

Review Article

The ecotoxicological consequences of microplastics and co-contaminants in aquatic organisms: a mini-review

 Farhan R. Khan¹, Ana I. Catarino² and Nathaniel J. Clark^{3,4}

¹Department of Climate & Environment, Norwegian Research Center (NORCE), Nygårdsporten 112, NO-5008 Bergen, Norway; ²Vlaams Instituut voor de Zee, Flanders Marine Institute InnovOcean site, Wandelaarkaai 7, 8400 Oostende, Belgium; ³School of Biological and Marine Sciences, University of Plymouth, Plymouth PL4 8AA, U.K.; ⁴School of Health Professionals, University of Plymouth, Plymouth PL4 8AA, U.K.

Correspondence: Farhan R. Khan (fakh@norce-research.no; farhan.khan@gmx.com)



Microplastics (MPs, <5 mm in size) are a grave environmental concern. They are a ubiquitous persistent pollutant group that has reached into all parts of the environment — from the highest mountain tops to the depths of the ocean. During their production, plastics have added to them numerous chemicals in the form of plasticizers, colorants, fillers and stabilizers, some of which have known toxicity to biota. When released into the environments, MPs are also likely to encounter chemical contaminants, including hydrophobic organic contaminants, trace metals and pharmaceuticals, which can sorb to plastic surfaces. Additionally, MPs have been shown to be ingested by a wide range of organisms and it is this combination of ingestion and chemical association that gives weight to the notion that MPs may impact the bioavailability and toxicity of both endogenous and exogenous co-contaminants. In this mini-review, we set the recent literature within what has been previously published about MPs as chemical carriers to biota, with particular focus on aquatic invertebrates and fish. We then present a critical viewpoint on the validity of laboratory-to-field extrapolations in this area. Lastly, we highlight the expanding ‘microplastic universe’ with the addition of anthropogenic particles that have gained recent attention, namely, tire wear particles, nanoplastics and, bio-based or biodegradable MPs, and highlight the need for future research in their potential roles as vehicles of co-contaminant transfer.

Introduction — the case for microplastics as a vehicle for chemicals

‘Microplastics’ (MPs) is used as a catch-all term to represent a complex variety of properties that arise during both the manufacturing process and following release into the environment [1]. Plastics are composed of different organic polymers to which an array of chemicals (termed here as ‘endogenous chemicals’, e.g. plasticizers, colorants, fillers and stabilizers) are added to enhance certain properties, such as rigidity, malleability, or thermal resistance and prolonging life [2]. Depending on use, plastics are produced within the MP size range of <5 mm (primary MPs such as microbeads) or into larger products that can subsequently breakdown releasing MPs (secondary MPs) [3,4]. MPs in the environment exist as a heterogeneous mixture of physical and chemical properties [1,4,5] and undergo several environmental transformations such as weathering, fragmentation, and biofilm and microbial colonization [6–8].

These transformations will affect how MPs interact within their environment — an environment that already contains a plethora of potential co-contaminants (termed here as ‘exogenous chemicals’). Tens of thousands of chemical entities are found on the global market with approximately 2000 new

Received: 13 June 2022
Revised: 28 July 2022
Accepted: 29 July 2022

Version of Record published:
16 August 2022

chemicals added each year [9]. The widespread ingestion of MPs, documented across aquatic taxa [10–12], provides a pathway for both endogenous and exogenous chemicals to enter the organism. Yet the role of MPs as chemical vectors not only relies upon the association with co-contaminants and the influence of the ambient environment, but considerations of the organism's biology and physiology are also important. For instance, feeding modes, gut retention times and digestive physiology will all play a role in whether the MP-associated chemical is bioaccessible and then bioavailable to the organism [5,13]. Toxicological consequences may result if the co-contaminants reach and become available at sites of biological activity.

The role of MPs as chemical carriers has been the subject of much investigation, debate and speculation (see reviews [5,14–18]), but consensus remains elusive. The interactions of MPs, environment, chemicals and biota are summarized in Figure 1, and whilst laboratory studies can only investigate a portion of this complexity at one time, it is precisely this complexity that has made the role MPs in co-contaminant transfer one of the most studied and divergent topics in MP research.

Biotic effects of microplastics and co-contaminant exposure

Endogenous chemicals

During the manufacturing of plastics various substances often termed as 'additives' are combined with the polymeric resins to improve the properties of final applications and a few other reaction by-products will be further accidentally incorporated [2,19,20]. When plastic debris reach aquatic environments, these endogenous substances, some of which are known to be toxic to biota, can migrate from the resin to the external medium, as substances are often physically, rather than chemically, bonded to the main polymer matrix [20]. The leaching of substances from MPs may occur at higher rates compared with macroplastic litter due to their increased surface area to volume ratio. The majority of organic additives have a low hydrophobicity, or low octanol–water partition coefficient (K_{ow}), and a low molecular mass [21]. Therefore, exposure of biota can be low due to the low diffusion of organic chemicals from the plastic to the water [21,22], as additives with a higher potential for toxicity and bioaccumulation have a higher K_{ow} [19].

Some endogenous chemicals found in aquatic environments are known to have toxicological properties, such as phthalates, bisphenol A (BPA), nonylphenol (NP) and brominated flame retardants (BFR) [19], as well as trace metals such as cadmium (Cd), lead (Pb), antimony (Sb) and tin (Sn) [2]. Of concern are the leaching rates of endogenous chemicals from weathered MPs [20], in particular when in gastrointestinal fluids during digestion where high levels of surfactants and lower pH may facilitate the migration process of compounds from the plastic resin [23]. Gut retention times of MPs vary amongst invertebrate species depending on physiology and relative MP size and shape and can last between a few hours and weeks. During this period, endogenous substances will increase in their bioaccessibility and bioavailability due to a low pH environment which enhances leaching from the polymeric matrix and further due to an affinity to fatty tissues of hydrophobic endogenous chemicals [22]. For example, Kühn et al. [24] demonstrated *in vitro* that environmental MPs will leach additives to stomach oil of northern fulmars. Tanaka et al. [25] demonstrated that feeding seabird chicks with MPs spiked in their resin with additives induced accumulation at 101–105 times above baseline in the tested individuals. However, a modeling exercise demonstrated that MPs may have a residual contribution to the accumulation and toxicity induced by leachates in the intestinal tracts of lugworms and in the North Sea cod when compared with sources of these contaminants such as water or sediments, but *in vivo* experimental validation is still required [22].

