

RESEARCH NOTE

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Emergence of G12 and G9 rotavirus genotypes in the Central African Republic, January 2014 to February 2016

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Abstract

Objectives: Rotavirus gastroenteritis is a major cause of death among children under 5 years globally. A rotavirus gastroenteritis surveillance program started in October 2011 in the Central African Republic (CAR) with the Surveillance Epidémiologique en Afrique Centrale (SURVAC) project. We present here genotyping results showing the emergence of G9 and G12 genotypes in Central African Republic.

Results: Among 222 children hospitalized with acute gastroenteritis who had a stool sample collected at the sentinel site, Complexe Pédiatrique de Bangui (CPB), Bangui, Central African Republic, 100 (45%) were positive for rotavirus between January 2014 and February 2016. During this period the most common rotavirus strains were G1P[8] (37%), G12P[6] (27%) and G9P[8] (18%).

Keywords: Rotavirus, Genotype, Emergence, CAR

Introduction

Rotavirus diarrhea is widespread with approximately 215,000 children less than 5 years of age dying each year due to severe dehydration caused by rotavirus [1]. Rotaviruses belong (RV) to the family Reoviridae, and the rotavirus genome consists of 11 double-stranded RNA gene segments that encode six structural (VP) and six non-structural proteins (NSP). Based on the two genes that encode the outer capsid proteins, VP4 (P-type) and VP7 (G-type), a widely used binary classification system was established for RV-A [2]. To date, at least 35 G and 50 P genotypes have been recognized in both mammalian and avian species. So far, 6 genotypes (G1P[8], G2P[4], G3P[8], G4P[8], G9P[8], and G12P[8]) are currently the most important genotypes in humans worldwide and are associated with 80–90% of the RVA associated disease

burden [3–5]. In Africa and sub-Saharan Africa in particular there is a greater genetic diversity and emergence of new and unusual strains [6]. In addition to the 6 common strains mentioned above, rotavirus strains with unusual combination, such as G8P[6], G3P[4], G8P[4], G2P[6] as well as high levels of mixed infections and strains that cannot be assigned any specific genotype have been reported in sub-Saharan Africa [7].

In 2009, WHO recommended the introduction of rotavirus vaccine in all countries [8]. In the Central African Republic, the sentinel surveillance of rotavirus gastroenteritis was established in 2011 by the Ministry of Health, with the support of the Surveillance Epidémiologique en Afrique Centrale (SURVAC) Project [9]. The main objective of the project was to assess the burden of rotavirus gastroenteritis and identify rotavirus strains circulating in CAR before the introduction of a rotavirus vaccine which was initially scheduled for 2017. The SURVAC project completed in 2014 but the surveillance continues with support from the WHO, the Institut Pasteur de Bangui (IPB) and The Centers for Diseases Control and

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prevention (CDC). The surveillance results between 2011 and 2013 have been published [10]. We present here the rotavirus surveillance results for the period January 2014 to February 2016.

Main text

Methods

Stool samples were collected from children less than 5 years of age who met the WHO rotavirus gastroenteritis case definition at the sentinel site, the Complex Pédiatrique de Bangui (CPB), in the capital of CAR [11]. CPB is the only pediatric hospital of the country where, all severe diarrhea cases originated from Bangui and surroundings are referred.

