



Matrix Metalloproteinases in Relation to Bone Mineral Density: A Two-Sample Mendelian Randomization Study

OPEN ACCESS

Edited by:

Guiyou Liu, Tianjin Institute of Industrial Biotechnology (CAS), China

Reviewed by:

Yingfang Ao, Peking University Third Hospital, China Biao Zeng, Icahn School of Medicine at Mount Sinai, United States Jianru Xiao, Shanghai Changzheng Hospital, China

*Correspondence:

Bing Wang wbxyeyy@csu.edu.cn

[†]These authors have contributed equally to this work and share first authorship

Specialty section:

This article was submitted to Statistical Genetics and Methodology, a section of the journal Frontiers in Genetics

> Received: 07 August 2021 Accepted: 02 November 2021 Published: 18 November 2021

Citation:

Lv X, Wu P, Xiao S, Zhang W, Li Y, Ren B, Li Z, Xia K and Wang B (2021) Matrix Metalloproteinases in Relation to Bone Mineral Density: A Two-Sample Mendelian Randomization Study. Front. Genet. 12:754795. doi: 10.3389/fgene.2021.754795 Xin Lv^{1†}, Pengfei Wu^{2,3†}, Shipeng Xiao^{1†}, Wan Zhang⁴, Yawei Li¹, Bolin Ren^{5,6}, Zhihong Li^{5,6}, Kun Xia^{2,3,7} and Bing Wang¹*

¹Department of Spine Surgery, The Second Xiangya Hospital, Central South University, Changsha, China, ²Center for Medical Genetics and Hunan Key Laboratory of Medical Genetics, School of Life Sciences, Central South University, Changsha, China, ³Hunan Key Laboratory of Animal Models for Human Diseases, Central South University, Changsha, China, ⁴Department of Biology, Boston University, Boston, MA, United States, ⁵Department of Orthopedics, The Second Xiangya Hospital, Central South University, Changsha, China, ⁶Hunan Key Laboratory of Tumor Models and Individualized Medicine, The Second Xiangya Hospital, Central South University, Changsha, China, ⁶Hunan Key Laboratory of Tumor Models and Individualized Medicine, The Second Xiangya Hospital, Central South University, Changsha, China, ⁷Hengyang Medical School, University of South China, Hengyang, China

Background: We aimed at investigating causal associations between matrix metalloproteinases (MMPs) and bone mineral density (BMD) by the Mendelian randomization (MR) analysis.

Methods: From genome-wide association studies of European ancestry, we selected instrumental variables for MMP-1, MMP-3, MMP-7, MMP-8, MMP-10, and MMP-12. Accordingly, we retrieved summary statistics of three site-specific BMD, namely, forearm, femoral neck, and lumbar spine. We conducted an inverse variance weighted MR as the primary method to compute overall effects from multiple instruments, while additional MR approaches and sensitivity analyses were implemented. Bonferroni-adjusted significance threshold was set at p < 0.05/18 = 0.003.

Results: Totally, there was no evidence for causal effects of genetically-predicted levels of MMPs on BMD measurement at three common sites. MR results indicated that there were no causal associations of circulating MMPs with forearm BMD (all $p \ge 0.023$) by the inverse variance weighted method. Similarly, there were no causal effects of MMPs on femoral neck BMD (all $p \ge 0.120$) and MR results did not support causal relationships between MMPs and lumbar spine BMD (all $p \ge 0.017$). Multiple sensitivity analyses suggested the robustness of MR results, which were less likely to be biased by unbalanced pleiotropy or evident heterogeneity.

Conclusion: We found no evidence for the causal relationship between MMPs and BMD in the European population.

