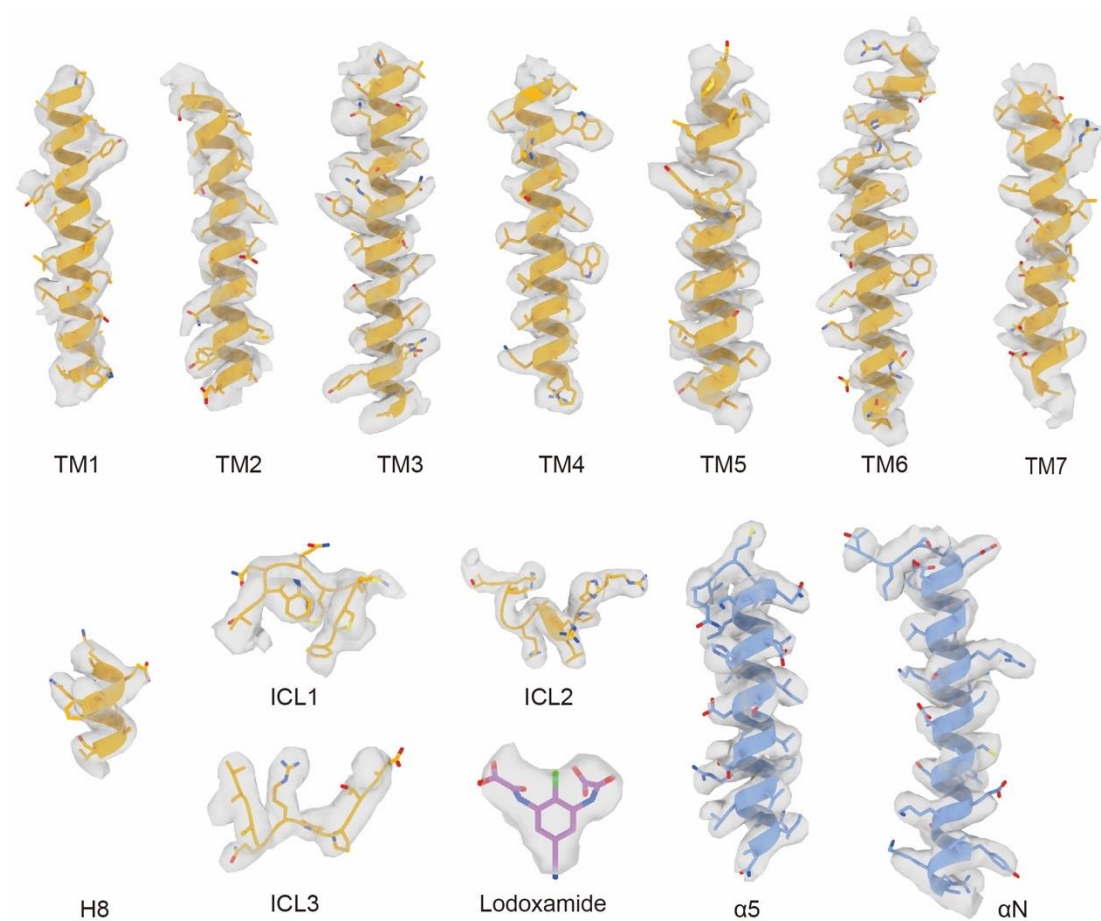