At the sentinel site laboratory, the samples were first screened for group A rotavirus antigen by enzyme immunoassay (EIA) using the ProSpecT™ Rotavirus Microplate Assay (Oxoid, Ltd., Basingstoke, Hampshire, UK). Aliquots of all the samples were then stored at - 20 °C before transport to the Institut Pasteur de Bangui where results were confirmed by EIA using the same kit and genotyping assays performed as previously described [12]. Briefly, RNA extracts were subjected to multiplex semi-nested reverse transcription polymerase chain reaction (RT-PCR). Two genes; VP7 (896 bp) and VP4 (876 bp) were reverse-transcribed and amplified with primer pairs 9Con1-L/VP7-R and Con3/Con2, respectively [13, 14]. Reverse transcription of double strand RNA (dsRNA) was carried out with the OneStep RT-PCR Kit (Qiagen, Inc., Valencia, CA USA). After a 5 min denaturation at 97 °C, the RNA was mixed with kit reagents and incubated at 42 °C for 30 min to obtain complementary DNA (cDNA), immediately followed by the PCR reaction (30 cycles: 94 °C for 30 s; 42 °C for 30 s, 72 °C for 45 s; one cycle at 72 °C for 7 min). These first round RT-PCR products then were used in a semi-nested PCR (30 cycles, 94 °C for 45 s, 42 °C for 30 s 72 °C for 1 min; and 1 cycle at 72 °C for 7 min) to identify G types (G1, G2, G3, G4, G9 and G12) and P types (P[4], P[6], P[8]) [13, 14]. All PCR products were analyzed by electrophoresis in 2% agarose gels containing Gel Red (Biotium) and visualized under UV illumination. Sample of RNA extracts were sent to CDC for genotyping quality control and sequencing confirmation.

Results and discussion

Between January 2014 and February 2016, 222 stools samples were collected from children, with an 8 months mean age (range 1–55 months) and analyzed for detection of rotavirus antigen. The annual sample distribution was: 115 in 2014, 62 in 2015 and 45 from January to February 2016. Rotavirus was detected in 45% (100/222) of stool specimens by EIA and this prevalence is in

agreement with the estimated 40% prevalence of rotavirus infection in other African countries [15] (Table 1). VP7 genotyping showed that G1 was the predominant strain 37% (37/100) followed by G12 and G9 with 27% (27/100) and 20%, (20/100), respectively, both of which have recently emerged in Africa [16, 17]. The other genotypes identified were G2 and G3 with 11% (11/100) and 3% (3/100), respectively. In a previous published study [10], G2 was found to be predominant in 66% (105/160) samples followed by G1, 28% (45/160) while G3 was not identified at all. In the same study the presence of G9 and G12 was for the first time reported at 3% (5/160) each. VP4 genotyping revealed that P[8] was predominant with 55% (55/100), followed by P[6] with 43% (43/100), compared to the 2011–2013 period when P[6] was predominant with 52% (83/106) with P[8] following at 35% (56/106). Two (2%) P[4] were identified, one of which was not confirmed by CDC. The annual distribution of G and P genotypes is shown in Fig. 1. The VP7/VP4 genotype combinations are shown in Fig. 2. G1P[8] was the predominant strain in 2014, 2015 and 2016 with 37.2% (16/43), 45.4% (10/22) and 29.4% (10/34), respectively. These data are consistent with our 2008 and 2011–2013 data and confirm data from other African countries [10, 18–20]. It was followed by G12P[6] with 25.5% (11/43), 31.8% (7/22) and 26.4% (9/34) in 2014, 2015 and 2016, respectively. G9P[8] was detected at 3% in 2013 [10], continued to circulate at 9.3% (4/43) in 2014, 13.6% (3/22) in 2015, and increased to 29.4% (10/34) in 2016. Other VP7/VP4 genotype combinations identified were: (i) G2P[6] which circulated in 2014 and 2015 at 23.2% (10/43) and 4.5% (1/22) respectively and 47% (75/106) during the 2011–2013 period; (ii) G1P[6] and G3P[6], which circulated in 2014 at 2.3% (1/43) each; (iii) G2P[4] which circulated in 2015 at 4.5% (1/22) and at 13% (21/106) from October 2011 to September 2013; and (iv) G9P[6] and G3P[8] with 8.8% (3/34) and 5.8% (2/34), respectively. Quality control was performed at the CDC on 102

Table 1 Annual breakdown of stool samples tested for the presence of Rotavirus antigens by EIA (ProSpecT™) at Institut Pasteur de Bangui, Central African Republic (CAR), January 2014 to February 2016

