# Ion Channel Selectivity through Stepwise Changes in Binding Affinity

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ABSTRACT Voltage-gated  $Ca^{2+}$  channels select  $Ca^{2+}$  over competing, more abundant ions by means of a high affinity binding site in the pore. The maximum off rate from this site is ~1,000× slower than observed  $Ca^{2+}$  current. Various theories that explain how high  $Ca^{2+}$  current can pass through such a sticky pore all assume that flux occurs from a condition in which the pore's affinity for  $Ca^{2+}$  transiently decreases because of ion interactions. Here, we use rate theory calculations to demonstrate a different mechanism that requires no transient changes in affinity to quantitatively reproduce observed  $Ca^{2+}$  channel behavior. The model pore has a single high affinity  $Ca^{2+}$ binding site flanked by a low affinity site on either side; ions permeate in single file without repulsive interactions. The low affinity sites provide steps of potential energy that speed the exit of a  $Ca^{2+}$  ion off the selectivity site, just as potential energy steps accelerate other chemical reactions. The steps could be provided by weak binding in the nonselective vestibules that appear to be a general feature of ion channels, by specific protein structures in a long pore, or by stepwise rehydration of a permeating ion. The previous ion-interaction models and this stepwise permeation model demonstrate two general mechanisms, which might well work together, to simultaneously generate high flux and high selectivity in single file pores.

KEY WORDS: Ca<sup>2+</sup> channel • ion channel • permeation • selectivity • single file pores

## INTRODUCTION

 $Ca^{2+}$  channels choose  $Ca^{2+}$  over  $Na^+$  at a ratio of 1,000:1 (Hess et al., 1986) because of an intrapore, high affinity  $Ca^{2+}$  binding site ( $K_d = 0.7 \mu M Ca^{2+}$ ). In the absence of Ca<sup>2+</sup>, Na<sup>+</sup> and other monovalent ions pass freely through Ca<sup>2+</sup> channels, but they are blocked when external Ca<sup>2+</sup> concentration rises above 1 µm (Kostyuk et al., 1983; Almers et al., 1984). Block depends on membrane voltage (Lansman et al., 1986; Fukushima and Hagiwara, 1985; Lux et al., 1990), as required of a site within the pore (Woodhull, 1973), and on the direction of ion movement (Kuo and Hess, 1993a, 1993b), as required of a site within a multi-ion pore (Hille and Schwarz, 1978). The maximum off rate from a micromolar binding site in free solution is  $\sim 10^3$  ions per second, yet picoampere currents (over 106 Ca2+ ions per second) pass through Ca2+ channels. This is a specific example of a general problem of channel permeation: how high flux can occur through a channel that selects its preferred ion with an intrapore binding site. This cannot happen in a rigid pore with just a single binding site because, as affinity for an ion increases, the ion's exit rate decreases in exact proportion to the increased occupancy of the site (Bezanilla and Armstrong, 1972).

The gross discrepancy between flux and binding in  $Ca^{2+}$  channels makes them a valuable system for exploring this problem. Several theories of  $Ca^{2+}$  channel permeation have been proposed and they share an essential feature: when  $Ca^{2+}$  ions bind to the channel, the pore's affinity for  $Ca^{2+}$  diminishes, thereby promoting high  $Ca^{2+}$  flux. Here, we describe a model that generates high flux without such changes in affinity, thereby demonstrating a second general mechanism.

The most widely known Ca<sup>2+</sup> channel model proposes a pair of high affinity intrapore binding sites through which ions move in single file (Hess and Tsien, 1984; Almers and McCleskey, 1984). Previously, single file models had successfully described permeation in K<sup>+</sup> channels (Hille and Schwarz, 1978), gramicidin channels (Andersen and Procopio, 1980), and Na<sup>+</sup> channels (Begenisich, 1987). In such models, ions hop sequentially from one site to the other following two rules: (a) an ion cannot bind to a site already occupied by another; (b) an ion cannot pass over a site without binding to it (Hille, 1992). The same rules govern movement of ions through semiconductor crystals (Shockley et al., 1952). Thus, such models explore whether this simple physical precedent can explain the relatively complex selectivity and permeation of ion channel proteins.

Although high fluxes are impossible with just a single high affinity binding site, they can occur when there are two sites because, when both are occupied, the adjacent ions can repel each other. Specifically, Ca<sup>2+</sup> affinity diminishes 20,000-fold when two ions are in the model pore compared with when one ion is present

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(Hess and Tsien, 1984; Almers and McCleskey, 1984). Electrostatic repulsion between  $Ca^{2+}$  ions ~10 Å apart is sufficient to generate this change in affinity, but it is not the only mechanism possible. Competition for binding moieties between two  $Ca^{2+}$  ions in a multiply occupied channel has been suggested as an alternate mechanism (Armstrong and Neyton, 1991; Yang et al., 1993), as has long-lived conformational changes triggered by  $Ca^{2+}$  binding in a single ion pore (Lux et al., 1990; Mironov, 1992).

Two observations indicate that the pore of the Ca<sup>2+</sup> channel contains only a single high affinity binding site, rather than the two suggested by the traditional model. (a) Single point mutations of critical glutamate residues simply shift the pore's affinity for Ca<sup>2+</sup>, whereas micromolar binding should still be present if there were two independent sites with the same affinity (Yang et al., 1993; Kim et al., 1993; Ellinor et al., 1995). (b) As judged from the voltage dependence of Ca<sup>2+</sup> block, Ca<sup>2+</sup> binds to the same pore location whether it arrives from the outside or the inside solution (Kuo and Hess, 1993a; but a different voltage dependence was found by Rosenberg and Chen, 1991). These experiments prompted us to quantitatively evaluate a model, mentioned briefly in Almers and McCleskey (1984), that uses only one high affinity intrapore site and needs no ion-ion interactions or transient decreases in Ca2+ affinity. Using rate theory calculations for single file pores, we test the ability of this mechanism to fit published data regarding: (a) selectivity of Ca2+ over monovalent ions, (b) selectivity of  $Ca^{2+}$  over  $Ba^{2+}$ , (c) sigmoidal current-voltage (I-V)<sup>1</sup> curves, and (d) block by  $Cd^{2+}$ .

