

Congenital leukemia of fetus with acquired *AML1* gene duplication

Sun-Young Kang¹, Young-Joo Lee¹, Kyung Hee Park², So-Eun Jun², Seung-Chul Kim¹, Jong-Kil Joo¹, In-Suk Kim³, Eun-Yup Lee³

Departments of ¹Obstetrics and Gynecology, ²Pediatrics, and ³Laboratory Medicine, Pusan National University School of Medicine, Busan, Korea

Congenital leukemia is very rare, and its prevalence according to recently published papers is from 1 to 5 per million live births. This can be often diagnosed in postpartum throughout bone marrow biopsy, showing abnormal proliferation of immature blasts and granulocytic precursors. Hepatosplenomegaly is the most common feature which is found during perinatal examinations, that diagnosing is difficult during perinatal period. Hepatosplenomegaly can occur not only in congenital leukemia but in many other cases such as infection which is the most common cause. In other words, congenital leukemia is the one of the rare causes of hepatosplenomegaly. However, this case shows the fetus with the features of hepatosplenomegaly during perinatal period and being diagnosed as congenital leukemia associated with acquired *AML1* gene duplication in postpartum through bone marrow biopsy. Due to its rare instance, we are to describe the case with a review of literatures.

Keywords: *AML1* gene; Duplication; Congenital leukemia; Fetal; Hepatosplenomegaly

Introduction

Congenital leukemia (CL) is a very rare disease that is seen from 1 to 5 per million live births, which develops in utero and usually is diagnosed at birth or within one month of life because of the rapid doubling time of leukemic cells [1,2]. It is a unique disease that most of them are of myeloid origin in contrast to general pediatric leukemia, which are usually lymphoid in origin [3]. Clinical manifestations of CL are similar to infectious diseases, therefore it is obscure to make a precise diagnosis, for which further evaluation and diagnostic tools are necessary [4]. Antenatally, diagnosis of CL cannot be made easily due to high risk of fetoscopy or cordocentesis, but fetal hepatosplenomegaly observed by ultrasonography can provide a clue for diagnosis of CL. Fetal hepatosplenomegaly may be presented in various conditions, most commonly in Rh isoimmunization and other forms of fetal anemia, congestive heart disease, infectious disorders such as cytomegalovirus, metabolic disorders such as Gaucher disease or pyruvate kinase deficiency, and transient myeloproliferative disease (TMD) which sometimes appears in fetuses with Down syndrome and overgrowth disorder [5]. In this report, we are to present a case of an incidental hepatosplenomegaly of fetus com-

plicated with acute myeloid leukemia associated with *AML1* gene duplication, which was diagnosed by bone marrow aspiration for identification of the cause of a newborn's leukemoid reaction.

Case report

A 36-year-old woman, had undergone in vitro fertilization and embryo transfer, was referred to our institution for evalu-

Received: 2014.2.3. Revised: 2014.3.13. Accepted: 2014.4.1.
Corresponding author: Seung-Chul Kim
Department of Obstetrics and Gynecology, Pusan National University School of Medicine, 305 Gudeok-ro, Seo-gu, Busan 602-739, Korea
Tel: +82-51-240-7282 Fax: +82-51-248-2384
E-mail: ksch0127@naver.com

Articles published in Obstet Gynecol Sci are open-access, distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © 2014 Korean Society of Obstetrics and Gynecology

ation of fetal hepatosplenomegaly at 37 weeks' gestation. The index pregnancy was otherwise uncomplicated. Her past history was uneventful, and the results of antenatal laboratory tests were normal. A fetal heart rate was about 150 beats per minute. Fluctuations in the fetal heart rate are called variability were normal without distress sign. A transabdominal ultrasonography with 3.5-5 MHz transducer (Voluson 730 pro, GE, Seoul, Korea) revealed a single, live fetus with measurements corresponding to the gestational age except abdominal circumference, due to hepatosplenomegaly. Findings of fetal hydrops also were not observed. A total amniotic fluid index measured 17.5 cm which is normal range. Hepatomegaly means that the liver reaches into the fetal pelvis adjacent to the fetal bladder. Length of liver in this period is normally varies from 40 to 60 mm. It can be measured from the dome of the diaphragm to the tip of the right lobe parallel to the long axis of the fetal trunk [6]. Splenomegaly is diagnosed by three findings. First, the stomach is displaced anteriorly and to the midline by the enlarged spleen. Second, the spleen is very close to the anterior abdominal wall. Third, color Doppler imaging depicts multiple ramifications of the splenic artery and vein. Length of spleen in this period is normally measured from 45 to 60 mm [7]. Echogenicity of both organs is usually isoechogenic. However, in this case, size of liver and spleen were above 95th percentile of the upper limit and also the Fig. 1 shows both organs adjacent to bladder, which indicates fetal hepatosplenomegaly. Although limited clinical informa-

