

# Decreased expression of *RASSF10* correlates with poor prognosis in patients with colorectal cancer

Junxun Ma, PhD, Sujie Zhang, PhD, Yi Hu, PhD\*, Xiaoyan Li, PhD, Fang Yuan, PhD, Danyang Sun, PhD, Lijie Wang, PhD, Fan Zhang, PhD, Guangying Chen, PhD, Pengfei Cui, PhD

## Abstract

Ras association domain protein 10 (*RASSF10*) was reported to act as a prognostic indicator in various types of cancer and it was proved to be tumor suppressor gene in colorectal cancer (CRC). The purpose of this study was to evaluate the prognostic significance of *RASSF10* in CRC.

Quantitative real-time polymerase chain reaction was used to detect the messenger RNA (mRNA) expression while enzyme-linked immunosorbent assay was taken to measure the protein expression of *RASSF10* in tumor tissues and adjacent normal tissues from 102 patients with CRC. The relationship between *RASSF10* expression level and clinical characteristics of CRC patients was analyzed by chi-squared test. In addition, the association between overall survival of CRC patients and *RASSF10* expression was estimated by Kaplan–Meier analysis. Cox regression analysis was used to evaluate the prognostic value of *RASSF10*.

The expression level of *RASSF10* in tumor tissues was significantly lower than that in the normal tissues both at mRNA and protein levels. Moreover, the expression level was correlated with lymph-node-metastasis and tumor-node-metastasis stage. Kaplan–Meier analysis suggested that patients with high expression level of *RASSF10* had a longer overall survival than those with low level (log-rank test,  $P < .001$ ). Besides, *RASSF10* might be a potential biomarker in the prognosis of CRC according to cox regression analysis.

The down regulated of *RASSF10* is found in CRC and it may be an ideal prognostic marker.

**Abbreviations:** 95% CI = 95% confidence interval, CRC = colorectal cancer, ELISA = enzyme-linked immunosorbent assay, GAPDH = glyceraldehyde-3-phosphate dehydrogenase, HR = hazard ratio, mRNA = messenger RNA, RASSF = Ras association domain protein, RT-PCR = real-time polymerase chain reaction, TNM = tumor node metastasis.

**Keywords:** colorectal cancer, prognosis, *RASSF10*

## 1. Introduction

Colorectal cancer (CRC) was the third most commonly diagnosed cancer in male and the second in female.<sup>[1,2]</sup> What's more, the morbidity of CRC was increasing in Asian, especially in China, due to the progressive “Westernization” of lifestyles.<sup>[3]</sup> CRC was the consequences of the accumulation of genetic and epigenetic alterations, so it was difficult to determine the risk factors for CRC.<sup>[4]</sup> At present time, the commonly used prognostic marker for CRC in clinical practice was tumor-node-metastasis (TNM) system.<sup>[5]</sup> However, TNM system could cause substantial under-treatment and over-treatment for CRC patients.<sup>[6]</sup> Therefore, it was urgently need to exploit novel and reliable biomarkers for the prognosis of CRC.

Editor: Somchai Amorniyotin.

JM and SZ contributed equally to this work.

The authors have no funding and conflicts of interest to disclose.

Department of Oncology, Chinese PLA General Hospital, Beijing, China.

\* Correspondence: Yi Hu, Department of Oncology, Chinese PLA General Hospital, Beijing 100853, China (e-mail: nv546dsdg@sina.com).

Copyright © 2017 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial License 4.0 (CCBY-NC), where it is permissible to download, share, remix, transform, and buildup the work provided it is properly cited. The work cannot be used commercially without permission from the journal.

Medicine (2017) 96:42(e7011)

Received: 13 March 2016 / Received in final form: 2 May 2017 / Accepted: 4 May 2017