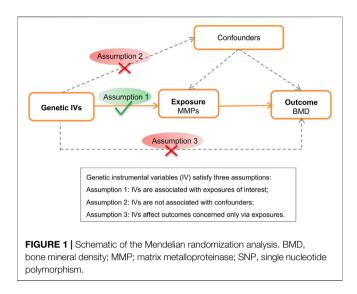
Keywords: matrix metalloproteinase, bone mineral density, mendelian randomization, genome-wide association study, summary statistics, causal inference

1

INTRODUCTION

Bone mineral density (BMD) is a key measurement of bone mass and an essential indicator of osteoporosis, which is prevalent in the aging society. In 1994, the World Health Organization gave the diagnosis standard of osteoporosis as 2.5 SD or more below the young adult average value (Kanis, 1994). The main characteristics of osteoporosis include loss of bone mass, deterioration of the bone microarchitecture, decrement of bone strength and increased risk of fractures, which lead to a systemic skeletal disorder with negative consequences on general health and quality of life in post menopause and in old age (Lane, 2006; Vidal et al., 2019; Capozzi et al., 2020). Fractures due to osteoporosis more likely occur on the hip, vertebral body and distal forearm, therefore, the BMD measurements of forearm (FA), lumbar spine (LS) and femoral neck (FN) are always taken by dual-energy X-ray absorptiometry (DXA) in patients to estimate the general risk of osteoporosis. With the continued ageing of the population worldwide, osteoporotic fractures could present an increasing prevalence and thus lead to higher rates of chronic pain, disability and even death in patients, as well as impose a major economic burden on healthcare systems (Sambrook and Cooper, 2006; Catalano et al., 2017). Current studies have found several risk factors that may decrease BMD (Kenny and Prestwood, 2000; Raisz, 2005; Li and Wang, 2018), but overall, the cause of osteoporosis still remains unclear, which brings difficulty in seeking for effective therapy for this disease.

Matrix metalloproteinases (MMPs) are a family of zincdependent neutral endopeptidases capable of degrading extracellular matrix components (Johansson et al., 2000). Previous studies have found that MMPs are expressed in bone tissue as key players in the digestion of bone matrix by osteoblasts, and are involved in bone-destructive lesions (Wahlgren et al., 2001; Azevedo et al., 2018; Fatemi et al., 2020), which indicates that MMPs may play a role in the pathogenesis of osteoporosis. It has been reported that the gene polymorphism of MMP-1 was associated with osteoporosis (Liang et al., 2019), and MMP-3 was negatively related to the osteoblast function markers of serum bone-specific alkaline phosphatase and osteocalcin while positively related to the resorptive function marker of serum cross-linked Ntelopeptides of type I collagen (Momohara et al., 2005). Increased levels of MMP-7 and 9 in osteoclasts were reported to be associated with rheumatic osteoporosis (Yang et al., 2013), while MMP-8 participated in the healing process as well as embryonic bone development, and may play an important role in the remodeling of extracellular matrix molecules during bone and cartilage formation (Sasano et al., 2002). MMP-10 was found strongly expressed in osteoclasts and most mononuclear cells within the marrow and produced in an active form with associated degradation (Bord et al., 1998). Meanwhile, recombinant MMP-12 cleaved the putative functional domains of osteopontin and bone sialoprotein, two bone matrix proteins that strongly influence osteoclast activities, such as attachment, spreading and resorption (Hou et al., 2004). These studies strongly suggested the possibility that MMPs are related to osteoporosis. However, due to current randomized controlled



trials which were based on either small samples or observational epidemiological studies, whether changes in MMP levels are correlated with BMD remains controversial.

Genome-wide association studies (GWAS) provide a new perspective for understanding genetic determinants that underlie complex disease. The technique of Mendelian randomization (MR), which employs single nucleotide polymorphism (SNPs) as instrumental variables, has been developed to identify causations between a wide range of risk factors and complex diseases. Unlike traditional observational studies, this analytical tool was less susceptible to confounding and reverse causation (Davey Smith and Hemani, 2014). MR has also been widely used these years to explore the causes of osteoporosis (Larsson et al., 2019; Zheng et al., 2019). Given that MMPs were hypothesized to participate in the development of osteoporosis, here we carried out an MR study to identify whether there existed causal associations between MMPs and BMD.

MATERIALS AND METHODS

The MR schematic was shown in **Figure 1**. There were three underlying assumptions: 1) relevance assumption, genetic instrumental variables are associated with the risk factor of interest; 2) independence assumption, genetic variants are not associated with confounders; and 3) exclusion-restriction assumption, instrumental SNPs influence the outcome concerned only through the risk factor (Burgess et al., 2019). This study utilized publicly accessible datasets from published studies wherein formal consent from participants and ethical approval by committees had been obtained.