tion was available to discuss the ideal mode of delivery with the parents, we recommended cesarean delivery next day to prompt evaluation of rapid progression of fetal hepatosplenomegaly. At delivery, the newborn, a male, weighted 3,070 g was born and Apgar scores were 5 and 7 at 1 and 5 minutes, respectively. When the newborn was physically examined, it revealed unstable respiration with tachypnea and desaturation due to abdominal distension. Intubation was immediately performed and the baby was mechanically ventilated in the neonatal intensive care unit. The abdomen was moderately distended and liver and spleen were palpable 4 and 2 cm below the costal margins. No skin lesions were present (e.g., twenty five to thirty percent of infants with CL have specific cutaneous infiltrates; leukemia cutis which usually appear as firm blue or red nodules called Blueberry Muffin) [8].

On laboratory tests performed at birth, the hemoglobin concentration was 13.3 g/dL, hematocrit was 42.9%, platelet cells count was $228 \times 10^3/\text{UL}$, and white blood cells count was $28.18 \times 10^3/\text{UL}$. A differential cell count revealed 25% neutrophils, 44% lymphocytes, 4% monocytes, 3% eosinophils, 0% basophils, 1% myelocytes, and 23% atypical lymphocyte. Few atypical cells were observed on peripheral blood smear and initial peripheral blood chromosome study reveals normal karyotype. Also, serology studies for TORCH syndrome, including serum antibody titers against toxoplasmosis, cytomegalovirus, rubella, and herpes simplex virus and venereal disease research laboratory tests were all negative ruling out infectious condition. Another work-up for septic condition and tandem mass for metabolic disorder were all negative. While further evaluation and monitoring were performed, aspects of hepatic failure were consistently shown with elevated total bilirubin (7.38 mg/dL) in spite of exchanged transfusion for prevention of kernicterus. On peripheral blood test performed on 28 days after birth, the hemoglobin concentration was 10.7 g/dL, hematocrit was 33.4%, platelet cells count was $123 \times 10^3/\text{UL}$, and white blood cells count was $67.94 \times 10^3/\text{UL}$. A differential cell count revealed 20% neutrophils, 6% lymphocytes, 22% monocytes, 19% eosinophils, 3% basophils, 0% myelocytes, and 30% atypical lymphocyte. Increased numbers of atypical cells which are large-sized immature blasts were seen in a peripheral blood smear (Fig. 2A), therefore bone marrow aspiration was performed. A bone marrow aspirate smear showed a moderately cellular marrow with abnormal proliferation of immature blasts ($\geq 35\%$) and granulocytic precursors and negative for myeloperoxidase, Sudan black B and periodic

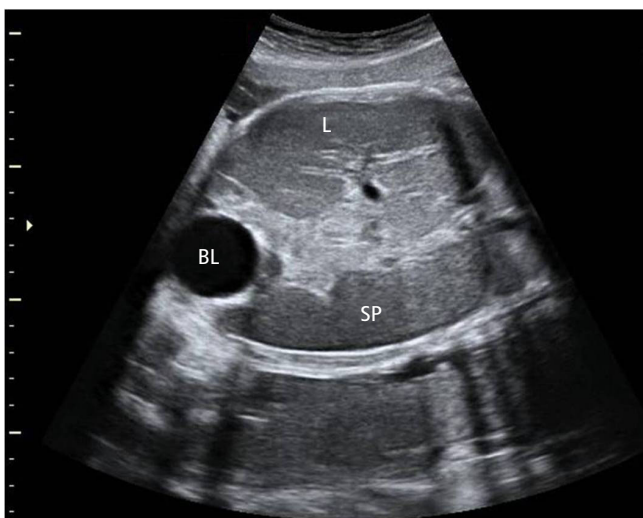


Fig. 1. Ultrasound scan shows liver and spleen adjacent to bladder, which indicates hepatosplenomegaly. L, liver; BL, bladder; SP, spleen.

