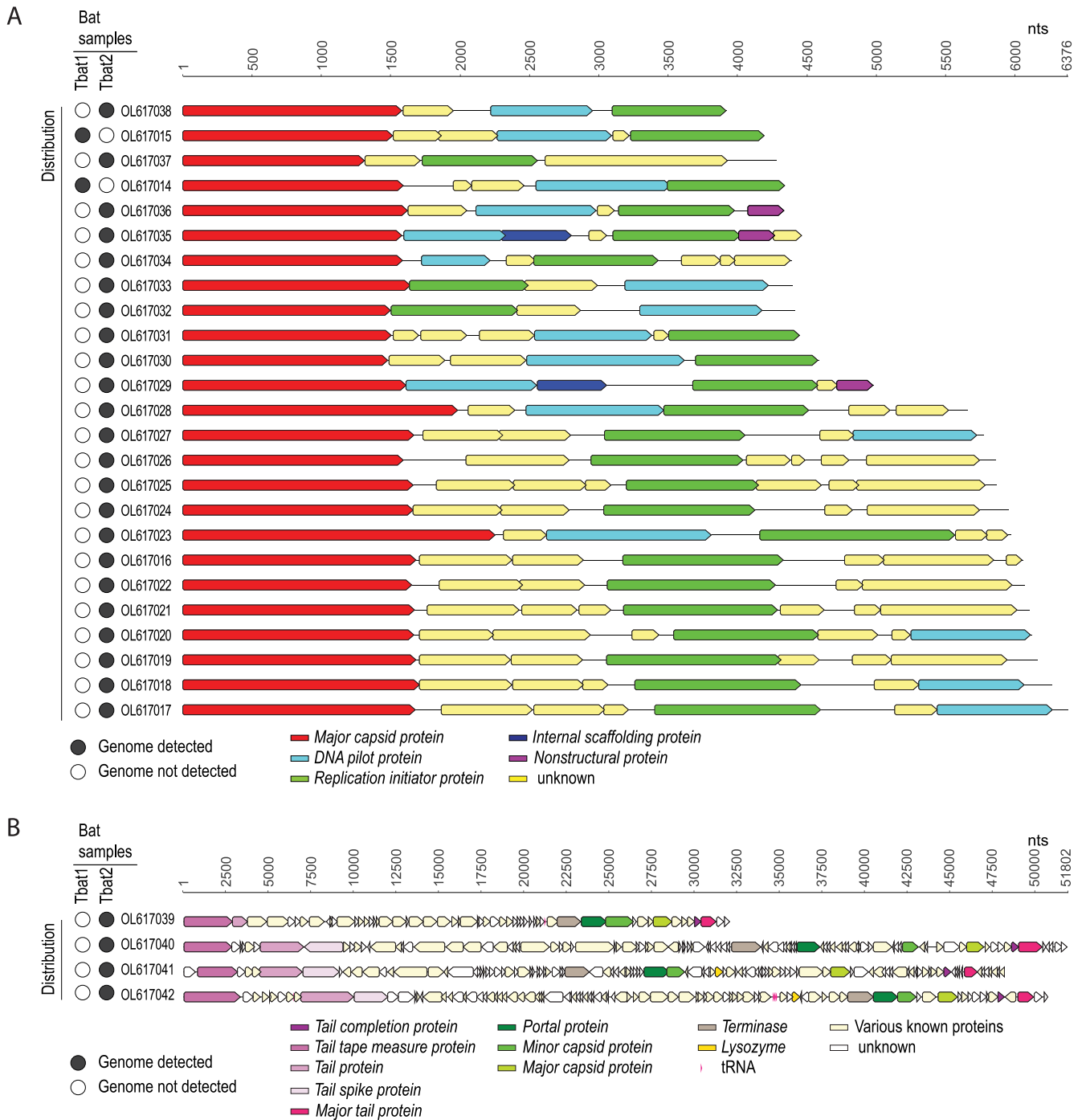
**Accepted** 1 February 2022

**Published** 17 February 2022

**TABLE 1** Summary of bacteriophage genomes determined from Tongan bat fecal samples and their top BLASTn hit information