<http://dx.doi.org/10.1097/MD.0000000000007011>

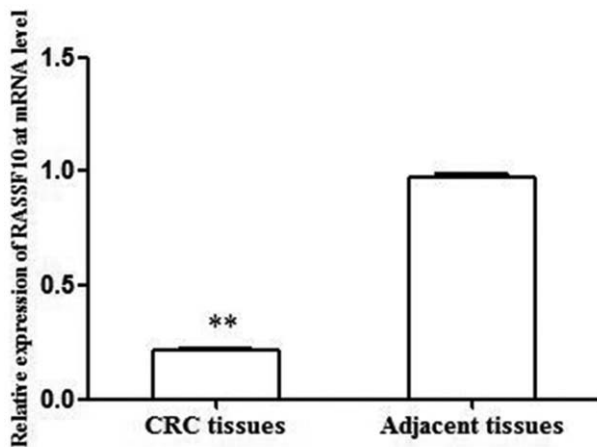
Ras association domain protein 10 (*RASSF10*) was a novel member of RASSF family and characterized by the inclusion of an N-terminus, which was described from a predicted sequence with homology to RASSF9/P-CIP1.<sup>[7]</sup> It located at chromosome 11p15.2 and contained a CpG island which was easy to be methylated leading to tumorigenesis.<sup>[8]</sup> In the previous studies, *RASSF10* was proved to act as a tumor suppressor in several cancers such as lung cancer, thyroid cancer, hepatocellular carcinoma, esophageal squamous cell carcinoma, and gastric cancer.<sup>[9–13]</sup> *RASSF10* was considered to be a tumor suppressor and could inhibit tumor growth by activating P53 signaling in CRC according to the study of Guo et al.<sup>[14]</sup> However, its clinical significance in CRC was never reported.

In this study, we aimed to detect the expression and its prognostic significance of *RASSF10* in patients with CRC. The expression level of *RASSF10* in CRC tissues both at messenger RNA (mRNA) and protein levels was detected. Meanwhile, the association between clinical characteristics and *RASSF10* expression was evaluated by chi-squared test. Besides, the overall survival of CRC patients according to the level of *RASSF10* was estimated and cox regression analysis was used to analyze the prognostic value of the gene in CRC, in order to find a novel indicator for CRC prognosis.

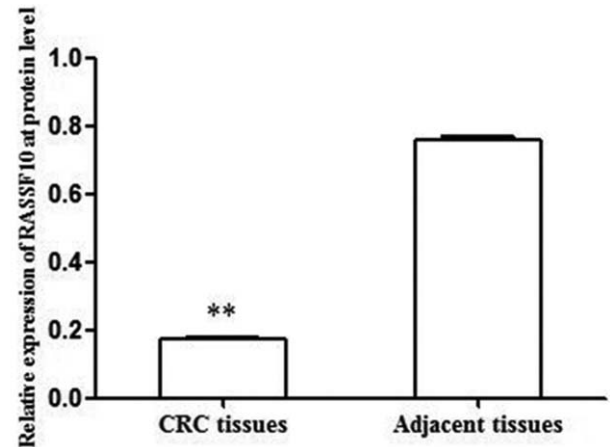
## 2. Materials and methods

### 2.1. Patients and tissue samples

One hundred two patients with CRC were enrolled in this study at Chinese PLA General Hospital from December 2008 to March 2010. The study was permitted by the Ethnic



**Figure 1.** Relative mRNA expression of *RASSF10* in CRC tissues and corresponding normal tissues. *RASSF10* mRNA expression was decreased in CRC tissues compared to that in corresponding normal tissues ( $P < .001$ ). CRC = colorectal cancer. RASSF10 = Ras association domain protein 10.



**Figure 2.** Relative protein expression of *RASSF10* in CRC tissues and corresponding normal tissues. *RASSF10* protein expression was lower in CRC tissues than that in corresponding normal tissues ( $P < .001$ ). CRC = colorectal cancer, RASSF10 = Ras association domain protein 10.

Committee of Chinese PLA General Hospital and all patients had signed written informed consents in advance. None of the patients had received any physical therapy and chemotherapy before sampling.

Pathological specimens and adjacent normal tissues were collected from CRC patients and frozen in liquid nitrogen immediately. Then all samples were stored at  $-80^{\circ}\text{C}$  until use, respectively. The detailed clinicopathologic characteristics of patients including age, gender, histological type, depth of invasion, location, lymph node metastasis, and TNM stage were recorded in database. A 5-year follow-up was conducted and patients who were died from unexpected events or other diseases were excluded from our study.