#### METHODS

Calculations were done using rate theory models written in Visual Basic 2.0 in which two different kinds of ions move through pores in single file via either two or three binding sites. In such models, the rate that an ion moves from one site to another equals the product of the rate constant for the transition and the probability that the site is occupied by that ion. The rate constant (r) diminishes exponentially as the energy of transition (G) increases relative to the average kinetic energy of an atom (kT): r = $\nu e^{-(G/kT)}$ , where k is the Boltzmann constant, T is the temperature, and  $\nu$ , the fastest possible reaction rate, is taken to be the atomic vibration frequency (5.8  $\times$  10<sup>12</sup> s<sup>-1</sup> = kT/h, where h is Plank's constant). G is the sum of the chemical and electrical potential energies that affect the ion movement:  $G = G_{\rm b} - G_{\rm s} +$  $z\delta V_m,$  where  $G_b$  is the energy of the barrier to be overcome,  $G_s$  is the energy of the present binding site, *z* is the ion's charge, and  $\delta$ is the fraction of the membrane voltage (V<sub>m</sub>) traversed by the ion in passing from the site to the barrier. If desired, rate constants can be adjusted with a "repulsion factor" to reflect negative interaction between ions that occupy adjacent sites. (We used the factor only to reproduce earlier repulsion-dependent models and raised the factor to an exponent of the product of the valences of the adjacent ions, as appropriate for electrostatic repulsion (see Almers and McCleskey, 1984).)

The energies of barriers and binding sites are all adjustable parameters, but critical ones are determined (or at least constrained) by known data. Outer barriers limit entry to the pore, so they must respect the diffusion limit. A typical diffusion limit for a small ion is  $10^9 \text{ M}^{-1} \text{ s}^{-1}$  (Hille, 1992); this corresponds to an energy barrier of 8.7 kT (=  $\ln[10^9/5.8 \times 10^{12}]$ ). The energies of some binding sites can be obtained from apparent dissociation constants ( $K_d$ ) by  $G = -kT \ln (K_d)$ .

The concentration of  $Ca^{2+}$  that causes half block of monovalent ion current provides a close approximation of the energy of the highest affinity  $Ca^{2+}$  binding site in the pore since block is caused by entry of a single  $Ca^{2+}$  into the pore; for example, a half-blocking concentration of 1  $\mu$ M suggests a binding energy of -14 kT (=  $\ln 10^{-6}$ ). The concentration of  $Ca^{2+}$  that causes half-maximal  $Ca^{2+}$  current provides an approximation of the energy of the lowest affinity site since saturation of current reflects filling of the pore; half saturation at 10 mM suggests a binding energy of -4.5 kT (=  $\ln 10^{-2}$ ). The locations of sites and barriers within the electric field can also be free parameters, but we just arranged them symmetrically.

The barrier and site energies determine the individual rate constants and these, together, determine the probabilities that sites are occupied by the different ions. Each site has three possible states: unoccupied or occupied by one of the two ions. Therefore, the two-site pore can exist in 9  $(3^2)$  possible states and the three-site pore can exist in 27 (33). The probabilities of occupancy of the different states of the pore were calculated through their steady state kinetic equations, in which the rates of creation and loss of the state are equal. For example, the steady state of a two-site channel having ion B in the left site and ion A in the right is described by  $\ddot{d}P_{BA}/dt = 0 = r_1P_{0A} + r_2P_{B0} - r_{-1}P_{BA}$  $r_{-2}P_{BA}$ , where  $r_1$  and  $r_{-1}$  are rate constants for entry and exit, respectively, of ion B to the left site, r<sub>2</sub> and r<sub>-2</sub> are rate constants for entry and exit of ion A to the right site, and  $P_{BA}$ ,  $P_{0A}$ , and  $P_{B0}$  are the probabilities that the channel exists in these three states (0 indicates an unoccupied site). Together with the conservation equation (the sum of all probabilities equals 1), there are 9 and 27 independent kinetic equations for the two- and three-site pores, respectively. These equations were solved for the state probabilities in  $9 \times 9$  and  $27 \times 27$  matrices by Gauss elimination. Numbering the states in base 3 greatly eased bookkeeping and troubleshooting for this operation.

Current for each ion was calculated as the net flux (rate constant  $\times$  state probability) over the second energy barrier times the ion's charge. Calculation of currents carried by extracellular Na<sup>+</sup> and Ca<sup>2+</sup> through a single site pore used the formula for a weighted rectangular hyperbola, as would be appropriate for competitive binding at a single site enzyme (Hille, 1992):

$$I = \frac{\left( \left[ \text{Na} \right]_{\text{o}} / K_{\text{Na}} \right) I_{\text{Na} \text{ max}} + \left( \left[ \text{Ca} \right]_{\text{o}} / K_{\text{Ca}} \right) I_{\text{Ca} \text{ max}}}{1 + \left( \left[ \text{Na} \right]_{\text{o}} / K_{\text{Na}} \right) + \left( \left[ \text{Ca} \right]_{\text{o}} / K_{\text{Ca}} \right)},$$

where  $I_{\text{max}}$  is the maximum current of the indicated ion and *K* is the ion's dissociation constant from the site.

#### RESULTS

Choosing Ca<sup>2+</sup> over Na<sup>+</sup>

From a physiological perspective, the selectivity of  $Ca^{2+}$ over Na<sup>+</sup> is the most important feature of the Ca<sup>2+</sup> channel pore; it allows the channel to preferentially pass Ca<sup>2+</sup> despite the 100-fold concentration advantage

<sup>&</sup>lt;sup>1</sup>Abbreviation used in this paper: I-V, current-voltage.



