

Fecal DNA Testing of *TWIST1* Methylation Identifies Patients With Advanced Colorectal Adenoma Missed by Fecal Immunochemical Test for Hemoglobin

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INTRODUCTION: We have reported previously that fecal DNA testing of *TWIST1* methylation in combination with the fecal immunochemical test for hemoglobin (FIT) (combination test) is useful for colorectal neoplasia screening. In this study, using larger sample sizes, we studied the clinical performance of the combination test for the detection of colorectal neoplasia and, especially, advanced colorectal adenoma.

METHODS: We performed a prospective study in which FIT, fecal DNA testing of *TWIST1* methylation, and colonoscopy were performed on 372 patients with colorectal neoplasia and 71 subjects without colorectal neoplasia. We assessed the individual clinical performance of each of FIT and fecal DNA testing of *TWIST1* methylation and of the combination test for the detection of colorectal neoplasia including advanced adenoma based on morphologic subtypes.

RESULTS: The FIT alone had a sensitivity of 7.5% (3/40) for nonadvanced adenoma, 32.3% (41/127) for advanced adenoma, and 93.7% (192/205) for colorectal cancer and a specificity of 87.3% (62/71). The combination test had a sensitivity of 35.0% (14/40) for nonadvanced adenoma, 68.5% (87/127) for advanced adenoma, and 95.6% (196/205) for colorectal cancer and a specificity of 80.3% (57/71). For morphological subtypes of advanced adenoma, the sensitivity of FIT was only 28.2% (20/71) for polypoid type and 16.1% (5/31) for nonpolypoid type, whereas the combination test increased the sensitivities to 64.8% (46/71) and 71.0% (22/31), respectively.

DISCUSSION: The combination of the fecal DNA test with FIT seemed to be useful to detect colorectal neoplasia and, especially, advanced adenoma of the nonpolypoid type.

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INTRODUCTION

Colorectal cancer is the second most commonly diagnosed cancer in females and the third most in males in the world (1). It is estimated that 1.8 million new cases and 861,663 deaths occurred worldwide in 2018 (1). Because more than 95% of patients with colorectal cancer would benefit from curative surgery if diagnosed at an earlier or precancerous stage (2), it

is important to develop highly sensitive and specific assays that are noninvasive, inexpensive, and easy to perform to detect precancerous lesions and colorectal cancer at the early stage.

The main approach to colorectal cancer screening throughout the world is the fecal immunochemical test for hemoglobin (FIT). Patients with positive FIT are referred for colonoscopy

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(3). Although the sensitivity of FIT for the diagnosis of colorectal neoplasia is 65.8%–73.8% for colorectal cancer, it decreases to 23.8%–27.1% for the detection of advanced adenoma (4,5). To improve the sensitivity of FIT for the detection of colorectal neoplasia, in 2014, the US Food and Drug Administration approved Cologuard, the first fecal-based colorectal screening test consisting of the combination of FIT and fecal DNA testing for the mutation of *KRAS* proto-oncogene, GTPase (*KRAS*) and methylation of *BMP3* and *NDRG4*. Cologuard improved the sensitivity for colorectal cancer to 92.3%. However, its sensitivity for advanced adenoma remained at a low level of only 42.4% (4,6).

Previously, we reported that *TWIST1* methylation is specific to colorectal neoplasia. Furthermore, we developed a new methylation assay without bisulfite treatment and methylated DNA immunoprecipitation—the combined restriction digital polymerase chain reaction assay (CORD assay). This assay has more than 100 times higher sensitivity for minute quantities of the target methylated gene than the conventional bisulfite-based methylation assay and, thus, overcomes the issue of limited input of DNA (7,8). In our previous study, fecal DNA testing of *TWIST1* methylation by the CORD assay in combination with FIT exhibited a sensitivity of 82.4% for the detection of advanced adenoma (7). However, because the sample size in the previous study was small (10 healthy subjects and 99 patients with colorectal neoplasia), we additionally performed this study using larger sample sizes to better evaluate the clinical performance of the combination test to detect colorectal neoplasias and morphological subtypes of advanced adenoma.

METHODS

We followed the Standards for the Reporting of Diagnostic Accuracy Studies (9).

Clinical materials

This prospective study designed to investigate the impact of the combination test for the detection of colorectal neoplasms was conducted in Yamaguchi University Hospital, IMSUT Hospital, St. Hill Hospital, and Ajisu Kyoritsu Hospital. The study protocol was approved by the institutional review board of each hospital (protocol number: H29-228). Potentially eligible participants were 20 years of age or over and consisted of asymptomatic persons ($n = 80$) and symptomatic patients who were scheduled to undergo colonoscopy for medical check-up and patients referred for further examinations and treatments of colorectal neoplasias ($n = 400$) from October 2007 to December 2019 in each hospital. Consecutive participants who provided a written informed consent for this study were eligible for enrollment (Figure 1). We excluded participants with a history of inflammatory bowel disease, colorectal cancer, or colorectal surgery. All participants were required to provide a fecal specimen for FIT and fecal DNA testing and to undergo colonoscopy. We excluded 30 participants because of a shortage of fecal sample for the DNA testing and 7 patients with colorectal lymphoma, carcinoid, endometriosis, or submucosal tumor as determined by colonoscopy and pathologic examination. The median interval between fecal sampling and colonoscopy was 1 day (range, 0–58 days). Although colonoscopists reported the location and size of all lesions, only the most advanced colorectal epithelial lesion and its location were used to categorize the participants for the analysis. The proximal colon was considered

to include the splenic flexure and all segments proximal to it; the distal colon was considered to include all other segments. We enrolled 480 participants of whom 443 showed results that could be fully evaluated, including 71 subjects without colorectal neoplasia as determined by colonoscopy (no neoplasia group), 40 patients with nonadvanced colorectal adenomas, 127 patients with advanced colorectal adenoma, and 205 patients with colorectal cancer of stages I–IV diagnosed by colonoscopy or surgical resection. Clinicopathologic features are summarized in Table 1. Criteria for advanced adenoma were defined as adenomas of 1 cm or greater in size, or with villous components (tubulovillous or villous), or with high-grade or severe dysplasia (4). Staging was classified according to the International Union Against Cancer (Union for International Cancer Control) (10).