Data Sources

Summary-level association data for MMPs were obtained from GWASs of European ancestry (Salminen et al., 2017; Folkersen et al., 2020). Folkersen et al. (Folkersen et al., 2020) recently conducted a large-scale mapping of protein quantitative trait loci.

Circulating levels of MMPs, including MMP-1 (n = 16,889), MMP-3 (n = 20,791), MMP-7 (n = 18,245), MMP-10 (n = 16,933), and MMP-12 (n = 19,178) were measured among a panel of 90 candidate biomarkers related to cardiovascular risk. Summary statistics were released by the SCALLOP consortium (http://www.scallop-consortium.com/scallop_downloads/).

Genetic variants associated with MMPs at genome-wide significant significance ($p = 5 \times 10^{-8}$) and clumped at the threshold ($r^2 = 0.001$ within ± 1 Mb, EUR 1000 Genomes phase 3) were selected as instrumental variables (**Supplementary Table S1**). Salminen et al. (Salminen et al., 2017) conducted a GWAS of MMP-8 concentrations in 6,049 Europeans and strongest associations were identified at locus 1q31.3. Two independent SNP associated with MMP-8 meeting the above criteria were utilized as instrumental variables in the ensuing MR analysis. Effect size was given in the unit of SD change in circulating concentration per additional effect allele (**Supplementary Table S2–7**).

Summary statistics for BMD used in this study were gained from the GWAS datasets released by the GEnetic Factors for OSteoporosis Consortium. Zheng et al. (Zheng et al., 2015) performed a large-scale meta-analysis in 2015 to identify genetic variants associated with BMD including FA-BMD (n = 8,143), FN-BMD (n = 32,735) and LS-BMD (n = 28,498) in individuals of European ancestry from the general population. It is the largest GWAS on DXA-measured BMD so far. The associations for BMD were derived from whole-genome sequencing, whole-exome sequencing, deep imputation, and de novo replication genotyping. The association of each SNP with BMD was tested and adjusted for sex, age, square of age and weight. When instrumental SNPs were not present in the BMD datasets, proxies ($r^2 > 0.8$) were searched and utilized if available. Effect size was given in SDs of BMD in association tests with the additive model (Supplementary Table S2-7). Summary statistics of MMPs and BMD were harmonized in terms of effect allele, and subsequent analyses were based on the merged exposureoutcome dataset.

Mendelian Randomization

The MR analysis was conducted using the TwoSampleMR (version 0.5.4) package (Hemani et al., 2018) in R 3.6.3 (R Foundation for Statistical Computing, Vienna, Austria). First, individual estimate of the causal effect MMPs on site-specific BMD mediated by each instrumental SNP was computed as the Wald ratio (Walker et al., 2019). Then, the primary method, the inverse variance weighted (IVW) MR was employed to generate overall estimates (Burgess et al., 2013). Two complementary approaches were implemented, considering that IVW estimates would be biased in the presence of invalid instruments or horizontal pleiotropy. Weighted median approach would give robust effect estimates when less than half instruments were invalid (Bowden et al., 2016). MR-Egger regression would serve as a tool to detect unbalanced horizontal pleiotropy, and generate estimates adjusted for pleiotropy (Burgess and Thompson, 2017). IVW estimates were generally more precise, whereas effect estimates given by weighted median and MR-Egger were accompanied by wide confidence intervals

(CIs) in the forest plots. Causal effects on BMD were presented in SD units per 1-SD increase in circulating levels of MMPs. The Bonferroni-corrected significance level at p < 0.05/18 = 0.003 was adopted in the scenario of multiple tests.

