

International Journal of Medical Sciences

2020; 17(17): 2751-2762. doi: 10.7150/ijms.46302

Research Paper

Terlipressin relieves intestinal and renal injuries induced by acute mesenteric ischemia via PI3K/Akt pathway

Zi-Meng Liu^{1*}, Han-Jin Lai^{1*}, Xiang-Dong Guan^{1*}, Shi-Hong Wen², Jian-Tong Shen², Yao Nie¹, Ning Liu¹, Xu-Yu Zhang^{2⊠}

1. Department of Critical Care Medicine, The First Affiliated Hospital, Sun Yat-sen University, No.58, Zhongshan 2nd Road, Guangzhou 510089, China.

2. Department of Anesthesiology, The First Affiliated Hospital, Sun Yat-sen University, No.58, Zhongshan 2nd Road, Guangzhou 510089, China.

*These authors contributed equally to this study.

🖂 Corresponding author: Xu-Yu Zhang, M.D., Ph.D. E-mail: zhangxuy@mail.sysu.edu.cn; Tel: +8620-87755766-8273; Fax: +8620-87755766-8271; Department of Anesthesiology, The First Affiliated Hospital, Sun Yat-sen University, No. 58, Zhongshan 2nd Road, Guangzhou 510089, China.

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2020.03.24; Accepted: 2020.09.16; Published: 2020.09.28

Abstract

Background: To date, the effect of vasopressin on organ damages after acute mesenteric ischemia (MI) remains poorly understood.

Aims: To investigate the effect of terlipressin, a selective vasopressin V1 receptor agonist, versus norepinephrine on the intestinal and renal injuries after acute MI, and to explore the underlying mechanism of terlipressin.

Methods: Acute MI model was produced by clamping the superior mesenteric artery for 1 hour. Immediately after unclamping, terlipressin or norepinephrine was intravenously administered for 2 hours. Meanwhile, *in vitro*, RAW264.7 cells were treated with lipopolysaccharide or lipopolysaccharide+terlipressin. In addition, wortmannin was used to determine the role of phosphoinositide 3-kinase (PI3K)/ protein kinase B (Akt) pathway in the potential impacts of terlipressin.

Results: MI led to severe hypotension, caused notable intestinal and renal impairments and resulted in high mortality, which were markedly improved by terlipressin or norepinephrine. Terlipressin increased mean arterial pressure, decreased intestinal epithelial cell apoptosis, inhibited the generation of MI macrophage in intestinal and renal tissues, and hindered the release of inflammatory cytokines after MI. Moreover, in cultured macrophages, terlipressin reduced the mRNA level of specific MI markers and the release of inflammatory cytokines caused by lipopolysaccharide challenge. Wortmannin decreased the expression of PI3K and Akt induced by terlipressin in cells and in tissues, and abolished the above protective effects conferred by terlipressin.

Conclusions: Terlipressin or norepinephrine could effectively improve organ damages and mortality after acute MI. Terlipressin elevates blood pressure and inhibits intestinal epithelial apoptosis and macrophage MI polarization via the PI3K/Akt pathway.

Key words: vasopressor; V1 receptor; ischemia reperfusion injury; intestine; apoptosis; macrophage polarization

Introduction

Acute mesenteric ischemia (MI), which can be subdivided into acute mesenteric arterial embolism and thrombosis, mesenteric venous thrombosis and nonocclusive mesenteric ischemia, according to the ischemic mechanism, is a life-threatening condition caused by impaired blood perfusion to the gut [1, 2]. Acute MI and succeeding reperfusion may lead to vasodilatory hypotension, exacerbation of bacterial translocation and inflammatory responses, distant organ impairment and eventually multiple organ dysfunction [3, 4]. As a consequence, acute MI is associated with high mortality for critically ill patients [1, 5]. A retrospective study reported that 24.0% of decedents caused by sepsis in ICU were diagnosed with validated MI [6]. Recently, another analysis showed that, in 214 ICU patients, acute MI was a secondary diagnosis in 58% of patients and the 30-day mortality rate was 71% [5]. To maintain hemodynamic stability, vasopressors are commonly prescribed in the ICU. However, the use of some vasopressors may further decrease mesenteric flow and then aggravate the organ damages caused by MI [5, 7]. Therefore, it is of clinical relevance to investigate the impacts of specific vasopressor on the injury of intestine and remote organs after the occurrence of acute MI.