Of the 127 patients with advanced adenoma, morphologic classification was available for 102 patients who underwent endoscopic resection in Yamaguchi University Hospital. The adenomas were independently classified into 2 types: polypoid type (subtype 0-I) and nonpolypoid type (subtype 0-II) according to the Paris classification of superficial neoplastic lesions (11) by 2 endoscopists with more than 9 years of experience in colonoscopy and without any previous information of the results of the FIT and fecal DNA test. Subtype 0-I includes the pedunculated type (0-Ip) and sessile type (0-Is) of which the elevation above the surface of mucosa is more than 2.5 mm (11). Subtype 0-II includes the slightly elevated type (0-IIa, the elevation above the surface of mucosa is less than 2.5 mm), flat type (0-IIb), and slightly depressed type (0-IIc) (11). When there was discordance between the classifications of the 2 endoscopists, the final classification of the morphology was determined after discussion by the 2 endoscopists.

Fecal specimens for DNA testing were collected before bowel preparation for colonoscopy or surgical treatment in Yamaguchi University Hospital, The Hospital of The Institute of Medical Science, IMSUT Hospital, Ajisu Kyoritsu Hospital, or St. Hill Hospital and were stored at -20°C until DNA extraction.

FIT

Fresh fecal specimens were collected into the sampling containers filled with 2 mL of a hemoglobin-stabilizing buffer solution (Eiken Chemical, Tokyo, Japan) before bowel preparation for the colonoscopy procedure. FIT was performed using OC-HEMODIA (Eiken Chemical). The OC-Sensor IO instrument HEMODIA (Eiken Chemical) processed and quantified the FIT results at the cutoff value of 20 μg hemoglobin (Hb)/g feces (100 ng Hb/mL buffer) for a positive test result (12,13).

CORD assay

Fecal samples were thawed from -20°C , and approximately 200 mg of each sample was used for DNA extraction using the QIAamp DNA Stool Mini kit (QIAGEN, Tokyo, Japan), as described previously (7). Eluted DNA (10 μL) was digested for 16 hours at 37°C by the addition of 10 units of HhaI, 10 units of HpaII, and 20 units of exonuclease I (Exo I) (all from Thermo Fisher Scientific, Tokyo, Japan). Additional digestion of DNA was performed for 16 hours at 60°C using 10 units of BstUI (New England Biolabs, Hitchin, United Kingdom). After the restriction was complete, the mixture was heated for 10 minutes at 98°C . We performed multiplex droplet digital polymerase chain reaction to simultaneously count the absolute copy numbers of human telomerase reverse transcriptase (*hTERT*) and the

Table 1. Clinicopathologic characteristics

	No neoplasia (n = 71)	Nonadvanced adenoma (n = 40)	Advanced adenoma (n = 127)	Carcinoma ^a (n = 205)
Age, yr, median (range)	51 (33–79)	58 (37–81)	69 (36–91)	69 (33–92)
Sex				
Men	30	25	81	109
Women	41	15	46	96
Tumor location				
Proximal		19	77	69
Distal		21	50	136
Tumor size, mm, median (range)		4 (1–8)	22 (5–100)	30 (6–150)

^apStage—I: 70, II: 52, III: 77, IV: 6. Pathologic diagnosis—tubular adenocarcinoma: well differentiated, 71; moderately differentiated, 116; poorly differentiated adenocarcinoma, 7; mucinous adenocarcinoma, 8; papillary adenocarcinoma, 2; and squamous cell carcinoma, 1.

methylated target gene (*TWIST1*), as described previously (7). Laboratory testing was performed without the knowledge of the results of either the comparator FIT or clinical findings.

Statistical analyses

To compare variables, the Mann-Whitney *U* test, Fisher test, and receiver operating characteristic curve analysis were used. A *P* value of less than 0.05 was considered statistically significant. Statistical analyses were performed with GraphPad InStat Ver. 3 and GraphPad Prism Ver. 6 statistical software (GraphPad Software, La Jolla, CA).

RESULTS

FIT

The criterion for a positive result of FIT was above a cutoff of 20 μ g Hb/g feces. FIT resulted in a sensitivity of 7.5% (3/40; 95% confidence interval [CI], 1.6%–20.4%) for nonadvanced adenoma, 32.3% (41/127; 95% CI, 24.2%–41.2%) for advanced adenoma, and 93.7% (192/205; 95% CI, 89.4%–96.6%) for colorectal cancer screening and in a specificity of 87.3% (62/71; 95% CI, 77.3%–94.0%) (Table 2).

Fecal DNA testing of *TWIST1* methylation

The distributions of copy numbers of *TWIST1* methylation and the methylation ratio of *TWIST1* are shown in Figure 2a, c, respectively. We set 20 copies of methylated *TWIST1* and a *TWIST1* methylation ratio of 19% as the cutoff points to discriminate between the no neoplasia group and the colorectal cancer group. Each cutoff value was determined so that the specificity would be approximately 95% (Figure 2b, d), which is compatible with the specificity of FIT in screening setting populations (4,14). The *TWIST1* methylation ratio was calculated according to the ratio of methylated *TWIST1* copy numbers to *hTERT* copy numbers. The criterion for a positive result with the fecal DNA testing of *TWIST1* is either the copy number of methylated *TWIST1* is above the cutoff point (≥ 20 copies) or the *TWIST1* methylation ratio is above the cutoff point ($\geq 19\%$), or both are above the cutoff points.

