





Draft Genome Sequence of Corynebacterium aurimucosum UMB7769, Isolated from the Female Urinary Tract

Samantha Eskandar, a Taylor Miller-Ensminger, b Adelina Voukadinova, b Alan J. Wolfe, c Catherine Putontia, b, c, d

- ^aDepartment of Biology, Loyola University Chicago, Chicago, Illinois, USA
- bBioinformatics Program, Loyola University Chicago, Chicago, Illinois, USA
- Department of Microbiology and Immunology, Stritch School of Medicine, Loyola University Chicago, Maywood, Illinois, USA
- ^dDepartment of Computer Science, Loyola University Chicago, Chicago, Illinois, USA

ABSTRACT Here, we present the draft genome sequence of Corynebacterium aurimucosum UMB7769, isolated from the female urinary tract. The size of the genome is 2,731,818 bp, assembled in 50 contigs, with an observed GC content of 60.9% and an N_{50} score of 129,518 bp. Annotation revealed 31 antibiotic resistance genes.

he prevalence of Corynebacterium in urine has been noted in both healthy and symptomatic male and female patients (1-4). Relative to other bacteria in the urobiome, relatively little is known about C. aurimucosum. Recently, the presence of C. aurimucosum in urine was associated with urinary tract infections (UTI) (5). A subsequent case study explored the possible infectious role of C. aurimucosum due to its presence in a patient with a UTI (6). Here, we report the genome sequence of C. aurimucosum UMB7769, isolated from the voided urine of a woman with a recurrent

C. aurimucosum UMB7769 was isolated from a prior institutional review board (IRB)-approved study (University of California, San Diego, IRB no. 170077AW) using the expanded quantitative urinary culture (EQUC) protocol (1). The sample was collected from a patient at the Women's Pelvic Medicine Center at the University of California, San Diego, in August 2017. One hundred microliters of urine was spread onto a 5% sheep blood agar plate (BAP) and incubated at 35°C in a 5% CO₂ environment for 24 h. Each distinct colony morphology on this plate was subcultured to obtain a pure culture for microbial identification. Matrix-assisted laser desorption ionization-time of flight (MALDI-TOF) mass spectrometry was used to determine the genus and species of this isolate, following protocols detailed previously (1). The isolate was then stored at -80° C until sequencing. From this freezer stock, the C. aurimucosum isolate was streaked onto a Columbia nalidixic acid (CNA) agar plate using the four-quadrant streaking method and incubated at 35°C with 5% CO₂ for 24 h. A single colony was selected and grown in LB overnight under the same conditions. DNA was extracted from this liquid culture using the Qiagen DNeasy blood and tissue kit with a few modifications to the Gram-positive bacterium protocol: in step 2, 230 μ l of lysis buffer (180 μ l of 20 mM Tris-Cl, 2 mM sodium EDTA, and 1.2% Triton X-100 and 50 μ l of lysozyme) was used to resuspend the pellet. The incubation time in step 5 was also shortened from 30 to 10 min. Purified DNA was quantified using a Qubit fluorometer. DNA was sent to the Microbial Genome Sequencing Center (MiGS) at the University of Pittsburgh for sequencing, where the DNA was first enzymatically fragmented using an Illumina tagmentation enzyme. Indices were attached using PCR and then sequenced using an Illumina NextSeq 550 flow cell. In total, 1,041,570 pairs of 150-bp reads were produced for the strain. The raw reads were trimmed using Sickle v1.33 (https://github.com/ najoshi/sickle). The reads were assembled using SPAdes v3.13.0 with the "only-

Citation Eskandar S, Miller-Ensminger T, Voukadinova A, Wolfe AJ, Putonti C. 2020. Draft genome sequence of Corynebacterium aurimucosum UMB7769, isolated from the female urinary tract. Microbiol Resour Announc 9:e00391-20. https://doi.org/10.1128/MRA

Editor David Rasko, University of Maryland School of Medicine

Copyright © 2020 Eskandar et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Catherine Putonti, cputonti@luc.edu.

Received 12 April 2020 Accepted 9 May 2020 Published 28 May 2020



assembler" option for k values of 55, 77, 99, and 127 (7). The genome was annotated using PATRIC v3.6.3 (8); however, the publicly available genome was annotated using Prokaryotic Genome Annotation Pipeline (PGAP) v4.11 (9). Unless previously noted, default parameters were used for each software tool.

The *C. aurimucosum* UMB7769 draft genome is 2,731,818 bp, assembled in 50 contigs, with a GC content of 60.9%. The assembly has a genome coverage of $99 \times$ and an N_{50} score of 129,518 bp. PATRIC annotation identified 31 antibiotic resistance genes, one of which encodes resistance to tetracycline. This resistance was also confirmed by ResFinder v3.2 (10). The PGAP annotation also identified 2,486 protein-coding genes, 52 tRNAs, and 4 rRNA operons. Through further isolation, study, and sequencing of *C. aurimucosum* strains from the urinary tract, we hope to ascertain whether this species should be considered an emerging pathogen.

Data availability. This whole-genome shotgun project has been deposited in GenBank under the accession no. JAAUWA00000000. The version described in this paper is the first version, JAAUWA010000000. The raw sequencing reads have been deposited in the SRA under the accession no. SRR11441031.

ACKNOWLEDGMENTS

This work was conducted as part of the Bacterial Genomics course at Loyola University Chicago's Department of Biology. For prior patient recruitment, we acknowledge the Loyola Urinary Education and Research Collaborative (LUEREC) and the patients who provided the samples for this study.

REFERENCES

- Hilt EE, McKinley K, Pearce MM, Rosenfeld AB, Zilliox MJ, Mueller ER, Brubaker L, Gai X, Wolfe AJ, Schreckenberger PC. 2014. Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. J Clin Microbiol 52:871–876. https://doi.org/10.1128/JCM.02876-13.
- Dong Q, Nelson DE, Toh E, Diao L, Gao X, Fortenberry JD, Van der Pol B. 2011. The microbial communities in male first catch urine are highly similar to those in paired urethral swab specimens. PLoS One 6:e19709. https://doi.org/10.1371/journal.pone.0019709.
- Kline KA, Lewis AL. 2016. Gram-positive uropathogens, polymicrobial urinary tract infection, and the emerging microbiota of the urinary tract. Microbiol Spectr 4. https://doi.org/10.1128/microbiolspec.UTI -0012-2012.
- Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, Kliethermes S, Schreckenberger PC, Brubaker L, Gai X, Wolfe AJ. 2014. The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. mBio 5:e01283-14. https://doi.org/10.1128/mBio.01283-14.
- Leal SM, Jr, Jones M, Gilligan PH. 2016. Clinical significance of commensal Gram-positive rods routinely isolated from patient samples. J Clin Microbiol 54:2928–2936. https://doi.org/10.1128/JCM.01393-16.
- 6. Lo S, Thiam I, Fall B, Ba-Diallo A, Diallo OF, Diagne R, Dia ML, Ka R, Sarr

- AM, Sow AI. 2015. Urinary tract infection with Corynebacterium aurimucosum after urethroplasty stricture of the urethra: a case report. J Med Case Rep 9:156. https://doi.org/10.1186/s13256-015-0638-0.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https:// doi.org/10.1038/srep08365.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi. org/10.1093/jac/dks261.

Volume 9 Issue 22 e00391-20