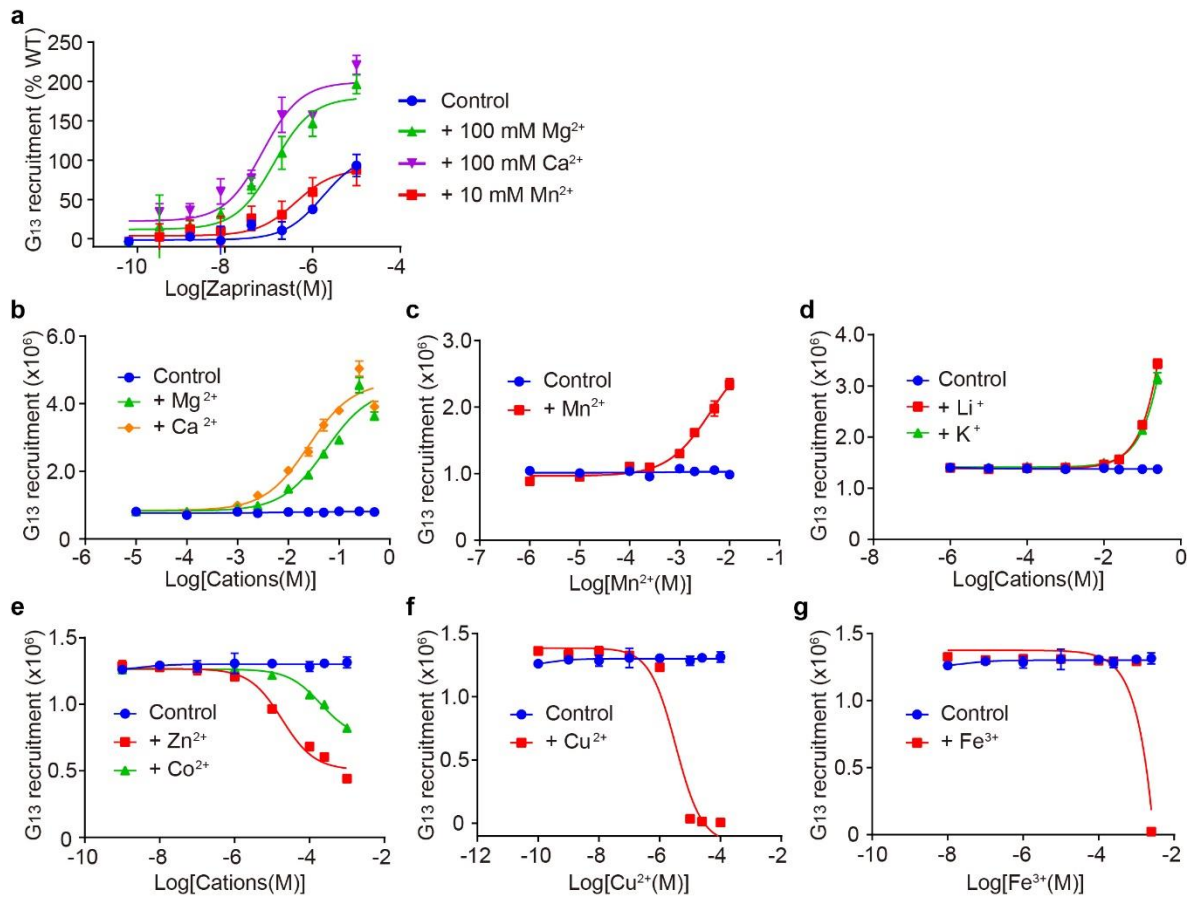