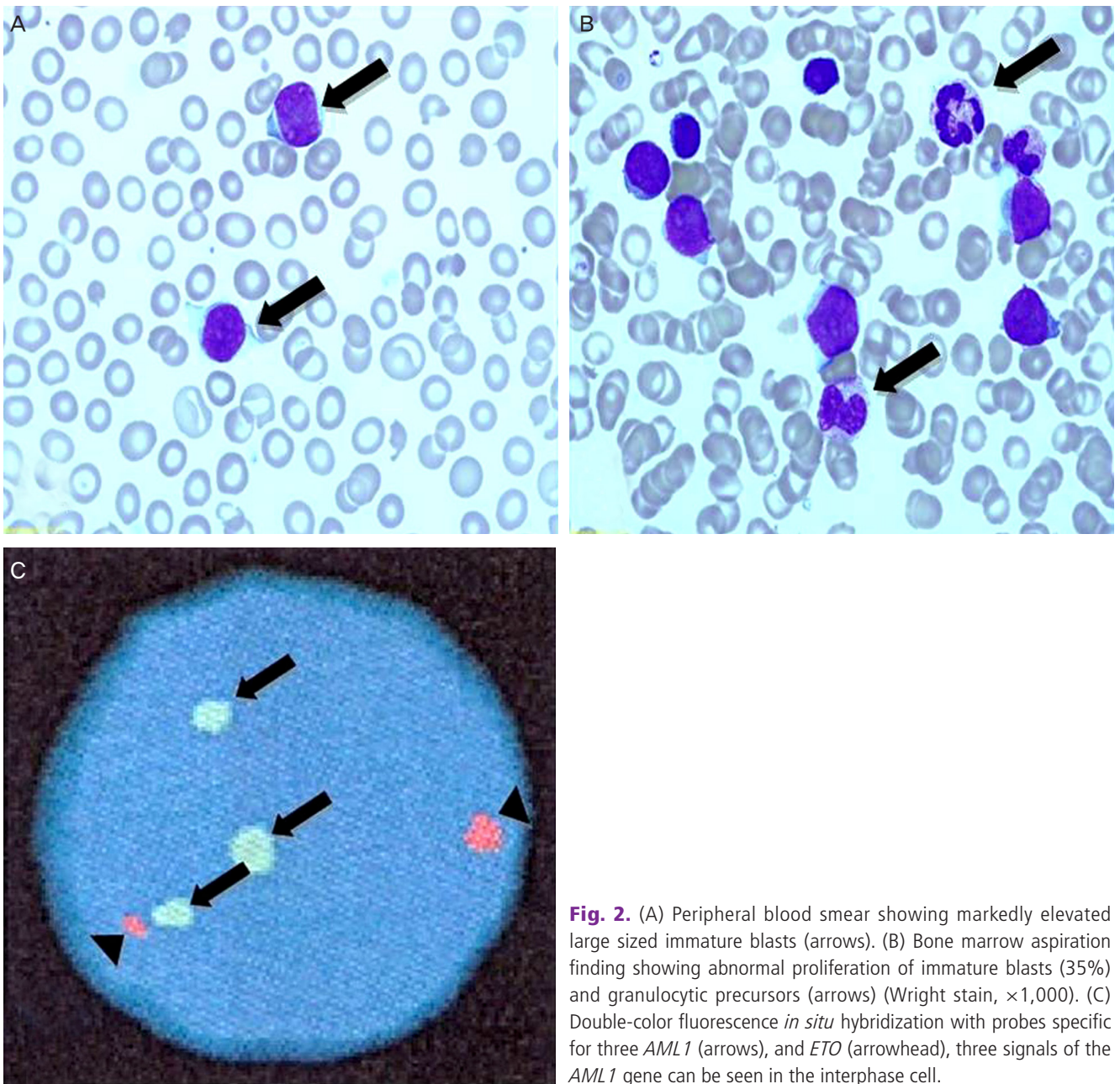


Fig. 2. (A) Peripheral blood smear showing markedly elevated large sized immature blasts (arrows). (B) Bone marrow aspiration finding showing abnormal proliferation of immature blasts (35%) and granulocytic precursors (arrows) (Wright stain, $\times 1,000$). (C) Double-color fluorescence *in situ* hybridization with probes specific for three *AML1* (arrows), and *ETO* (arrowhead), three signals of the *AML1* gene can be seen in the interphase cell.

acid Schiff stain. The erythroid precursors and megakaryocytes were decreased in number (Fig. 2B). Immunophenotyping revealed that the 75% of blasts were positive for myeloid leukemic marker of CD33+. The final diagnosis was acute myeloid leukemia. Work-up for detecting metastasis of leukemic cell was done, and there were no specific finding except small ventricular septal defect in computed tomography angiography.