FIGURE 1. Ca<sup>2+</sup> blocks and permeates in two distinct Ca<sup>2+</sup> channel models. (A) Chemical potential energies experienced by Ca<sup>2+</sup> (solid line) or Na<sup>+</sup> (dashed line) in passing through the repulsion model pore (left, from Almers and McCleskey, 1984) or the step model (right). Axes: potential energy (divided by kT, the average kinetic energy per atom) versus fractional distance across the membrane electric field. In the repulsion model, off rates are increased 20,000-fold when two Ca2+ ions simultaneously occupy the pore; in the step model, off rates are unaffected by occupancy. Exact values of parameters are below. (B) Fits to scaled data points (from Almers et al., 1984) describing whole cell Na<sup>+</sup> and Ca<sup>2+</sup> currents through skeletal muscle L-type Ca2+ channels as a function of log extracellular [Ca<sup>2+</sup>]. The three curves describe average current for single channels predicted by the repulsion model (broken line), step model (solid line), and a single-site model (dashed line). (To offset it from the step model curve, 0.005 pA has been added to the repulsion curve.)  $[Ca^{2+}]_i = 0$ ,  $[Na^+]_i = [Na^+]_0 = 32$  mM, and

exhibits appropriate block of Na<sup>+</sup> current by micromolar Ca<sup>2+</sup>, but no significant Ca<sup>2+</sup> flux as  $[Ca^{2+}]$  rises further (Fig. 1 *B*, *dashed curve*). As discussed above, high Ca<sup>2+</sup> flux occurs in the repulsion model because off rates increase when both sites are simultaneously occupied by Ca<sup>2+</sup> ions. In contrast, off rates are unaffected by multiple occupancy in the other model. This new model has a single high affinity site, two flanking low affinity sites, and allows no repulsion between ions. We will call this the "step model" because the low affinity sites essentially provide stairsteps of potential energy

channel data.

that speed exit from the pore (see DISCUSSION). The inset shows the contributions of the individual ions to the total current in the step model in the vicinity of  $10^{-4}$  M Ca<sup>2+</sup>. Fig. 1 *C* plots the probability that the step model pore is occupied in various ways as Ca<sup>2+</sup> concentration changes.

Block of Na<sup>+</sup> current occurs as occupancy of the high

held by Na<sup>+</sup>. The two essential features of this selectiv-

ity are: (a) Na<sup>+</sup> (and other inorganic monovalents) pass freely through the channel when  $Ca^{2+}$  is absent,

but their currents are blocked by micromolar  $Ca^{2+}$ ; (*b*) high  $Ca^{2+}$  flux occurs at millimolar  $Ca^{2+}$  and there is

no inward Na<sup>+</sup> flux at these concentrations. These

properties are evident in the data points in Fig. 1 B

(scaled and replotted from Almers et al., 1984), which show combined  $Na^+$  and  $Ca^{2+}$  current through  $Ca^{2+}$ 

channels over a 1,000,000-fold range of extracellular  $[Ca^{2+}]$ . There is a broad minimum between  $10^{-6}$  and  $10^{-3}$  M Ca<sup>2+</sup>; Na<sup>+</sup> carries the current below and Ca<sup>2+</sup>

carries it above this range. In practice, fitting this data

is the primary challenge of the modeling; once success-

ful, the same model easily fits a variety of other Ca<sup>2+</sup>

model (Fig. 1 A, left) and the new model (Fig. 1 A, right)

fit the data in Fig. 1 *B* (*broken* and *solid curves*). However, a pore with only a single high affinity binding site

Both the traditional, two-site repulsion-dependent

 $V_m = -20$  mV for data and calculations. Inset shows current carried by Na<sup>+</sup> and Ca<sup>2+</sup> in the step model in the region where the pore switches from  $Na^+$  to  $Ca^{2+}$  permeability. (C) Probability that the repulsion model pore exists in the indicated states. (Empty) No ion in the pore; (Na only) only Na<sup>+</sup> occupies the pore; (1 Ca) Ca<sup>2+</sup> occupies the high affinity site and no other site; (2-3 Ca) two or three Ca<sup>2+</sup> ions occupy the pore. Parameters for Ca<sup>2+</sup> in the repulsion model: outer barriers, 10.3; middle barrier, 0; wells, -14.5. Parameters for Ca<sup>2+</sup> in the step model: outer barriers, 10; inner barriers, -2; outer wells, -4; central well, -15.5. Na<sup>+</sup> in the repulsion model: barriers, 10; wells, -2. Na<sup>+</sup> in the step model: outer barriers, 10; inner barriers, 9.2; wells, -2. Barriers and wells are arranged symmetrically within the membrane field with the outer barriers set at 0.05 Vm. Repulsion factors are 11.89 (repulsion model) and 1 (step model). Single-site model:  $K_{dCa} = 0.3 \,\mu M$  (chosen to provide a small offset;  $0.7 \ \mu M$  gives data that exactly aligns with the other models),  $K_{dNa} = 135$  mM,  $I_{Namax} = 1.5$  pA,  $I_{Camax} =$ 0.32 fA ( $10^3 \text{ Ca}^{2+}/\text{s}$ , the maximum off rate for a 1  $\mu$ M  $K_d$  binding site).