Years	Rota positive (%)	Rota negative (%)	Total
2014 ^a	41 (35.6)	74 (64.4)	115
2015 ^a	22 (35.4)	40 (64.6)	62
2016 ^b	37 (82.2)	8 (17.8)	45
Total	100 (45.0)	111 (55.0)	222

^a CAR experienced military and political crises, especially in Bangui, where the civilian population remained cloistered at home for days to weeks

^b January to February, high season for rotavirus transmission

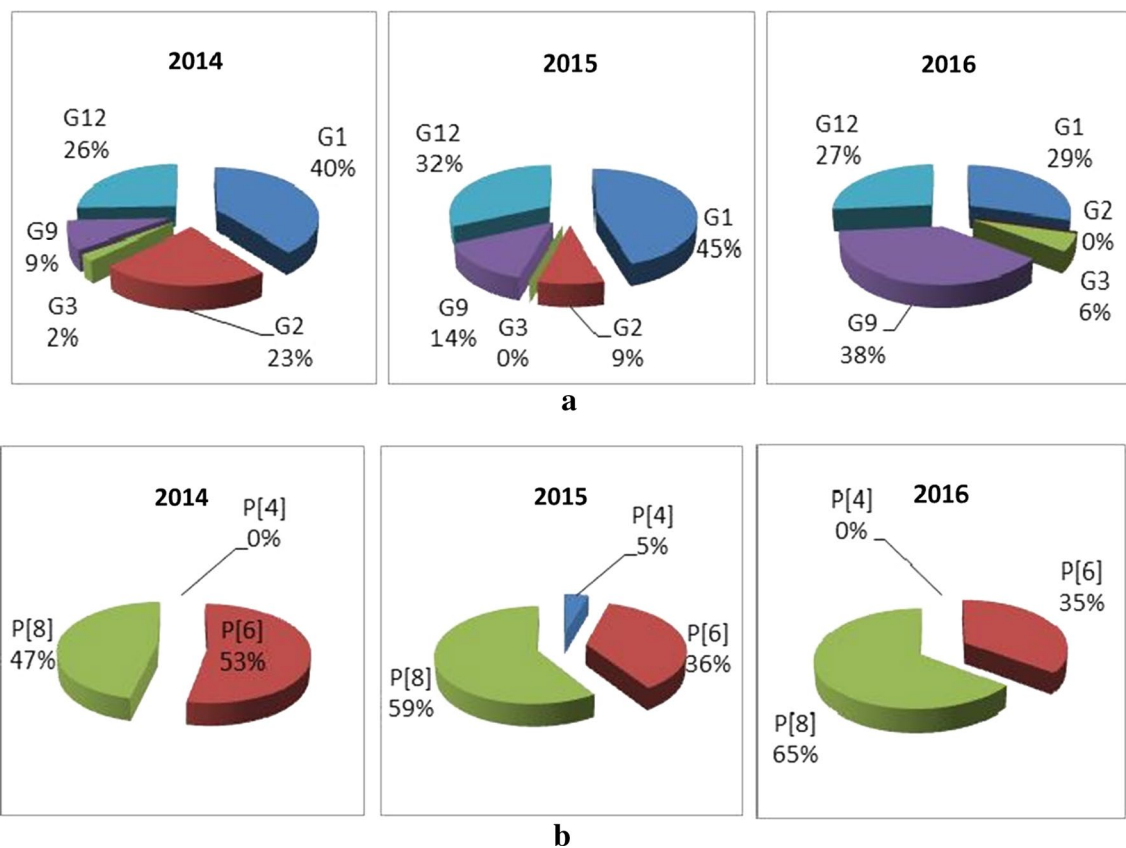


Fig. 1 Annual G and P genotype distribution, January 2014–February 2016. **a** VP7(G) genotypes; **b** VP4 (P) genotypes. The total number of samples genotyped by year is 41, 22 and 37 in 2014, 2015 and 2016 (January to February only), respectively

samples, including two EIA negatives. The correlation of genotyping results between the two laboratories was 57%. Based on the low correlation between the genotyping results from CAR compared to results from CDC, the genotyping results reported in this study were CDC results only. Due to the low degree of correlation, as a corrective action, a new set of primers from the CDC was sent to the CAR laboratory to help improve genotyping results. One of the samples bearing genotype P[4] was confirmed at the CDC, however, due to a sample leak during transportation, the tube containing the other sample arrived empty at the CDC and therefore could not be confirmed.