To figure out the cause of CL, especially for recurrent genet-

ic abnormalities in de novo *AML*, fluorescence *in situ* hybridization (FISH) for core-binding factor subunit beta, myeloid lymphoid lineage, *AML1/ETO* gene rearrangement were done. Although recurrent genetic abnormalities were not detected, 94% of three *AML1* gene signals which could be interpreted as trisomy 21 were detected in this bone marrow specimen. The *AML1* (*CBFA2*) gene, located in the chromosomal band 21q22, has recently attracted a lot of interest in terms of its role in leukemogenesis [9]. Retrospectively, to investigate

the three *AML1* signals on initial peripheral blood specimen shown normal karyotype, we performed the FISH, *AML1/ETO* gene rearrangement. FISH results showed 50% three *AML1* signals on initial peripheral blood specimen. Based on the presence of three *AML1* signals with normal karyotype in initial study, this patient was thought to have a clonal change of cryptic three *AML1* gene signals carrying normal karyotype, and in other words, this result can confirm the presence of CL with *AML1* gene duplication. Three *AML1* clonal progressions were noted in bone marrow specimen (Fig. 2C). Therefore, this patient had very low possibility of Down syndrome except the case of very low level of mosaic Down syndrome.

Even though the infant was treated with chemotherapy (idarubicin, cytarabine, etoposide, and daunorubicin), ventilator care and inotropic agents, the baby developed increasing respiratory distress and hypotension. He died of respiratory distress with pneumonia at 6 weeks after birth.

Discussion

CL becomes clinically evident at birth or within the first month of life because of the rapid doubling time of leukemic cells. It is a rare entity, with reported incidence that ranges from 1 to 5 per million live births [1,2].

To fulfill the criteria of CL, first, there must be manifest at or shortly after birth (<30 days) and second, the blood should reflect an alteration of the bone marrow by showing the presence of immature cell proliferation of myelomonocytic, lymphoid, or erythroid series or in extrahematopoietic tissue. Third, there should be absence of any other condition that might cause leukemoid reactions mimicking CL, such as, congenital infections (TORCH), hemolytic disease of the newborn (HDN), hereditary spherocytosis, twin to twin transfusion, other neoplastic infiltrates and TMD, a nonmalignant leukocytosis seen usually in neonates with Down syndrome [8,10]. Therefore, to perform accurate diagnosis, differential diagnosis is very important. In this case, the infant could be diagnosed as congenital leukemia satisfying all three conditions mentioned above.

The causes of CL are still unknown, but it has been associated with exposure to radiation, dietary compounds called bioflavonoid, use of tobacco and illicit drugs, and other substances, as well as coexisting congenital anomalies and the possibility of an underlying genetic predisposition in infants

born with leukemia [10]. In this case, this patient was thought to have a clonal change of cryptic three *AML1* signals carrying normal karyotype. Therefore, this result can confirm the presence of CL with *AML1* gene duplication.

CL is often associated with hepatosplenomegaly, petechia, ecchymosis, lethargy, pallor and poor feeding at or shortly after birth and most infants die of respiratory distress which is a result of pulmonary leukostasis and bronchopneumonia [11]. In this case, there was no specific skin lesion found except hepatosplenomegaly at birth and the baby died of respiratory distress cause by pneumonia. Furthermore, autopsy was needed for diagnosis of pulmonary leukostasis and bronchopneumonia. However, it could not be done due to parents' rejection.

The differential diagnosis should include sepsis, intra-uterine infections (TORCH), HDN and TMD which is seen usually in association with Down syndrome. Historically, TMD has been described as a benign condition association with transient polycythemia and thrombocytosis, because the leukocytosis and its secondary clinical complications typically disappear without treatment over the first 2 months of life [4,10]. In this case, infectious diseases were ruled out through serologic studies and negative bacterial cultures, and HDN was also excluded through decreased numbers of erythroid precursors in peripheral blood. In addition, TMD which sometimes appears in fetuses with Down syndrome was excluded through identifying the presence of three *AML1* signals carrying normal karyotype.