Exogenous chemicals

MPs enter an environment that already contain a chemical cocktail, including hydrophobic organic contaminants (HOCs i.e. polycyclic aromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs)) [26,27], trace metals [28,29] and pharmaceuticals [30–32], all of which have been measured on MPs collected from the environment. These exogenous chemicals can 'sorb' (covering both surface adsorption and internal partition) to the MP based on the structure of the polymer (i.e. its ratio of crystalline and amorphous regions) [33]. The ingestion of the MPs can thus change the route of uptake compared with the dissolved form of the exogenous chemical from waterborne to dietary exposure [34] and following desorption within an organisms' digestive system there is potentially a greater level of chemical exposure [35]. Accordingly, this so-called 'vector-effect' [3] has been the subject of much research and speculation, but even the earliest investigations demonstrated

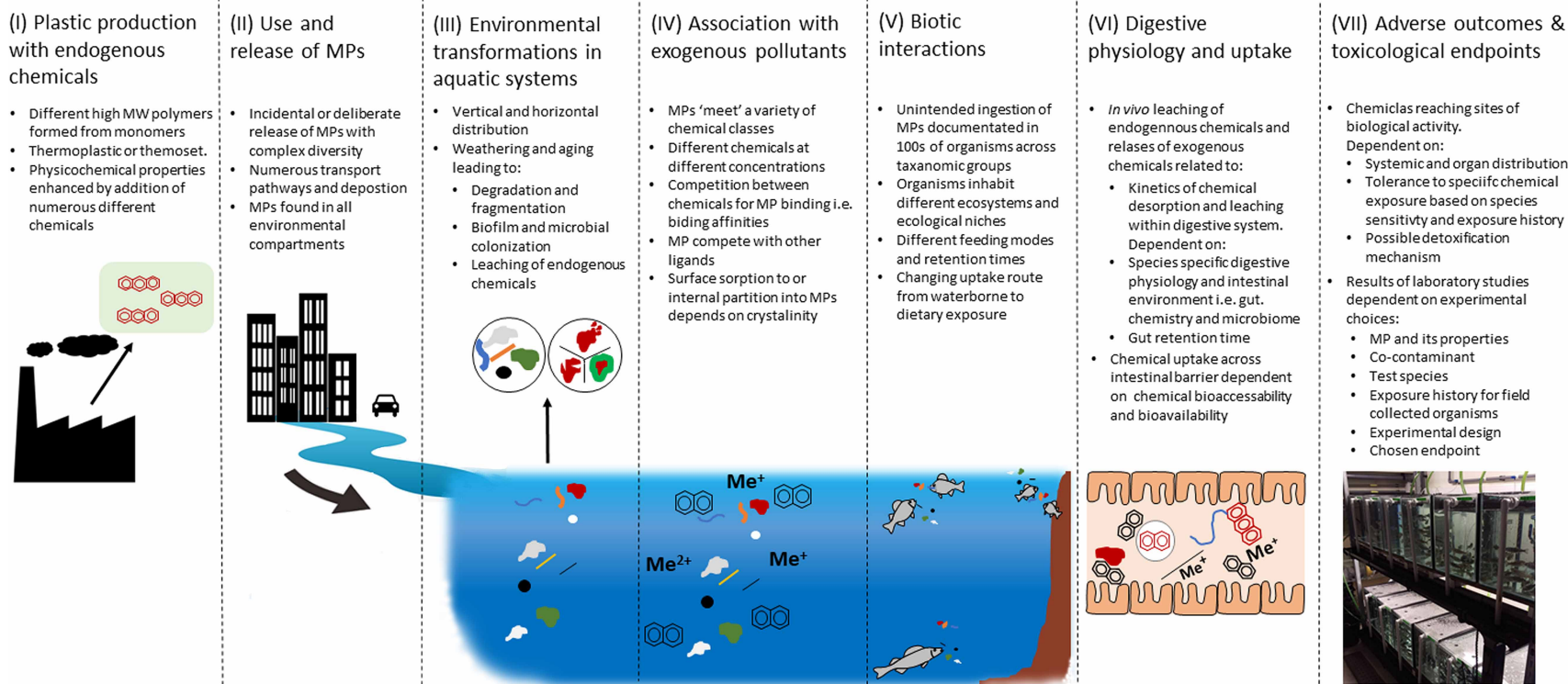


Figure 1. Schematic diagram of the complexity involved in the association of MPs with co-contaminants.

Starting at the production stage where a range of endogenous chemical 'additives' (schematic aromatic rings in red) are incorporated into the polymeric resin (green) (I). MPs entering or produced through breakdown in the environment are a complex suite of physico-chemical properties (II) which are subject to the environmental transformations (III). MPs are known to sorb of exogenous pollutants, such as hydrophobic organic pollutants (schematic aromatic rings, black) and metals (Me^+ and Me^{2+} black) (IV) and be ingested by biota (V). Within the digestive system endogenous additives (red) leach out of the MP and sorbed exogenous pollutants (black) desorb. The released chemicals may then be available for uptake (VI). The onset of toxicological outcomes will depend on the further transport of the chemicals to sites of biological activity and the ecotoxicological assessments of effects will depend on experimental choices made (VII). The complexity and array of variables presented here highlights why the role of MPs as chemical carriers is still debated. Schematic based on Khan et al. [5].

the varying nature of the vector phenomenon where MPs enhanced the bioavailability and toxicity of co-contaminants [36,37], where the addition of MPs to the exposure scenario resulted in negligible impacts [38], and cases where MPs reduced pollutant bioavailability [39].

The scientific literature in this area is too vast to comprehensively cover in this short review (see recent reviews [5,14–18]); however, recent descriptions of vector effects continue to vary. Two approaches are used to determine vector effects, (i) to directly measure chemicals in tissues to determine the influence of MPs to the exposure scenario and (ii) measure a toxicological marker of exposure as an indicator of chemical bioavailability and biological reactivity. Numerous laboratory studies have reported that exogenous contaminants sorbed to MPs are bioavailable and lead to a measurable transfer into the tissue [40–42] and toxicological impacts based upon the assessment of biomarkers [42–44], even if the tissue burdens did not correspondingly increase [45–47]. Conversely, several studies have demonstrated that at least for some measured endpoints, the role of MPs on chemical-induced negative impacts is not significant [48–52]. Using a novel ‘feeding tube’ method to directly introduce polyethylene and polystyrene MPs loaded with PCB-153 into the digestive tract of fish larvae showed no transfer of the PCB from MP into the tissue [53]. In the exposures of *Talitrus saltator* MPs were shown to carry HOCs into the tissue following ingestion, but when uncontaminated MPs were fed to sand hoppers, then the MP scavenged the chemicals and reduced the tissue burden [54]. Thus, under some circumstances, MP ingestion can potentially perform a ‘cleaning effect’ [54,55]. The transfer of PCBs from MPs under simulated gut fluid conditions was demonstrated to be biphasic and reversible [55].