## 2.2. RNA extraction and quantitative RT-PCR analysis

Total RNA was extracted from the collected specimens using Trizol agent (Invitrogen, Carlsbad, CA) following the manufacturer's instructions. DNase I was used to treat the residual DNA in RNA samples. The concentration and quality of the RNA samples were detected by UV absorbance ( $A_{260}/A_{280}$ ) and 1% agarose gel electrophoresis, respectively. The first chain of cDNA was compounded through a Prime Scrip RT reagent kit (Takara Biotechnology Co, Ltd). Real-time polymerase chain reaction (RT-PCR) was performed with SYBR Green I assay (Takara) in the Applied Biosystems 7900 Fast Real-Time PCR system (Applied Biosystems, Foster City, CA). Glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*) acted as internal control. The relative expression of *RASSF10* was calculated with  $2^{-\Delta\Delta\text{Ct}}$  method. The sequences of primers used in this study were as followed. *RASSF10*: forward 5'-CCATGACCCAGGAGAAACAG-3'; reverse 5'-GCTGGCGAATTGTGTGGTC-3'. *GAPDH*: forward 5'-CATGAGAAGTATGACAACAGCCT-3'; reverse 5'-AGTCCTTCCACGATACCAAAGT-3'.

## 2.3. ELISA analysis

Total protein was extracted from all samples. The expression of RASSF10 protein was measured by enzyme-linked immunosorbent assay (ELISA) kits (DSA00-R&D systems) according to the manufacturer's protocol. Each experimental was in triplicate.

## 2.4. Statistical analysis

SPSS 18.0 software was used for all statistical analysis and GraphPad Prism 5 was used for designing the figures in this study. All quantitative variables were shown as mean  $\pm$  standard deviation. According to the average expression level of *RASSF10*, the patients were divided into high expression group and low expression group. The difference of the *RASSF10* expression in collected specimens was analyzed by Student *t* test. Chi-squared test was used to analyze the relationship between the gene expression level and clinical characteristics of CRC patients. The association between *RASSF10* expression and overall survival was analyzed with Kaplan-Meier analysis. Hazard ratios (HRs) with the corresponding 95% confidence intervals (95% CIs). Besides, the prognostic significance of *RASSF10* was evaluated by cox regression analysis.  $P < .05$  was considered as statistical significance.

## 3. Results

### 3.1. Relative mRNA expression of RASSF10 in collected specimens

Quantitative RT-PCR was used to detect the relative mRNA expression of *RASSF10* in CRC tissues and adjacent normal tissues. As shown in Fig. 1, the expression level of *RASSF10* in tumor tissues was significantly lower than that in the adjacent normal tissues ( $0.215 \pm 0.093$  vs.  $0.974 \pm 0.126$ ,  $P < .001$ ).

### 3.2. Relative protein expression of RASSF10 in collected specimens

The protein expression of *RASSF10* in collected specimens was measured by ELISA analysis. The result demonstrated that *RASSF10* protein expression was decreased in tumor tissues compared to that in adjacent normal tissues ( $0.172 \pm 0.075$  vs.  $0.759 \pm 0.098$ ,  $P < .001$ , Fig. 2).

### 3.3. Clinical characteristics of CRC patients and their correlation with RASSF10 expression level

In order to analyze the correlation between *RASSF10* expression and clinical characteristics, the patients were divided into high

**Table 1**  
**Relationship between RASSF10 expression and clinical characteristics of patients with CRC.**

Characteristics	Cases, n	RASSF10 expression		$\chi^2$	P
		High, n	Low, n		
Gender				1.522	.217
Male	53	16	37		
Female	49	30	19		
Age				0.103	.748
>65	47	25	22		
≤65	55	21	34		
Histological type				0.813	.367
Well, moderate	56	23	33		
Poor, mucinous	46	23	23		
Depth of invasion				0.002	.969
T1 + T2	49	27	22		
T3 + T4	53	19	34		
Location				0.010	.919
Colon	56	21	35		
Rectum	46	25	21		
Lymph node metastasis				6.796	.009
Yes	50	34	16		
No	52	12	40		
TNM stage				5.511	.019
I + II	58	16	42		
III + IV	44	30	14		

CRC = colorectal cancer, RASSF10 = Ras association domain protein 10, TNM = tumor node metastasis.

expression group and low expression group according to RASSF10 average expression level ( $0.172 \pm 0.075$ ). Chi-squared test suggested that RASSF10 level was significantly correlated with lymph node metastasis ( $P = .009$ ) and TNM stage ( $P = .019$ ) (Table 1). However, there were no significant relationship between RASSF10 expression level and age, gender, histological differentiation, the depth of invasion, and tumor location.

