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RESEARCH ARTICLE

# Nitrogen and boron nutrition in grafted watermelon II: Impact on nutrient accumulation in fruit rind and flesh

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# Abstract

Turkey ranks second in watermelon (Citrullus lunatus L.) production globally and the highest production is witnessed for Cukurova plains the country. Although watermelon is extensively cultivated in the Çukurova region, studies on optimum nitrogen (N) and boron (B) doses for watermelon cultivation are guite limited. This study, evaluated the impact of increasing N (0, 90, 180 and 270 kg ha<sup>-1</sup>) and B (0 and 2 kg ha<sup>-1</sup> B) doses on nutrient uptake in rind (exocarp) and flesh (endocarp) of watermelon fruit. Grafted watermelon variety 'Starburst', widely cultivated in the region was used as experimental material. The concentrations of different macro and micronutrients were analyzed from fruit rind and flesh. Individual and interactive effect of N and B doses significantly altered macro and micronutrients' uptake in rind and flesh. Higher amounts of macro and micronutrients were accumulated in rind than flesh. Nutrients' uptake was increased with increasing N doses, whereas B had limited impact. The accumulated nutrients were within the safe limits for human consumption. The N concentrations of rind and flesh increased with increasing N dose. Similarly, B concentration in rind and flesh and N concentration in rind significantly increased, while N concentration in flesh decreased with B application. It was concluded that 270 kg ha<sup>-1</sup> N and 2 kg ha<sup>-1</sup> B are optimum for better nutrient uptake in watermelon fruit. Thus, these doses must be used for watermelon cultivation in Çukurova plains of the country.

# Introduction

Watermelon (*Citrullus lanatus* L.) is globally important fruit vegetable cultivated on commercial level. China is the leading watermelon producer in the world followed by Turkey [1]. Watermelon is extensively cultivated in Turkey and the country follows China with 10% share in global watermelon production. According to Turkish Ministry of Food, Agriculture and Livestock, annual watermelon production in Turkey is 3.9 million tons. Watermelon production is adversely affected by numerous factors and mineral nutrition is among the major reasons of low yield [1,2]. Nitrogen (N) and boron (B) are critical nutrients required for optimum watermelon production [1–5]. Nitrogen is a macronutrient and required in large amount for normal growth and development of crop plants. Numerous metabolic and biochemical process in plants require N for proper development and higher yield. Chlorophyll formation and photosynthates' assimilation are directly influenced by N [6–10]. Low N availability hampers plant growth as it is an important constituent of amino acids, nucleic acid, proteins, chlorophyll and hormones [11]. Nonetheless, plant architecture, photosynthesis, flowering and fruit development are positively influenced by optimum N availability resulting in higher yields [6,12,13]. Plant roots absorb N either in nitrate (NO<sub>3</sub><sup>-</sup>) or in ammonium (NH<sub>4</sub><sup>+</sup>) form. The NO<sub>3</sub><sup>-</sup> is transformed to NH<sub>4</sub><sup>+</sup> and subsequently NH<sub>4</sub><sup>+</sup> is converted to glutamine or glutamate. The compounds that are synthesized in this process are utilized as a precursor in the formation of amino acids, proteins and other N-containing metabolites [6,7]. Nitrogen is the most deficient nutrient in plant production and important for increasing yield. Nonetheless, excess N supply causes late ripening of fruits, leading to a decreased resistance to certain diseases [14,15]. Therefore, determining and supplying optimum N is imperative for successful crop production and higher economic returns.

Boron is required at all developmental stages of crop plants; however, fruit development is the most critical stage [16]. The cultivated soils of the world are very low in B [17]. Boron fertilizers are frequently used to overcome B-deficiency; however, their excessive application could cause B toxicity. Boron deficiency causes vegetative and reproductive defects in plants; therefore, it must be supplied in sufficient quantities. Chlorosis and thick curled leaves with water soaked black spots are typical B-deficiency symptoms in watermelon [5]. The plants capable of accumulating B under B-deficit conditions are well adapted to the soils low in B [18].

Special attention should be given to B nutrition in areas with high relative humidity [19]. Boron deficiency symptoms gradually increase and become fully visible during flowering phase in watermelon [20]. It is well-known that B plays a critical role during reproductive phase compared to vegetative period of plants [21]. Boron must be supplied to plants during flowering and fruit/grain formation in order to harvest higher yields [22].

Nutrient uptake is critical for the proper growth and development of crop plants. Nutrient use efficiency (NUE) can be improved through several approaches. These approaches include modifications in root architecture [23-25], efficient fertilizer application method and soil microorganism [10,26]. Nonetheless, rootstocks are utilized to improve NUE in fruit and vegetable crops [10,26-28]. Rootstock has improved ion uptake in several species [6,27,29-31]. Therefore, selection of an efficient rootstock is important to get high yields [32,33]. Plant biologists are currently working to identify nutrient-specific rootstocks to overcome the deficiency of a particular nutrient [26,32,33]. Watermelon cultivars in Turkey are grafted and well-adapted to Çukurova region. However, limited is known for their optimum N and B requirements.

Watermelon is cultivated in Turkey in open fields and low tunnels. Çukurova region is important watermelon producer [34] and Adana province shares 20% production in the region. Conscious and balanced fertilization is required to obtain higher yield and quality. This study was conducted to optimize N and B doses for nutrient uptake in watermelon. It was hypothesized that nutrient uptake will linearly increase with increasing N and B doses. The optimized doses would help to improve nutrient uptake and productivity of watermelon in the region.

#### Materials and methods

#### Studied species

Watermelon, a Cucurbitaceae member is a xerophytic tropical fruit. It is widely cultivated in warm regions [35]. Watermelon fruit has a thick rind (exocarp) that has variable pigmentation

with a solid or striped appearance, a fleshy mesocarp and an endocarp which varies in color from white to yellow or red [36,37].

#### **Experimental site**

This study was conducted at experimental fields of Research and Application Center, Çukurova University, Faculty of Agriculture, Department of Soil Science and Plant Nutrition, Turkey during 2018 and 2019. Grafted watermelon cultivar 'Starburst', widely cultivated in Çukurova region was used in the experiments. The experiments were set up according to split plot design keeping N as main factor and B as sub-factor. All experimental treatments had four replications and edge effect was excluded to possible extent.

Seedlings were planted keeping 4 m distance between rows, 1.2 m between plants and 6 plants were transplanted in each replication. The soil was analyzed prior to experimentation and depending on the results of the soil analysis 25 kg phosphorus ( $P_2O_5$ ) was applied per hectare at the time of planting.

Four different N doses, i.e.,  $N_0$  (0 kg N ha<sup>-1</sup>),  $N_1$  (90 kg N ha<sup>-1</sup>),  $N_2$  (180 kg N ha<sup>-1</sup>) and  $N_3$  (270 kg N ha<sup>-1</sup>) and two different B doses, i.e.,  $B_0$  (0 kg ha<sup>-1</sup>) and  $B_2$  (2 kg ha<sup>-1</sup> B) were used in the study. Nitrogen was applied by using ammonium sulfate as a source and applied in three equal splits (i.e., at sowing, flowering and fruiting). Etidot67-B was used as B source and all B was applied at sowing.