The combination of MPs and co-contaminants may be viewed as similar to the interactions within a chemical mixture — independent or dependent action, or additivity, synergism or antagonism [56]. The joint exposure of MPs and the pharmaceutical triclosan to marine microalgae induced antagonistic effects with increasing MP concentrations reducing the triclosan toxicity based on the adsorption of the chemical to the plastic surface [57]. However, the co-exposure of the marine copepod *Acartia tonsa* to polyethylene microbeads and triclosan resulted in a relatively obscure mixture effect known as potentiation in which the MP, without itself being toxic, enhanced the toxicity of triclosan [56]. Thus, even when assessed through a recognized framework designed to disentangle the effects of single components within a mixture, the MP vector effect does not provide consistent outcomes. Though little used in MP research, the analysis of mixtures may provide a mechanistic insight into the individual roles of each competent within the MP-co-contaminant combination and further attention with this approach would be warranted.

Focus on digestive physiology

Recognizing that there are important differences between all the studies described in the preceding sections that impact the outcome — choices relating to MP properties, co-contaminant, species, experimental design and biological endpoint (see Figure 1) — there remains disparity in descriptions of the MPs a chemical carrier which goes beyond the interactions and sorptive behavior in the test media and needs to consider physiology, particularly that of the intestinal environment. If MP ingestion is the assumed route of entry, then there are two possibilities for MPs and co-contaminants to enter the gastrointestinal tract (GIT) of aquatic animals — independently or with the co-contaminant associated (sorbed) to the MP [58]. In the GIT the fate of the MP and co-contaminant to remain independent, remain sorbed or desorb is largely driven by the gut lumen environment. Perhaps the earliest study to investigate HOC desorption in simulated gut conditions showed pH and temperature were important factors in determining desorption rates, suggesting that warm-blooded animals could be of the greater threat of MP-facilitated HOC transfer, but desorption also occurred in cold-blooded conditions representative of fish and invertebrates [59]. Recent follow-up studies have also demonstrated that both endogenous and exogenous chemicals separate from the MP within intestinal and biological fluids and conditions [13,60,61].

However, the lumen of the gastrointestinal tract (GIT) is a dynamic environment that varies between species and within species. For instance, the luminal pH of the polychaete worms *Lumbriculus variegatus* and *Arenicola marina* are 5.4–6.5 and 6.8–7.2, respectively, whereas carnivorous fish, such as rainbow trout (*Oncorhynchus mykiss*) exhibit a wider range pH 2.0–8.5 [62]. In the latter, the GIT is compartmentalized into different anatomical regions, with an acidic lumen in the stomach and alkali lumen in the intestinal regions. The pH is a main driver for determining the partitioning of chemicals onto the surface of MPs for ionizable organic chemicals [63] and dissolved metals [64,65], typically with lower pH values causing less chemical to bind to the surface of the MPs. For fish, at least, the variation in pH along the lumen of the GIT creates the potential for the cycling of chemicals on and off the MP [55]. Temperature, salinity and ionic strength have

been shown to affect the sorption behavior of co-contaminants to MPs [15]. However, determining the relative contribution of each GIT parameter to the potential for vector effects and co-contaminant transfer is difficult *in vivo*, but a greater understanding of the role of species-specific digestive physiology is paramount to better understand the toxicological effects of MP and co-contaminant exposures.

Laboratory-to-field extrapolation of MP co-contaminant studies

The ecotoxicological consequences of MPs and co-contaminants have largely been studied within laboratory settings. In extrapolating those findings to the natural world, two pertinent questions need to be addressed: (i) do laboratory studies realistically reflect the complexity of MPs in the environment and (ii) are MPs relevant chemical carriers compared with other potential sorbents? It is now established that in the environment, MPs are neither just pristine nor just contaminated, but rather exist in a continuum as a class of complex pollutants from different polymer types, shapes and sizes, at different levels of environmental transformations, and which can leach or sorb a multitude of chemicals (Figure 1) [1,5,18,66]. Despite this, most co-contaminant experimental studies employ aspects that lack environmental relevance; the use of pristine MPs, single polymers and MP types (e.g. the overuse of polystyrene spheres [67]) at levels above field concentrations coupled with single pollutants, short equilibrium times or methods to artificially hasten sorption kinetics [5,33]. Thus, studies with different MP morphologies are needed to reflect environmental prevalence [68] and as different types may exhibit different gut passage times which may affect chemical transfer from MP to tissue. Natural ageing (i.e. weathering) of MPs increases their adsorption affinity towards contaminants, but this parameter has seldom been considered in the effect assessments of MPs. The weathering of plastics in environmental settings is affected by exposure to UV radiation (sunlight), temperature shifts, humidity, and oxygen and ozone levels [69–71], and in turn the weathering of MPs can further play an important role in the leachates released and toxicity to organisms [20,72,73]. Furthermore, as climatic conditions shift due to global change (e.g. lower pH, increased temperature and fluctuating salinities) the impact of such parameters should be better linked to plastic pollution [66,74].

The aspect of relevance has been most comprehensively addressed by Koelmans et al. [22]. Briefly, the authors modeled analysis considered the whole mass of various compartments of the ocean including plastics, and then in which compartment exogenous HOCs may preferentially reside based upon partition coefficients. Ocean water would hold 98.3% of HOCs in the ocean and plastics just 0.0002% — in last place of the nine compartments included in the model [22]. Thus, when assessing the relative importance of MPs as chemical carriers, other compartments namely food and water, may be of greater importance as contaminant vectors; however, it is not possible to entirely disregard the link between MP ingestion and chemical availability [18]. Similarly, the transfer of endogenous chemicals is not accounted for.

Expanding the microplastics universe

As the MP field progresses, new classes of anthropogenic particles are coming into focus and the same questions regarding chemical transfer are being asked. Nanoplastics (defined as $<1\ \mu\text{m}$ by ISO [75]) have been shown to be taken up by invertebrates [76,77] and translocated across the gastrointestinal membrane of fish in an *ex vivo* gut sac model [78]. Nano-sized particles have the potential to achieve cellular internalization via endocytotic mechanism and with this exists the possibility that endogenous and exogenous chemicals associated to nanoplastics may be carried into the cell. Coupled with the greater biological reactivity at the nano-size, the overall hazard of nanoplastics may be greater than MPs [79]. However, clear demonstrations of this potential are currently absent from the literature.

Concerns about plastic pollution and greater environmental sustainability have promoted ‘bioplastics’ as an alternative to conventional fossil-fuel-based polymers. The term ‘bioplastics’ may encompass both bio-based plastics made from renewable or natural sources (i.e. plant material) and biodegradable plastics that are made from materials which can be subject to enzymatic degradation of the polymeric matrix [20]. Thus, whilst conventional wisdom would say that bioplastics are designed to degrade faster than conventional plastics, there is specificity to the conditions of degradation, such as the right medium (water, soil, compost), and the absence of such conditions may result in a longer than expected residence time in the environment [80]. Though generally considered ‘green’ the bioplastic polyhydroxybutyrate (PHB) still contained a wide variety of exogenous chemicals and showed slight toxicity to sea urchin larvae [81]. Also using PHB as a test bioplastic, Magara

et al. [82] compared the effects of polyethylene and PHB MPs as a vector of fluoranthene to *Mytilus edulis* with the two polymers exhibiting similar minimal differences to fluoranthene-only exposures. Thus, whether such materials constitute toxicologically safer alternatives is not yet verified as the literature is limited.