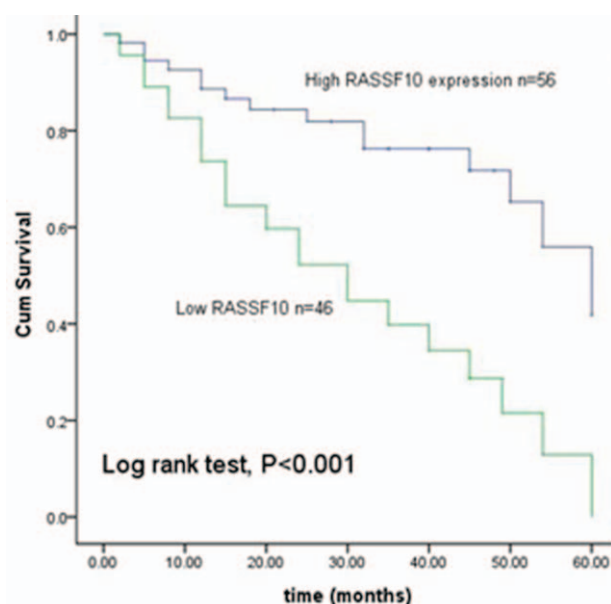
### 3.4. Prognostic value of RASSF10 in CRC

The overall survival of CRC patients with different expression of RASSF10 was analyzed by Kaplan–Meier analysis with log-rank test. The results indicated that patients with high expression level of RASSF10 had a longer overall survival than those with low expression level (48.9 vs. 35.9 months, log-rank test,  $P < .001$ , Fig. 3). Cox regression analysis was used to analyze the prognostic significance of RASSF10 in CRC and the results were listed in Table 2. The results of univariate analysis indicated that RASSF10 expression level was significantly correlated with CRC prognosis (HR = 3.060, 95% CI = 1.608–5.821,  $P = .001$ ). Then the multivariate cox regression analysis demonstrated that RASSF10 could act as an independent biomarker in the prognosis of CRC (HR = 3.333, 95% CI = 1.823–6.095,  $P < .0001$ ).

## 4. Discussion

CRC is one of the most common malignancies and the third-leading cause of cancer-related death, causing over 600,000 deaths every year all over the world.<sup>[15,16]</sup> Although great advance has been got in the treatment of CRC, the prognosis of CRC is still poor due to its frequently metastasis.<sup>[2]</sup> Therefore, it is necessary to explore effective molecular marker for the prognosis of CRC.

RASSF10 belongs to the RASSF family which play important roles in various pathological pathways such as microtubule



**Figure 3.** Kaplan–Meier analysis for estimating the overall survival of patients with different expression of RASSF10 in colorectal cancer. Patients with high RASSF10 expression had a longer overall survival than those with low expression (log-rank test,  $P < .001$ ). RASSF10 = Ras association domain protein 10.

stability, cell division, migration, apoptosis and adhesion, and modulating NF $\kappa$ B activity and the duration of inflammation.<sup>[17]</sup> According to the previous studies, we found that many of the family members were tumor suppressor genes, which were easily methylated then leading to the silencing of the according transcript in neoplasia.<sup>[18]</sup> In the study of Zhang et al,<sup>[19]</sup> the promote methylation and silencing of RASSF2 was detected in cervical cancer tissues which indicated that abnormal methylation of RASSF2 might involved in cervical carcinogenesis. Calvisi et al<sup>[20]</sup> had indicated that RASSF1A, RASSF2, and RASSF5 were significantly correlated with human hepatocellular carcinoma and inactivating these genes would inhibit the treatment of the cancer. Other RASSF family associated with cancers including neuroendocrine tumors of the lung, bladder cancer, gastric cancer, melanoma, CRC, nonsmall cell lung cancer, and so on were also covered.<sup>[21–24]</sup> As respect to the role of RASSF family in CRC, there were also some studies. For instance, in the study of Akino et al,<sup>[25]</sup>RASSF2 was proved to be a tumor inhibitor in CRC which played a pivotal role in the early stage of CRC via regulating Ras signaling. The aberrant promoter hypermethylation of RASSF5 was detected in colorectal tumorigenesis and the results suggested that the gene was correlated with colorectal tumor.<sup>[26]</sup> Fernandes et al<sup>[27]</sup> revealed that RASSF1A, RASSF2, and RASSF5 took part in CRC development, although the mechanisms of action remained poorly understood. Guo et al<sup>[14]</sup> indicated that reduced expression of RASSF10 was associated with RASSF10 promoter region methylation significantly in CRC leading to loss of expression. Despite RASSF10 was detected in CRC, its prognostic value was still unclear.