FIGURE 2.  $Ca^{2+}$  and  $Ba^{2+}$  permeation in the step model. (A) Fits to data (from Hess et al., 1986) describing unitary currents for single cardiac L-type  $Ca^{2+}$  channels as a function of  $[Ca^{2+}]_{out}$  or  $[Ba^{2+}]_{out}$  at  $V_m = 0$  mV.  $[Ca^{2+}]_{in} = [Ba^{2+}]_{in} = [Na^+]_{out} = 0$ .  $[Na^+]_{in} = 140$  mM. (*inset*) Energy profiles for  $Ca^{2+}$  and  $Ba^{2+}$ ; exact parameters below. (B) Anomalous mole fraction behavior.  $Ca^{2+}$ current is roughly proportional to  $[Ca^{2+}]$ , but  $Ba^{2+}$  current is lost upon addition of a few millimoles  $Ca^{2+}$ . Consequently, net current passes through a minimum. Sum of extracellular  $[Ca^{2+}]$  and  $[Ba^{2+}] = 10$  mM throughout; intracellular concentrations = 0. V<sub>m</sub> = 0 mV. Parameters for  $Ca^{2+}$  in the model: outer barriers, 9.6; inner barriers, -3; outer wells, -4.5; central well, -15.5.  $Ba^{2+}$ : barriers same as  $Ca^{2+}$ ; outer wells, -3.5; central well, -10.3. Na<sup>+</sup> (not shown): same as in Fig. 1.

affinity site by  $Ca^{2+}$  (one Ca curve) increases when  $Ca^{2+}$ rises to the micromolar range.  $Ca^{2+}$  flux occurs at still higher  $Ca^{2+}$  concentrations as the probability increases that two or three  $Ca^{2+}$  ions simultaneously occupy the pore. Most directly, inward  $Ca^{2+}$  flux parallels the occupancy of the internal low affinity site. This occurs as the external  $Ca^{2+}$  concentration becomes significant compared with the site's dissociation constant, which is 18 mM  $Ca^{2+}$ . Although the occupancy plots for the step model are qualitatively similar to those for the repulsion model (Almers and McCleskey, 1984), the effects of multiple  $Ca^{2+}$  occupancy are different. An ion at one site of the step model does not affect an ion at another site through remote forces.

## Choosing $Ca^{2+}$ over $Ba^{2+}$

 $Ca^{2+}$  channels generally pass greater  $Ba^{2+}$  currents than  $Ca^{2+}$ , but they choose  $Ca^{2+}$  when both ions are present. The key features of Ca<sup>2+</sup>/Ba<sup>2+</sup> permeation are: (a) pure  $Ba^{2+}$  currents are larger and saturate at higher concentrations than pure Ca<sup>2+</sup> currents (Hess et al., 1986); (b) Ba<sup>2+</sup> blocks monovalent currents at  $\sim 50 \times$  higher concentration than does Ca<sup>2+</sup> (Kostyuk et al., 1983); (c) Ca<sup>2+</sup> blocks Ba<sup>2+</sup> current, and does so more strongly than would occur through simple competition for a single site. The powerful block of Ba<sup>2+</sup> current by Ca<sup>2+</sup> can manifest itself as an "anomalous mole fraction effect" in which current carried by mixtures of Ca<sup>2+</sup> and Ba<sup>2+</sup> having constant total divalent ion concentration exhibit a minimum at low  $[Ca^{2+}]$ (Hess and Tsien, 1984; Almers and McCleskey, 1984; Friel and Tsien, 1989; but Yue and Marban, 1990, saw no  $Ca^{2+}/Ba^{2+}$  anomalous mole fraction effect).

The data points in Fig. 2 A (replotted from Hess et al., 1984) give unitary Ca<sup>2+</sup> ( $\bigcirc$ ) and Ba<sup>2+</sup> ( $\bigcirc$ ) current amplitudes from cardiac Ca2+ channels as a function of divalent ion concentration. Being actual single channel amplitudes (rather than the scaled whole cell currents of Fig. 1 B), these data test whether the step model can quantitatively fit observed single channel flux. The  $Ca^{2+}$  (solid curve) and  $Ba^{2+}$  (dashed curve) currents calculated from the model (inset) reproduce the key features of the data—Ba<sup>2+</sup> current has higher amplitude and higher half-saturation than Ca2+ current. The energies for Ca<sup>2+</sup> in the model were adjusted to give a good fit to this cardiac Ca2+ channel data and they differ only slightly from those used to fit skeletal muscle data in Fig. 1. The Ba<sup>2+</sup> and Ca<sup>2+</sup> models differ only in the energy of the binding sites. Ba<sup>2+</sup> energies for the high and low affinity sites correspond to Ba<sup>2+</sup> concentrations that cause half-block of monovalent currents (35 µM, Kostyuk et al., 1983) and half-saturation of Ba<sup>2+</sup> current (28 mM, Hess et al., 1986), respectively.

Fig. 2 *B* shows the predicted anomalous mole fraction behavior when the total divalent ion concentration is 10 mM. Net current passes through a minimum as the Ca<sup>2+</sup>:Ba<sup>2+</sup> ratio changes because Ca<sup>2+</sup> is so effective at blocking Ba<sup>2+</sup> current. For example, there is virtually no current carried by 5 mM Ba<sup>2+</sup> when 5 mM Ca<sup>2+</sup> is present. Fig. 1 *B* shows a similar, but far more dramatic, U shape because the difference in affinities between Ca<sup>2+</sup> and Na<sup>+</sup> is far greater than between Ca<sup>2+</sup> and Ba<sup>2+</sup>. (As seen experimentally [Friel and Tsien, 1989], the mole fraction curve no longer passes through a minimum at high divalent ion concentrations [data not shown]. In this respect, a nearly saturated multi-site pore behaves like a single site pore; presumably, this is



FIGURE 3. Effects of  $Ca^{2+}$  and  $Na^+$  on I-V curves of the step model. (A) I-V curves with the indicated (millimolar) concentrations of extracellular  $Ca^{2+}$ . Current becomes more sigmoid near the reversal potentials as  $Ca^{2+}$  rises. (*inset*) Current carried by Na<sup>+</sup> and  $Ca^{2+}$  near the reversal potential when  $[Ca^{2+}]_{out} = 10$  mM. For all curves,  $[Ca^{2+}]_{in} = 0$ ,  $[Na^+]_{in} = [Na^+]_{out} = 140$  mM. Energies as in Fig. 1 *A*, *right*. (*B*) I-V curves with the indicated internal and external  $[Na^+]$  ( $[Ca^{2+}]_{out} = 3$  mM;  $[Ca^{2+}]_{in} = 0$ ). Reversal potentials: 43.5 mV with symmetrical Na<sup>+</sup>; 41.5 mV with low external Na<sup>+</sup>; 72 mV with low internal Na<sup>+</sup>.

because only a single site at a time is unoccupied in the near-saturated pore.)