Tire wear particles (TWPs), tire and road wear particles (TRWPs), recycled crumb rubber (RTC) and tire-repair-polished debris (TRD) are rubber-related additions to the MP field [83–85]. Of these TWP is perhaps the most discussed with estimates of release suggesting that TWP is a significant component of MP pollution [86]. The chemicals added to tires during manufacturing have been shown to readily be released from the tire under laboratory conditions [87]. This complex ‘leachate’ has been shown to be toxic to a variety of aquatic organisms [87,88] with some specific chemicals now being pinpointed as known toxic agents. For instance, 6PPD-quinone was responsible for the acute toxicity of Pacific Northwest coho salmon observed in the field [89]. Recent studies with TWP have focussed on the particle and the leachate with several species ingesting TWP [84,90,91] and the two fractions showing distinct toxicities [91,92]. Thus, the role of the rubber particle delivering leachate *in vivo* requires greater attention.

Conclusions

The role of MPs in effecting the bioavailability and toxicological consequences of endogenous and exogenous co-contaminants has been a much-debated aspect of plastic pollution. There is a wealth of in-depth literature on the subject (see reviews [2,5,15,16,18,19], but experimental studies often display inconsistencies. This is not surprising since the delivery of chemicals by MPs is dependent on multiple inter-connected factors (Figure 1) [5,47]). Thus, it remains difficult to judge whether MPs are realistic carriers of chemicals and furthermore, based on modeled analysis, whether MPs are relevant to study in this context given their relative contribution to oceanic mass compared with other sorbents [22]. The expansion of the field to include a greater range of particles, namely, nanoplastics, TWP and ‘bioplastics’, will increase the focus to cellular-level vector effects, leachate-related toxicity and ‘benign-by-design’, but future research should also consider the complex processes involved in MP-facilitated chemical transfer (Figure 1) with greater attention needed for biological parameters. Overall, greater environmental and physiological realism is needed to bridge the gap between the laboratory and the real world.

Summary

- The transfer of endogenous or exogenous co-contaminants from MPs to biota is one of the most studied aspects of plastic pollution.
- Consensus as to the validity and relevance of MPs as chemical carriers is still debated.
- A multitude of inter-connected factors from production and release, environmental transformations to biological and physiological interactions need to be considered.
- Greater environmental realism is needed to bridge the gap between laboratory studies and the real world.
- New particles such as nanoplastics, TWPs and bioplastics expand the scope for chemical transfer.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

F.R.K.’s contribution is supported by the North Atlantic Microplastic Centre (NAMC) funded by Sparebanken Vest Foundation. A.I.C.’s contribution is supported by the resources of the Flanders Marine Institute (VLIZ,

Ostend, Belgium). N.J.C. was funded by the NERC Current and Future Effects of Microplastics on Marine Ecosystems (MINIMISE) project, reference NE/S003967/1.

Author Contribution

All authors contributed to all aspects of this publication.

Abbreviations

GIT, gastrointestinal tract; HOCs, hydrophobic organic contaminants; MPs, microplastics; PCBs, polychlorinated biphenyls; PHB, polyhydroxybutyrate; TWP, tire wear particles.