In this study, we detected the expression level of RASSF10 in CRC tissues and correspondingly normal tissues both at mRNA and protein levels. The present data indicated that RASSF10 expressed lower in CRC tissues compared with normal tissues. What's more, the expression level was significantly correlated

**Table 2****Univariate and multivariate analyses with cox regression analysis adjusted for the clinical factors for the prognostic value of RASSF10 in CRC patients.**

	Univariate analysis			Multivariate analysis		
	HR	95% CI	P	HR	95% CI	P
RASSF10 (low vs. high)	3.060	1.608–5.821	.001	3.333	1.823–6.095	<.0001
Age (>65 vs. ≤65)	1.109	0.611–2.014	.733	—	—	—
Gender (male vs. female)	1.131	0.595–2.150	.707	—	—	—
Histological differentiation (poor vs. well)	1.267	0.667–2.406	.470	—	—	—
The depth of invasion						
(T3 + T4 vs. T1 + T2)	1.115	0.597–2.083	.733	—	—	—
Location (colon vs. rectum)	0.783	0.420–1.459	.441	—	—	—
Lymph node metastasis (yes vs. no)	0.607	0.315–1.170	.136	—	—	—
Tumor stage (III + IV vs. I + II)	1.213	1.035–3.682	.045	—	—	—

CI = confidence interval, CRC = colorectal cancer, HR = hazard ratio, RASSF10 = Ras association domain protein 10.

with lymph node metastasis and TNM stage. These results suggested that RASSF10 might involved in the development of CRC.

In the study of Hill et al,<sup>[28]</sup> RASSF10 was proved to act as a prognostic marker for gliomagenesis. While according to the research of Deng et al,<sup>[29]</sup> we found that methylated RASSF10 promoter was an independent predictor for the survival of patients with gastric cancer. Therefore, we investigated the prognostic value of RASSF10 in CRC. First, we analyzed the overall survival of patients with CRC according to the expression level of RASSF10 through Kaplan–Meier analysis with log-rank test. The results showed that patients with low expression level of RASSF10 had a shorter overall survival than those with high expression level which revealed RASSF10 might be related to the prognosis of CRC. Then cox regression analysis was conducted to estimate the prognostic value of RASSF10 in CRC and the outcome showed it could be an independent prognostic indicator.

In conclusion, the expression level of RASSF10 is decreased in CRC tissues, compared with correspondingly normal tissues. In addition, the expression level is significantly associated with lymph node metastasis and TNM stage. Besides, we prove that RASSF10 may be a potential prognostic marker for patients with CRC.