## Sigmoid I-V Curves

The current-voltage curve of Ca<sup>2+</sup> channels exposed to physiological ionic concentrations has a rich array of features that further illustrate the role of Ca<sup>2+</sup> block in selectivity against monovalent ions: (a) current reverses at positive potentials, but not nearly as positive as the equilibrium potential for Ca<sup>2+</sup> (Fenwick et al., 1982; Lee and Tsien, 1984); (b) the outward current is carried by intracellular monovalent cations (Lee and Tsien, 1984; Hess et al., 1986), and these are far more permeant than extracellular monovalents (Yamashita et al., 1990); (c) despite their impermeability, extracellular monovalents at physiological concentrations block currents carried by divalent ions (Polo-Parada and Korn, 1997); (d) the I-V curve is sigmoid in the vicinity of the reversal potential (Fenwick et al., 1982; Lee and Tsien, 1984). The step model generates each of these features (Fig. 3).

In the absence of  $Ca^{2+}$ , the single  $Ca^{2+}$  channel I-V curve is linear and reverses at 0 mV (Rosenberg and Chen, 1991); the model does the same (not shown). As

extracellular  $Ca^{2+}$  concentration increases, the reversal potential becomes more positive and the curve becomes more and more sigmoid near the reversal potential (Fig. 3 *A*). Sigmoidicity, a region of low conductance, occurs when the pore is more likely occupied by just a single  $Ca^{2+}$  ion (occupancy plot not shown). When sufficiently positive voltages force the final  $Ca^{2+}$ ion outward from the pore, monovalent cations pass freely outward. Thus, sigmoidicity in the I-V curve reflects block of the pore by single  $Ca^{2+}$  ions.

Fig. 3 A (inset) shows how much of the net current is carried by each of the individual ions near the reversal potential when [Ca<sup>2+</sup>]<sub>out</sub> is 10 mM (0 [Ca<sup>2+</sup>]<sub>in</sub>). Although Na<sup>+</sup> is at the same concentration (140 mM) inside and out, it only carries outward current; clearly, internal Na<sup>+</sup> is more permeant than external in the model. This point was demonstrated in elegant experiments showing that internal Na<sup>+</sup> affects the reversal potential of Ca2+ channels far more than external Na+ (Yamashita et al., 1990). In the step model, a change in Na<sup>+</sup> concentration that causes a 2-mV shift if made extracellularly causes a 27-mV shift if made intracellularly (Fig. 3 B). With respect to monovalent cations,  $Ca^{2+}$ channels are outward rectifiers because of the presence of an extracellular blocking ion, Ca2+. Intracellular Na<sup>+</sup> is more permeant than extracellular because of the difference in Ca<sup>2+</sup> concentration inside and out. As shown previously by Hille and Schwarz (1978), such asymmetry requires neither an asymmetric energy profile nor ionic repulsion.

Block of inward  $Ca^{2+}$  current by 140 mM extracellular Na<sup>+</sup> is reproduced (compare *solid* and *dashed curves*, Fig. 3 *B*). As was suggested by Polo-Parada and Korn (1997), this block is caused by competition between Na<sup>+</sup> and Ca<sup>2+</sup> at the external, low affinity binding site.

## Block by Cadmium

Cd<sup>2+</sup> is one of several polyvalent cations that block Ca<sup>2+</sup> channels at very low concentrations (Hess et al., 1986). The data points in Fig. 4, replotted from Ellinor et al. (1995), show that Cd<sup>2+</sup> blocks monovalent ion currents at ~200-fold lower concentrations than divalent ion currents. The data is fit by assuming that Cd<sup>2+</sup> binds to the selectivity site far more strongly than does Ca<sup>2+</sup> (-24 kT for Cd<sup>2+</sup> [Fig. 4, *inset*] vs. -15.5 kT for Ca<sup>2+</sup>). The difference in blocking concentrations for Li<sup>+</sup> and Ba<sup>2+</sup> currents is due to their differences in binding to this site (-10.3 kT for Ba<sup>2+</sup> and -5 kT for Li<sup>+</sup>).

#### DISCUSSION

This paper asks how a million  $Ca^{2+}$  ions can pass through a  $Ca^{2+}$  channel in a second when its pore contains a binding site having a maximum off rate of a thousand per second. This is a specific example of a



FIGURE 4. Block of monovalent and divalent currents by Cd<sup>2+</sup>. Data points from Ellinor et al. (1995) are normalized inward whole cell currents carried by Li<sup>+</sup> or Ba<sup>2+</sup> at the indicated extracellular [Cd<sup>2+</sup>]. [Ba<sup>2+</sup>]<sub>out</sub> = 40 mM; [Li<sup>+</sup>]<sub>out</sub> = [Li<sup>+</sup>]<sub>in</sub> = 100 mM. V<sub>m</sub> = -20 mV for data and calculations. Calculated Ba<sup>2+</sup> and Li<sup>+</sup> currents normalized to their values at 10<sup>-10</sup> and 10<sup>-12</sup> M Cd<sup>2+</sup>, respectively. (*inset*) Energy profile for Cd<sup>2+</sup> (5 kT per tick). Cd<sup>2+</sup> energies are the same as for Ca<sup>2+</sup> in Fig. 2, except the central well, which was set at -24 kT (absolute  $K_d = 40$  pM) to give a good fit to the data. Ba<sup>2+</sup> energies same as in Fig. 2. Li<sup>+</sup> energies: outer barriers, 9; inner barriers, 7; wells, -5.

general problem: how channels pass high flux if they select their preferred ion with a binding site. All previous models assumed that one Ca<sup>2+</sup> ion diminishes the affinity of the pore for others and that Ca2+ flux ensued from this low affinity state. The suggested mechanisms for this negative interaction between ions are electrostatic repulsion (Hess and Tsien, 1984; Almers and Mc-Cleskey, 1984), competition for binding moieties (Armstrong and Neyton, 1991; Yang et al., 1993; Ellinor et al., 1995), and Ca<sup>2+</sup>-induced conformational changes (Mironov, 1992; Lux et al., 1990). The step model described here does not allow one ion to alter another's binding to other sites, yet it fits the wide variety of Ca<sup>2+</sup> channel permeation data; most importantly, submicromolar Ca<sup>2+</sup> blocks current carried by monovalent ions and Ca<sup>2+</sup> above a millimolar generates picoampere, saturable Ca<sup>2+</sup> flux. The calculations demonstrate that ion-ion interactions are not required to generate high flux in pores that tightly bind their preferred ion. Kiss et al. (1998) demonstrates that the same principle fits data from K<sup>+</sup> channels. Below, we describe the mechanism that underlies the model, evidence that supports its plausibility, and alternate interpretations of it.