References

- Rochman, C.M., Brookson, C., Bikker, J., Djuric, N., Earn, A., Bucci, K. et al. (2019) Rethinking microplastics as a diverse contaminant suite. *Environ. Toxicol. Chem.* **38**, 703–711 <https://doi.org/10.1002/etc.4371>
- Hahladakis, J.N., Velis, C.A., Weber, R., Iacovidou, E. and Purnell, P. (2018) An overview of chemical additives present in plastics: migration, release, fate and environmental impact during their use, disposal and recycling. *J. Hazard. Mater.* **344**, 179–199 <https://doi.org/10.1016/j.jhazmat.2017.10.014>
- Syberg, K., Khan, F.R., Selck, H., Palmqvist, A., Banta, G.T., Daley, J. et al. (2015) Microplastics: addressing ecological risk through lessons learned. *Environ. Toxicol. Chem.* **34**, 945–953 <https://doi.org/10.1002/etc.2914>
- Andrady, A.L. (2017) The plastic in microplastics: a review. *Mar. Pollut. Bull.* **119**, 12–22 <https://doi.org/10.1016/j.marpolbul.2017.01.082>
- Khan, F.R., Patsiou, D. and Catarino, A.I. (2021) Pollutants bioavailability and toxicological risk from microplastics. In *Handbook of Microplastics in the Environment* (Rocha-Santos, T., Costa, M. and Mouneyrac, C., eds.), pp. 1–40, Springer International Publishing, Cham https://doi.org/10.1007/978-3-030-10618-8_19-1
- Corcoran, P.L. (2022) Degradation of microplastics in the environment. In *Handbook of Microplastics in the Environment* (Rocha-Santos, T., Costa, M. and Mouneyrac, C., eds.), pp. 531–542, Springer International Publishing, Cham https://doi.org/10.1007/978-3-030-39041-9_10
- Wu, X., Liu, P., Shi, H., Wang, H., Huang, H., Shi, Y. et al. (2021) Photo aging and fragmentation of polypropylene food packaging materials in artificial seawater. *Water Res.* **188**, 116456 <https://doi.org/10.1016/j.watres.2020.116456>
- Amaral-Zettler, L.A., Zettler, E.R. and Mincer, T.J. (2020) Ecology of the plastisphere. *Nat. Rev. Microbiol.* **18**, 139–151 <https://doi.org/10.1038/s41579-019-0308-0>
- Brander, S.M. (2022) Rethinking our chemical legacy and reclaiming our planet. *One Earth* **5**, 316–319 <https://doi.org/10.1016/j.oneear.2022.03.020>
- Phuong, N.N., Zalouk-Vergnoux, A., Poirier, L., Kamari, A., Châtel, A., Mouneyrac, C. et al. (2016) Is there any consistency between the microplastics found in the field and those used in laboratory experiments? *Environ. Pollut.* **211**, 111–123 <https://doi.org/10.1016/j.envpol.2015.12.035>
- Du, S., Zhu, R., Cai, Y., Xu, N., Yap, P.S., Zhang, Y. et al. (2021) Environmental fate and impacts of microplastics in aquatic ecosystems: a review. *RSC Adv.* **11**, 15762–15784 <https://doi.org/10.1039/D1RA00880C>
- Courteney-Jones, W., Clark, N.J., Fischer, A.C., Smith, N.S. and Thompson, R.C. (2022) Ingestion of microplastics by marine animals. In *Plastics and the Ocean* (Andrady, A.L., ed.), pp. 349–366, John Wiley & Sons inc., Hoboken, New Jersey, U.S.
- Bao, Z.Z., Chen, Z.F., Lu, S.Q., Wang, G., Qi, Z. and Cai, Z. (2021) Effects of hydroxyl group content on adsorption and desorption of anthracene and anthrol by polyvinyl chloride microplastics. *Sci. Total Environ.* **790**, 148077 <https://doi.org/10.1016/j.scitotenv.2021.148077>
- Hartmann, N.B., Rist, S., Bodin, J., Jensen, L.H.S., Schmidt, S.N., Mayer, P. et al. (2017) Microplastics as vectors for environmental contaminants: exploring sorption, desorption, and transfer to biota. *Integr. Environ. Assess. Manag.* **13**, 488–493 <https://doi.org/10.1002/ieam.1904>
- Fred-Ahmadu, O.H., Bhagwat, G., Oluoye, I., Benson, N.U., Ayejuyo, O.O. and Palanisami, T. (2020) Interaction of chemical contaminants with microplastics: principles and perspectives. *Sci. Total Environ.* **706**, 135978 <https://doi.org/10.1016/j.scitotenv.2019.135978>
- Santos, L.H.M.L.M., Rodríguez-Mozaz, S. and Barceló, D. (2021) Microplastics as vectors of pharmaceuticals in aquatic organisms – an overview of their environmental implications. *Case Stud. Chem. Environ. Eng.* **3**, 100079 <https://doi.org/10.1016/j.csee.2021.100079>
- Huang, W., Song, B., Liang, J., Niu, Q., Zeng, G., Shen, M. et al. (2021) Microplastics and associated contaminants in the aquatic environment: a review on their ecotoxicological effects, trophic transfer, and potential impacts to human health. *J. Hazard. Mater.* **405**, 124187 <https://doi.org/10.1016/j.jhazmat.2020.124187>
- Koelmans, A.A., Diepens, N.J. and Mohamed Nor, N.H. (2022) Weight of evidence for the microplastic vector effect in the context of chemical risk assessment. In *Microplastic in the Environment: Pattern and Process* (Banks, M.S., ed.), pp. 155–197, Springer, London, United Kingdom https://doi.org/10.1007/978-3-030-78627-4_6
- Hermabessiere, L., Dehaut, A., Paul-Pont, I., Lacroix, C., Jezequel, R., Soudant, P. et al. (2017) Occurrence and effects of plastic additives on marine environments and organisms: a review. *Chemosphere* **182**, 781–793 <https://doi.org/10.1016/j.chemosphere.2017.05.096>
- Curto, M., le Gall, M., Catarino, A.I., Niu, Z., Davies, P., Everaert, G. et al. (2021) Long-term durability and ecotoxicity of biocomposites in marine environments: a review. *RSC Adv.* **11**, 32917–32941 <https://doi.org/10.1039/D1RA03023J>
- Kwon, J.H., Chang, S., Hong, S.H. and Shim, W.J. (2017) Microplastics as a vector of hydrophobic contaminants: importance of hydrophobic additives. *Integr. Environ. Assess. Manag.* **13**, 494–499 <https://doi.org/10.1002/ieam.1906>
- Koelmans, A.A., Bakir, A., Burton, G.A. and Janssen, C.R. (2016) Microplastic as a vector for chemicals in the aquatic environment: critical review and model-supported reinterpretation of empirical studies. *Environ. Sci. Technol.* **50**, 3315–3326 <https://doi.org/10.1021/acs.est.5b06069>
- Bakir, A., O'Connor, I.A., Rowland, S.J., Hendriks, A.J. and Thompson, R.C. (2016) Relative importance of microplastics as a pathway for the transfer of hydrophobic organic chemicals to marine life. *Environ. Pollut.* **219**, 56–65 <https://doi.org/10.1016/j.envpol.2016.09.046>
- Kühn, S., Booth, A.M., Sørensen, L., van Oyen, A. and van Franeker, J.A. (2020) Transfer of additive chemicals from marine plastic debris to the stomach oil of northern fulmars. *Front. Environ. Sci.* **8**, 138 <https://doi.org/10.3389/fenvs.2020.00138>
- Tanaka, K., Watanuki, Y., Takada, H., Ishizuka, M., Yamashita, R., Kazama, M. et al. (2020) In vivo accumulation of plastic-derived chemicals into seabird tissues. *Curr. Biol.* **30**, 723–728.e3 <https://doi.org/10.1016/j.cub.2019.12.037>