## References

- [1] Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015;136:E359–86.
- [2] Jemal A, Bray F, Center MM, et al. Global cancer statistics. *CA Cancer J Clin* 2011;61:69–90.
- [3] Zhong R, Chen X, Zhu B, et al. MAD1L1 Arg558His and MAD2L1 Leu84Met interaction with smoking increase the risk of colorectal cancer. *Sci Rep* 2015;5:12202.
- [4] Grady WM, Carethers JM. Genomic and epigenetic instability in colorectal cancer pathogenesis. *Gastroenterology* 2008;135:1079–99.
- [5] Wolpin BM, Mayer RJ. Systemic treatment of colorectal cancer. *Gastroenterology* 2008;134:1296–310.
- [6] Reimers MS, Zeestraten EC, Kuppen PJ, et al. Biomarkers in precision therapy in colorectal cancer. *Gastroenterol Rep* 2013;1:166–83.
- [7] Sherwood V, Manboddh R, Sheppard C, et al. RASSF7 is a member of a new family of RAS association domain-containing proteins and is required for completing mitosis. *Mol Biol Cell* 2008;19:1772–82.
- [8] Richter AM, Pfeifer GP, Dammann RH. The RASSF proteins in cancer; from epigenetic silencing to functional characterization. *Biochim Biophys Acta* 2009;1796:114–28.
- [9] Wang Y, Ma T, Bi J, et al. RASSF10 is epigenetically inactivated and induces apoptosis in lung cancer cell lines. *Biomed Pharmacother* 2014;68:321–6.
- [10] Schagdarsurengin U, Richter AM, Wohler C, et al. Frequent epigenetic inactivation of RASSF10 in thyroid cancer. *Epigenetics* 2009;4:571–6.
- [11] Jin Y, Cao B, Zhang M, et al. RASSF10 suppresses hepatocellular carcinoma growth by activating P53 signaling and methylation of RASSF10 is a docetaxel resistant marker. *Genes Cancer* 2015;6:231–40.
- [12] Lu D, Ma J, Zhan Q, et al. Epigenetic silencing of RASSF10 promotes tumor growth in esophageal squamous cell carcinoma. *Discov Med* 2014;17:169–78.
- [13] Li Z, Chang X, Dai D, et al. RASSF10 is an epigenetically silenced tumor suppressor in gastric cancer. *Oncol Rep* 2014;31:1661–8.
- [14] Guo J, Yang Y, Linghu E, et al. RASSF10 suppresses colorectal cancer growth by activating P53 signaling and sensitizes colorectal cancer cell to docetaxel. *Oncotarget* 2015;6:4202–13.
- [15] Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. *CA Cancer J Clin* 2012;62:10–29.
- [16] Siegel R, Desantis C, Jemal A. Colorectal cancer statistics, 2014. *CA Cancer J Clin* 2014;64:104–17.
- [17] Volodko N, Gordon M, Salla M, et al. RASSF tumor suppressor gene family: biological functions and regulation. *FEBS Lett* 2014;588:2671–84.
- [18] Richter AM, Zimmermann T, Haag T, et al. Promoter methylation status of Ras-association domain family members in pheochromocytoma. *Front Endocrinol* 2015;6:21.
- [19] Zhang X, Ma Y, Wu Y, et al. Aberrant promoter methylation and silencing of RASSF2A gene in cervical cancer. *J Obstet Gynaecol Res* 2014;40:1375–81.
- [20] Calvisi DF, Evert M, Dombrowski F. Pathogenetic and prognostic significance of inactivation of RASSF proteins in human hepatocellular carcinoma. *Mol Biol Int* 2012;2012:849874.
- [21] Pelosi G, Fumagalli C, Trubia M, et al. Dual role of RASSF1 as a tumor suppressor and an oncogene in neuroendocrine tumors of the lung. *Anticancer Res* 2010;30:4269–81.
- [22] Meng W, Huebner A, Shabsigh A, et al. Combined RASSF1A and RASSF2A promoter methylation analysis as diagnostic biomarker for bladder cancer. *Mol Biol Int* 2012;2012:701814.
- [23] Maruyama R, Akino K, Toyota M, et al. Cytoplasmic RASSF2A is a proapoptotic mediator whose expression is epigenetically silenced in gastric cancer. *Carcinogenesis* 2008;29:1312–8.
- [24] van der Weyden L, Adams DJ. The Ras-association domain family (RASSF) members and their role in human tumorigenesis. *Biochim Biophys Acta* 2007;1776:58–85.
- [25] Akino K, Toyota M, Suzuki H, et al. The Ras effector RASSF2 is a novel tumor-suppressor gene in human colorectal cancer. *Gastroenterology* 2005;129:156–69.
- [26] Lee CK, Lee JH, Lee MG, et al. Epigenetic inactivation of the NORE1 gene correlates with malignant progression of colorectal tumors. *BMC Cancer* 2010;10:577.
- [27] Fernandes MS, Carneiro F, Oliveira C, et al. Colorectal cancer and RASSF family—a special emphasis on RASSF1A. *Int J Cancer* 2013;132:251–8.
- [28] Hill VK, Underhill-Day N, Krex D, et al. Epigenetic inactivation of the RASSF10 candidate tumor suppressor gene is a frequent and an early event in gliomagenesis. *Oncogene* 2011;30:978–89.
- [29] Deng J, Liang H, Ying G, et al. Methylation of Ras association domain protein 10 (RASSF10) promoter negative association with the survival of gastric cancer. *Am J Cancer Res* 2014;4:916–23.