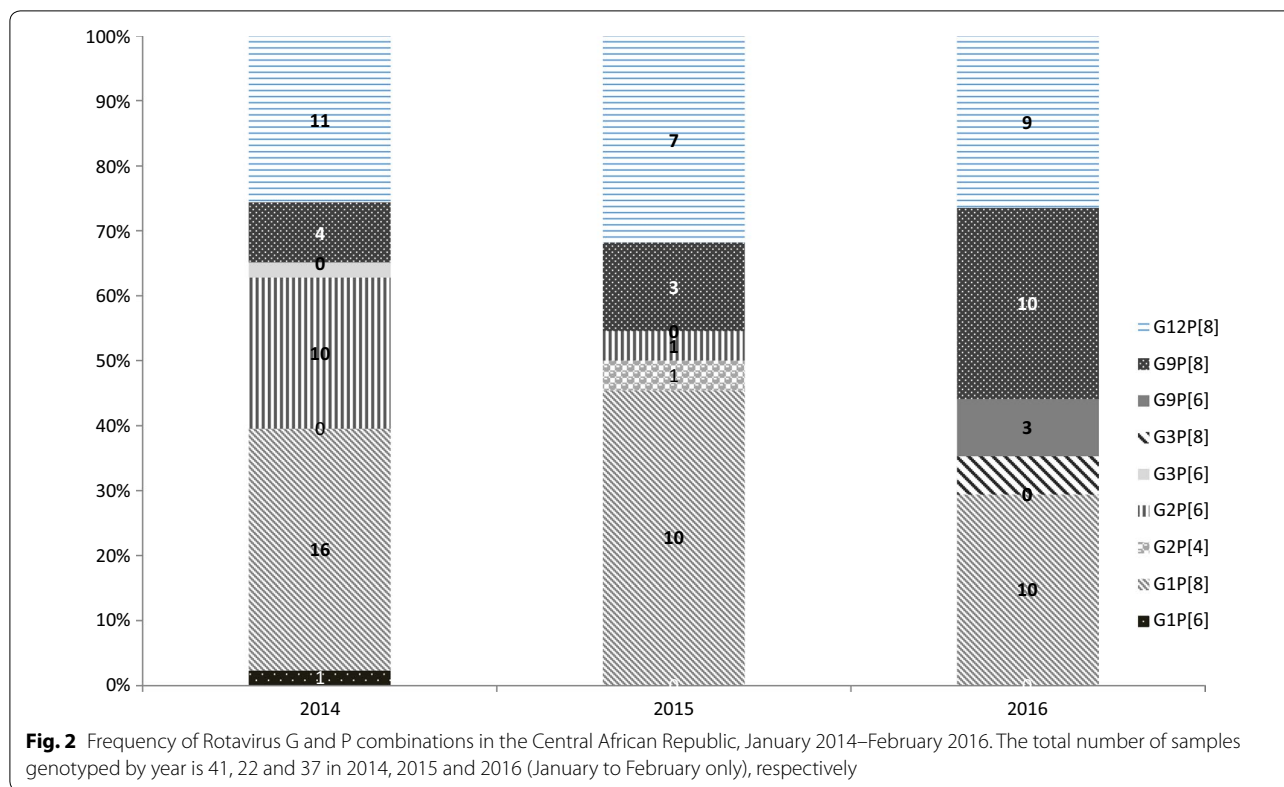
Conclusion

Genotyping data showed that genotypes G1P[8], G12P[6] and G9P[8] were the most common rotavirus strains circulating in Bangui during the study period. The detection of G12 and G9 strains contributes to the body of evidence that these genotypes are becoming increasingly dominant on the African continent and worldwide [15, 16, 21]. Currently, rotavirus vaccination