Once the diagnosis of leukemia is established, an intensive multi-agent chemotherapeutic regimen should be instituted. There is no specific therapeutic protocol for the treatment of either neonatal acute lymphoid leukemia or AML. Whether they undergo chemotherapy or not, neonates with leukemia usually require treatment in an intensive care unit [12].

CL has a poor prognosis, with only 23% surviving at 24 months [13,14]. However, a few patients were described that they have undergone spontaneous remission of CL without chemotherapy [15].

In conclusion, if during perinatal period, hepatosplenomegaly, hydrops and polyhydramnios by ultrasonography can be detected, perinatologist should consider various possibilities such as infectious disease, hemolytic disorder, metabolic disorder and TMD associated with Down syndrome. If it is possible, fetoscopy and umbilical blood sampling, which are not preferred to be performed, is needed. Finally perinatologist can

then establish the diagnosis of leukemia beyond performing differential diagnosis.

We would like to report this case, which shows the fetus with the features of hepatosplenomegaly during perinatal period, on 37 weeks' of gestational age, and being diagnosed as CL associated with acquired *AML1* gene duplication after birth through bone marrow aspiration.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

This work was supported by clinical research grant from Pusan National University Hospital (2012).

References

1. Bader JL, Miller RW. US cancer incidence and mortality in the first year of life. *Am J Dis Child* 1979;133:157-9.
2. Badhe PB, Sane SY. Congenital leukemia: organ involvement in six autopsy cases. *J Postgrad Med* 1992;38:127-9.
3. Bargoitra R, Suri J, Gupta Y. Congenital leukemia. *JK Sci* 2010;12:201-2.
4. Prakash KP, Rau AT, Bhat ST, Rau AR. Congenital leukemia: a diagnostic dilemma. *Indian J Med Paediatr Oncol* 2008;29:41-3.
5. Bronshtein M, Blazer S, Zimmer EZ. The gastrointestinal tract and abdominal wall. In: Callen PW, editor. *Ultrasonography in obstetrics and gynecology*. 5th ed. Philadelphia: Saunders Elsevier; 2008. p.615-23.
6. Vintzileos AM, Neckles S, Campbell WA, Andreoli JW Jr, Kaplan BM, Nochimson DJ. Fetal liver ultrasound measurements during normal pregnancy. *Obstet Gynecol* 1985;66:477-80.
7. Schmidt W, Yarkoni S, Jeanty P, Grannum P, Hobbins JC. Sonographic measurements of the fetal spleen: clinical implications. *J Ultrasound Med* 1985;4:667-72.
8. Choi JH, Lee HB, Park CW, Lee CH. A case of congenital leukemia cutis. *Ann Dermatol* 2009;21:66-70.
9. Harewood L, Robinson H, Harris R, Al-Obaidi MJ, Jalali GR, Martineau M, et al. Amplification of *AML1* on a duplicated chromosome 21 in acute lymphoblastic leukemia: a study of 20 cases. *Leukemia* 2003;17:547-53.
10. Sande JE, Arceci RJ, Lampkin BC. Congenital and neonatal leukemia. *Semin Perinatol* 1999;23:274-85.
11. Nathan DG, Oski FA. Congenital leukemia. In: Nathan DG, Oski FA, editors. *Hematology of infancy and childhood*. 3rd ed. Philadelphia: Saunders; 1987. p.1052-3.
12. Van der Linden MH, Creemers S, Pieters R. Diagnosis and management of neonatal leukaemia. *Semin Fetal Neonatal Med* 2012;17:192-5.
13. Bresters D, Reus AC, Veerman AJ, van Wering ER, van der Does-van den Berg A, Kaspers GJ. Congenital leukaemia: the Dutch experience and review of the literature. *Br J Haematol* 2002;117:513-24.
14. Isaacs H Jr. Fetal and neonatal leukemia. *J Pediatr Hematol Oncol* 2003;25:348-61.
15. Grundy RG, Martinez A, Kempfski H, Malone M, Atherton D. Spontaneous remission of congenital leukemia: a case for conservative treatment. *J Pediatr Hematol Oncol* 2000;22:252-5.