RESULTS

Mendelian Randomization Analyses of Matrix Metalloproteinases on FA-Bone Mineral Density

MR results demonstrated that genetically-predicted levels of MMPs were not associated with changes in FA-BMD (Figure 2). By the primary method, causal effects on FA-BMD were 0.024 SD (-0.018-0.402, p = 0.402) per 1-SD increase in MMP-1 levels, -0.005 SD (-0.074-0.065; p = 0.896) per 1-SD increase in MMP-3 levels, -0.218 SD (-0.461-0.025; *p* = 0.079) per 1-SD increase in MMP-7 levels, -0.252 SD (-0.535-0.032; p =0.082) per 1-SD increase in MMP-8 levels, -0.271 SD (-0.504--0.038; p = 0.023) per 1-SD increase in MMP-10 levels, and -0.016 SD (-0.070-0.039; p = 0.575) per 1-SD increase in MMP-10 levels. MR results were generally consistent among causal estimates given by IVW methods and two additional approaches (Supplementary Table S8). In MR analyses with three or more instrumental variables (except for MMP-8), no horizontal pleiotropy was detected according to MR-Egger intercepts and no evident heterogeneity was identified (Supplementary Table S8).

Mendelian Randomization Analyses of Matrix Metalloproteinases on FN-Bone Mineral Density

Overall, MR estimates suggested that circulating concentrations of MMPs were not associated with FN-BMD. As shown in **Figure 2**, there was no evidence for causal effects of MMP-1 (-0.018 SD; -0.059-0.024; p = 0.402), MMP-3 (0.006 SD; -0.027-0.040; p = 0.708), MMP-7 (0.017 SD; -0.070 to 0.104; p = 0.697), MMP-8 (-0.073 SD; -0.168-0.023; p = 0.135), MMP-10 (-0.145 SD; -0.327-0.038; p = 0.120) and MMP-12 (-0.016 SD; -0.042-0.010; p = 0.238) by the IVW approach. Complimentary methods further verified the robustness of MR results by the primary method, and there was no evidence for the existence of unbalanced horizontal pleiotropy or heterogeneity (**Supplementary Table S9**).

Mendelian Randomization Analyses of Matrix Metalloproteinases on LS-Bone Mineral Density

MR analyses showed that genetically-predicted MMPs were not in relation to LS-BMD (**Figure 2**). Causal relationships between circulating levels of MMP-1 (-0.007 SD; -0.046-0.032; p = 0.718), MMP-3 (0.013 SD; -0.020-0.047; p = 0.430), MMP-7 (0.028 SD; -0.077-0.134; p = 0.599), MMP-8 (-0.107 SD; -0.194--0.019; p = 0.017), MMP-10 (-0.099 SD; -0.223-0.025; p = 0.118) and MMP-12 (-0.006 SD; -0.036-0.025; p = 0.721) and measurement in LS-BMD were not significant by the IVW method.

MMPs	BMD	SNPs		Effect (95% CI)	<i>P</i> -value
		-		0.004 (0.040, 0.004)	0.540
MMP-1	FA-BMD FN-BMD	7		0.021 (-0.048, 0.091)	0.549 0.402
		7		-0.018 (-0.059, 0.024)	
	LS-BMD	7		-0.007 (-0.046, 0.032)	0.718
	FA-BMD	8	· · · · · · · · · · · · · · · · · · ·	-0.005 (-0.074, 0.065)	0.896
MMP-3	FN-BMD	8		0.006 (-0.027, 0.040)	0.708
	LS-BMD	8		0.013 (-0.020, 0.047)	0.430
	FA-BMD	5	• — •	-0.218 (-0.461, 0.025)	0.079
MMP-7	FN-BMD	5		0.017 (-0.070, 0.104)	0.697
	LS-BMD	5	·	0.028 (-0.077, 0.134)	0.599
	FA-BMD	2	• •	-0.252 (-0.535, 0.032)	0.082
MMP-8	FN-BMD	2	·	-0.073 (-0.168, 0.023)	0.135
	LS-BMD	2	·•	-0.107 (-0.194, -0.019)	0.017
	FA-BMD	3	• •	-0.271 (-0.504, -0.038)	0.023
MMP-10	FN-BMD	2	·	-0.145 (-0.327, 0.038)	0.120
	LS-BMD	2	·•	-0.099 (-0.223, 0.025)	0.118
	FA-BMD	6	, 	-0.016 (-0.070, 0.039)	0.575
MMP-12	FN-BMD	6		-0.016 (-0.042, 0.010)	0.238
	LS-BMD	6	- 	-0.006 (-0.036, 0.025)	0.721
			r		
			-0.3 -0.2 -0.1 0 0.1 Beta (95% CI)		

According to sensitivity analyses (**Supplementary Table S10**), MR results by different methods were consistent; besides, unbalanced horizontal pleiotropy or obvious heterogeneity was not present.