The fruits were manually harvested at harvest maturity. For nutrient analysis, samples were washed with distilled water, 0.1% HCl and tap water. After washing, rind and flesh were separated. Separated samples were chopped into small pieces and dried in an oven at 70°C until constant weight. The dried samples were ground in an agate mill, separately for analysis. The ground samples were burnt in ash furnace according to dry burning method [38]. Boron concentration was analyzed on spectrophotometer following Bingham [39]. Nitrogen was analyzed according to Kjeldahl method [40]. The Ca, Mg and K were analyzed by semi-micro wet digestion method [41]. The concentrations of Zn, Fe, Mn and Cu in the digested solutions were determined by inductively coupled plasma atomic emission spectroscopy (ICP-AES, OPTIMA 3300 DV, Perkin-Elmer, USA) [42].

#### Statistical analysis

The collected data for nutrient uptake were tested for normality by Shapiro-Wilk normality test [43]. The data were normally distributed; therefore, original data were used in statistical analysis. The differences among years were analyzed by paired t test, which were significant. Therefore, data of both years were analyzed and presented separately. Two-way analysis of variance (ANOVA) was used to test the differences among N and B doses, and their interaction [44]. Least significant difference at 5% probability was used to separate means where ANOVA indicated significant differences. Finally, spearman correlation was computed among nutrient acquisition in rind and flesh, separately. The correlation was computed on PAST software [45].

#### Results

The nutrients accumulated by the rind and flesh were divided into macro and micronutrients based on human consumption. Calcium (Ca), magnesium (Mg) and potassium (K) are required in large quantities by humans; therefore, referred as macro elements, whereas iron (Fe), copper (Cu), manganese (Mn), boron (B), zinc (Zn) and N are required in trace/minor amounts; thus, regarded as micro elements.

## Macronutrients' accumulation in rind

Different N doses significantly (p<0.05) altered macro elements' concentration in rind during both years, except for K concentration during 1<sup>st</sup> year (<u>Table 1</u>). Similarly, Mg during 1<sup>st</sup> year and Ca and K during 2<sup>nd</sup> year were significantly (p<0.05) affected by B doses, while rest of the macro elements were not affected (p>0.05). Nonetheless, interactive effects of N and B were significant for all of the macro elements in the rind during both years (<u>Table 1</u>).

The highest concentrations of all macronutrients in rind were noted for  $N_3$ , whereas the lowest values were recorded for  $N_0$  during both years (Table 2). Similarly, higher amount of Mg was accumulated under  $B_2$ , during first year compared to no B application. However, higher K accumulation was recorded under no B application during 2<sup>nd</sup> year, while higher Ca was acquired under  $B_2$  (Table 2).

Regarding N by B interaction, the highest Ca and Mg were accumulated in rind with  $N_3$  and  $B_2$  interaction, while plants grown under  $N_2$  and  $B_2$  combination acquired the highest amount of K during  $1^{st}$  year. The lowest macro elements' accumulation in rind was observed

Table 1. Analysis of variance of different mineral u	mtales tusits of an	بمقدما مسموه مسموا مستسط	anarun un dan maniarua nitua a	an and honon dooro
Table 1. Analysis of variance of different mineral u	diake traits of gr	alled watermelon rind s	erown under various nitrog	en and boron doses.

				Year-1			Year-2			
Mineral	SOV	DF	SS	MS	F value	P value	SS	MS	F value	P value
Ca	Ν	3	0.16	0.05	2.21	0.003*	0.13	0.04	7.50	0.00*
	В	1	0.05	0.05	2.09	0.152 <sup>NS</sup>	0.04	0.04	7.63	0.01*
	$N \times B$	3	0.06	0.02	0.88	0.453 <sup>NS</sup>	0.02	0.01	0.94	0.43 <sup>NS</sup>
Mg	Ν	3	0.54	0.18	3.94	0.011 <sup>NS</sup>	0.05	0.02	11.12	0.0001*
	В	1	0.23	0.23	5.01	0.028*	0.01	0.01	3.84	0.05*
	$N \times B$	3	0.29	0.10	2.12	0.014*	0.02	0.01	4.42	0.01*
К	Ν	3	2.39	0.80	0.96	0.414 <sup>NS</sup>	6.61	2.20	8.06	0.0001*
	В	1	0.36	0.36	0.44	0.510 <sup>NS</sup>	1.20	1.20	4.40	0.04*
	$N \times B$	3	2.98	0.99	1.21	0.031 <sup>NS</sup>	1.24	0.41	1.51	0.22 <sup>NS</sup>
Fe	Ν	3	9338.66	3112.89	2.57	0.059 <sup>NS</sup>	774.95	258.32	2.35	0.08 <sup>NS</sup>
	В	1	1463.95	1463.95	1.21	0.274 <sup>NS</sup>	1467.24	1467.24	13.36	0.00*
	$N \times B$	3	14254.82	4751.61	3.93	0.011*	876.87	292.29	2.66	0.05*
Mn	Ν	3	7035.30	2345.10	2.05	0.113 <sup>NS</sup>	57.25	19.08	3.06	0.03*
	В	1	141.06	141.06	0.12	0.726 <sup>NS</sup>	14.90	14.90	2.39	0.13 <sup>NS</sup>
	$N \times B$	3	7217.67	2405.89	2.10	0.105 <sup>NS</sup>	119.65	39.88	6.39	0.00*
Cu	Ν	3	445.06	148.35	4.31	0.007*	36.22	12.07	1.61	0.19 <sup>NS</sup>
	В	1	0.21	0.21	0.01	0.937 <sup>NS</sup>	67.40	67.40	8.97	0.00*
	$N \times B$	3	395.95	131.98	3.83	0.013*	38.60	12.87	1.71	0.17 <sup>NS</sup>
В	Ν	3	652.51	217.50	6.90	0.000*	145.63	48.54	5.06	0.00*
	В	1	134.39	134.39	4.26	0.042*	57.57	57.57	6.01	0.02*
	$N \times B$	3	32.30	10.77	0.34	0.795 <sup>NS</sup>	7.15	2.38	0.25	0.86 <sup>NS</sup>
Zn	Ν	3	4873.05	1624.35	4.86	0.004	140.71	46.90	3.05	0.03*
	В	1	34.83	34.83	0.10	0.748 <sup>NS</sup>	13.19	13.19	0.86	0.36 <sup>NS</sup>
	$N \times B$	3	2844.74	948.25	2.83	0.043*	19.88	6.63	0.43	0.73 <sup>NS</sup>
N	N	3	4.21	1.40	9.23	0.0001*	4.31	1.44	10.78	0.0001*
	В	1	0.42	0.42	2.77	0.09 <sup>NS</sup>	0.85	0.85	6.36	0.013*
	$N \times B$	3	0.04	0.01	0.08	0.97 <sup>NS</sup>	0.41	0.14	1.03	0.385 <sup>NS</sup>

SOV = source of variation, DF = degree of freedom, SS = sum of squares, MS = mean squares

\* = significant (p<0.05)

NS = non-significant (p>0.05).