Positive results of the fecal DNA testing of *TWIST1* were found in 8.5% (6/71) of the individuals in the no neoplasia group (specificity of 91.5%; 95% CI, 82.5%–96.8%), in 27.5% (11/40; 95% CI, 14.6%–43.8%) of the nonadvanced adenoma group, in 47.2% (60/127; 95% CI, 38.4%–56.4%) of the advanced adenoma group, and in 44.4% (91/205; 95% CI, 37.4%–51.5%) of the colorectal cancer group (Table 2 and Figure 3a).

Combination of FIT and fecal DNA testing of *TWIST1* methylation

The criterion for a positive result with the combination of FIT and the fecal DNA testing of *TWIST1* methylation (combination test) is either FIT (a cutoff of 20 μ g Hb/g feces) or fecal DNA testing of *TWIST1* is positive or both are positive. The combination test resulted in a sensitivity of 35.0% (14/40; 95% CI,

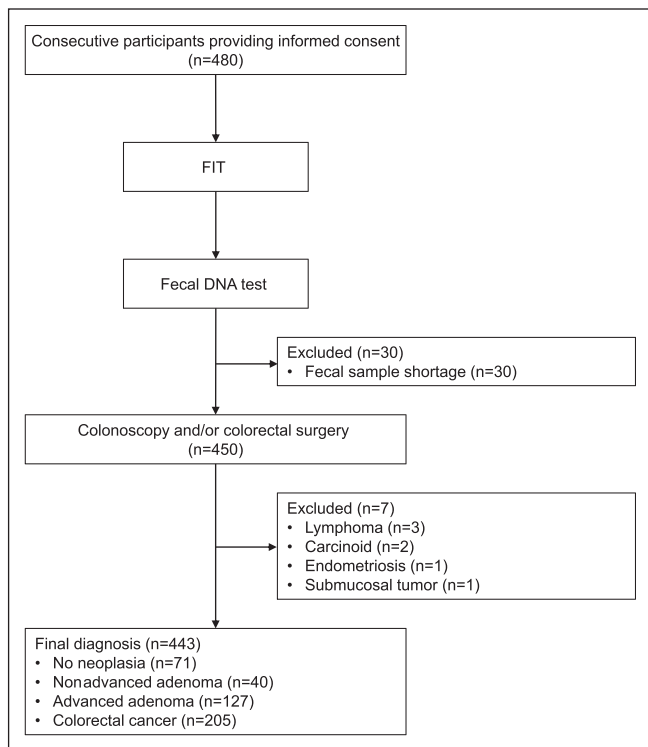


Figure 1. Flow diagram of enrollment. Of the 480 participants who provided an informed consent, 37 were excluded because of a diagnosis of lymphoma, carcinoid, endometriosis, or submucosal tumor or because of failure in fecal sampling. FIT, fecal immunochemical test for hemoglobin.

Table 2. Sensitivity and specificity of fecal DNA test of methylated *TWIST1* and FIT for the findings of colonoscopy

Most advanced findings	Colonoscopy (n = 443), No.	FIT				Fecal DNA test				Combination			
		Positive results, ^a No.	Specificity (95% CI)	P Value	OR (95% CI)	Positive results, ^b No.	Specificity (95% CI)	P Value	OR (95% CI)	Positive results, ^c No.	Specificity (95% CI)	P Value	OR (95% CI)
Negative results on colonoscopy	71	9	87.3% (77.3–94.0)		1.0 (reference)	6	91.5% (82.5–96.8)		1.0 (reference)	14	80.3% (69.1–88.8)		1.0 (reference)
Most advanced findings	Colonoscopy (n = 443), No.	Positive result, ^a No.	Sensitivity (95% CI)	P Value	OR (95% CI)	Positive result, ^b No.	Sensitivity (95% CI)	P Value	OR (95% CI)	Positive result, ^c No.	Sensitivity (95% CI)	P Value	OR (95% CI)
Nonadvanced adenoma	40	3	7.5% (1.6–20.4)	0.5316	0.56 (0.14–2.2)	11	27.5% (14.6–43.8)	0.0122	4.1 (1.4–12.2)	14	35.0% (20.1–51.7)	0.1101	2.2 (0.9–5.3)
Advanced adenoma	127	41	32.3% (24.2–41.2)	0.0021	3.3 (1.5–7.3)	60	47.2% (38.4–56.4)	<0.0001	9.7 (3.9–24.0)	87	68.5% (59.7–76.4)	<0.0001	8.9 (4.4–17.7)
Colorectal cancer	205	192	93.7% (89.4–96.6)	<0.0001	101.7 (41.5–249.5)	91	44.4% (37.4–51.5)	<0.0001	8.7 (3.6–20.1)	196	95.6% (91.8–98.0)	<0.0001	88.7 (36.5–215.5)

CI, confidence interval; FIT, fecal immunochemical test for hemoglobin; OR, odds ratio.
^aCriterion for a positive result of FIT is above a cutoff of 20 µg Hb/g feces.
^bCriterion for a positive result of the fecal DNA test is either 20 or more copy numbers of methylated *TWIST1* or *TWIST1* methylation ratio of 19% or more or both.
^cCriterion for a positive result with the combination of FIT and fecal DNA test is either a positive FIT or fecal DNA test or both are positive.

20.1%–51.7%) for nonadvanced adenoma, 68.5% (87/127; 95% CI, 59.7%–76.4%) for advanced adenoma, and 95.6% (196/205; 95% CI, 91.8%–98.0%) for colorectal cancer, and the specificity was 80.3% (57/71; 95% CI, 69.1%–88.8%) (Table 2 and Figure 3a, b).