DISCUSSION

Osteoporosis is a common cause of morbidity and mortality worldwide especially in people aged over 60 years. Studies have shown that for decrease per 10 percent in bone mineral density, the risk of fracture increases 2–3 folds (Nguyen et al., 1993), and the mortality rate of patients caused by hip and spine fractures increases to 10–20% (Ioannidis et al., 2009). The causes of decrease in BMD have always been discussed in order to benefit for seeking effective therapy, and more and more risk factors are being identified to better predict the occurrence of osteoporosis and therefore avoid the severe complications of fracture.

The family of matrix metalloproteinases have been considered involved in basic pathological processes of osteoporosis for acting as key roles in the digestion of bone matrix by osteoblasts (Azevedo et al., 2018). However, different studies showed conflict results. For example, Zuo et al. (Zuo et al., 2020) found that MMP-8 was involved in the 17β -Estradiol replacement therapy for

postmenopausal osteoporosis, while Viljakainen et al. (Viljakainen et al., 2017) found that there was no significant correlation between MMP-8 levels and low BMD. MR is an effective tool for identifying the causal association between certain exposure and disease while circumventing confounders, which might be the main cause of these inconsistent results. In the recent 3 years, a lot of factors that had been reported related to osteoporosis before have been re-evaluated by MR. Some were further confirmed to be associated with BMD, for instance, serum calcium (Sun et al., 2021), sex hormone-binding globulin (Qu et al., 2021) and age at menarche (Magnus et al., 2020), while others such as vascular endothelial growth factor, uric acid and serum vitamin D got no evidence for their correlations with osteoporosis (Lee and Song, 2019; Sun et al., 2019; Keller-Baruch et al., 2020). In a previous MR study of heel-ultrasound estimated BMD (Folkersen et al., 2020), there were no causal effects of MMP-1, 3, 7, 10, 12 in the European population. In this study, we found no evidence for the causal relationship between MMPs and DXA measured BMD at three common sites.

There are some limitations in present study. First, we could not identify the non-linear relationship between MMPs and BMD. Second, we only evaluated the effect of a small set of MMPs on BMD, but missed such types as MMP-2, -9 and -13, which might be in relation to osteoporosis according to previous studies (Bolton et al., 2009; Zheng et al., 2018). Further MR studies were warranted when relevant datasets are available. Third, it is noteworthy that our study was limited to the effect of circulating MMP levels on BMD, but the intracellular function of MMPs cannot be denied. Forth, both association data of MMPs and BMD were obtained from Europeans in this study. We

should be cautious when generalizing the conclusion to other populations. In this study, we found no evidence for causal relationships

between MMPs (MMP-1, 3, 7, 8, 10, 12) and BMD in the European population.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

XL, PW, KX, and BW contributed to the conceptualization of the study. XL, PW, SX, and WZ played a part in the acquisition and