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Treatments		Year-1			Year-2	
	Calcium (%)	Magnesium (%)	Potassium (%)	Calcium (%)	Magnesium (%)	Potassium (%)
			Factor A-Nitrogen (	N)		
0 kg ha <sup>-1</sup> (N <sub>0</sub> )	0.38 b	0.55 b	6.29	0.28 bc	0.20 c	3.80 b
90 kg ha <sup>-1</sup> (N <sub>1</sub> )	0.42 ab	0.64 ab	6.21	0.26 c	0.23 b	4.39 a
180 kg ha <sup>-1</sup> (N <sub>2</sub> )	0.44 ab	0.65 ab	6.28	0.31 b	0.24 b	4.40 a
270 kg ha <sup>-1</sup> (N <sub>3</sub> )	0.49 a	0.76 a	5.90	0.36 a	0.27 a	4.43 a
LSD 0.05	0.03	0.10	NS	0.04	0.03	0.22
			Factor B-Boron (E	3)		
0 kg ha <sup>-1</sup> (B <sub>0</sub> )	0.41	0.60 b	6.11	0.28 b	0.23	4.37 a
2 kg ha <sup>-1</sup> (B <sub>2</sub> )	0.45	0.70 a	6.23	0.32 a	0.24	4.14 b
LSD 0.05	NS	0.09	NS	0.03	NS	0.21
			N × B interaction			
N <sub>0</sub> B <sub>0</sub>	0.38 b	0.57 bc	6.14 ab	0.27 cd	0.19 d	4.01 cd
N <sub>1</sub> B <sub>0</sub>	0.41 b	0.61 bc	6.21 ab	0.22 d	0.21 d	4.42 abc
$N_2B_0$	0.42 b	0.57 bc	5.97 ab	0.29 bc	0.22 cd	4.66 a
N <sub>3</sub> B <sub>0</sub>	0.43 b	0.64 bc	6.09 ab	0.35 ab	0.29 a	4.40 abc
N <sub>0</sub> B <sub>2</sub>	0.37 b	0.52 c	6.44 ab	0.29 bc	0.21 d	3.60 d
N <sub>1</sub> B <sub>2</sub>	0.43 b	0.66 bc	6.21 ab	0.30 bc	0.26 ab	4.37 abc
$N_2B_2$	0.46 ab	0.72 ab	6.56 a	0.34 ab	0.25 bc	4.16 bc
N <sub>3</sub> B <sub>2</sub>	0.56 a	0.87 a	5.72 b	0.37 a	0.25 bc	4.45 ab
LSD 0.05	0.10	0.15	0.35	0.03	0.04	0.29

#### Table 2. The impact of different nitrogen and boron doses and their interaction on macro mineral contents in grafted watermelon rind.

Means followed by similar letters within a column are statistically non-significant (p>0.05). NS = non-significant.

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for  $N_0$  and  $B_0$  combination (Table 2). During  $2^{nd}$  year,  $N_3$  and  $B_2$  combination recorded the highest concentration of Ca, whereas  $N_3B_1$  combination acquired the highest amount of Mg. Similarly, the highest K uptake was recorded for  $N_2B_0$  combination. The lowest concentration of these nutrients was recorded for  $N_0B_0$  interaction during  $2^{nd}$  year of the study (Table 2).

#### Macronutrients' accumulation in flesh

The Mg concentration was significantly (p<0.05) affected by N doses during 1<sup>st</sup> year, whereas Ca and K were not affected (Table 3). Nitrogen doses had significant effect on Mg and K accumulation during 2<sup>nd</sup> year, while had non-significant on Ca. Different B does had non-significant impact on K uptake during 1<sup>st</sup> year and Ca accumulation during 2<sup>nd</sup> year, whereas remaining macro elements were significantly altered by B doses during both years. Nonetheless, interactive effects of N and B were significant for all macro elements except Ca during both years (Table 3).

The highest Mg concentration was noted for  $N_3$  during  $1^{st}$  year, whereas N doses were nonsignificant for the rest of macro elements. Similarly,  $N_1$  and  $N_2$  recorded the highest concentrations of Mg and K, respectively during  $2^{nd}$  year of the study (Table 4). Similarly, the highest Ca and Mg concentrations during  $1^{st}$  year and Mg and K concentrations during  $2^{nd}$  year were noted for  $B_2$  (Table 4).

Regarding N × B interaction, the highest Mg and K concentrations were recorded for  $N_3B_2$ and  $N_3B_1$ , respectively during 1<sup>st</sup> year. The lowest accumulation of macro elements in rind was observed for  $N_0B_0$  (Table 4). During 2<sup>nd</sup> year,  $N_1B_2$  and  $N_2B_2$  recorded the highest concentrations of Mg and K, respectively. The lowest concentration of these nutrients was recorded for  $N_0B_0$  during 2<sup>nd</sup> year (Table 4).

				Y	ear-I		Year-II			
Mineral	SOV	DF	SS	MS	F value	P value	SS	MS	F value	P value
Ca	N	3	0.000	0.000	1.04	0.381 <sup>NS</sup>	0.000	0.000	0.817	0.488 <sup>NS</sup>
	В	1	0.003	0.003	36.72	0.0001*	0.000	0.000	1.751	0.189 <sup>NS</sup>
	$N \times B$	3	0.000	0.000	0.85	0.469 <sup>NS</sup>	0.001	0.000	6.567	0.000*
Mg	N	3	0.71	0.24	21.75	0.0001*	0.01	0.00	7.65	0.00*
	В	1	0.26	0.26	23.38	0.0001*	0.00	0.00	9.70	0.00*
	$N \times B$	3	0.74	0.25	22.59	0.0001*	0.00	0.00	2.45	0.07 <sup>NS</sup>
K	N	3	0.008	0.003	0.71	0.551 <sup>NS</sup>	2.79	0.93	7.77	0.00*
	В	1	0.002	0.002	0.55	0.461 <sup>NS</sup>	2.48	2.48	20.72	0.0001*
	$N \times B$	3	0.019	0.006	1.65	0.184 <sup>NS</sup>	0.85	0.28	2.35	0.08 <sup>NS</sup>
Fe	N	3	197.43	65.81	0.99	0.403 <sup>NS</sup>	3978.58	1326.19	5.09	0.00*
	В	1	0.72	0.72	0.01	0.917 <sup>NS</sup>	1246.96	1246.96	4.78	0.03*
	$N \times B$	3	465.38	155.13	2.33	0.080 <sup>NS</sup>	1457.46	485.82	1.86	0.14 <sup>NS</sup>
Mn	Ν	3	32.65	10.88	1.04	0.379 <sup>NS</sup>	51.38	17.13	4.18	0.01*
	В	1	15.93	15.93	1.52	0.221 <sup>NS</sup>	31.44	31.44	7.66	0.01*
	$N \times B$	3	76.09	25.36	2.42	0.071 <sup>NS</sup>	14.24	4.75	1.16	0.33 <sup>NS</sup>
Cu	Ν	3	1.87	0.62	0.78	0.51 <sup>NS</sup>	1.32	0.44	0.21	0.89 <sup>NS</sup>
	В	1	10.93	10.93	13.63	0.000*	0.54	0.54	0.25	0.62 <sup>NS</sup>
	$N \times B$	3	2.21	0.74	0.92	0.436 <sup>NS</sup>	9.86	3.29	1.53	0.21 <sup>NS</sup>
В	Ν	3	20.52	6.84	1.18	0.324 <sup>NS</sup>		1.12	0.74	0.53 <sup>NS</sup>
	В	1	9.56	9.56	1.64	0.203 <sup>NS</sup>	8.25	8.25	5.47	0.02*
	$N \times B$	3	0.93	0.31	0.05	0.984 <sup>NS</sup>	3.54	1.18	0.78	0.51 <sup>NS</sup>
Zn	Ν	3	15.86	5.29	0.62	0.604 <sup>NS</sup>	36.06	12.02	1.58	0.20 <sup>NS</sup>
	В	1	28.30	28.30	3.32	0.072 <sup>NS</sup>	14.55	14.55	1.91	0.17 <sup>NS</sup>
	$N \times B$	3	40.17	13.39	1.57	0.202 <sup>NS</sup>	11.75	3.92	0.51	0.67 <sup>NS</sup>
N	Ν	3	2.30	0.77	19.01	0.0001*	1.57	0.52	12.79	0.0001*
	В	1	0.02	0.02	0.38	0.539 <sup>NS</sup>	0.92	0.92	22.58	0.0001*
	$N \times B$	3	0.41	0.14	3.36	0.022*	0.01	0.00	0.11	0.95 <sup>NS</sup>