- 26 Syberg, K., Knudsen, C.M.H., Tairova, Z., Khan, F.R., Shashoua, Y., Geertz, T. et al. (2020) Sorption of PCBs to environmental plastic pollution in the North Atlantic Ocean: importance of size and polymer type. *Case Stud. Chem. Environ. Eng.* **2**, 100062 <https://doi.org/10.1016/j.csee.2020.100062>
- 27 Fred-Ahmadu, O.H., Tenebe, I.T., Ayejuyo, O.O. and Benson, N.U. (2022) Microplastics and associated organic pollutants in beach sediments from the Gulf of Guinea (SE Atlantic) coastal ecosystems. *Chemosphere* **298**, 134193 <https://doi.org/10.1016/j.chemosphere.2022.134193>
- 28 Vedolin, M.C., Teophilo, C.Y.S., Turra, A. and Figueira, R.C.L. (2018) Spatial variability in the concentrations of metals in beached microplastics. *Mar. Pollut. Bull.* **129**, 487–493 <https://doi.org/10.1016/j.marpolbul.2017.10.019>
- 29 Carbery, M., MacFarlane, G.R., O'Connor, W., Afrose, S., Taylor, H. and Palanisami, T. (2020) Baseline analysis of metal(loid)s on microplastics collected from the Australian shoreline using citizen science. *Mar. Pollut. Bull.* **152**, 110914 <https://doi.org/10.1016/j.marpolbul.2020.110914>
- 30 Santana-Viera, S., Montesdeoca-Esponda, S., Torres-Padrón, M.E., Sosa-Ferrera, Z. and Santana-Rodríguez, J.J. (2021) An assessment of the concentration of pharmaceuticals adsorbed on microplastics. *Chemosphere* **266**, 129007 <https://doi.org/10.1016/j.chemosphere.2020.129007>
- 31 Wagstaff, A., Lawton, L.A. and Petrie, B. (2022) Polyamide microplastics in wastewater as vectors of cationic pharmaceutical drugs. *Chemosphere* **288**, 132578 <https://doi.org/10.1016/j.chemosphere.2021.132578>
- 32 McDougall, L., Thomson, L., Brand, S., Wagstaff, A., Lawton, L.A. and Petrie, B. (2022) Adsorption of a diverse range of pharmaceuticals to polyethylene microplastics in wastewater and their desorption in environmental matrices. *Sci. Total Environ.* **808**, 152071 <https://doi.org/10.1016/j.scitotenv.2021.152071>
- 33 Velez, J.F.M., Shashoua, Y., Syberg, K. and Khan, F.R. (2018) Considerations on the use of equilibrium models for the characterisation of HOC-microplastic interactions in vector studies. *Chemosphere* **210**, 359–365 <https://doi.org/10.1016/j.chemosphere.2018.07.020>
- 34 Khan, F.R., Syberg, K., Shashoua, Y. and Bury, N.R. (2015) Influence of polyethylene microplastic beads on the uptake and localization of silver in zebrafish (*Danio rerio*). *Environ. Pollut.* **206**, 73–79 <https://doi.org/10.1016/j.envpol.2015.06.009>
- 35 Sørensen, L., Rogers, E., Altin, D., Salaberria, I. and Booth, A.M. (2020) Sorption of PAHs to microplastic and their bioavailability and toxicity to marine copepods under co-exposure conditions. *Environ. Pollut.* **258**, 113844 <https://doi.org/10.1016/j.envpol.2019.113844>
- 36 Oliveira, M., Ribeiro, A., Hylland, K. and Guilhermino, L. (2013) Single and combined effects of microplastics and pyrene on juveniles (0 + group) of the common goby *Pomatoschistus microps* (Teleostei, Gobiidae). *Ecol. Indic.* **34**, 641–647 <https://doi.org/10.1016/j.ecolind.2013.06.019>
- 37 Rochman, C.M., Kurobe, T., Flores, I. and Teh, S.J. (2014) Early warning signs of endocrine disruption in adult fish from the ingestion of polyethylene with and without sorbed chemical pollutants from the marine environment. *Sci. Total Environ.* **493**, 656–661 <https://doi.org/10.1016/j.scitotenv.2014.06.051>
- 38 Besseling, E., Wegner, A., Foekema, E.M., van den Heuvel-Greve, M.J. and Koelmans, A.A. (2013) Effects of microplastic on fitness and PCB bioaccumulation by the lugworm *Arenicola marina* (L.). *Environ. Sci. Technol.* **47**, 593–600 <https://doi.org/10.1021/es302763x>
- 39 Chua, E.M., Shimeta, J., Nuggeoda, D., Morrison, P.D. and Clarke, B.O. (2014) Assimilation of polybrominated diphenyl ethers from microplastics by the marine amphipod, *Allochrestes compressa*. *Environ. Sci. Technol.* **48**, 8127–8134 <https://doi.org/10.1021/es405717z>
- 40 González-Soto, N., Hatfield, J., Katsumiti, A., Duroudier, N., Lacave, J.M., Bilbao, E. et al. (2019) Impacts of dietary exposure to different sized polystyrene microplastics alone and with sorbed benzo[a]pyrene on biomarkers and whole organism responses in mussels *Mytilus galloprovincialis*. *Sci. Total Environ.* **684**, 548–566 <https://doi.org/10.1016/j.scitotenv.2019.05.161>
- 41 O'Donovan, S., Mestre, N.C., Abel, S., Fonseca, T.G., Carteny, C.C., Cormier, B. et al. (2018) Ecotoxicological effects of chemical contaminants adsorbed to microplastics in the Clam *Scrobicularia plana*. *Front. Mar. Sci.* **5**, 143
- 42 Lu, K., Qiao, R., An, H. and Zhang, Y. (2018) Influence of microplastics on the accumulation and chronic toxic effects of cadmium in zebrafish (*Danio rerio*). *Chemosphere* **202**, 514–520 <https://doi.org/10.1016/j.chemosphere.2018.03.145>
- 43 Rainieri, S., Conlledo, N., Larsen, B.K., Granby, K. and Barranco, A. (2018) Combined effects of microplastics and chemical contaminants on the organ toxicity of zebrafish (*Danio rerio*). *Environ. Res.* **162**, 135–143 <https://doi.org/10.1016/j.envres.2017.12.019>
- 44 Batel, A., Borchert, F., Reinwald, H., Erdinger, L. and Braunbeck, T. (2018) Microplastic accumulation patterns and transfer of benzo[a]pyrene to adult zebrafish (*Danio rerio*) gills and zebrafish embryos. *Environ. Pollut.* **235**, 918–930 <https://doi.org/10.1016/j.envpol.2018.01.028>
- 45 Granby, K., Rainieri, S., Rasmussen, R.R., Kotterman, M.J.J., Sloth, J.J., Cederberg, T.L. et al. (2018) The influence of microplastics and halogenated contaminants in feed on toxicokinetics and gene expression in European seabass (*Dicentrarchus labrax*). *Environ. Res.* **164**, 430–443 <https://doi.org/10.1016/j.envres.2018.02.035>
- 46 Magara, G., Elia, A.C., Syberg, K. and Khan, F.R. (2018) Single contaminant and combined exposures of polyethylene microplastics and fluoranthene: accumulation and oxidative stress response in the blue mussel, *Mytilus edulis*. *J. Toxicol. Environ. Health - A: Curr. Issues* **81**, 761–773 <https://doi.org/10.1080/15287394.2018.1488639>
- 47 Ašmonaitė, G., Tivefälv, M., Westberg, E., Magnér, J., Backhaus, T. and Carney Almroth, B. (2020) Microplastics as a vector for exposure to hydrophobic organic chemicals in fish: a comparison of two polymers and silica particles spiked with three model compounds. *Front. Environ. Sci.* **8**, 87 <https://doi.org/10.3389/fenvs.2020.00087>
- 48 Beiras, R. and Tato, T. (2019) Microplastics do not increase toxicity of a hydrophobic organic chemical to marine plankton. *Mar. Pollut. Bull.* **138**, 58–62 <https://doi.org/10.1016/j.marpolbul.2018.11.029>
- 49 Beiras, R., Muniategui-Lorenzo, S., Rodil, R., Tato, T., Montes, R., López-Ibáñez, S. et al. (2019) Polyethylene microplastics do not increase bioaccumulation or toxicity of nonylphenol and 4-MBC to marine zooplankton. *Sci. Total Environ.* **692**, 1–9 <https://doi.org/10.1016/j.scitotenv.2019.07.106>
- 50 Wen, B., Jin, S.R., Chen, Z.Z., Gao, J.Z., Liu, Y.N., Liu, J.H. et al. (2018) Single and combined effects of microplastics and cadmium on the cadmium accumulation, antioxidant defence and innate immunity of the discus fish (*Symphysodon aequifasciatus*). *Environ. Pollut.* **243**, 462–471 <https://doi.org/10.1016/j.envpol.2018.09.029>
- 51 Schmieg, H., Burmester, J.K.Y., Krais, S., Ruhl, A.S., Tisler, S., Zwiener, C. et al. (2020) Interacting effects of polystyrene microplastics and the antidepressant amitriptyline on early life stages of brown trout (*Salmo trutta f. fario*). *Water (Switzerland)* **12**, 2361 <https://doi.org/10.3390/w12092361>
- 52 Guven, O., Bach, L., Munk, P., Dinh, K.V., Mariani, P. and Nielsen, T.G. (2018) Microplastic does not magnify the acute effect of PAH pyrene on predatory performance of a tropical fish (*Lates calcarifer*). *Aquat. Toxicol.* **198**, 287–293 <https://doi.org/10.1016/j.aquatox.2018.03.011>
- 53 Norland, S., Vorkamp, K., Bøgevik, A.S., Koelmans, A.A., Diepens, N.J., Burgerhout, E. et al. (2021) Assessing microplastic as a vector for chemical entry into fish larvae using a novel tube-feeding approach. *Chemosphere* **265**, 129144 <https://doi.org/10.1016/j.chemosphere.2020.129144>
- 54 Scopetani, C., Cincinelli, A., Martellini, T., Lombardini, E., Ciofini, A., Fortunati, A. et al. (2018) Ingested microplastic as a two-way transporter for PBDEs in *Talitrus saltator*. *Environ. Res.* **167**, 411–417 <https://doi.org/10.1016/j.envres.2018.07.030>