**Fig. S1 The lodoxamide-GPR35-G<sub>13</sub>-scFv16 complex purification and cryo-EM data processing.**

**a** Representative elution profile of the lodoxamide-GPR35-G<sub>13</sub>-scFv16 complex and SDS-PAGE of the size-exclusion chromatography peak. **b, c** Representative cryo-EM micrographs of the lodoxamide-GPR35-G<sub>13</sub>-scFv16 complex (**b**, scale bar: 50 nm) and 2D class averages (**c**, scale bar: 5 nm). **d** Flow chart of cryo-EM data processing of the lodoxamide-GPR35-G<sub>13</sub>-scFv16 complex. **e** 3D density map of the GPR35 complex colored according to local resolution (Å). **f** Gold-standard Fourier Shell Correlation (FSC) curve of the global non-uniform refinement of the complex.



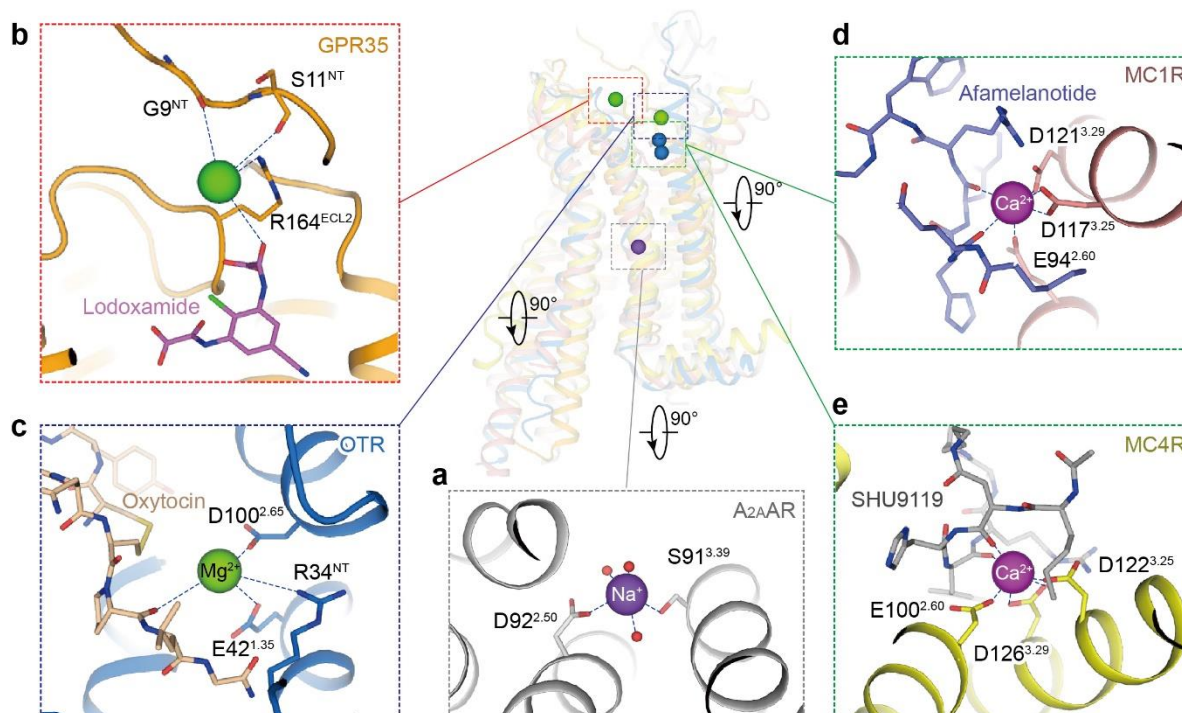
**Fig. S2 Representative EM density and coordinate of the lodoxamide-GPR35-G<sub>13</sub>-scFv16 complex.**



**Fig. S3 Allosteric agonism effects of cations on zaprinast for GPR35 and their effects on the activity of apo GPR35.**

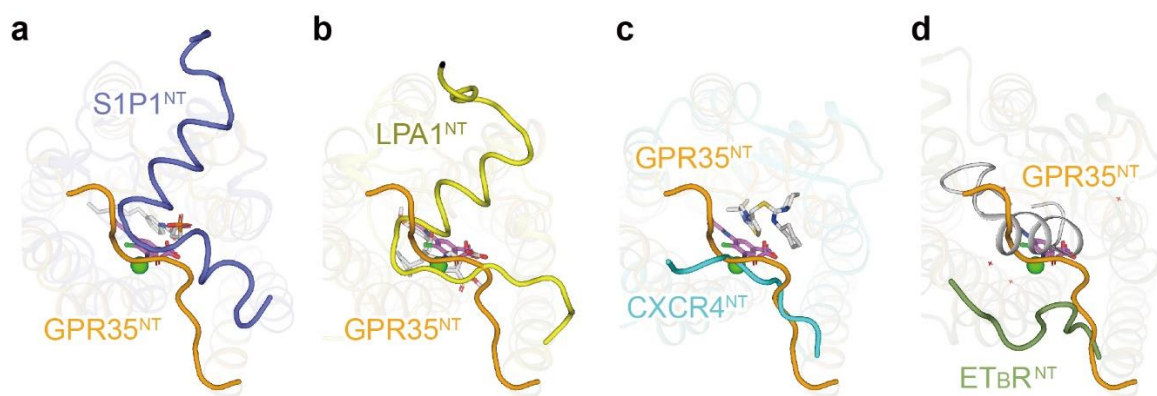
**a** Allosteric agonism effects of divalent cations ( $Mg^{2+}$ ,  $Ca^{2+}$ , and  $Mn^{2+}$ ) on zaprinast for GPR35.