has not been introduced into CAR; the introduction of Rotarix vaccine has been rescheduled for 2018. Rotarix vaccine has been shown to provide a good level of protection against severe rotavirus gastroenteritis in multiple African countries [22]. This vaccine is derived from a genotype G1P[8] strain but induces immunity against both homotypic and heterotypic rotavirus strains [22]. We therefore are expecting a drop in the number of gastroenteritis cases due to rotavirus infection after the introduction of the Rotarix vaccine [23]. Continued rotavirus surveillance is necessary to monitor the impact of the vaccine on the overall number of hospitalizations for severe gastroenteritis and for the emergence of new genotypes against which the available rotavirus vaccines may be less effective [24].

Limitations

The study limitations are linked to the fact that CAR has only one sentinel site and did not reach the WHO recommendation to collect and test 250 samples per year. This is mainly due to the military and political problems that CAR has been experiencing since 2013.



Authors' contributions

All authors read and approved the manuscript before submission. UAEM, VBM and JF performed the EIA and genotyping, contributed in data analysis and writing of the article; GIV coordinated the project at IPB and was a major contributor in writing the manuscript; DWK, MDE and MDB performed the RT-PCR quality control and sequencing and were major contributors in writing the manuscript; JCG, JMM, CM and TAK coordinated the rotavirus surveillance and contributed to writing the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

Consent for publication

Not applicable.

Ethics approval and consent to participate

The Rotavirus Gastroenteritis surveillance program is a Public Health program ethically approved by the expert committee for Integrated diseases surveillance of the Ministry of Health in CAR (Arrêté No. 0277/MSPP/CAB/DGSP/DMPP/SMEE du 05 Août 2002) it is a routine surveillance activity and genotyping is part it. The ethical committee in CAR is "Comité Scientifique Chargé De La Validation Des Protocoles D'étude Et Des Résultats" (Scientific comite in charge of validation of studies's protocols and results) of the University of Bangui [25]. It stated that for all MoH surveillance programs supported by partners a consent to participate is not required.