REFERENCES

- Azevedo, A., Prado, A. F., Feldman, S., de Figueiredo, F. A. T., Dos Santos, M. C. G., and Issa, J. P. M. (2018). MMPs Are Involved in Osteoporosis and Are Correlated with Cardiovascular Diseases. *Cpd* 24, 1801–1810. doi:10.2174/ 1381612824666180604112925
- Bolton, C., Stone, M., Edwards, P., Duckers, J., Evans, W., and Shale, D. (2009). Circulating Matrix Metalloproteinase-9 and Osteoporosis in Patients with Chronic Obstructive Pulmonary Disease. *Chron. Respir. Dis.* 6, 81–87. doi:10.1177/1479972309103131
- Bord, S., Horner, A., Hembry, R. M., and Compston, J. E. (1998). Stromelysin-1 (MMP-3) and Stromelysin-2 (MMP-10) Expression in Developing Human Bone: Potential Roles in Skeletal Development. *Bone* 23, 7–12. doi:10.1016/ s8756-3282(98)00064-7
- Bowden, J., Davey Smith, G., Haycock, P. C., and Burgess, S. (2016). Consistent Estimation in Mendelian Randomization with Some Invalid Instruments Using a Weighted Median Estimator. *Genet. Epidemiol.* 40, 304–314. doi:10.1002/ gepi.21965
- Burgess, S., and Thompson, S. G. (2017). Interpreting Findings from Mendelian Randomization Using the MR-Egger Method. *Eur. J. Epidemiol.* 32, 377–389. doi:10.1007/s10654-017-0255-x
- Burgess, S., Butterworth, A., and Thompson, S. G. (2013). Mendelian Randomization Analysis with Multiple Genetic Variants Using Summarized Data. *Genet. Epidemiol.* 37, 658–665. doi:10.1002/gepi.21758
- Burgess, S., Davey Smith, G., Davies, N. M., Dudbridge, F., Gill, D., Glymour, M. M., et al. (2019). Guidelines for Performing Mendelian Randomization Investigations. Wellcome Open Res. 4, 186. doi:10.12688/ wellcomeopenres.15555.1
- Capozzi, A., Scambia, G., and Lello, S. (2020). Calcium, Vitamin D, Vitamin K2, and Magnesium Supplementation and Skeletal Health. *Maturitas* 140, 55–63. doi:10.1016/j.maturitas.2020.05.020
- Catalano, A., Martino, G., Morabito, N., Scarcella, C., Gaudio, A., Basile, G., et al. (2017). Pain in Osteoporosis: From Pathophysiology to Therapeutic Approach. *Drugs Aging* 34, 755–765. doi:10.1007/s40266-017-0492-4
- Davey Smith, G., and Hemani, G. (2014). Mendelian Randomization: Genetic Anchors for Causal Inference in Epidemiological Studies. *Hum. Mol. Genet.* 23, R89–R98. doi:10.1093/hmg/ddu328

analysis of data, and validation and visualization of results. SX, YL, BR, and ZL participated in drafting and reviewing the main manuscript. ZL, KX, and BW contributed to the project administration, and funding acquisition. All authors contributed to the article and approved the final version of the manuscript.

ACKNOWLEDGMENTS

The authors are grateful to Dr. Folkersen, Dr. Salminen, Dr. Zheng, et al. for making summary-level association statistics of MMPs and BMD possible and accessible. We also want to acknowledge all participants and investigators in these studies. PW has received a visiting PhD scholarship from the China Scholarship Council.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2021.754795/full#supplementary-material

- Fatemi, K., Rezaee, S. A., Banihashem, S. A., Keyvanfar, S., and Eslami, M. (2020). Importance of MMP-8 in Salivary and Gingival Crevicular Fluids of Periodontitis Patients. *Iran J. Immunol.* 17, 236–243. doi:10.22034/ iji.2020.81170.1512
- Folkersen, L., Gustafsson, S., Wang, Q., Hansen, D. H., Hedman, Å. K., Schork, A., et al. (2020). Genomic and Drug Target Evaluation of 90 Cardiovascular Proteins in 30,931 Individuals. *Nat. Metab.* 2, 1135–1148. doi:10.1038/ s42255-020-00287-2
- Hemani, G., Zheng, J., Elsworth, B., Wade, K. H., Haberland, V., Baird, D., et al. (2018). The MR-Base Platform Supports Systematic Causal Inference across the Human Phenome. *Elife* 7, e34408. doi:10.7554/eLife.34408
- Hou, P., Troen, T., Ovejero, M. C., Kirkegaard, T., Andersen, T. L., Byrjalsen, I., et al. (2004). Matrix Metalloproteinase-12 (MMP-12) in Osteoclasts: New Lesson on the Involvement of MMPs in Bone Resorption. *Bone* 34, 37–47. doi:10.1016/j.bone.2003.08.011
- Ioannidis, G., Papaioannou, A., Hopman, W. M., Akhtar-Danesh, N., Anastassiades, T., Pickard, L., et al. (2009). Relation between Fractures and Mortality: Results from the Canadian Multicentre Osteoporosis Study. *Can. Med. Assoc. J.* 181, 265–271. doi:10.1503/cmaj.081720
- Johansson, N., Ahonen, M., and Kähäri, V. M. (2000). Matrix Metalloproteinases in Tumor Invasion. Cell Mol. Life Sci. (Cmls) 57, 5-15. doi:10.1007/s000180050495
- Kanis, J. A., and Kanis, J. A. (1994). Assessment of Fracture Risk and its Application to Screening for Postmenopausal Osteoporosis: Synopsis of a WHO Report. Osteoporos. Int. 4, 368–381. doi:10.1007/bf01622200
- Keller-Baruch, J., Forgetta, V., Manousaki, D., Zhou, S., and Richards, J. B. (2020). Genetically Decreased Circulating Vascular Endothelial Growth Factor and Osteoporosis Outcomes: A Mendelian Randomization Study. J. Bone Miner Res. 35, 649–656. doi:10.1002/jbmr.3937
- Kenny, A. M., and Prestwood, K. M. (2000). Osteoporosis. Rheum. Dis. Clin. North Am. 26, 569–591. doi:10.1016/s0889-857x(05)70157-5
- Lane, N. E. (2006). Epidemiology, Etiology, and Diagnosis of Osteoporosis. Am. J. Obstet. Gynecol. 194, S3–S11. doi:10.1016/j.ajog.2005.08.047
- Larsson, S. C., Michaëlsson, K., and Burgess, S. (2019). Mendelian Randomization in the Bone Field. Bone 126, 51–58. doi:10.1016/j.bone.2018.10.011
- Lee, Y. H., and Song, G. G. (2019). Uric Acid Level, Gout and Bone mineral Density: A Mendelian Randomization Study. *Eur. J. Clin. Invest.* 49, e13156. doi:10.1111/eci.13156