**b-g** Effects of cations on apo GPR35 activity, including  $Mg^{2+}$ ,  $Ca^{2+}$  (**b**),  $Mn^{2+}$  (**c**),  $Li^+$ ,  $K^+$  (**d**),  $Zn^{2+}$ ,  $Co^{2+}$  (**e**),  $Cu^{2+}$  (**f**), and  $Fe^{3+}$  (**g**). The GPR35 pretreated with EDTA and without the addition of any cations is defined as control.



**Fig. S4 Comparison of cation coordination sites in class A GPCRs.**

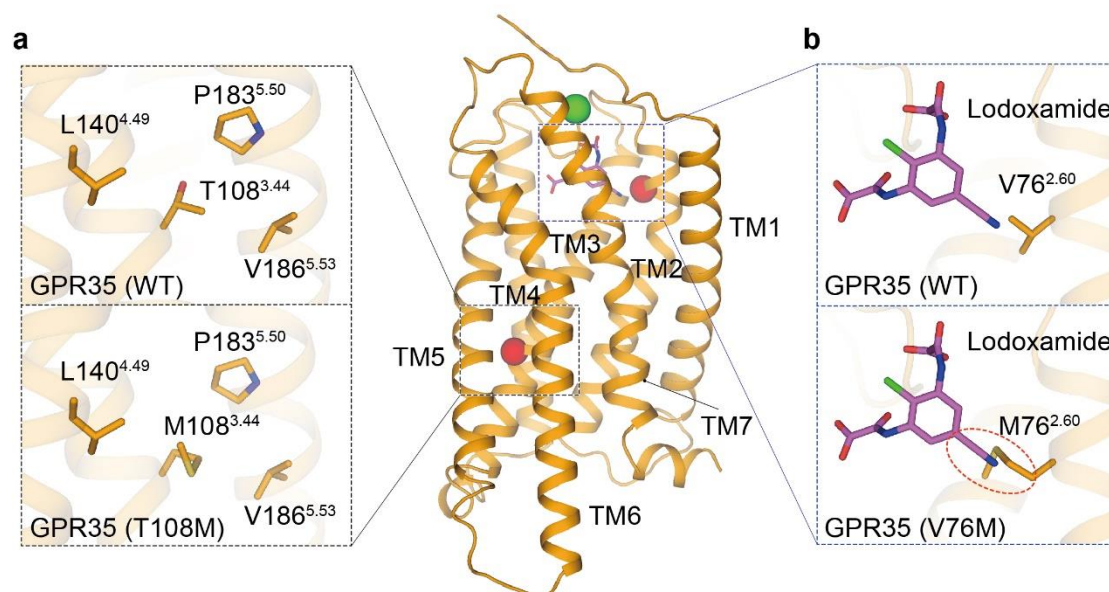
**a**  $\text{Na}^+$  coordination site buried in the TMD helices of  $\text{A}_{2\text{A}}\text{AR}$  (PDB: 4EII). The  $\text{Na}^+$  site is highly conserved across class A GPCRs. **b** The divalent cation binding site in GPR35. **c**  $\text{Mg}^{2+}$  coordination site in oxytocin bound oxytocin receptor (OTR, PDB: 7RYC). **d**  $\text{Ca}^{2+}$  coordination site in afamelanotide bound melanocortin receptor 1 (MC1R, PDB: 7F4H). **e**  $\text{Ca}^{2+}$  coordination site in SHU9119 bound melanocortin receptor 4 (MC4R, PDB: 6W25). Polar interactions between cations and their coordinate residues are indicated by blue dashed lines.