- 55 Nor NH, M. and Koelmans, A.A. (2019) Transfer of PCBs from microplastics under simulated gut fluid conditions is biphasic and reversible. *Environ. Sci. Technol.* **53**, 1874–1883 <https://doi.org/10.1021/acs.est.8b05143>
- 56 Syberg, K., Nielsen, A., Khan, F.R., Banta, G.T., Palmqvist, A. and Jepsen, P.M. (2017) Microplastic potentiates triclosan toxicity to the marine copepod *Acartia tonsa* (Dana). *J. Toxicol. Environ. Health - A: Curr. Issues* **80**, 1369–1371 <https://doi.org/10.1080/15287394.2017.1385046>
- 57 Zhu, Z.-L., Wang, S.-C., Zhao, F.-F., Wang, S.G., Liu, F.F. and Liu, G.Z. (2019) Joint toxicity of microplastics with triclosan to marine microalgae *Skeletonema costatum*. *Environ. Pollut.* **246**, 509–517 <https://doi.org/10.1016/j.envpol.2018.12.044>
- 58 Khan, F.R., Boyle, D., Chang, E. and Bury, N.R. (2017) Do polyethylene microplastic beads alter the intestinal uptake of Ag in rainbow trout (*Oncorhynchus mykiss*)? Analysis of the MP vector effect using in vitro gut sacs. *Environ. Pollut.* **231**, 200–206 <https://doi.org/10.1016/j.envpol.2017.08.019>
- 59 Bakir, A., Rowland, S.J. and Thompson, R.C. (2014) Enhanced desorption of persistent organic pollutants from microplastics under simulated physiological conditions. *Environ. Pollut.* **185**, 16–23 <https://doi.org/10.1016/j.envpol.2013.10.007>
- 60 Coffin, S., Huang, G.Y., Lee, I. and Schlenk, D. (2019) Fish and seabird gut conditions enhance desorption of estrogenic chemicals from commonly-ingested plastic items. *Environ. Sci. Technol.* **53**, 4588–4599 <https://doi.org/10.1021/acs.est.8b07140>
- 61 Wu, P., Tang, Y., Jin, H., Song, Y., Liu, Y. and Cai, Z. (2020) Consequential fate of bisphenol-attached PVC microplastics in water and simulated intestinal fluids. *Environ. Sci. Ecotechnol.* **2**, 100027 <https://doi.org/10.1016/j.ese.2020.100027>
- 62 van der Zande, M., Jemec Kokalj, A., Spurgeon, D.J., Loureiro, S., Silva, P.V., Khodaparast, Z. et al. (2020) The gut barrier and the fate of engineered nanomaterials: a view from comparative physiology. *Environ. Sci.: Nano* **7**, 1874–1898 <https://doi.org/10.1039/D0EN00174K>
- 63 Zhang, H., Wang, J., Zhou, B., Zhou, Y., Dai, Z., Zhou, Q. et al. (2018) Enhanced adsorption of oxytetracycline to weathered microplastic polystyrene: kinetics, isotherms and influencing factors. *Environ. Pollut.* **243**, 1550–1557 <https://doi.org/10.1016/j.envpol.2018.09.122>
- 64 Ahechti, M., Benomar, M., El Alami, M. and Mendiguchia, C. (2022) Metal adsorption by microplastics in aquatic environments under controlled conditions: exposure time, pH and salinity. *Int. J. Environ. Anal. Chem.* **102**, 1118–1125 <https://doi.org/10.1080/03067319.2020.1733546>
- 65 Wang, Y., Liu, C., Wang, F. and Sun, Q. (2022) Behavior and mechanism of atrazine adsorption on pristine and aged microplastics in the aquatic environment: kinetic and thermodynamic studies. *Chemosphere* **292**, 1118–1125 <https://doi.org/10.1016/j.chemosphere.2021.133425>
- 66 Catarino, A.I., Asselman, J., Niu, Z. and Everaert, G. (2022) Micro- and nanoplastics effects in a multiple stressed marine environment. *J. Hazard. Mater. Adv.* **7**, 100119 <https://doi.org/10.1016/j.hazadv.2022.100119>
- 67 Coffin, S., Bouwmeester, H., Brander, S., Damdimopoulou, P., Gouin, T., Hermabessiere, L. et al. (2022) Development and application of a health-based framework for informing regulatory action in relation to exposure of microplastic particles in California drinking water. *Microplastics Nanoplastics* **2**, 12 <https://doi.org/10.1186/s43591-022-00030-6>
- 68 Suaria, G., Achtypi, A., Perold, V., Lee, J.R., Pierucci, A., Bornman, T.G. et al. (2020) Microfibers in oceanic surface waters: a global characterization. *Sci. Adv.* **6**, eaay8493 <https://doi.org/10.1126/sciadv.aay8493>
- 69 Bhagat, K., Barrios, A.C., Rajwade, K., Kumar, A., Oswald, J., Apul, O. et al. (2022) Aging of microplastics increases their adsorption affinity towards organic contaminants. *Chemosphere* **298**, 134238 <https://doi.org/10.1016/j.chemosphere.2022.134238>
- 70 Liu, G., Zhu, Z., Yang, Y., Sun, Y., Yu, F. and Ma, J. (2019) Sorption behavior and mechanism of hydrophilic organic chemicals to virgin and aged microplastics in freshwater and seawater. *Environ. Pollut.* **246**, 26–33 <https://doi.org/10.1016/j.envpol.2018.11.100>
- 71 Cormier, B., Gambardella, C., Tato, T., Perdrat, Q., Costa, E., Vecclin, C. et al. (2021) Chemicals sorbed to environmental microplastics are toxic to early life stages of aquatic organisms. *Ecotoxicol. Environ. Saf.* **208**, 111665 <https://doi.org/10.1016/j.ecoenv.2020.111665>
- 72 Bejjani, S., MacLeod, M., Bogdal, C. and Breitholtz, M. (2015) Toxicity of leachate from weathering plastics: an exploratory screening study with *Nitocra spinipes*. *Chemosphere* **132**, 114–119 <https://doi.org/10.1016/j.chemosphere.2015.03.010>
- 73 Jiang, X., Lu, K., Tunnell, J.W. and Liu, Z. (2021) The impacts of weathering on concentration and bioaccessibility of organic pollutants associated with plastic pellets (nurdles) in coastal environments. *Mar. Pollut. Bull.* **170**, 112592 <https://doi.org/10.1016/j.marpolbul.2021.112592>
- 74 Ford H, V., Jones, N.H., Davies, A.J., Godley, B.J., Jambeck, J.R., Napper, I.E. et al. (2022) The fundamental links between climate change and marine plastic pollution. *Sci. Total Environ.* **806**, 150392 <https://doi.org/10.1016/j.scitotenv.2021.150392>
- 75 Allan, J., Belz, S., Hoeveler, A., Hugas, M., Okuda, H., Patri, A. et al. (2021) Regulatory landscape of nanotechnology and nanoplastics from a global perspective. *Regul. Toxicol. Pharmacol.* **122**, 104885 <https://doi.org/10.1016/j.yrtph.2021.104885>
- 76 Lahive, E., Cross, R., Saarloos, A.I., Horton, A.A., Svendsen, C., Hufenus, R. et al. (2022) Earthworms ingest microplastic fibres and nanoplastics with effects on egestion rate and long-term retention. *Sci. Total Environ.* **807**, 151022 <https://doi.org/10.1016/j.scitotenv.2021.151022>
- 77 Redondo-Hasselerharm, P.E., Vink, G., Mitrano, D.M. and Koelmans, A.A. (2021) Metal-doping of nanoplastics enables accurate assessment of uptake and effects on *Gammarus pulex*. *Environ. Sci.: Nano* **8**, 1761–1770 <https://doi.org/10.1039/D1EN00068C>
- 78 Clark, N.J., Khan, F.R., Mitrano, D.M., Boyle, D. and Thompson, R.C. (2022) Demonstrating the translocation of nanoplastics across the fish intestine using palladium-doped polystyrene in a salmon gut-sac. *Environ. Int.* **159**, 106994 <https://doi.org/10.1016/j.envint.2021.106994>
- 79 Koelmans, A.A., Besseling, E. and Shim, W.J. (2015) Nanoplastics in the aquatic environment. Critical review. In *Marine Anthropogenic Litter* (Bergman, M., Gutow, L., and Klages, M., eds.), pp. 325–340, Cham: Springer International Publishing
- 80 Tong, H., Zhong, X., Duan, Z., Yi, X., Cheng, F., Xu, W. et al. (2022) Micro- and nanoplastics released from biodegradable and conventional plastics during degradation: formation, aging factors, and toxicity. *Sci. Total Environ.* **833**, 155275 <https://doi.org/10.1016/j.scitotenv.2022.155275>
- 81 Uribe-Echeverria, T. and Beiras, R. (2022) Acute toxicity of bioplastic leachates to *Paracentrotus lividus* sea urchin larvae. *Marine Environ. Res.* **176**, 105605 <https://doi.org/10.1016/j.marenvres.2022.105605>
- 82 Magara, G., Khan, F.R., Pinti, M., Syberg, K., Inzirillo, A. and Elia, A.C. (2019) Effects of combined exposures of fluoranthene and polyethylene or polyhydroxybutyrate microplastics on oxidative stress biomarkers in the blue mussel (*Mytilus edulis*). *J. Toxicol. Environ. Health - A: Curr. Issues* **82**, 616–625 <https://doi.org/10.1080/15287394.2019.1633451>
- 83 Wagner, S., Hüffer, T., Klöckner, P., Wehrhahn, M., Hofmann, T. and Reemtsma, T. (2018) Tire wear particles in the aquatic environment - a review on generation, analysis, occurrence, fate and effects. *Water Res.* **139**, 83–100 <https://doi.org/10.1016/j.watres.2018.03.051>
- 84 Halle, L.L., Palmqvist, A., Kampmann, K. and Khan, F.R. (2020) Ecotoxicology of micronized tire rubber: past, present and future considerations. *Sci. Total Environ.* **706**, 135694 <https://doi.org/10.1016/j.scitotenv.2019.135694>