Table 3. Analysis of variance of	different mineral uptake traits	s of grafted watermelon fles	sh grown under variou	s nitrogen and boron doses.
		8		

SOV = source of variation, DF = degree of freedom, SS = sum of squares, MS = mean squares

\* = significant (p<0.05)

NS = non-significant (p>0.05).

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#### Microelements' accumulation in rind

The concentration of all microelements in rind was significantly affected by different N doses during both years except non-significant effect for Cu uptake during  $2^{nd}$  year (Table 1). All microelements, except B were not affected by B doses during first year; however, B doses significantly altered all microelements during  $2^{nd}$  year except Mn and Zn. The N × B interaction had significant effect on the concentration of all microelements during both years (Table 1).

The highest concentration of Fe, Mn and Cu was noted with  $N_1$ , whereas  $N_3$  recorded the highest concentration of B, Zn and N during 1<sup>st</sup> year (Table 5). The highest concentration of all microelements was observed for  $N_2$  and  $N_3$  during 2<sup>nd</sup> year. The highest concentration of B was recorded under  $B_2$ , whereas B application had no impact on rest of the microelements during 1<sup>st</sup> year. Regarding interaction  $N_3$  with both B doses observed the highest concentration of all microelements, while  $N_0B_0$  had the lowest values of these traits during both years (Table 5).

Treatments		Year-1			Year-2	
	Calcium (%)	Magnesium (%)	Potassium (%)	Calcium (%)	Magnesium (%)	Potassium (%)
			Factor A-Nitrogen (N)			
0 kg ha <sup>-1</sup> (N <sub>0</sub> )	0.09	0.14 b	0.82	0.04	0.15 b	1.65 c
90 kg ha <sup>-1</sup> (N <sub>1</sub> )	0.09	0.13 b	0.83	0.04	0.18 a	1.93 b
180 kg ha <sup>-1</sup> (N <sub>2</sub> )	0.09	0.15 b	0.83	0.04	0.16 b	2.13 a
270 kg ha <sup>-1</sup> (N <sub>3</sub> )	0.09	0.34 a	0.81	0.04	0.16 b	1.90 b
LSD 0.05	NS	0.11	NS	NS	0.02	
			Factor B-Boron (B)			
0 kg ha <sup>-1</sup> (B <sub>0</sub> )	0.08 b	0.13 b	0.82	0.04	0.16 b	1.73 b
2 kg ha <sup>-1</sup> (B <sub>2</sub> )	0.09 a	0.24 a	0.82	0.04	0.17 a	2.06 a
LSD 0.05	0.01	0.09	NS	NS	0.01	0.23
			N × B interaction			
N <sub>0</sub> B <sub>0</sub>	0.08	0.14 b	0.81 ab	0.04	0.15 b	1.59 e
N <sub>1</sub> B <sub>0</sub>	0.08	0.13 b	0.83 ab	0.04	0.16 b	1.81 cde
N <sub>2</sub> B <sub>0</sub>	0.08	0.13 b	0.83 ab	0.04	0.15 b	1.97 bcd
N <sub>3</sub> B <sub>0</sub>	0.08	0.13 b	0.84 a	0.05	0.16 b	1.58 e
N <sub>0</sub> B <sub>2</sub>	0.09	0.13 b	0.82 ab	0.05	0.16 b	1.71 de
N <sub>1</sub> B <sub>2</sub>	0.10	0.12 b	0.83 ab	0.04	0.20 a	2.06 abc
$N_2B_2$	0.09	0.16 b	0.82 ab	0.04	0.17 b	2.28 a
N <sub>3</sub> B <sub>2</sub>	0.09	0.54 a	0.78 b	0.04	0.16 b	2.21 ab
LSD 0.05	NS	0.25	0.04	NS	0.02	0.05

Table 4. The impact of different nitrogen and boron doses and their interaction on macro mineral contents in watermelon flesh.

Means followed by similar letters within a column are statistically non-significant (p>0.05). NS = non-significant.

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#### Microelement accumulation in flesh

The concentration of all microelements except N in flesh was not affected by N doses during  $1^{st}$  year, whereas Fe, Mn and N were significantly altered by N doses during  $2^{nd}$  year (Table 2). All microelements, except Cu were not affected by B doses during first year; however, B doses significantly altered all microelements during  $2^{nd}$  year except Cu and Zn. The N × B interaction had significant effect on the concentration of all microelements during both years except for B during  $1^{st}$  year and Cu during  $2^{nd}$  year (Table 2).

The highest concentration of N was noted with  $N_2$  and  $N_3$  during 1<sup>st</sup> year (Table 6). The highest concentration of all microelements was observed for  $N_3$  during 2<sup>nd</sup> year. The highest concentration of Cu was recorded under B<sub>0</sub>, whereas B application had no impact on rest of the microelements during 1<sup>st</sup> year. Regarding interaction,  $N_3$  with both B doses observed the highest concentration of all microelements, while  $N_0B_0$  had the lowest values of these traits during both years (Table 6).

#### Correlation among mineral uptake traits of rind

Most of the nutrient uptake traits had non-significant correlations with each other during both years. The only significant and strong positive correlation was noted for Ca and Mg uptake with N accumulation and B during 1<sup>st</sup> year (Fig 1). Similarly, Fe had significant positive correlation with Zn, and B accumulation was positively correlated with N uptake. Similar correlations were recorded during 2<sup>nd</sup> year. The only negative correlation was noted among Ca and Cu accumulation during 2<sup>nd</sup> year (Fig 1).