In our recently published multicenter clinical study, we found that severe diarrhea was more common in the septic shock patients receiving terlipressin, a highly selective vasopressin V1 receptor than in those receiving agonist, traditional norepinephrine, although the suspected MI rates in two groups were similar [8]. Consistent with our study, several case reports have demonstrated that V1 receptor agonists could cause severe bowel ischemic complications in the clinical settings [9-11]. However, most of the experimental studies did not demonstrate any detrimental effect of vasopressin V1 agonist on splanchnic macro- and microcirculation [12]. We previously found that terlipressin could protect intestinal epithelial cells against oxygen-glucose deprivation attack in vitro [13]. Qiu et al. have demonstrated that terlipressin could promote intestinal microvascular flow in rats with endotoxic shock [14]. So far, the definite effect of terlipressin on intestinal and other splanchnic injuries following acute MI insult has not been clarified. It is rather significant to investigate whether terlipressin increases MI-induced organ damages when compared with traditional vasopressor.

Therefore, we aimed to determine the effects of terlipressin on intestinal and renal injury with an acute MI model and to explore its underlying mechanisms. Norepinephrine was set as the control vasopressor.

Materials and Methods

Animal and cell line

The current animal experiments were approved by the Institutional Review Board of our university [2015A-059; President: X.Q. Yu] and was performed in accordance with National Institutes of Health guidelines for the experimental animal. Adult specific pathogen free (SPF) male Sprague-Dawley rats (300-350 g) were purchased and acclimated for 1 week before experiment entry.

The mouse RAW264.7 macrophage cell lines (ATCC, MD, USA) were maintained in Dulbecco's modified Eagle's medium containing fetal bovine serum, streptomycin and penicillin, and incubated at $37 \,^{\circ}$ C in a humidified 5% CO₂ incubator.

Experimental Protocol

The present in vivo and in vitro experimental

protocol was determined according to previous literatures and our pilot results.

The rats were anesthetized with intraperitoneal injection of 30 mg kg-1 pentobarbital and were mechanically ventilated with a standard ventilation protocol. Then, a catheter was inserted into the carotid artery and connected to a digital monitoring system for measuring hemodynamics, and the femoral vein was cannulated for drug infusion, as previously described [15]. After midline laparotomy, the superior mesenteric artery (SMA) was identified. The rats were randomly allocated into 6 groups. Sham group (Sham): the animals underwent laparotomy without occlusion of the SMA, and received 0.5 mL normal saline (NS) injection followed by 0.75 mL h⁻¹ continuous intravenous infusion for 2 hours. Acute MI group (MI): the SMA was clamped with a microaneurysm clip for 1 hour, and the animals received 0.5 mL normal saline (NS) immediately after unclamping of SMA followed by 0.75 mL h⁻¹ for 2 hours. Terlipressin group (TP): Terlipressin (Hybio Pharmaceutical Co., Shenzhen, China) was dissolved in NS to 8 µg mL-1, and 0.5 mL terlipressin solution was given immediately after ischemia followed by 0.75 mL h⁻¹ for 2 hours. Norepinephrine group (NE): Norepinephrine was dissolved in NS to 100 µg mL⁻¹, and 0.5 mL norepinephrine solution was given immediately after ischemia followed by 0.75 mL h-1 for 2 hours. Wortmannin group (WT): 1.5 mg kg-1 3-kinase (PI3K) inhibitor phosphoinositide wortmannin (MedChem Express, Shanghai, China) dissolved in 10% dimethyl sulfoxide (DMSO) 100 µL was administered intraperitoneally after unclamping, and then NS was administered. Telipressin+ Wortmannin group (T+W): isometric wortmannin and terlipressin were administered after unclamping according to the above-mentioned regimen. At 2nd hour after unclamping, the animals were euthanized by overdose of pentobarbital, and then the biological samples were harvested. No equivalent dose of terlipressin compared to norepinephrine has been reported. Thus, based on our preliminary hemodynamic data, the current dosing regimens of terlipressin and norepinephrine were designed to keep MAP within 15% of baseline level after ischemia. And each rat received equivalent volume of infusing solution (2 mL) during experimental period.

Confluent RAW264.7 cells were randomly assigned to 4 groups. Control group (Control): the cells received 0.1% DMSO and were incubated for 24 hours. Lipopolysaccharide group (LPS): the cells received 100 ng mL⁻¹ LPS (E. coli O111:B4, Sigma Aldrich, MO, USA) and were incubated for 24 hours. Terlipressin group (Terlipressin): 50 nM terlipressin was administered to the medium immediately after LPS treatment, and then the cells were incubated for 24 hours. Telipressin+Wortmannin group (Telipressin+Wortmannin): 5 μ M wortmannin was given at 1 hour before terlipressin treatment in the presence of LPS for 24 hours. After 24 hours incubation, the samples in each group were collected and detected.