Sensitivity for polypoid and nonpolypoid advanced adenomas

The morphologic classification available for 102 of the 127 patients with advanced adenoma was either polypoid type or nonpolypoid type. As shown in Figure 4, the sensitivity of FIT was only 28.2% (20/71; 95% CI, 18.1%–40.2%) for polypoid type and 16.1% (5/31; 95% CI, 5.4%–33.7%) for nonpolypoid type, whereas that of fecal DNA testing of *TWIST1* methylation was 47.9% (34/71; 95% CI, 35.9%–60.1%) and 54.8% (17/31; 95% CI, 36.0%–72.7%), respectively. The combination test resulted in an increase in the sensitivities to 64.8% (46/71; 95% CI, 52.6%–75.7%) and 71.0% (22/31; 95% CI, 52.0%–85.8%), respectively. The results of FIT and fecal DNA testing were considerably mutually exclusive in polypoid type and completely mutually exclusive in nonpolypoid type (Figure 4).

DISCUSSION

Regarding the cutoff point of the CORD assay of *TWIST1* methylation, we had set 5 copies of methylated *TWIST1* in our previous study as a tentative cutoff point to discriminate between the no neoplasia group and the colorectal cancer group, resulting in a specificity of 100% in the small sample size (10 subjects without colorectal neoplasia and 99 patients with colorectal neoplasia) (7). In this study, using a larger sample size, we changed the cutoff point from 5 to 20 copies so that the specificity would be approximately 95%. Furthermore, we found that setting of the cutoff point by the copy number alone is not enough because the copy number is influenced by a substantial amount of water in the fecal samples (fecal forms). Watery feces classified as types 6 and 7 on the Bristol Stool Form Scale (15) results in a low concentration of extracted DNA that leads to low copies of methylated *TWIST1*

and might be assessed as a false-negative result even if the tumor has hypermethylated *TWIST1*. To solve this problem, we added another cutoff point calculated according to the ratio of methylated *TWIST1* copy numbers to *hTERT* copy numbers so that the methylation level of *TWIST1* can be assessed even if the fecal DNA contains only a small amount of DNA. By contrast, there is a disadvantage in assessment by the *TWIST1* methylation ratio. When normal DNA (normal cells) is increased in a fecal sample for reasons including inflammation of the gastrointestinal tract, an increase in *hTERT* copies reduces the ratio of *TWIST1* methylation copies/*hTERT* copies, which might lead to a false-negative result even if the individual has colorectal neoplasm with *TWIST1* hypermethylation. To solve this problem, the assessment by absolute copy number of methylated *TWIST1* is also needed. Therefore, we determined the criterion for a positive result with the fecal DNA testing of *TWIST1* to be either that the copy number of methylated *TWIST1* is above the cutoff point (≥ 20 copies) or the *TWIST1* methylation ratio is above the cutoff point ($\geq 19\%$), or both are above their respective cutoff points.

Colorectal cancer screening

FIT is inevitably used for colorectal cancer detection because of its considerably high sensitivity (65.8%–78.6%) and very high specificity (92.8%–96.4%) (4,5,16). The use of FIT in symptomatic patients presenting in primary care can help rule out colorectal cancer rather than rule it in (17). In this study, the sensitivity of 93.7% was higher than those in reports by other investigators, as mentioned earlier (4,5,16). The difference in sensitivity between this study and those of reports by other groups seems to be because of the difference in the target population of each study. Our study comprised asymptomatic persons, symptomatic patients, and patients referred for further examinations and treatments, whereas the studies by other investigator were limited to asymptomatic persons who were at average risk for colorectal cancer (screening settings) (4,5,16). In this study, although most of the patients with colorectal cancer had positive results of FIT, 13 patients with