Year-I									
Treatments	Iron (mg kg <sup>-1</sup> )	Manganese (mg kg <sup>-1</sup> )	Copper (mg kg <sup>-1</sup> )	Boron (mg kg <sup>-1</sup> )	Zinc (mg kg <sup>-1</sup> )	Nitrogen (mg kg <sup>-1</sup> )			
			Factor A-Nitrogen (N)						
0 kg ha <sup>-1</sup> (N <sub>0</sub> )	112.61 a	82.07 ab	13.04 ab	32.98 b	66.41 bc	2.32 b			
90 kg ha <sup>-1</sup> (N <sub>1</sub> )	118.02 a	89.98 a	16.38 a	33.84 b	73.05 ab	2.38 b			
180 kg ha <sup>-1</sup> (N <sub>2</sub> )	91.36 b	65.99 b	10.25 b	34.21 b	61.04 c	2.48 b			
270 kg ha <sup>-1</sup> (N <sub>3</sub> )	109.41 ab	78.62 ab	13.02 ab	39.61 a	80.21 a	2.86 a			
LSD 0.05	7.76	8.12	3.40	4.44	12.21	0.34			
			Factor B-Boron (B)						
0 kg ha <sup>-1</sup> (B <sub>0</sub> )	104.23	78.21	13.19	33.98 b	69.76	2.44			
2 kg ha <sup>-1</sup> (B <sub>2</sub> )	111.73	80.37	13.22	36.34 a	70.78	2.57			
LSD 0.05	NS	NS	NS	2.34	NS	NS			
			N × B interaction						
N <sub>0</sub> B <sub>0</sub>	100.92 bc	88.79 ab	13.02 bc	31.52 c	62.16 c	2.26 c			
N <sub>1</sub> B <sub>0</sub>	98.81 bc	75.49 b	13.40 bc	33.08 bc	67.34 bc	2.28 c			
$N_2B_0$	95.49 bc	62.32 b	10.31 c	33.67 bc	60.25 c	2.41 c			
N <sub>3</sub> B <sub>0</sub>	120.99 ab	84.94 ab	15.78 ab	37.61 ab	88.50 a	2.81 ab			
N <sub>0</sub> B <sub>0</sub>	124.31 ab	75.36 b	13.06 bc	34.43 bc	70.67 bc	2.38 c			
N <sub>1</sub> B <sub>0</sub>	137.23 a	104.46 a	19.37 a	34.59 bc	78.76 ab	2.48 c			
N <sub>2</sub> B <sub>0</sub>	87.57 c	69.36 b	10.19 c	34.71 bc	61.76 c	2.54 bc			
N <sub>3</sub> B <sub>2</sub>	97.82 bc	72.30 b	10.26 c	41.61 a	71.92 bc	2.90 a			
LSD 0.05	17.80	26.23	8.98	4.23	8.78	0.09			
			Year-II						
			Factor A-Nitrogen (N)						
0 kg ha <sup>-1</sup> (N <sub>0</sub> )	63.69 b	10.55 b	25.44	31.58 b	26.09 b	1.22 b			
90 kg ha <sup>-1</sup> (N <sub>1</sub> )	69.00 ab	12.12 a	26.72	33.53 a	29.44 a	1.38 b			
180 kg ha <sup>-1</sup> (N <sub>2</sub> )	71.63 a	11.31 ab	25.26	34.56 a	27.35 ab	1.66 a			
270 kg ha <sup>-1</sup> (N <sub>3</sub> )	67.67 ab	12.57 a	25.24	34.65 a	28.07 ab	1.75 a			
LSD 0.05	7.80	2.01	NS	2.21	2.28	0.38			
		·	Factor B-Boron (B)						
0 kg ha <sup>-1</sup> (B <sub>0</sub> )	71.89 a	12.04	26.53 a	32.77 b	28.12	1.40 b			
2 kg ha <sup>-1</sup> (B <sub>2</sub> )	64.11 b	11.25	24.83 b	34.35 a	27.37	1.60 a			
LSD 0.05	3.34	NS	1.12	1.90	NS	0.18			
		·	N × B interaction						
N <sub>0</sub> B <sub>0</sub>	63.14 b	10.35 bc	25.90 b	30.65 d	27.21 ab	1.05 d			
N <sub>1</sub> B <sub>0</sub>	74.06 a	11.93 bc	28.61 a	32.41 cd	29.56 a	1.25 cd			
$N_2B_0$	75.19 a	10.91 bc	25.50 b	34.14 abc	27.69 ab	1.57 ab			
$N_3B_0$	75.46 a	14.89 a	26.00 b	34.01 abc	27.99 ab	1.76 a			
N <sub>0</sub> B <sub>0</sub>	64.25 b	10.74 bc	24.99 b	32.50 bcd	24.97 b	1.39 bc			
N <sub>1</sub> B <sub>0</sub>	63.94 b	12.32 b	24.83 b	34.66 abc	29.32 a	1.51 abc			
N <sub>2</sub> B <sub>0</sub>	68.37 ab	11.68 bc	25.04 b	34.94 ab	27.03 ab	1.74 a			
N <sub>3</sub> B <sub>2</sub>	59.88 b	10.24 c	24.48 b	35.29 a	28.15 a	1.74 a			
LSD 0.05	11.02	1.87	3.32	1.34	2.12	0.45			

#### Table 5. The impact of different nitrogen and boron doses and their interaction on micro mineral contents in watermelon rind.

Means followed by similar letters within a column are statistically non-significant (p>0.05). NS = non-significant.

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			Year-I			
Treatments	Iron (mg kg <sup>-1</sup> )	Manganese (mg kg <sup>-1</sup> )	Copper (mg kg <sup>-1</sup> )	Boron (mg kg <sup>-1</sup> )	Zinc (mg kg <sup>-1</sup> )	Nitrogen (mg kg <sup>-1</sup> )
			Factor A-Nitrogen (N)			
0 kg ha <sup>-1</sup> (N <sub>0</sub> )	37.13	16.94	4.46	17.27	14.34	1.39 c
90 kg ha <sup>-1</sup> (N <sub>1</sub> )	37.20	15.82	4.25	16.42	13.89	1.60 b
180 kg ha <sup>-1</sup> (N <sub>2</sub> )	40.49	17.41	4.46	16.52	15.05	1.72 a
270 kg ha <sup>-1</sup> (N <sub>3</sub> )	39.43	16.96	4.65	17.49	14.37	1.79 a
LSD 0.05	NS	NS	NS	NS	NS	0.14
			Factor B-Boron (B)			
0 kg ha <sup>-1</sup> (B <sub>0</sub> )	38.61	17.18	4.80 a	16.61	14.95	1.61
2 kg ha <sup>-1</sup> (B <sub>2</sub> )	38.48	16.38	4.12 b	17.24	13.87	1.64
LSD 0.05	NS	NS	0.54	NS	NS	NS
			N × B interaction			
N <sub>0</sub> B <sub>0</sub>	36.80 ab	16.11 b	4.62 abc	17.09	15.19 a	1.47 b
N <sub>1</sub> B <sub>0</sub>	34.56 b	15.86 b	4.54 abc	15.96	15.35 a	1.51 b
N <sub>2</sub> B <sub>0</sub>	40.28 ab	19.01 a	4.81 ab	16.16	14.99 a	1.72 a
N <sub>3</sub> B <sub>0</sub>	42.94 a	17.90 ab	5.22 a	17.20	14.28 ab	1.75 a
N <sub>0</sub> B <sub>2</sub>	37.46 ab	17.77 ab	4.30 bc	17.46	13.50 ab	1.30 c
N <sub>1</sub> B <sub>2</sub>	39.84 ab	15.78 b	3.97 c	16.88	12.43 b	1.68 a
N <sub>2</sub> B <sub>2</sub>	40.68 ab	15.94 b	4.14 bc	16.85	15.10 a	1.73 a
$N_3B_2$	35.92 b	16.01 b	4.07 bc	17.77	14.46 ab	1.84 a
LSD 0.05	2.87	3.45	1.76	NS	2.34	0.75
			Year-II			1
			Factor A-Nitrogen (N)			
0 kg ha <sup>-1</sup> (N <sub>0</sub> )	107.85 c	17.84 b	7.32	18.32	13.26	1.50 b
90 kg ha <sup>-1</sup> (N <sub>1</sub> )	120.10 ab	19.28 a	7.02	17.96	13.60	1.75 a
180 kg ha <sup>-1</sup> (N <sub>2</sub> )	113.80 bc	19.63 a	7.24	18.32	14.37	1.76 a
270 kg ha <sup>-1</sup> (N <sub>3</sub> )	124.93 a	19.62 a	7.10	18.47	14.82	1.84 a
LSD 0.05	4.56	3.45	NS	NS	NS	0.30
			Factor B-Boron (B)			1
0 kg ha <sup>-1</sup> (B <sub>0</sub> )	113.07 b	19.66 a	7.25	17.97 b	14.40	1.81 a
$2 \text{ kg ha}^{-1} (B_2)$	120.26 a	18.52 b	7.10	18.56 a	13.63	1.61 b
LSD 0.05	8.78	2.21	NS	3.45	NS	0.34
			N × B interaction			1
N <sub>0</sub> B <sub>1</sub>	97.77 b	18.05 cd	7.63	18.06 ab	14.25 ab	1.58 de
N <sub>1</sub> B <sub>1</sub>	116.90 a	19.86 ab	7.06	17.93 b	13.79 ab	1.86 ab
$N_2B_1$	112.86 a	20.90 a	6.77	17.97 b	14.64 a	1.87 ab
N <sub>3</sub> B <sub>1</sub>	124.75 a	19.94 ab	7.48	17.90 b	14.94 a	1.93 a
N <sub>0</sub> B <sub>2</sub>	117.93 a	17.64 d	7.01	18.57 ab	12.27 b	1.42 e
N <sub>1</sub> B <sub>2</sub>	123.31 a	18.70 bcd	6.98	17.99 b	13.42 ab	1.64 cd
$N_2B_2$	114.67 a	18.46 bcd	7.67	18.63 ab	14.12 ab	1.66 cd
N <sub>3</sub> B <sub>2</sub>	125.12 a	19.29 abc	6.73	19.04 a	14.69 a	1.74 bc
LSD 0.05	30.90	1.03	NS	2.21	0.56	0.54