- Li, L., and Wang, Z. (2018). Ovarian Aging and Osteoporosis. *Adv. Exp. Med. Biol.* 1086, 199–215. doi:10.1007/978-981-13-1117-8_13
- Liang, L., Zhu, D. P., Guo, S. S., Zhang, D., and Zhang, T. (2019). MMP-1 Gene Polymorphism in Osteoporosis. *Eur. Rev. Med. Pharmacol. Sci.* 23, 67–72. doi:10.26355/eurrev_201908_18631
- Magnus, M. C., Guyatt, A. L., Lawn, R. B., Wyss, A. B., Trajanoska, K., Küpers, L. K., et al. (2020). Identifying Potential Causal Effects of Age at Menarche: a Mendelian Randomization Phenome-wide Association Study. *BMC Med.* 18, 71. doi:10.1186/ s12916-020-01515-y
- Momohara, S., Okamoto, H., Yago, T., Furuya, T., Nanke, Y., Kotake, S., et al. (2005). The Study of Bone mineral Density and Bone Turnover Markers in Postmenopausal Women with Active Rheumatoid Arthritis. *Mod. Rheumatol.* 15, 410–414. doi:10.1007/s10165-005-0435-510.3109/s10165-005-0435-5
- Nguyen, T., Sambrook, P., Kelly, P., Jones, G., Lord, S., Freund, J., et al. (1993). Prediction of Osteoporotic Fractures by Postural Instability and Bone Density. *Bmj* 307, 1111–1115. doi:10.1136/bmj.307.6912.1111
- Qu, Z., Jiang, J., Yang, F., Huang, J., Zhao, J., and Yan, S. (2021). Genetically Predicted Sex Hormone-Binding Globulin and Bone Mineral Density: A Mendelian Randomization Study. *Calcif Tissue Int.* 108, 281–287. doi:10.1007/s00223-020-00770-8
- Raisz, L. G. (2005). Pathogenesis of Osteoporosis: Concepts, Conflicts, and Prospects. J. Clin. Invest. 115, 3318–3325. doi:10.1172/jci27071
- Salminen, A., Vlachopoulou, E., Havulinna, A. S., Tervahartiala, T., Sattler, W., Lokki, M.-L., et al. (2017). Genetic Variants Contributing to Circulating Matrix Metalloproteinase 8 Levels and Their Association with Cardiovascular Diseases. *Circ. Cardiovasc. Genet.* 10. doi:10.1161/circgenetics.117.001731
- Sambrook, P., and Cooper, C. (2006). Osteoporosis. Lancet 367, 2010–2018. doi:10.1016/s0140-6736(06)68891-0
- Sasano, Y., Zhu, J.-X., Tsubota, M., Takahashi, I., Onodera, K., Mizoguchi, I., et al. (2002). Gene Expression of MMP8 and MMP13 during Embryonic Development of Bone and Cartilage in the Rat Mandible and Hind Limb. *J. Histochem. Cytochem.* 50, 325–332. doi:10.1177/002215540205000304
- Sun, J.-y., Zhao, M., Hou, Y., Zhang, C., Oh, J., Sun, Z., et al. (2019). Circulating Serum Vitamin D Levels and Total Body Bone mineral Density: A Mendelian Randomization Study. J. Cel Mol Med. 23, 2268–2271. doi:10.1111/jcmm.14153
- Sun, J.-y., Zhang, H., Zhang, Y., Wang, L., Sun, B.-I., Gao, F., et al. (2021). Impact of Serum Calcium Levels on Total Body Bone mineral Density: A Mendelian Randomization Study in Five Age Strata. *Clin. Nutr.* 40, 2726–2733. doi:10.1016/ j.clnu.2021.03.012
- Vidal, M., Thibodaux, R. J., Neira, L. F. V., and Messina, O. D. (2019). Osteoporosis: a Clinical and Pharmacological Update. *Clin. Rheumatol.* 38, 385–395. doi:10.1007/ s10067-018-4370-1
- Viljakainen, H. T., Koistinen, H. A., Tervahartiala, T., Sorsa, T., Andersson, S., and Mäkitie, O. (2017). Metabolic Milieu Associates with Impaired Skeletal