**Fig. S5 Structural comparison of N-terminus in representative class A GPCRs.**

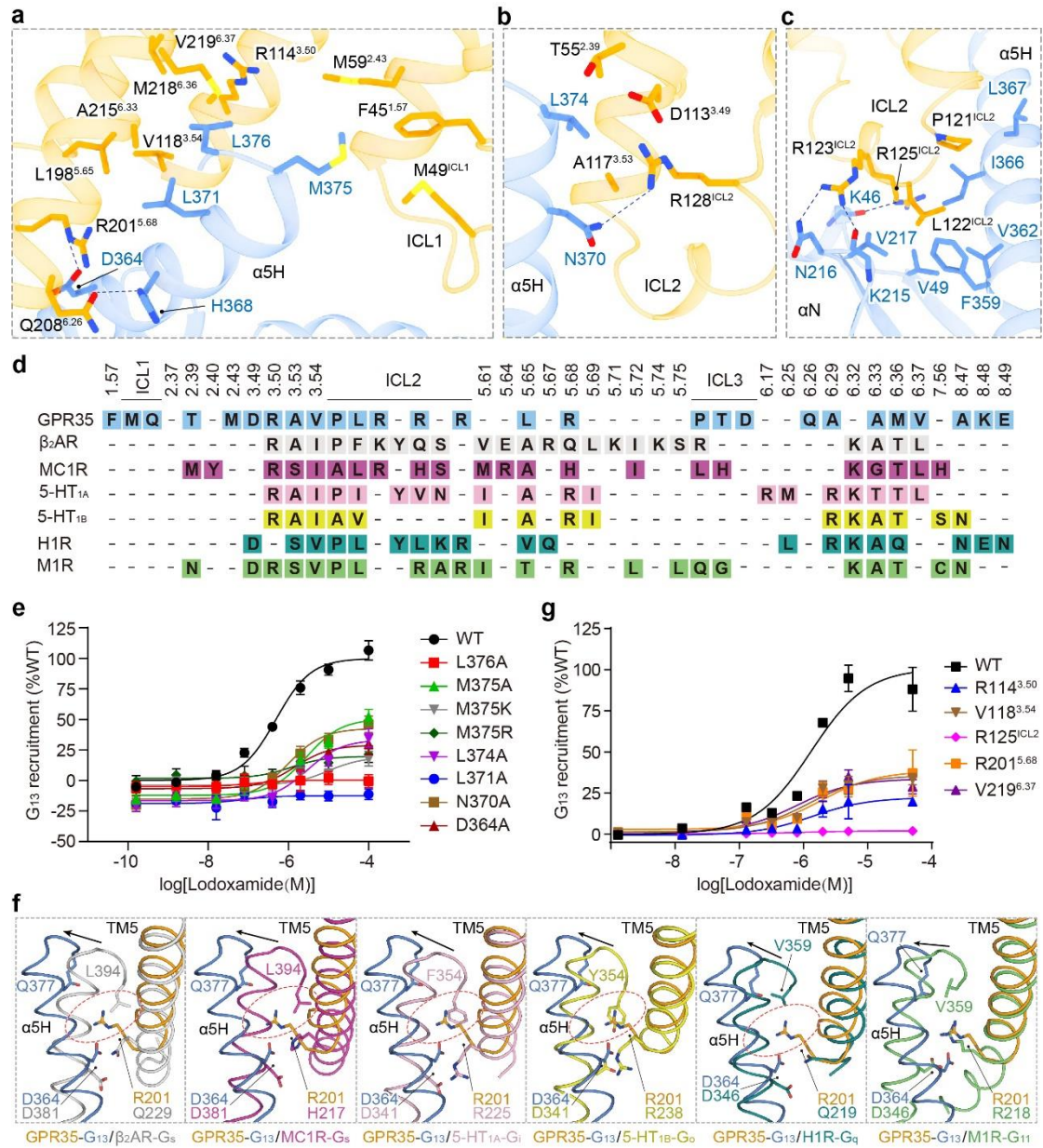
Structural comparison of N-terminus (NT) of GPR35 with that of sphingosine-1-phosphate receptor subtype 1 (S1P1, PDB: 3V2Y, **a**), lysophospholipid receptor 1 (LPA1, PDB: 4Z34, **b**), C-X-C chemokine receptor type 4 (CXCR4, PDB: 3ODU, **c**), and endothelin B receptor (ETBR, PDB: 5GLH, **d**).





**Fig. S6 Disease-associated mutations on GPR35.**

Two disease-associated mutations T108<sup>3.44</sup>M (a) and V76<sup>2.60</sup>M (b) are shown as two red spheres (middle panel). Upper panel, WT GPR35; bottom panel, two disease-associated GPR35 mutants. The potential steric hindrance between lodoxamide and M76<sup>2.60</sup> is highlighted in a red circle.