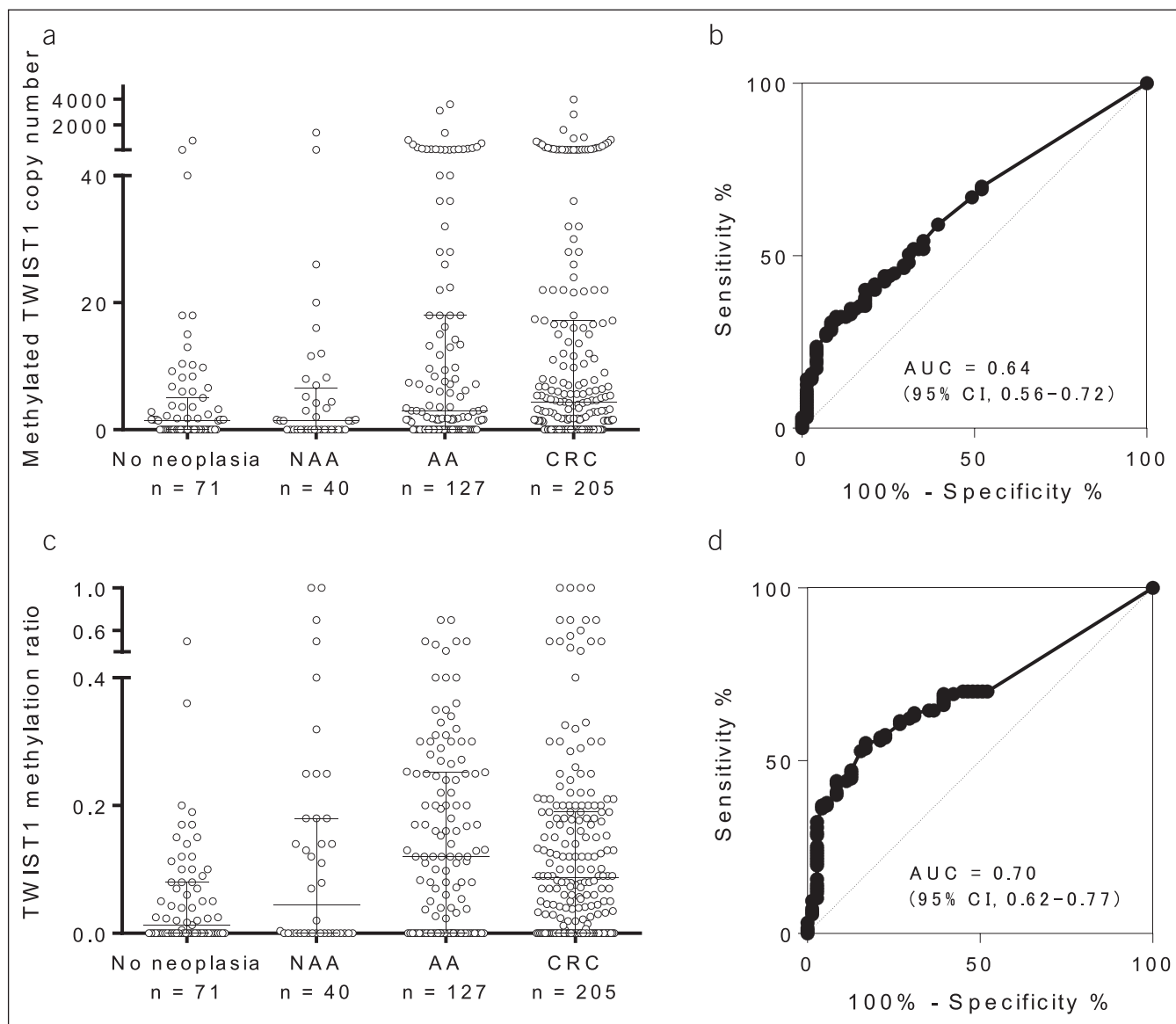


Figure 2. Fecal DNA testing of *TWIST1* methylation. Distribution of methylated *TWIST1* copy numbers (a) and *TWIST1* methylation ratios (ratio of methylated target gene to *human telomerase reverse transcriptase* copy numbers) (c) by fecal combined restriction digital polymerase chain reaction assay in each group are shown. Each sample is indicated by an open circle. The horizontal lines show the median with interquartile range (25th and 75th percentiles). ROC curve analyses of methylated *TWIST1* copy number (b) and *TWIST1* methylation ratio (d) to discriminate between the no neoplasia group and the advanced adenoma group are shown. AA, advanced adenoma; AUC, area under the curve; CI, confidence interval; CRC, colorectal cancer; NAA, nonadvanced adenoma; ROC, receiver operating characteristic.

colorectal cancer had negative results of FIT, suggesting screening by FIT alone may occasionally miss the detection of colorectal cancer. The advantage of fecal DNA testing of *TWIST1* methylation in addition to FIT is to provide a chance to identify some patients with colorectal neoplasia who are missed by FIT. Indeed, of the 13 patients with colorectal cancer who had a negative result of FIT, 4 patients with stage I colorectal cancer were positive for fecal DNA testing of *TWIST1* methylation (Figure 3b). Thus, the combination of the fecal DNA testing of *TWIST1* methylation with FIT may be more useful for colorectal cancer detection than FIT alone. Further studies using a screening setting population are needed to support our findings.

Advanced adenoma detection

In this study, although screening tests for advanced adenoma by independent use of the FIT and fecal DNA testing of *TWIST1* methylation had low or moderate performance (sensitivities of 31.5% and 47.2%, respectively), the combination test increased the sensitivity to 68.5%, showing about twice higher sensitivities than that of FIT alone and an absolute difference of approximately 20 percentage points compared with the sensitivity by the fecal DNA test alone. The synergistic effect of both tests may be because of the mutual exclusiveness of FIT and fecal DNA testing for advanced adenoma screening. Of the 87 patients with positive result(s) of FIT and/or fecal DNA testing of *TWIST1* methylation, 27 (31.0%) had a positive result of FIT alone, 46 (52.9%) had