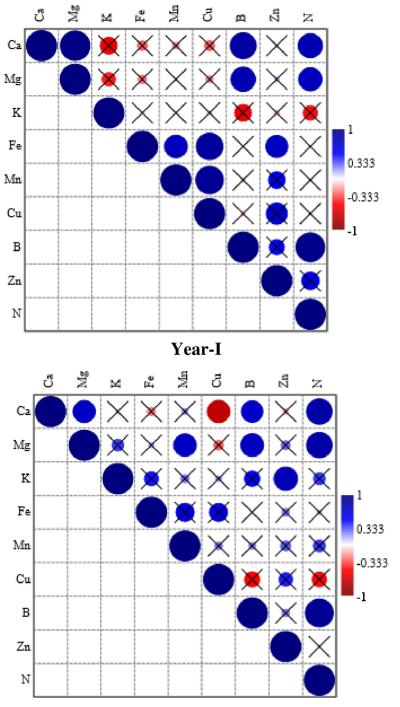
#### Table 6. The impact of different nitrogen and boron doses and their interaction on micro mineral contents in watermelon flesh.

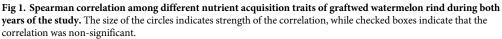
Means followed by similar letters within a column are statistically non-significant (p>0.05). NS = non-significant.

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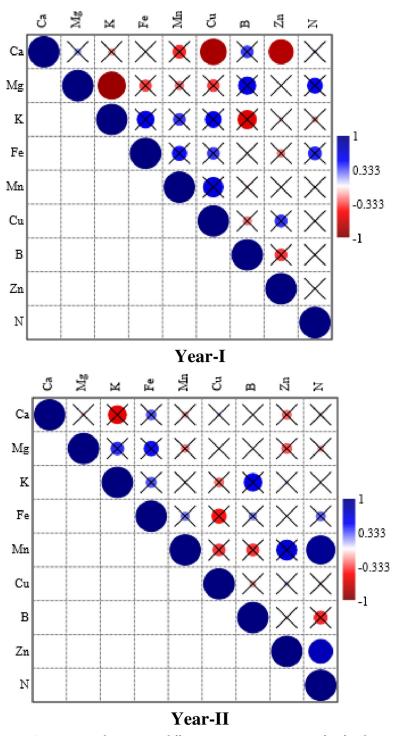
# Correlation among mineral uptake traits of flesh

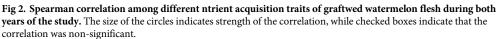
Most of the nutrient uptake traits had non-significant correlations with each other during both years. The only significant and strong negative correlation was noted for Ca with Cu and Zn uptake during 1<sup>st</sup> year (Fig 2). The only positive correlation was noted among Mn and N accumulation during 2<sup>nd</sup> year (Fig 2).





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## Discussion

Different nutrient uptake traits were significantly altered by N and B doses. As hypothesized, concentration of most of the nutrients was increased with increasing N and B doses. It was noted that N and other minerals' concentration increased with N application compared to control treatment. Torun [3] reported that yield, weight, diameter and TSS content of the fruit increased with N application. Similar results have been demonstrated by other researchers [2,46]. Colla et al. [2] also reported that N use efficiency and N uptake efficiency were significantly affected by combinations of N fertilization and grafting. Wehner [47] reported that TSS content in watermelon should be at least 10% for an ideal flavor.

Watermelon is an important fruit vegetable commercially cultivated worldwide. Boron deficiency is common in cultivated areas, globally [17]. Boron fertilizers are used to overcome B-deficiency, which increase input cost. Boron deficiency restricts plant growth and a wide range of symptoms, including chlorosis and thick curled leaves with water soaked black spots appear on watermelon [5]. The adaptability of crops under limited B availability can be attributed to plant ability to absorb B under B-deficient conditions [18].

Nitrogen is required by plants in large amounts for normal growth and development. Numerous metabolic and biochemical process require N for the proper development and yield [6-10]. Low N availability hampers plant growth as it is an important constituent of amino acids, nucleic acid, proteins, chlorophyll and hormones [11].

Boron is widely distributed in earth crust and equally important for plants and animals. The involvement of B in several physiological processes of plants has been reported [21,27,48]. Sufficient B availability in soil solution is important for proper physiological functioning of plants. Principally, B is involved in cell wall structural integration and linkage of B with pectic polysaccharide rhamnogalacturonan II (RGII) controls porosity and tensile strength of cell wall [49]. Considering plant requirement on molar basis, B requirement for dicots is higher compared with any other microelement [48]. However, limitation or excess of B adversely affect plant growth. Interestingly, the range between deficiency and toxicity of B is narrow [50-53]. In soils, the concentration of B varies from 10 mg kg<sup>-1</sup> to 300 mg kg<sup>-1</sup> depending on the soil type, amount of organic matter and precipitation [54]. In heavy textured soils B reaches to toxic level that adversely affects plant growth and yield [55,56]. However, in acidic soils B-deficiency is commonly observed because of ion leaching. Boron deficiency alters plant metabolic, cellular, biological and molecular processes such as photosynthesis, cell wall and membrane integration, cell division, carbohydrate metabolism, sugar and hormonal transport, protein biosynthesis and nucleic acid metabolism [27,57]. The obvious response of B-deficiency in several crops is inhibition of root growth because of reduced cell division [58]. Moreover, long-term B-deficiency provokes lipid peroxidation and reduces antioxidant enzymes' activities because of increased production of reactive oxygen species [48,52].