**Fig. S7  $G_{13}$ -coupling of GPR35.**

**a-c** Detail interactions between the cytoplasmic cavity of receptor helices and  $\alpha 5$  helix of the  $G_{\alpha 13}$  subunit (**a**, **b**), and a hydrophobic interface between ICL2 of the receptor and  $\alpha N$  and  $\alpha 5$  of the  $G_{\alpha 13}$  subunit (**c**). **d** Comparison of the GPR35- $G_{13}$  coupling profile with that of GPCRs coupled to other G protein subtypes. **e** Concentration-response curves of lodoxamide on  $G_{\alpha 13}$  mutants at GPR35- $G_{13}$  interfaces. **f** The interactions between D (-14,  $\alpha 5$  helix numbering start with -1 from the terminal residue) of  $\alpha 5$  helix of  $G_{\alpha}$  subunits and R/Q/H at position 5.68 of receptors. The potential steric hindrance between R/Q/H<sup>5.68</sup> and the C-terminal residue (-1) and entire  $\alpha 5$  helices are highlighted in red dashed circles. The movement of the C-terminus of  $\alpha 5$  helix of  $G_{\alpha}$  subunit for GPR35- $G_{13}$  complex compared to other GPCR-G protein complexes are shown as black arrows. **g** Concentration-response curves of lodoxamide on GPR35 mutants at GPR35- $G_{13}$  interfaces.

**Table S1 Cryo-EM data collection, model refinement, and validation statistics**

Lodoxamide-GPR35-G <sub>13</sub> -scFv16 complex	
<b>Data collection and processing</b>	
Magnification	81,000
Voltage (kV)	300
Electron exposure (e <sup>-</sup> /Å <sup>2</sup> )	50
Defocus range (μm)	-1.0~-2.0
Pixel size (Å)	1.04
Symmetry imposed	C1
Initial particle projections (no.)	32,292,277
Final particle projections (no.)	591246
Map resolution (Å)	3.20
Map resolution range (Å)	2.7-4.0
FSC threshold	0.143
<b>Model Refinement</b>	
Refinement package	PHENIX-1.17.1-3660
Real or reciprocal space	Real space
Model-Map CC (mask)	0.74
Model resolution (Å)	3.3
FSC threshold	0.5
B factors (Å <sup>2</sup> , mean value)	
Protein residues	65.2
Ligands	89.06
<b>Model composition</b>	
Non-hydrogen atoms	8,452
Protein residues	1,138
R.m.s. deviations	
Bond lengths (Å)	0.002 (0)
Bond angles (°)	0.522(4)
<b>Validation</b>	
MolProbity score	1.58
Clashscore	9.63
Rotamer outliers (%)	0.31
Ramachandran plot	
Favored (%)	97.68
Allowed (%)	2.32
Disallowed (%)	0
<b>Data availability</b>	
EMDB entry	EMD-34549
PDB entry	8H8J



**Table S2 Allosteric effects of divalent cations on ligands for GPR35 mutants.** NanoBiT assay was performed to evaluate the effects of divalent cations on the G protein recruitment of GPR35 and 5-HT<sub>1A</sub> in the presence of Iodexamide and 5-HT, respectively. The physiological concentrations of Co<sup>2+</sup>, Zn<sup>2+</sup>, Cu<sup>2+</sup>, and Fe<sup>3+</sup> are much lower than that of Mg<sup>2+</sup> and Ca<sup>2+</sup> (WHO Vitamin and Mineral Nutrition Information System, VMNIS). The maximum cell safety concentrations of these cations under our experimental conditions were used. Data are presented as means  $\pm$  S.E.M. of three independent experiments (n=3). All data were analyzed by two-side, one-way ANOVA with Tukey's test. \*\*\**P* < 0.001 vs. control.