- 85 Luo, Z., Zhou, X., Su, Y., Wang, H., Yu, R., Zhou, S. et al. (2021) Environmental occurrence, fate, impact, and potential solution of tire microplastics: similarities and differences with tire wear particles. *Sci. Total Environ.* **795**, 148902 <https://doi.org/10.1016/j.scitotenv.2021.148902>
- 86 Boucher, J. and Friot, D. (2017) Primary microplastics in the oceans: A global evaluation of sources. IUCN International Union for Conservation of Nature
- 87 Marwood, C., McAtee, B., Kreider, M., Ogle, R.S., Finley, B., Sweet, L. et al. (2011) Acute aquatic toxicity of tire and road wear particles to alga, daphnid, and fish. *Ecotoxicology* **20**, 2079–2089 <https://doi.org/10.1007/s10646-011-0750-x>
- 88 Wik, A., Nilsson, E., Källqvist, T., Tobiesen, A. and Dave, G. (2009) Toxicity assessment of sequential leachates of tire powder using a battery of toxicity tests and toxicity identification evaluations. *Chemosphere* **77**, 922–927 <https://doi.org/10.1016/j.chemosphere.2009.08.034>
- 89 Tian, Z., Zhao, H., Peter, K.T., Gonzalez, M., Wetzel, J., Wu, C. et al. (2021) A ubiquitous tire rubber-derived chemical induces acute mortality in coho salmon. *Science* **371**, 185–189 <https://doi.org/10.1126/science.abd6951>
- 90 Redondo-Hasselerharm, P.E., de Ruijter, V.N., Minterig, S.M., Verschoor, A. and Koelmans, A.A. (2018) Ingestion and chronic effects of car tire tread particles on freshwater benthic macroinvertebrates. *Environ. Sci. Technol.* **52**, 13986–13994 <https://doi.org/10.1021/acs.est.8b05035>
- 91 Khan, F.R., Halle, L.L. and Palmqvist, A. (2019) Acute and long-term toxicity of micronized car tire wear particles to *Hyalella azteca*. *Aquat. Toxicol.* **213**, 105216 <https://doi.org/10.1016/j.aquatox.2019.05.018>
- 92 Halle, L.L., Palmqvist, A., Kampmann, K., Jensen, A., Hansen, T. and Khan, F.R. (2021) Tire wear particle and leachate exposures from a pristine and road-worn tire to *Hyalella azteca*: comparison of chemical content and biological effects. *Aquat. Toxicol.* **232**, 105769 <https://doi.org/10.1016/j.aquatox.2021.105769>