#### Mechanism

The model has a simple mathematical explanation and a physical mechanism that is part of everyday experience. Fig. 5 shows the potential energies of  $Ca^{2+}$  in a single-site pore (*A*), the repulsion model (*B*), and the step model (*C*). The energy of the strongest binding site in each model (*arrow 1*) determines block of foreign ions by micromolar  $Ca^{2+}$ . The energy of the high-



FIGURE 5.  $Ca^{2+}$  energies in three models. The potential energies that determine  $Ca^{2+}$  block (1) and limit  $Ca^{2+}$  flux (2) are interlinked in a one-site pore (A); since both energies increase together, strong binding and high flux cannot both occur. In contrast, the block and flux energies in the repulsion model (B) and the step model (C) can be independently manipulated. In the repulsion model, multiple  $Ca^{2+}$  occupancy ( $\bullet$ ) forces a change to a low affinity, high flux state due to the negative interaction between the ions. Low affinity sites are built into the step model, making ion–ion interactions unnecessary for high flux. The energy of the low affinity (escape) site in both successful models is ~100 millielectronvolts (scale bar in A).

est barrier to exit the pore (*arrow 2*) limits  $Ca^{2+}$  flux. In a single-site pore, the rate-limiting exit barrier increases as the selecting binding well deepens. This pictorially represents why an increase in selectivity is expected to decrease flux.

The repulsion and step models provide two ways of separating the energies that control binding and flux. In the repulsion model (Fig. 5 *B*), double occupancy (*right*) of the pore diminishes affinity of the binding sites. Dissociation constants for Ca<sup>2+</sup> change from 0.5  $\mu$ M to 10 mM when the pore goes from singly occupied

to doubly occupied. Whether the mechanism is electrostatic repulsion, competition, or conformational changes,  $Ca^{2+}$  flux occurs from this state of shallow binding wells.

In the step model (Fig. 5 C), flux occurs from low affinity sites that are built into the energy profile; therefore, there is no need for multiple occupancy to alter binding affinities. The essence of the mathematics is trivial: because the energies that control binding and flux are similar in the two viable models, binding and flux are similar. The physical mechanisms of permeation differ, but with the arrows having roughly similar length in the two models, the calculations come out approximately the same. In both the repulsion and step models, exit from the pore occurs from a site of  $\sim -4$ kT. This is  $\sim 100$  millielectronvolts (Fig. 5 A, scale bar), the energy imparted to a single charge by the 100-mV driving force typical of biological membranes. Regardless of the permeation mechanism, it seems that the final site before escape from a pore must have an energy no more negative than such a "biological" magnitude.

The step used in the model more or less cuts the exit barrier into two halves. The physical mechanism that causes the profound increase in flux with just a twofold change in barrier height is analogous to stairsteps. Imagine a wall, a gravitational potential energy barrier, 2 m high. Flux of people over it is exceedingly low because only the most talented can leap 2 m. Now, place a 1-m step in front of it. Most of us can easily step up that meter, and then the next, so flux over the wall increases dramatically. The increase in flux is disproportional to the change in limiting barrier height because the number of people that can hurdle a gravitational barrier decreases nonlinearly with increasing height. Similarly, the number of atoms with sufficient kinetic energy to surpass a chemical potential energy barrier decreases exponentially with increasing height. Once it has reached the intermediate step, an atom faces the next barrier with no memory of the energy expended in reaching there because atoms thermally reequilibrate at the atomic vibration frequency (kT/h =  $5.8 \times$  $10^{12} \text{ s}^{-1}$ ). Therefore, steps of chemical potential energy speed reaction rates much as stairsteps of gravitational potential energy speed traffic over obstacles.

Although remote interactions between ions do not occur in the step model, multiple occupancy by  $Ca^{2+}$  is required for significant  $Ca^{2+}$  flux. The rate constant for exit of  $Ca^{2+}$  from one of the low affinity sites in Fig. 1 over the exit barrier to the outside is  $5 \times 10^6$  per second (=[kT/h]exp - [14]). If ions went directly from the high affinity site to the exterior, the rate constant would be 50 per second (=[kT/h]exp - [25.5]). As large as this difference is, it is insignificant if the pore is singly occupied. With just a single ion in the pore, the probability of occupying one of the low affinity sites is  $10^{-5}$  that of occupying the high affinity site ( $P_{low}/P_{high} =$ 

 $\exp - [G_{low} - G_{high}])$ . This exactly negates the advantage in rate constant since flux equals the product of the occupancy probability and the rate constant. Significant flux occurs only when there is high occupancy of the escape site by Ca<sup>2+</sup>. This occurs when extracellular [Ca<sup>2+</sup>] becomes significant compared with the site's dissociation constant (18 mM). Ca<sup>2+</sup> flux is entirely driven by Ca<sup>2+</sup> concentration and its effect on occupancy of the pore.