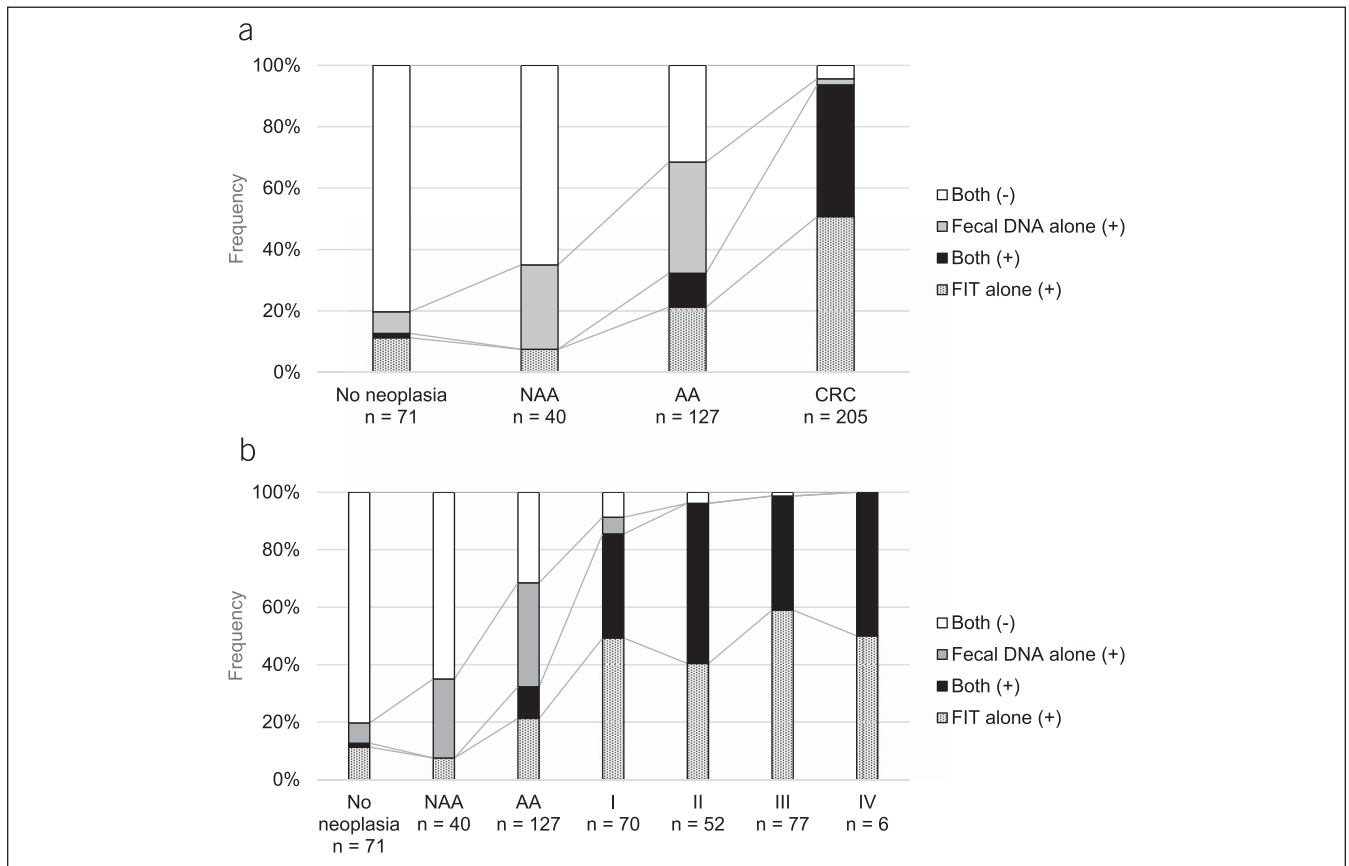


Figure 3. FIT and fecal DNA testing of *TWIST1* methylation for colorectal neoplasia screening. Distribution of the results of FIT and fecal DNA testing of *TWIST1* methylation in the no neoplasia, NAA, AA, and CRC groups is shown (a). Distribution of the results of FIT and fecal DNA testing of *TWIST1* methylation according to colorectal cancer stages is shown (b). Roman numerals indicate stages of colorectal cancer. AA, advanced adenoma; both (+), both FIT and fecal DNA testing are positive; both (-), both FIT and fecal DNA testing are negative; CRC, colorectal cancer; fecal DNA alone (+), fecal DNA test alone is positive; FIT, fecal immunochemical test for hemoglobin; FIT alone (+), FIT alone is positive; NAA, nonadvanced adenoma.

a positive result of fecal DNA testing of *TWIST1* methylation alone, and only 14 (16.1%) had positive results of both FIT and fecal DNA testing of *TWIST1* methylation. Thus, fecal testing with the combination of different modalities would increase the sensitivity for the detection of advanced adenoma. The sensitivity of our combination test for advanced adenoma is superior to that of Cologuard, which also uses a combination of FIT and fecal DNA markers (68.5% vs 42.4%) (4,6). However, because Cologuard is currently not available in Japan, we cannot directly compare the clinical performance of both tests. Therefore, further examination outside Japan will be required to compare clinical performance between the 2 tests.

The sensitivities of FIT for the morphologies of polypoid type and nonpolypoid type were 28.2% and 16.1%, respectively, similar to those of another report in which the sensitivities were 30.1% and 18.5%, respectively (16). In this study, the combination test resulted in a higher sensitivity for each tumor type, especially for the nonpolypoid type. The sensitivity of the combination test for polypoid type was 2.3 times higher than that of FIT alone (64.8% vs 28.2%) and that for nonpolypoid type was 4.4 times higher than that of FIT alone (71.0% vs 16.1%). The synergistic effect of both tests also seems to be because of the mutual exclusiveness of FIT and fecal DNA

testing as mentioned earlier. Interestingly, the results of FIT and fecal DNA testing in nonpolypoid type were completely mutually exclusive. Thus, fecal DNA testing of *TWIST1* methylation would complement FIT for the detection of advanced adenoma. To our knowledge, this is the first report to show the usefulness of the combination of FIT and fecal DNA testing of *TWIST1* methylation for the detection of advanced adenoma based on morphological findings.

Although we set a cutoff of 20 μg Hb/g feces for FIT and our combination test, a threshold of 10 μg Hb/g feces for FIT could decrease the number of false negatives of FIT. Indeed, other investigators reported changes of the cutoff point from 20 to 10 μg Hb/g feces decreased false negatives of FIT for advanced adenoma detection from 72.8% (67/92) to 67.4% (62/92) (18). However, changing the cutoff value resulted in a slight decline in specificity from 97.8% (515/527) to 94.7% (499/527) (18). Because fecal hemoglobin concentration is directly related to the severity of colorectal neoplastic disease (19), further studies are needed to confirm the best cutoff point of FIT for our combination test.

Regarding other methylated genes, 2 blood-based tests (detection of circulating tumor DNA in plasma or serum samples) are reported. One is COLVERA and the other is the Epi