<b>Divalent Cations</b>	<b>GPR35</b>		<b>5-HT<sub>1A</sub></b>
	<i>pEC<sub>50</sub></i> (Iodexamide)	<i>pEC<sub>50</sub></i> (zaprinast)	<i>pEC<sub>50</sub></i> (5-HT)
Control	6.55 $\pm$ 0.02	5.88 $\pm$ 0.05	7.63 $\pm$ 0.02
10 mM Mg <sup>2+</sup>	6.99 $\pm$ 0.05***	/	/
100 mM Mg <sup>2+</sup>	7.09 $\pm$ 0.12***	6.44 $\pm$ 0.03**	7.39 $\pm$ 0.12
10 mM Ca <sup>2+</sup>	7.22 $\pm$ 0.15***	/	/
100 mM Ca <sup>2+</sup>	7.16 $\pm$ 0.16***	7.53 $\pm$ 0.18***	7.71 $\pm$ 0.18
1 mM Mn <sup>2+</sup>	6.94 $\pm$ 0.09***	/	/
10 mM Mn <sup>2+</sup>	7.44 $\pm$ 0.03***	6.56 $\pm$ 0.04**	7.62 $\pm$ 0.11
1 $\mu$ M Co <sup>2+</sup>	6.55 $\pm$ 0.02	/	/
1 $\mu$ M Zn <sup>2+</sup>	6.46 $\pm$ 0.04	/	/
0.1 $\mu$ M Cu <sup>2+</sup>	6.48 $\pm$ 0.05	/	/
10 mM K <sup>+</sup>	6.52 $\pm$ 0.03	/	/
100 mM K <sup>+</sup>	6.55 $\pm$ 0.02	/	/
10 mM Li <sup>+</sup>	6.53 $\pm$ 0.04	/	/
100 mM Li <sup>+</sup>	6.59 $\pm$ 0.07	/	/
100 $\mu$ M Fe <sup>3+</sup>	6.56 $\pm$ 0.08	/	/

**Table S3 Effects of Iodexamide on GPR35 mutants.** NanoBiT assay was performed to evaluate the effects of Iodexamide on  $\beta$ -arrestin 2 or G protein recruitment of GPR35. The surface expression of each GPR35 mutant was normalized to wild-type (WT) receptor, which was set to 100%. Data are presented as means  $\pm$  S.E.M. of three independent experiments (n=3). All data were analyzed by two-side, one-way ANOVA with Tukey's test. \* $P$  < 0.05, \*\* $P$  < 0.01, \*\*\* $P$  < 0.001, vs. WT receptor. U.D., undetectable.

GPR35 Mutants	G protein recruitment	Surface Expression (%WT)
	$pEC_{50}$ (Iodexamide)	
WT	6.02 $\pm$ 0.04	100
V76A	< 5.00 $\pm$ 0.00	50.57 $\pm$ 1.91
L80A	5.07 $\pm$ 0.16**	75.15 $\pm$ 3.62
Y96A	U.D.	65.52 $\pm$ 5.16
R100A	U.D.	147.03 $\pm$ 12.54
F163A	U.D.	154.82 $\pm$ 8.23
R240A	5.15 $\pm$ 0.18**	122.99 $\pm$ 2.42
F45A/M49A/M59A	U.D.	164.90 $\pm$ 8.11
R114A	6.03 $\pm$ 0.14	77.12 $\pm$ 4.12
V118A	5.74 $\pm$ 0.13	89.74 $\pm$ 2.00
R125A	U.D.	111.26 $\pm$ 6.30
R201A	5.74 $\pm$ 0.28*	76.86 $\pm$ 3.59
V219A	6.06 $\pm$ 0.08	118.25 $\pm$ 4.94
S265G	U.D.	170.67 $\pm$ 3.20
S265F	U.D.	169.29 $\pm$ 7.48

**Table S4 Effects of Iodoxamide on  $G\alpha_{13}$  mutants.** NanoBiT assay was performed to evaluate the effects of Iodoxamide on  $G\alpha_{13}$  mutants recruitment by GPR35. Data are presented as means  $\pm$  S.E.M. of three independent experiments (n=3). All data were analyzed by two-side, one-way ANOVA with Tukey's test. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , vs. WT  $G\alpha_{13}$  subunit. U.D., undetectable.

<b><math>G\alpha_{13}</math> Mutants</b>	<b><math>pEC_{50}</math></b>
WT	6.36 $\pm$ 0.03
D364A	6.03 $\pm$ 0.10*
N370A	5.97 $\pm$ 0.07*
L371A	U.D.
L374A	5.67 $\pm$ 0.08***
M375A	5.66 $\pm$ 0.06***
M375K	5.44 $\pm$ 0.22***
M375R	5.79 $\pm$ 0.13**
L376A	U.D.