Data for top BLASTn hit											
Family	GenBank accession no.	Length (nt)	GC content (%)	Read depth (x)	No. of mapped reads	GenBank accession no.	Virus type <sup>a</sup>	Query coverage (%)	E value	Identity (%)	
Microviridae	OL617014	4,315	36.80	21.1775	1,019	MH617689	Microviridae sp. isolate ctbe975	17	1.00E-49	68.61	
Microviridae	OL617015	4,179	55.20	26.3508	1,228	MH992220	Apis mellifera-associated microvirus 41 INH_SP_302	15	2.00E-48	69.41	
Microviridae	OL617016	6,049	54.40	150.1405	10,117	MT310373	Microvirus sp. isolate 1712115_248	56	0	68.49	
Microviridae	OL617017	6,376	54.80	53.9305	3,828	MN988476	Rhizobium phage RHph_TM1_7A	51	0	72.17	
Microviridae	OL617018	6,260	55.30	133.1529	9,282	MT310373	Microvirus sp. isolate 1712115_248	60	0	68.05	
Microviridae	OL617019	6,153	53.90	93.6808	6,414	MT310373	Microvirus sp. isolate 1712115_248	59	0	68.24	
Microviridae	OL617020	6,111	59.40	422.1753	28,704	MK765588	Tortoise microvirus 38 isolate 38_SP_76	53	0	70.08	
Microviridae	OL617021	6,096	55.40	10.9934	747	MK765589	Tortoise microvirus 39 isolate 39_SP_82	18	2.00E-69	69.44	
Microviridae	OL617022	6,062	57.50	41.7634	2,820	MK765589	Tortoise microvirus 39 isolate 39_SP_82	20	2.00E-76	71.33	
Microviridae	OL617023	5,964	38.10	38.0241	2,525	BK033080	TPA: Microviridae sp. isolate ctr3K1	86	0	86.82	
Microviridae	OL617024	5,949	55.50	8,094.895	536,306	MT310362	Microvirus sp. isolate 1712115_301	69	0	70.69	
Microviridae	OL617025	5,857	58.00	24.46565	1,592,983	MH616837	Microviridae sp. isolate ctcf_4	36	3.00E-111	71.96	
Microviridae	OL617026	5,849	56.40	29.7458	1,940	MT310362	Microvirus sp. isolate 1712115_301	73	0	69.02	
Microviridae	OL617027	5,767	56.80	34.6105	2,228	MK765635	Tortoise microvirus 82 isolate 82_SP_77	28	2.00E-86	70.32	
Microviridae	OL617028	5,648	40.70	11.9102	749	MH616763	Microviridae sp. isolate ctbb565	46	1.00E-133	67.60	
Microviridae	OL617029	4,969	56.60	25.4431	1,409	BK015202	TPA: Microviridae sp. ct2OM3	30	6.00E-144	68.48	
Microviridae	OL617030	4,580	55.40	16.5537	845	MZ364279	Robin microvirus RP_139	37	3.00E-135	68.06	
Microviridae	OL617031	4,440	47.60	31.5509	1,566	MH992217	Apis mellifera-associated microvirus 42 INH_SP_292	34	3.00E-102	66.49	
Microviridae	OL617032	4,405	44.40	22.8904	1,123	KM589510	Microviridae Fen7786_21	2	2.00E-17	77.69	
Microviridae	OL617033	4,389	47.70	58.9834	2,870	MT309935	Microvirus sp. isolate BS1_501	12	2.00E-11	64.20	
Microviridae	OL617034	4,380	37.50	15.1317	741	MH649004	Microviridae sp. isolate ctbd002	11	1.00E-32	71.60	
Microviridae	OL617035	4,362	48.30	28.903	1,404	MN582079	Microviridae sp. ctODW36	31	8.00E-85	69.94	
Microviridae	OL617036	4,325	44.30	16.5563	800	MT309934	Microvirus sp. isolate BS1_502	14	9.00E-46	67.53	
Microviridae	OL617037	4,275	60.60	276.2742	13,132	MT310281	Microvirus sp. isolate 1712115_698	21	8.00E-104	69.70	
Microviridae	OL617038	3,911	41.40	19.495	851	MT310293	Microvirus sp. isolate 1712115_653	10	1.00E-30	73.93	
Myoviridae	OL617039	31,988	35.60	4741.802	1,517,943	MN855801	Bacteriophage sp. isolate 103	4	0	82.23	
Podoviridae	OL617040	51,802	41.70	290.3623	161,065	MN840487	Proteus phage 2207-N35	60	0	74.50	
Siphoviridae	OL617041	48,082	47.70	388.254	203,214	BK017157	TPA: Siphoviridae sp. isolate ctEwt2	70	0	94.03	
Siphoviridae	OL617042	50,667	48.50	22.8497	12,658	BK057309	TPA: Siphoviridae sp. isolate ctWpt2	24	0	72.15	

<sup>a</sup> TPA, third party annotation.



**FIG 1** (A) Genome organization of the 25 microviruses from the Tongan bat fecal samples. (B) Genome organization of the 2 siphoviruses, 1 myovirus, and 1 podovirus from the Tongan bat fecal samples. Solid circles on the left indicate full genome coverage of the mapped reads in either the pooled sample of April 2014 or that of January 2015.

protein and replication initiator protein, with the majority also encoding a recognizable DNA pilot protein (Fig. 1). BLASTn analysis against the nonredundant nucleotide database revealed that the microvirus genome with GenBank accession number [OL617023](#) has the greatest nucleotide identity in this group, i.e., 86.8% (86% genome coverage) with respect to a microvirus identified in human samples (GenBank accession number [BK033080](#)) (13), whereas that with GenBank accession number [OL617032](#) has the lowest nucleotide identity, i.e., 77.69% (2% genome coverage) with respect to a microvirus from soil (GenBank accession number [KM589510](#)) (14) (Table 1).