#### Plausibility

Ca<sup>2+</sup> channels clearly have only a single high affinity binding site within their pore (Yang et al., 1993; Kim et al., 1993; Ellinor et al., 1995). This selectivity site appears to be flanked by other binding sites because: (a) extracellular monovalent ions inhibit exit of Ca<sup>2+</sup> from the high affinity site to the outside and intracellular ions inhibit exit to the inside (Kuo and Hess, 1993b); (b) extracellular Na<sup>+</sup> diminishes inward Ca<sup>2+</sup> and Ba<sup>2+</sup> current, suggesting that the ions compete for a low affinity entry site that is on the outside of the selectivity site (Polo-Parada and Korn, 1997); (c) in the presence of extracellular Ca2+, monovalent ions are far more permeant from the inside than the outside (Yamashita et al., 1990), suggesting an internal entry site at which monovalents compete favorably with Ca<sup>2+</sup> because of the low intracellular Ca2+ concentration. These findings suggest that the Ca2+ channel is a multi-site pore that has a single high affinity site that is flanked by others of lower affinity. This image of the Ca<sup>2+</sup> channel pore has potential energy steps built into it. Any such step of potential energy must contribute to generating high Ca<sup>2+</sup> flux.

Although our calculations show that appropriately designed potential energy steps are sufficient to fit critical  $Ca^{2+}$  channel data, the steps need not be the only factor. Ion–ion interactions are physically reasonable because both electrostatic repulsion and competition for binding moieties are plausible when identical ions simultaneously occupy a pore. The ionic repulsion model and the step model both quantitatively fit the same  $Ca^{2+}$  channel data; at most, this demonstrates that these mechanisms make sense, not that they are real. The reality of  $Ca^{2+}$  channel permeation might involve a combination of the mechanisms that have been theoretically explored in isolation, or, perhaps, some mechanism that has been overlooked.

#### Steps Might Not Be Sites

The literal interpretation of an energy diagram is that each well corresponds to a discrete binding site that should be evident in the three-dimensional structure of the protein's pore. Surely, this must be the case with the high affinity site, which consists of a cage of four glutamates (Yang et al., 1993; Kim et al., 1993). However, physical interpretation of the low affinity sites is ambiguous. It may be that the Ca<sup>2+</sup> ion can experience steps of potential energy without explicit binding sites built into the protein. For example, ion channels appear generally to have wide entrance vestibules that may be on either side of a narrow, short pore (Miller, 1996). Weak ion binding within the vestibule might be able to provide the same favorable energy steps provided by discrete sites in a long pore. Indeed, placement of the model's low affinity sites at the very edge of the electric field, the location of the vestibules, gives qualitatively similar results as those shown. If one literally interprets the steps as sites in the vestibules, the model suggests that a vestibule serves as an energy "plateau" from which Ca<sup>2+</sup> can rapidly enter the pore over a low energy barrier. However, literal interpretation is dangerous; the low affinity sites are just a mathematical artifice for showing that steps of potential energy, no matter what their cause, can speed flux.

A different interpretation of the low affinity sites is suggested by a previous case in which stairsteps of potential energy were invoked in the ion transport literature. Eigen and Winkler (1971) and Chock et al. (1977) asked how entry of ions to binding sites on carriers could be diffusion limited when the ions had to undergo complete dehydration, which is normally an exceedingly slow process. The paradox is solved through stepwise replacement of water molecules by coordinating ligands on the carrier; in essence, the carrier catalyzes dehydration of the ion, step by step. This same phenomenon must occur with channels, where entry is generally diffusion limited (e.g., Lansman et al., 1986) despite the need for nearly complete dehydration (Hille, 1971).

Might stepwise rehydration of an exiting ion also occur? This seems probable, not just possible. A Ca<sup>2+</sup> ion coordinates to seven or eight electronegative oxygen ligands at a time (Falke et al., 1994). Outside the pore, water molecules provide this coordination. At the selectivity site, negative carboxyl oxygens on four glutamates contribute to coordination, thereby forming a stronger bond (Yang et al., 1993; Kim et al., 1993). Unless the Ca<sup>2+</sup> ion bursts from the glutamate cage immediately into water, there must be intermediate lower affinity states in which the glutamates are replaced by less negative moieties. One possibility would be stepwise replacement of the four carboxyls by water molecules, a mechanism consistent with the proposed flexibility of the glutamate residues (Yang et al., 1993; Ellinor et al., 1995). Any such event would provide a step of potential energy in the Ca<sup>2+</sup> ion's travel, even though it need not be a discrete binding site in the channel structure. In this manner, stepwise rehydration might provide a path for rapid exit from a pore just as stepwise dehydration surely provides the path for rapid entry. By showing that even a single energy step can be sufficient, our calculations suggest the feasibility of this simple mechanism of permeation.

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#### REFERENCES

- Almers, W., and E.W. McCleskey. 1984. Non-selective conductance in calcium channels of frog muscle: calcium selectivity in a singlefile pore. *J. Physiol. (Camb.).* 353:585–608.
- Almers, W., E.W. McCleskey, and P.T. Palade. 1984. A non-selective cation conductance in frog muscle membrane blocked by micromolar external calcium ions. J. Physiol. (Camb.). 353:565–583.
- Andersen, O.S., and J. Procopio. 1980. Ion movement through gramicidin A channels. Acta Physiol. Scand. 481(Suppl.):27–35.
- Armstrong, C.M., and J. Neyton. 1991. Ion permeation through calcium channels: a one-site model. *Ann. NY Acad. Sci.* 635:18–25.
- Begenisich, T. 1987. Molecular properties of ion permeation through sodium channels. Annu. Rev. Biophys. Biophys. Chem. 16: 247–263.
- Bezanilla, F., and C.M. Armstrong. 1972. Negative conductance caused by entry of sodium and cesium ions into the potassium channels of squid giant axons. J. Gen. Physiol. 60:588–608.
- Chock, P.B., F. Eggers, M. Eigen, and R. Winkler. 1977. Relaxation studies on complex formation of macrocyclic and open chain an-

tibiotics with monovalent cations. *Biophys. Chem.* 6:239–251.