Characteristics in Obesity. PLoS One 12, e0179660. doi:10.1371/journal.pone.0179660

- Wahlgren, J., Maisi, P. i., Sorsa, T., Sutinen, M., Tervahartiala, T., Pirilä, E., et al. (2001). Expression and Induction of Collagenases (MMP-8 and -13) in Plasma Cells Associated with Bone-Destructive Lesions. J. Pathol. 194, 217–224. doi:10.1002/path.854
- Walker, V. M., Davies, N. M., Hemani, G., Zheng, J., Haycock, P. C., Gaunt, T. R., et al. (2019). Using the MR-Base Platform to Investigate Risk Factors and Drug Targets for Thousands of Phenotypes. *Wellcome Open Res.* 4, 113. doi:10.12688/ wellcomeopenres.15334.2
- Yang, P. T., Meng, X. H., Yang, Y., and Xiao, W. G. (2013). Inhibition of Osteoclast Differentiation and Matrix Metalloproteinase Production by CD4+CD25+ T Cells in Mice. Osteoporos. Int. 24, 1113–1114. doi:10.1007/s00198-012-2014-x
- Zheng, H. F., Forgetta, V., Hsu, Y. H., Estrada, K., Rosello-Diez, A., Leo, P. J., et al. (2015). Whole-genome Sequencing Identifies EN1 as a Determinant of Bone Density and Fracture. *Nature* 526, 112–117. doi:10.1038/nature14878
- Zheng, X., Zhang, Y., Guo, S., Zhang, W., Wang, J., and Lin, Y. (2018). Dynamic Expression of Matrix Metalloproteinases 2, 9 and 13 in Ovariectomy-Induced Osteoporosis Rats. *Exp. Ther. Med.* 16, 1807–1813. doi:10.3892/etm.2018.6356
- Zheng, J., Frysz, M., Kemp, J. P., Evans, D. M., Davey Smith, G., and Tobias, J. H. (2019). Use of Mendelian Randomization to Examine Causal Inference in Osteoporosis. *Front. Endocrinol.* 10, 807. doi:10.3389/fendo.2019.00807
- Zuo, H.-L., Xin, H., Yan, X.-N., Huang, J., Zhang, Y.-P., and Du, H. (2020). 17β-Estradiol Improves Osteoblastic Cell Function through the Sirt1/NFκB/MMP-8 Pathway. *Climacteric* 23, 404–409. doi:10.1080/ 13697137.2020.1758057

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Lv, Wu, Xiao, Zhang, Li, Ren, Li, Xia and Wang. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.