In the Tbat2 pooled sample, we identified four genomes in the order *Caudovirales*, i.e., two siphovirus genomes (48,082 to 50,667 nt, with GC contents of 47.7 to 48.5%), one myovirus (31,988 nt, with a GC content of ~35%), and one podovirus (51,802 nt, with a GC content of ~41%) (Fig. 1 and Table 1). Conserved tail and capsid protein coding regions were identified in these genomes (Fig. 1). The two siphoviruses share 72.15 and 94.03% nucleotide identity (70% and 24% genome coverage, respectively) with other siphoviruses from humans (GenBank accession numbers [BK017157](#) and [BK057309](#)) (13). The myovirus (GenBank accession number [OL617039](#)) shares 82% nucleotide identity (4% genome coverage) with a genome from honeybees (GenBank accession number [MN855801](#)) (15), and the podovirus (GenBank accession number [OL617040](#)) shares 74.5% nucleotide identity (60% genome coverage) with a genome from *Proteus mirabilis* (GenBank accession number [MN840487](#)) available in GenBank (Table 1).

**Data availability.** The bacteriophage sequences have been deposited in the NCBI SRA under BioProject accession number [PRJNA780525](#) (SRA accession numbers [SRX13144068](#) and [SRX13144069](#)) and in GenBank under accession numbers [OL617014](#) to [OL617042](#).

## REFERENCES

- Banack SA. 1998. Diet selection and resource use by flying foxes (genus *Pteropus*). *Ecology* 79:1949–1967. <https://doi.org/10.2307/176701>.
- Miller CA, Wilson DE. 1997. *Pteropus tonganus*. *Mammalian Species* <https://doi.org/10.2307/3504121>.
- Nelson SL, Kunz TH, Humphrey SR. 2005. Folivory in fruit bats: leaves provide a natural source of calcium. *J Chem Ecol* 31:1683–1691. <https://doi.org/10.1007/s10886-005-5920-y>.
- Pierson ED, Rainey WE. 1992. The biology of flying foxes of the genus *Pteropus*: a review. *Biol Rep* 90(23):1–17. <https://apps.dtic.mil/sti/pdfs/ADA322812.pdf>.
- Male MF, Kraberger S, Stainton D, Kami V, Varsani A. 2016. Cycloviruses, gemycircularviruses and other novel replication-associated protein encoding circular viruses in Pacific flying fox (*Pteropus tonganus*) faeces. *Infect Genet Evol* 39:279–292. <https://doi.org/10.1016/j.meegid.2016.02.009>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Roux S, Enault F, Hurwitz BL, Sullivan MB. 2015. VirSorter: mining viral signal from microbial genomic data. *PeerJ* 3:e985. <https://doi.org/10.7717/peerj.985>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
- Potter SC, Luciani A, Eddy SR, Park Y, Lopez R, Finn RD. 2018. HMMER web server: 2018 update. *Nucleic Acids Res* 46:W200–W204. <https://doi.org/10.1093/nar/gky448>.
- Tisza MJ, Belford AK, Dominguez-Huerta G, Bolduc B, Buck CB. 2021. Cento-Taker 2 democratizes virus discovery and sequence annotation. *Virus Evol* 7:veaa100. <https://doi.org/10.1093/ve/veaa100>.
- Bushnell B. 2014. BMAP: a fast, accurate, splice-aware aligner. Lawrence Berkeley National Laboratory, Berkeley, CA.
- Tisza MJ, Buck CB. 2021. A catalog of tens of thousands of viruses from human metagenomes reveals hidden associations with chronic diseases. *Proc Natl Acad Sci U S A* 118:e2023202118. <https://doi.org/10.1073/pnas.2023202118>.
- Quaiser A, Dufresne A, Ballaud F, Roux S, Zivanovic Y, Colombet J, Sime-Ngando T, Francez AJ. 2015. Diversity and comparative genomics of *Microviridae* in *Sphagnum*-dominated peatlands. *Front Microbiol* 6:375. <https://doi.org/10.3389/fmicb.2015.00375>.
- Deboutte W, Beller L, Yinda CK, Maes P, de Graaf DC, Matthijnsens J. 2020. Honey-bee-associated prokaryotic viral communities reveal wide viral diversity and a profound metabolic coding potential. *Proc Natl Acad Sci U S A* 117:10511–10519. <https://doi.org/10.1073/pnas.1921859117>.