- Eigen, M., and R. Winkler. 1971. Carriers and specificity in membranes. II. Characteristics of carriers. Alkali ion carriers: specificity, architecture, and mechanisms: an essay. *Neurosci. Res. Prog. Bull.* 9:330–338.
- Ellinor, P.T., J. Yang, W.A. Sather, J.-F. Zhang, and R.W. Tsien. 1995. Ca<sup>2+</sup> channel selectivity at a single locus for high-affinity Ca<sup>2+</sup> interactions. *Neuron.* 15:1121–1132.
- Falke, J.J., S.K. Drake, A.L. Hazard, and O.B. Peersen. 1994. Molecular tuning of ion binding to calcium signaling proteins. *Q. Rev. Biophys.* 27:219–290.
- Fenwick, E.M., A. Marty, and E. Neher. 1982. Sodium and calcium channels in bovine chromaffin cells. J. Physiol. (Camb.). 331:599– 635.
- Friel, D.D., and R.W. Tsien. 1989. Voltage-gated calcium channels: direct observation of the anomalous mole fraction effect at the single-channel level. *Proc. Natl. Acad. Sci. USA*. 86:5207–5211.
- Fukushima, Y., and S. Hagiwara. 1985. Currents carried by monova-

lent cations through calcium channels in mouse neoplastic B lymphocytes. J. Physiol. (Camb.). 358:255–284.

- Hess, P., and R.W. Tsien. 1984. Mechanism of ion permeation through calcium channels. *Nature*. 309:453–456.
- Hess, P., J.B. Lansman, and R.W. Tsien. 1986. Calcium channel selectivity for divalent and monovalent cations: voltage and concentration dependence of single channel current in guinea pig ventricular heart cells. *J. Gen. Physiol.* 88:293–319.
- Hille, B. 1971. The hydration of sodium ions crossing the nerve membrane. *Proc. Natl. Acad. Sci. USA*. 68:280–282.
- Hille, B. 1992. Ionic Channels in Excitable Membranes. 2nd ed. Sinauer Associates, Inc. Sunderland, MA. 607 pp.
- Hille, B., and W. Schwarz. 1978. Potassium channels as multi-ion single-file pores. J. Gen. Physiol. 72:409–442.
- Kim, M.-S., T. Mori, L.-X. Sun, K. Imoto, and Y. Mori. 1993. Structural determinants of ion selectivity in brain calcium channel. *FEBS Lett.* 318:145–148.
- Kiss, L., D. Immke, J. LoTurco, and S.J. Korn. 1998. The interaction of Na<sup>+</sup> and K<sup>+</sup> in voltage-gated potassium channels: evidence for cation binding sites of different affinity. *J. Gen. Physiol.* 111:195– 206.
- Kostyuk, P.G., S.L. Mironov, and Ya.M. Shuba. 1983. Two ion-selecting filters in the calcium channel of the somatic membrane of mollusc neurons. *J. Membr. Biol.* 76:83–93.
- Kuo, C.-C., and P. Hess. 1993*a*. Ion permeation through the L-type Ca<sup>2+</sup> channel in rat phaeochromocytoma cells: two sets of ion binding sites in the pore. *J. Physiol. (Camb.)*. 466:629–655.
- Kuo, C.-C., and P. Hess. 1993*b*. Characterization of the high-affinity Ca<sup>2+</sup> binding sites in the L-type Ca<sup>2+</sup> channel pore in rat phaeochromocytoma cells. *J. Physiol. (Camb.)*. 466:657–682.
- Lansman, J.B., P. Hess, and R.W. Tsien. 1986. Blockade of current through single Ca channels by Cd<sup>2+</sup>, Mg<sup>2+</sup>, and Ca<sup>2+</sup>: voltage and concentration dependence of Ca entry into the pore. *J. Gen. Physiol.* 88:321–347.

- Lee, K.S., and R.W. Tsien. 1984. High selectivity of calcium channels in single dialysed heart cells of the guinea-pig. J. Physiol. (Camb.). 354:253–272.
- Lux, H.D., E. Carbone, and H. Zucker. 1990. Na<sup>+</sup> currents through low-voltage-activated Ca<sup>2+</sup> channels of chick sensory neurons: block by external Ca<sup>2+</sup> and Mg<sup>2+</sup>. J. Physiol. (Camb.). 430:159–188.
- Miller, C. 1996. The long pore gets molecular. J. Gen. Physiol. 107: 445–447.
- Mironov, S.L. 1992. Conformational model for ion permeation in membrane channels: a comparison with multi-ion models and applications to calcium channel permeability. *Biophys. J.* 63:485– 496.
- Polo-Parada, L., and S. Korn. 1997. Block of N-type calcium channels in chick sensory neurons by external sodium. J. Gen. Physiol. 109:693–702.
- Rosenberg, R.L., and X.-H. Chen. 1991. Characterization and localization of two ion-binding sites within the pore of cardicac L-type calcium channels. *J. Gen. Physiol.* 97:1207–1225.
- Shockley, W., J.H. Hollomon, R. Maurer, and F. Seitz. 1952. Imperfections in nearly perfect crystals. John Wiley & Sons Inc., New York. 490 pp.
- Woodhull, A.M. 1973. Ionic blockage of sodium channels in nerve. J. Gen. Physiol. 61:687–708.
- Yamashita, N., S. Ciani, and S. Hagiwara. 1990. Effects of internal Na<sup>+</sup> on the Ca channel outward current in mouse neoplastic B lymphocytes. *J. Gen. Physiol.* 96:559–579.
- Yang, J., P.T. Ellinor, W.A. Sather, J.-F. Zhang, and R.W. Tsien. 1993. Molecular determinants of Ca<sup>2+</sup> channel selectivity and ion permeation in L-type Ca<sup>2+</sup> channels. *Nature*. 366:158–161.
- Yue, D.T., and E. Marban. 1990. Permeation in the dihydropyridine-sensitive calcium channel: multi-ion occupancy but no anomalous mole-fraction effect between Ba<sup>2+</sup> and Ca<sup>2+</sup>. *J. Gen. Physiol.* 95:911–939.