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References

1. Tate JE, Burton AH, Boschi-Pinto C, Parashar UD. Global, regional, and national estimates of rotavirus mortality in children < 5 years of age, 2000–2013. *Clin Infect Dis*. 2016;62(Suppl 2):S96–105.
2. Estes MK, Greenberg HB, Knipe DM, Howley PM, Cohen JI, Griffin DE, Lamb RA, Martin MA, Racaniello VR, Roizman B. *Rotaviruses*. In: Knipe DM, Howley PM, Cohen JI, Griffin DE, Lamb RA, Martin MA, Racaniello VR, Roizman B, editors. *Fields virology*. 6th ed. Philadelphia: Walters Kluwer Health/Lippincott Williams & Wilkins; 2013. p. 1347–401.
3. Banyai K, et al. Systematic review of regional and temporal trends in global rotavirus strain diversity in the pre rotavirus vaccine era: insights for understanding the impact of rotavirus vaccination programs. *Vaccine*. 2012;30(Suppl 1):A122–30.
4. Gentsch JR, Laird AR, Bielfelt B, Griffin DD, Banyai K, Ramachandran M, Jain V, Cunliffe NA, Nakagomi O, Kirkwood CD, Fischer TK, Parashar UD, Bressee JS, Jiang B, Glass RI. Serotype diversity and reassortment between human and animal rotavirus strains: implications for rotavirus vaccine programs. *J Infect Dis*. 2005;192(Supplement):S146–59.
5. Santos N, Hoshino Y. Global distribution of rotavirus serotypes/genotypes and its implication for the development and implementation of an effective rotavirus vaccine. *Rev Med Virol*. 2005;15(1):29–56.
6. Kl K. The burden and etiology of diarrheal illness in developing countries. *Pediatr Clin North Am*. 2017;64(4):799–814.
7. Todd S, et al. Rotavirus strain types circulating in Africa: review of studies published during 1997–2006. *J Infect Dis*. 2010;202(Suppl):S34–42.
8. World Health Organization. Meeting of the immunization strategic advisory group of experts, April 2009—conclusions and recommendations. *Wkly Epidemiol Rec*. 2009;84:220–36.
9. Waku-Kouomou D, et al. Strengthening laboratory capacity through the surveillance of rotavirus gastroenteritis in Central Africa: the Surveillance Epidemiologique en Afrique Centrale (SURVAC) Project. *Trop Med Int Health*. 2016;21(1):122–30.
10. Banga-Mingo V, et al. Molecular surveillance of rotavirus infection in Bangui, Central African Republic, October 2011–September 2013. *Infect Genet Evol*. 2014;28:476–9.
11. WHO. Manual of rotavirus detection and characterization methods, ed. WHO/IVE. 2008.
12. Pukuta ES, Esona MD, Nkongolo A, Seheri M, Makasi M, Nyembwe M, Mondonge V, Dahl BA, Mphahlele MJ, Cavallaro K, Gentsch J. Molecular surveillance of rotavirus infection in the Democratic Republic of the Congo August 2009 to June 2012. *Pediatr Infect Dis J*. 2014;33(4):355–9.
13. Gentsch JR, et al. Identification of group A rotavirus gene 4 types by polymerase chain reaction. *J Clin Microbiol*. 1992;30(6):1365–73.
14. Das BK, et al. Characterization of rotavirus strains from newborns in New Delhi, India. *J Clin Microbiol*. 1994;32(7):1820–2.
15. Mwenda JM, et al. Burden and epidemiology of rotavirus diarrhea in selected African countries: preliminary results from the African Rotavirus Surveillance Network. *J Infect Dis*. 2010;202(Suppl):S5–11.
16. Page NA, et al. The detection and molecular characterization of human G12 genotypes in South Africa. *J Med Virol*. 2009;81(1):106–13.
17. Steele AD, Ivanoff B. Rotavirus strains circulating in Africa during 1996–1999: emergence of G9 strains and P[6] strains. *Vaccine*. 2003;21(5–6):361–7.
18. Gouandijka-Vasilache I, et al. Rotavirus epidemiology in Bangui, Central African Republic, 2008. *Emerg Infect Dis*. 2014;20(7):1254–5.
19. Boula A, et al. Molecular surveillance of rotavirus strains circulating in Yaounde, Cameroon, September 2007–December 2012. *Infect Genet Evol*. 2014;28:470–5.
20. Seheri M, et al. Update of rotavirus strains circulating in Africa from 2007 through 2011. *Pediatr Infect Dis J*. 2014;33(Suppl 1):S76–84.
21. Rahman M, et al. Evolutionary history and global spread of the emerging g12 human rotaviruses. *J Virol*. 2007;81(5):2382–90.
22. O’Ryan M, Giaquinto C, Benninghoff B. Human rotavirus vaccine (Rotarix): focus on effectiveness and impact 6 years after first introduction in Africa. *Expert Rev Vaccines*. 2015;14(8):1099–112.
23. Tsolenyanu E, et al. Early evidence of impact of monovalent rotavirus vaccine in Togo. *Clin Infect Dis*. 2016;62(Suppl 2):S196–9.
24. Parashar UD, et al. Health impact of rotavirus vaccination in developing countries: progress and way forward. *Clin Infect Dis*. 2016;62(Suppl 2):S91–5.
25. Ministère de l’Hygiène Publique, d.I.Pe.d.I.S.P. Guide Technique de Surveillance Intégrée de la Maladie et la Riposte en République Centrafricaine (SMIR). 2011.

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