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Supervision: Kemal Yalçın Gülüt, Ayfer Alkan Torun.

Validation: Kemal Yalçın Gülüt, Ebru Duymuş.

Visualization: Kemal Yalçın Gülüt, Ebru Duymuş.

Writing - original draft: Kemal Yalçın Gülüt.

Writing – review & editing: Ebru Duymuş, İlknur Solmaz, Ayfer Alkan Torun.

#### References

- Shireen F, Nawaz MA, Xiong M, Ahmad A, Sohail H, Chen Z, et al. Pumpkin rootstock improves the growth and development of watermelon by enhancing uptake and transport of boron and regulating the gene expression. Plant Physiol Biochem. 2020; 154: 204–218. https://doi.org/10.1016/j.plaphy.2020. 06.003 PMID: 32563044
- 2. Colla G, Rouphael Y, Mirabelli C, Cardarelli M. Nitrogen-use efficiency traits of mini-watermelon in response to grafting and nitrogen-fertilization doses. J Plant Nutr Soil Sci. 2011; 174: 933–941.
- Torun AA, Solmaz İ, Duymuş E, Aydın O, Cenkseven Ş, Yalçınkaya A, et al. Effect of different doses of nitrogen and potassium fertilization on yield and nutrient uptake in grafted watermelon growing in çukurova region conditions. Int J Agric Nat Sci. 2018; 1: 228–232.
- Nawaz MA, Wang L, Jiao Y, Chen C, Zhao L, Mei M, et al. Pumpkin rootstock improves nitrogen use efficiency of watermelon scion by enhancing nutrient uptake, cytokinin content, and expression of nitrate reductase genes. Plant Growth Regul. 2017; 82: 233–246. https://doi.org/10.1007/s10725-017-0254-7
- Moustafa-Farag M, Bingsheng F, Malangisha Guy K, Hu Z, Yang J, Zhang M. Activated antioxidant enzymes-reduced malondialdehyde concentration, and improved mineral uptake-promoted watermelon seedlings growth under boron deficiency. J Plant Nutr. 2016; 39: 1989–2001.
- Sarwar N., Farooq O., Wasaya A., Hussain M., El-Shehawi A. M., Ahmad S., et al. Integrated nitrogen management improves productivity and economic returns of wheat-maize cropping system. Journal of King Saud University-Science, 2021;101475.
- Luo J, Li H, Liu T, Polle A, Peng C, Luo Z-B. Nitrogen metabolism of two contrasting poplar species during acclimation to limiting nitrogen availability. J Exp Bot. 2013; 64: 4207–4224. <u>https://doi.org/10.1093/jxb/ert234 PMID: 23963674</u>
- Takei K, Sakakibara H, Taniguchi M, Sugiyama T. Nitrogen-dependent accumulation of cytokinins in root and thetranslocation to leaf: Implication of cytokinin species that induces geneexpression of maize responseregulator. Plant Cell Physiol. 2001; 42: 85–93. <u>https://doi.org/10.1093/pcp/pce009</u> PMID: <u>11158447</u>
- Tischner R. Nitrate uptake and reduction in higher and lower plants. Plant Cell Environ. 2000; 23: 1005– 1024.
- Colla G, Suarez CMC, Cardarelli M, Rouphael Y. Improving nitrogen use efficiency in melon by grafting. HortScience. 2010; 45: 559–565.
- Masclaux-Daubresse C, Daniel-Vedele F, Dechorgnat J, Chardon F, Gaufichon L, Suzuki A. Nitrogen uptake, assimilation and remobilization in plants: challenges for sustainable and productive agriculture. Ann Bot. 2010; 105: 1141–1157. https://doi.org/10.1093/aob/mcq028 PMID: 20299346
- Prinsi B, Negri AS, Pesaresi P, Cocucci M, Espen L. Evaluation of protein pattern changes in roots and leaves of Zea mays plants in response to nitrate availability by two-dimensional gel electrophoresis analysis. BMC Plant Biol. 2009; 9: 1–17. https://doi.org/10.1186/1471-2229-9-1 PMID: 19123941
- Curci PL, Cigliano RA, Zuluaga DL, Janni M, Sanseverino W, Sonnante G. Transcriptomic response of durum wheat to nitrogen starvation. Sci Rep. 2017; 7: 1–14. <u>https://doi.org/10.1038/s41598-016-0028-x</u> PMID: 28127051
- 14. Fageria NK. The use of nutrients in crop plants. CRC press; 2016.
- Bolat İ, Kara Ö. Plant nutrients: sources, functions, deficiencies and redundancy. Bartın Orman Fakültesi Derg. 2017; 19: 218–228.
- Gormus O, Barutcular C. Boron Nutrition Studies with Cotton and Sunflower in Southern Turkey. Commun Soil Sci Plant Anal. 2016; 47: 915–929.