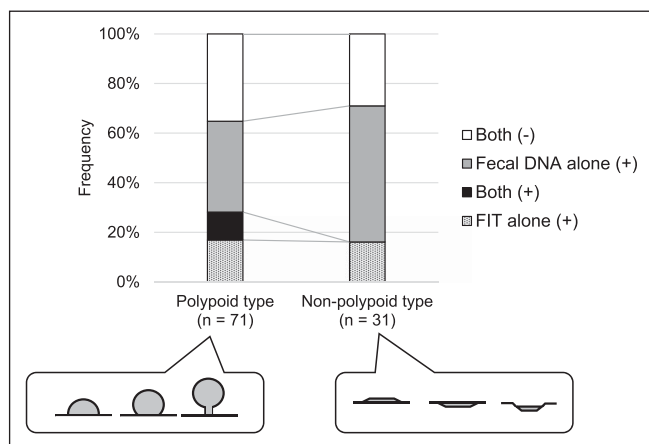


Figure 4. FIT and fecal DNA testing of *TWIST1* methylation for the screening of polypoid and nonpolypoid advanced adenoma. Distribution of the results of FIT and fecal DNA testing of *TWIST1* methylation in polypoid and nonpolypoid advanced adenoma groups is shown. Both (+), both FIT and fecal DNA testing are positive; both (-), both FIT and fecal DNA testing are negative; fecal DNA alone (+), fecal DNA test alone is positive; FIT, fecal immunochemical test for hemoglobin; FIT alone (+), FIT alone is positive.

proColon 2.0 test. COLVERA detects 2 methylated genes (BCAT1 and IKZF1), and Epi proColon detects 1 methylated gene (septin 9). COLVERA is not intended for screening but can be used with carcinoembryonic antigen for surveillance of recurrent colorectal cancer after primary treatment (20). The Epi proColon 2.0 test can be used for screening, and its sensitivity for colorectal cancer is 61%–83% with specificity of 82%–98% (21). However, the sensitivity falls to 27% for advanced adenoma (22). We previously reported that the combination of circulating methylated septin 9 level by the CORD assay with FIT moderately increases the sensitivity for advanced adenoma from 24% to 44% with a slight decline in the specificity from 92% to 80% compared with the serum methylated septin 9 test alone. Thus, the combination of the fecal DNA test of *TWIST1* methylation with FIT seems to offer a better clinical performance to detect advanced adenoma compared with the blood-based test alone and the combination of the blood-based test with FIT. Further studies with larger sample sizes are warranted to confirm our findings.

Specificity

In cancer-screening tests, specificity is as important as sensitivity because specificity affects the number of persons who may have false-positive test results. In this study, the specificity of FIT alone was superior to that of the combination test by a difference of 7.0 percentage points (87.3% vs 80.3%, respectively), resembling another report in which the specificity of FIT alone was superior to that of the combination of FIT with fecal DNA testing of the K-ras mutation and methylation of *NDRG4* and *BMP3* by 6.6 percentage points (96.4% vs 89.8%, respectively) (4).

The baselines of the specificity of the FIT between this study (87.3%) and the reports by other investigator (92.8%–96.4%) (4,5,16) are somewhat different because the target population in each study is different as mentioned earlier. Therefore, further studies using a screening setting

population are needed to assess the clinical performance of the combination test.

Implications for future study

Removing adenomatous polyps of the colon and rectum by colonoscopic polypectomy reduces the incidence of colorectal cancer by up to 90% (23). Furthermore, colonoscopic removal of adenomatous polyps prevents death from colorectal cancer through a 53% reduction in mortality (24). Colonoscopy is the best available method to detect and remove colonic polyps and can, therefore, be considered the gold standard for this purpose (25,26). However, the miss rate of colonoscopy for adenomas of any size is 20.9%–24% and that by size is 26%–27% for adenomas of 1–5 mm, 13% for adenomas of 6–9 mm, and 2.1%–6% for adenomas ≥ 10 mm (27–29). Furthermore, the overall miss rate for adenomas of the flat type is higher than that for the protruding type (44.3% vs 15.1%) (27). The miss rate for flat adenoma by size is 57.8% for adenomas of 1–5 mm, 48.3% for adenomas of 6–9 mm, and 7.6% for adenomas ≥ 10 mm (27). Because the combination of FIT and fecal DNA testing of *TWIST1* methylation by the CORD assay seems to be useful to predict individuals with advanced colorectal adenoma, especially advanced adenoma of the nonpolypoid type, a positive result of the combination test will motivate individuals to undergo colonoscopy. Furthermore, performing the combination test before colonoscopy may predict the presence of colorectal neoplasia, which may lead to more intensive examination during colonoscopy. Confirmatory studies are needed to support our hypothesis.

CONFLICTS OF INTEREST

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Study Highlights

WHAT IS KNOWN

- ✓ The main approach to colorectal cancer screening is the FIT for which the sensitivity for the detection of advanced colorectal adenoma is low, ranging from 23.8% to 27.1%. For morphological subtypes of advanced adenoma, its sensitivity is 30.1% for polypoid type but only 18.5% for nonpolypoid type.
- ✓ Colonoscopy is the gold standard for the detection of colonic polyps. However, the miss rate of colonoscopy is 20.9%–24% for adenomas of any size, and the overall miss rate for adenomas of the flat type is higher than that for the protruding type (44.3% vs 15.1%).
- ✓ Fecal DNA testing in combination with FIT has recently been introduced for colorectal neoplasia detection. However, its sensitivity for advanced adenoma remains at a low level of only 42.4%.
- ✓ We reported that *TWIST1* methylation is specific to colorectal neoplasia and developed a new methylation assay without bisulfite treatment and methylated DNA immunoprecipitation, the CORD assay.

WHAT IS NEW HERE

- ✓ Fecal DNA testing of *TWIST1* methylation by the CORD assay in combination with the FIT increased sensitivity to 68.5% for the detection of advanced adenoma.
- ✓ For morphological subtypes of advanced adenoma, the sensitivity of the combination test for polypoid type was 2.3 times higher than that of FIT alone (64.8% vs 28.2%), and that for nonpolypoid type was 4.4 times higher than that of FIT alone (71.0% vs 16.1%).

TRANSLATIONAL IMPACT

- ✓ Performing fecal DNA testing of *TWIST1* methylation by the CORD assay in combination with the FIT before colonoscopy might predict the presence of colorectal neoplasia, which might lead to more intensive examination during colonoscopy.

REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68:394–424.
2. Pawa N, Arulampalam T, Norton JD. Screening for colorectal cancer: Established and emerging modalities. *Nat Rev Gastroenterol Hepatol* 2011;8:711–22.
3. Young GP, Symonds EL, Allison JE, et al. Advances in fecal occult blood tests: The FIT revolution. *Dig Dis Sci* 2015;60:609–22.
4. Imperiale TF, Ransohoff DF, Itzkowitz SH, et al. Multitarget stool DNA testing for colorectal-cancer screening. *N Engl J Med* 2014;370:1287–97.
5. Morikawa T, Kato J, Yamaji Y, et al. A comparison of the immunochemical fecal occult blood test and total colonoscopy in the asymptomatic population. *Gastroenterology* 2005;129:422–8.
6. Ahlquist DA. Multi-target stool DNA test: A new high bar for noninvasive screening. *Dig Dis Sci* 2015;60:623–33.
7. Suehiro Y, Zhang Y, Hashimoto S, et al. Highly sensitive faecal DNA testing of *TWIST1* methylation in combination with faecal immunochemical test for haemoglobin is a promising marker for detection of colorectal neoplasia. *Ann Clin Biochem* 2018;55:59–68.
8. Suehiro Y, Hashimoto S, Higaki S, et al. Blood free-circulating DNA testing by highly sensitive methylation assay to diagnose colorectal neoplasias. *Oncotarget* 2018;9:16974–87.
9. Bossuyt PM, Reitsma JB, Bruns DE, et al. STARD 2015: An updated list of essential items for reporting diagnostic accuracy studies. *BMJ* 2015;351:h5527.
10. Sobin LH, Gospodarowicz MK, Wittekind C (eds). *TNM Classification of Malignant Tumours*. 7th edn. Wiley-Blackwell: Oxford, 2009.
11. The Paris endoscopic classification of superficial neoplastic lesions: Esophagus, stomach, and colon: November 30 to December 1, 2002. *Gastrointest Endosc* 2003;58:S3–43.
12. Fraser CG, Allison JE, Halloran SP, et al. A proposal to standardize reporting units for fecal immunochemical tests for hemoglobin. *J Natl Cancer Inst* 2012;104:810–4.
13. Chiang TH, Chuang SL, Chen SL, et al. Difference in performance of fecal immunochemical tests with the same hemoglobin cutoff concentration in a nationwide colorectal cancer screening program. *Gastroenterology* 2014;147:1317–26.
14. Gies A, Cuk K, Schrotz-King P, et al. Fecal immunochemical test for hemoglobin in combination with fecal transferrin in colorectal cancer screening. *United European Gastroenterol J* 2018;6:1223–31.
15. O'Donnell LJ, Virjee J, Heaton KW. Detection of pseudodiarrhoea by simple clinical assessment of intestinal transit rate. *BMJ* 1990;300:439–40.
16. Chiu HM, Lee YC, Tu CH, et al. Association between early stage colon neoplasms and false-negative results from the fecal immunochemical test. *Clin Gastroenterol Hepatol* 2013;11:832–8.e1–2.
17. Westwood M, Lang S, Armstrong N, et al. Faecal immunochemical tests (FIT) can help to rule out colorectal cancer in patients presenting in primary care with lower abdominal symptoms: A systematic review conducted to inform new NICE DG30 diagnostic guidance. *BMC Med* 2017;15:189.
18. Bosch LJW, Melotte V, Mongera S, et al. Multitarget stool DNA test performance in an average-risk colorectal cancer screening population. *Am J Gastroenterol* 2019;114:1909–18.
19. Digby J, Fraser CG, Carey FA, et al. Faecal haemoglobin concentration is related to severity of colorectal neoplasia. *J Clin Pathol* 2013;66:415–9.
20. Young GP, Pedersen SK, Mansfield S, et al. A cross-sectional study comparing a blood test for methylated *BCAT1* and *IKZF1* tumor-derived DNA with CEA for detection of recurrent colorectal cancer. *Cancer Med* 2016;5:2763–72.
21. Hu J, Hu B, Gui YC, et al. Diagnostic value and clinical significance of methylated *SEPT9* for colorectal cancer: A meta-analysis. *Med Sci Monit* 2019;25:5813–22.
22. Jin P, Kang Q, Wang X, et al. Performance of a second-generation methylated *SEPT9* test in detecting colorectal neoplasm. *J Gastroenterol Hepatol* 2015;30:830–3.
23. Winawer SJ, Zauber AG, Ho MN, et al. Prevention of colorectal cancer by colonoscopic polypectomy: The National Polyp Study Workgroup. *N Engl J Med* 1993;329:1977–81.
24. Zauber AG, Winawer SJ, O'Brien MJ, et al. Colonoscopic polypectomy and long-term prevention of colorectal-cancer deaths. *N Engl J Med* 2012;366:687–96.
25. Winawer S, Fletcher R, Rex D, et al. Colorectal cancer screening and surveillance: Clinical guidelines and rationale-update based on new evidence. *Gastroenterology* 2003;124:544–60.
26. Mandel JS, Bond JH, Church TR, et al. Reducing mortality from colorectal cancer by screening for fecal occult blood. Minnesota Colon Cancer Control Study. *N Engl J Med* 1993;328:1365–71.
27. Xiang L, Zhan Q, Zhao XH, et al. Risk factors associated with missed colorectal flat adenoma: A multicenter retrospective tandem colonoscopy study. *World J Gastroenterol* 2014;20:10927–37.
28. Rex DK, Cutler CS, Lemmel GT, et al. Colonoscopic miss rates of adenomas determined by back-to-back colonoscopies. *Gastroenterology* 1997;112:24–8.
29. van Rijn JC, Reitsma JB, Stoker J, et al. Polyp miss rate determined by tandem colonoscopy: A systematic review. *Am J Gastroenterol* 2006;101:343–50.

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