- Sheng O, Song S, Peng S, Deng X. The effects of low boron on growth, gas exchange, boron concentration and distribution of 'Newhall'navel orange (*Citrus sinensis* Osb.) plants grafted on two rootstocks. Sci Hortic. 2009; 121: 278–283.
- Brown PH, Hu H. Phloem mobility of boron is species dependent: evidence for phloem mobility in sorbitol-rich species. Ann Bot. 1996; 77: 497–506.
- Krug BA, Whipker BE, McCall I, Frantz J. Elevated relative humidity increases the incidence of distorted growth and boron deficiency in bedding plant plugs. HortScience. 2013; 48: 311–313.
- 20. Lordkaew S, Dell B, Jamjod S, Rerkasem B. Boron deficiency in maize. Plant Soil. 2011; 342: 207–220.
- 21. Dell B, Huang L. Physiological response of plants to low boron. Plant Soil. 1997; 193: 103–120.
- 22. Güneş NT, Horzum O, Güneş E. Economic and technical evaluation of fruit sector in Turkey. Balk Near East J Soc Sci. 2017; 3: 37–49.
- Meister R, Rajani MS, Ruzicka D, Schachtman DP. Challenges of modifying root traits in crops for agriculture. Trends Plant Sci. 2014; 19: 779–788. https://doi.org/10.1016/j.tplants.2014.08.005 PMID: 25239776
- Rogers ED, Benfey PN. Regulation of plant root system architecture: implications for crop advancement. Curr Opin Biotechnol. 2015; 32: 93–98. https://doi.org/10.1016/j.copbio.2014.11.015 PMID: 25448235
- Wissuwa M, Kretzschmar T, Rose TJ. From promise to application: root traits for enhanced nutrient capture in rice breeding. J Exp Bot. 2016; 67: 3605–3615. https://doi.org/10.1093/jxb/erw061 PMID: 27036129
- 26. Nawaz MA, Imtiaz M, Kong Q, Cheng F, Ahmed W, Huang Y, et al. Grafting: a technique to modify ion accumulation in horticultural crops. Front Plant Sci. 2016; 7: 1457. <u>https://doi.org/10.3389/fpls.2016</u>. 01457 PMID: 27818663
- Shireen F, Nawaz MA, Chen C, Zhang Q, Zheng Z, Sohail H, et al. Boron: functions and approaches to enhance its availability in plants for sustainable agriculture. Int J Mol Sci. 2018; 19: 1856. <u>https://doi.org/ 10.3390/ijms19071856</u> PMID: 29937514
- Bie Z, Nawaz MA, Huang Y, Lee J-M, Colla G. Introduction of vegetable grafting. Veg Grafting Princ Pract Wallingford, UK CABI Publ. 2017; 1–21.
- 29. Pulgar G, Villora G, Moreno DA, Romero L. Improving the mineral nutrition in grafted watermelon plants: Nitrogen metabolism. Biol Plant. 2000; 43: 607–609.
- Nawaz MA, Shireen F, Huang Y, Zhilong B, Ahmed W, Saleem BA. Perspectives of vegetable grafting in Pakistan, current status, challenges and opportunities. Int J Agric Biol. 2017; 19: 1165–1174.
- Nawaz MA, Chen C, Shireen F, Zheng Z, Jiao Y, Sohail H, et al. Improving vanadium stress tolerance of watermelon by grafting onto bottle gourd and pumpkin rootstock. Plant Growth Regul. 2018; 85: 41– 56.
- Gregory PJ, Atkinson CJ, Bengough AG, Else MA, Fernández-Fernández F, Harrison RJ, et al. Contributions of roots and rootstocks to sustainable, intensified crop production. J Exp Bot. 2013; 64: 1209–1222. https://doi.org/10.1093/jxb/ers385 PMID: 23378378
- Albacete A, Martínez-Andújar C, Martínez-Pérez A, Thompson AJ, Dodd IC, Pérez-Alfocea F. Unravelling rootstock× scion interactions to improve food security. J Exp Bot. 2015; 66: 2211–2226. <u>https://doi.org/10.1093/jxb/erv027 PMID: 25754404</u>
- Özmen S, Kanber R, Nebahat S, Mustafa Ü. Damla sulama koşullarında aşılı ve aşısız karpuzlarda bitki, su ve verim ilişkilerinin irdelenmesi. Düzce Üniversitesi Bilim ve Teknol Derg. 2014; 2: 141–153.
- Boualem A, Lemhemdi A, Sari M-A, Pignoly S, Troadec C, Abou Choucha F, et al. The andromonoecious sex determination gene predates the separation of Cucumis and Citrullus genera. PLoS One. 2016; 11: e0155444. https://doi.org/10.1371/journal.pone.0155444 PMID: 27171236
- Bahari M, Rafii MY, Saleh GB, Latif MA. Combining ability analysis in complete diallel cross of watermelon (*Citrullus lanatus* (Thunb.) Matsum. & Nakai). Sci World J. 2012; 2012. <u>https://doi.org/10.1100/</u> 2012/543158 PMID: 22566772
- Munisse P, Jensen BD, Andersen SB. Genetic differentiation of watermelon landraces in Mozambique using microsatellite markers. African J Biotechnol. 2013; 12.
- 38. Kacar B, İnal A. Plant Analysis. Nobel Publisher; 2008.
- 39. Bingham FT. Boron. Methods Soil Anal Part 2 Chem Microbiol Prop. 1983; 9: 431–447.
- 40. Bremner JM. Determination of nitrogen in soil by the Kjeldahl method. J Agric Sci. 1960; 55: 11–33.
- **41.** Kelley OJ, Hunter AS, Sterges AJ. Determination of nitrogen, phosphorus, potassium, calcium, and magnesium in plant tissue. Semimicro wet-digestion method for large numbers of samples. Ind Eng Chem Anal Ed. 1946; 18: 319–322.

- Xue Y, Yue S, Zhang W, Liu D, Cui Z, Chen X, et al. Zinc, iron, manganese and copper uptake requirement in response to nitrogen supply and the increased grain yield of summer maize. PLoS One. 2014; 9: e93895. https://doi.org/10.1371/journal.pone.0093895 PMID: 24705926
- **43.** Shapiro SS, Wilk MB. An analysis of variance test for normality (complete samples). Biometrika. 1965; 52: 591–611.
- 44. Steel R., Torrei J, Dickey D. Principles and Procedures of Statistics A Biometrical Approach. A Biometrical Approach. 1997.
- **45.** Hammer Ø, Harper DAT, Ryan PD. PAST: Paleontological statistics software package for education and data analysis. Palaeontol Electron. 2001; 4: 9.
- Demirbas A. The Effects of different fertigation treatments on yield and nutrient uptake of watermelon plants grown as second crop in Cukurova region. Horticulture. 2017; 61: 327–332.
- 47. Wehner TC. Watermelon. Vegetables I. Springer; 2008. pp. 381-418.
- Brown PH, Bellaloui N, Wimmer MA, Bassil ES, Ruiz J, Hu H, et al. Boron in plant biology. Plant Biol. 2002; 4: 205–223.
- 49. O'Neill MA, Ishii T, Albersheim P, Darvill AG. Rhamnogalacturonan II: structure and function of a borate cross-linked cell wall pectic polysaccharide. Annu Rev Plant Biol. 2004; 55: 109–139. https://doi.org/10. 1146/annurev.arplant.55.031903.141750 PMID: 15377216
- 50. Gupta U, Solanki H. Impact of boron deficiency on plant growth. Int J Bioassays. 2013; 2: 1048–1050.
- Landi M, Margaritopoulou T, Papadakis IE, Araniti F. Boron toxicity in higher plants: an update. Planta. 2019; 250: 1011–1032. https://doi.org/10.1007/s00425-019-03220-4 PMID: 31236697
- Landi M, Degl'Innocenti E, Pardossi A, Guidi L. Antioxidant and photosynthetic responses in plants under boron toxicity: a review. Am J Agric Biol Sci. 2012; 7: 255–270.
- Tombuloglu H, Tombuloglu G, Sakcali MS, Turkan A, Hakeem KR, Alharby HF, et al. Proteomic analysis of naturally occurring boron tolerant plant *Gypsophila sphaerocephala* L. in response to high boron concentration. J Plant Physiol. 2017; 216: 212–217. https://doi.org/10.1016/j.jplph.2017.06.013 PMID: 28732263
- Ozturk M, Sakcali S, Gucel S, Tombuloglu H. Boron and plants. Plant adaptation and phytoremediation. Springer; 2010. pp. 275–311.
- Sakcali MS, Kekec G, Uzonur I, Alpsoy L, Tombuloglu H. Randomly amplified polymorphic-DNA analysis for detecting genotoxic effects of Boron on maize (*Zea mays* L.). Toxicol Ind Health. 2015; 31: 712– 720. https://doi.org/10.1177/0748233713483202 PMID: 23546396
- 56. Tombuloglu H, Kekec G, Sakcali MS, Unver T. Transcriptome-wide identification of R2R3-MYB transcription factors in barley with their boron responsive expression analysis. Mol Genet genomics. 2013; 288: 141–155. https://doi.org/10.1007/s00438-013-0740-1 PMID: 23539153
- 57. Goldbach HE, Wimmer MA. Boron in plants and animals: is there a role beyond cell-wall structure? J Plant Nutr Soil Sci. 2007; 170: 39–48.
- Goldbach HE, Yu Q, Wingender R, Schulz M, Wimmer M, Findeklee P, et al. Rapid response reactions of roots to boron deprivation. J Plant Nutr Soil Sci. 2001; 164: 173–181.