Histological analysis

Tissues were collected from a 0.5 cm ileum fragment (2 cm next to ileocecal valve) and upper pole of the left kidney. The segments were fixed in 4% paraformaldehyde and embedded in paraffin. The fixed tissues were sectioned and stained with hematoxylin-eosin. The histological damage was assessed by two independent and blinded pathologists using respective scoring system based on similar literatures (Table S1).

Enzyme linked immunosorbent assay (ELISA) and biochemical analysis

Rat's tumor necrosis factor-a (TNF-a), interleukin (IL)-1 β and IL-6 concentrations in rat's serum, the supernatants of tissue homogenates and culture medium were determined with commercial ELISA kits (R&D systems, MN, USA and Mlbio, 8-isoprostane, Shanghai, China). an accurate biomarker of lipid peroxidation, in rat's serum was detected with ELISA kit (ab175819, Abcam, Shanghai, China). The results were expressed as pg mL⁻¹.

Rat's serum blood urea nitrogen (BUN) and creatinine levels were detected with an autoanalyzer (Hitachi, Tokyo, Japan). Diamine oxidase (DAO), an indicator of the integrity and functional mass of the intestine [16], in intestinal mucosal tissues was detected by a chemical assay kit (Jiancheng Bioengineering Institute, Nanjing, China). Results were expressed as U L⁻¹.

Western blot analysis

The proteins in the RAW264.7 cells, intestinal epithelia and kidney were detected by western blot. Briefly, the proteins were extracted from cells and tissues, and subjected to SDS-PAGE and electrophoretically transferred to nitrocellulose membranes. Membranes were blocked and incubated overnight with anti-PI3K antibody (4257, Cell Signaling Technology, MA, USA), anti-phospho-protein kinase B (p-Akt) antibody (13038, Cell Signaling Technology) or anti-cleaved caspase-3 antibody (9664, Cell Signaling Technology). After being washed three times, the membrane was incubated with a secondary antibody (7074, Cell Signaling Technology). Antibody dilutions used were according to user manual. Finally, the bands were imaged and quantified by Image J software (Ver 1.51,

National Institutes of Health, MA, USA) and normalized with the glyceraldehyde-3-phosphate dehydrogenase (GAPDH).

Quantitative polymerase chain reaction (qPCR)

The mRNA expression in RAW264.7 cells was detected through qPCR. In brief, total RNA was isolated by using TRIzol reagent (Invitrogen, CA, USA). Based on a similar study [17], quantitative PCR was performed by a C1000 Touch Thermal cycler (Bio-Rad Laboratories Inc, CA, USA) with SYBR Green Supermix to measure inducible nitric oxide synthase (iNOS), IL-1 β , Arg1 and FIZZ1 mRNA levels. The relative expressions of mRNA transcripts were normalized by the internal control GAPDH.

Immunohistochemistry and terminal deoxynucleotidyl transferase dUTP nick end labeling staining

Immunohistochemical examination was conducted to investigate the change of macrophages polarization [18]. Briefly, the slices of intestine and kidney sections were incubated with anti-iNOS antibody (1:400, ab15323, Abcam, Shanghai, China) and CD163 antibody (1:400, ab182422, Abcam), followed by a secondary HRP-labeled antibody (1:200, ab205718, Abcam) for 1 hour at room temperature. Images were taken by using a microscope (BX51, Olympus Corporation, Tokyo, Japan) at а magnification of 400×. Finally, the number of positive cell (brown yellow) was counted by using Image-Pro Plus software. Quantification of cells expressing the specified marker was performed by calculating positive cells in five randomly chosen 400× fields of each sample.

The severity of apoptosis in the small intestine and kidney was respectively detected by terminal deoxynucleotidyl transferase biotin-dUTP nick end-labeling assay (TUNEL). The number of apoptotic cells was counted using Image-Pro Plus software (ver. 6.0, Media Cybernetics, MD, USA) and the apoptotic index was calculated accordingly (the number of apoptotic cell nuclei/the number of total cell nuclei ×100).

Survival analysis

Briefly, the animals were treated with the above interventions and then transferred to the individual cage. Each rat was monitored via video recording for 24 hours [15].

Statistical analysis

SPSS 18.0 software (SPSS Inc, IL, USA) was used for the data analysis. Hemodynamic data of rats was expressed as the mean ± standard error of the mean (SEM) and was analyzed by two-way ANOVA with repeated measures followed by Bonferroni posttest. Survival time from the beginning of unclamping were expressed as median (range), and results were compared by Kaplan Meier log-rank test. The survival rate was analyzed by Fisher exact test. The rest of the data such as biochemistry, histologic, western blot analysis and qPCR were expressed as mean \pm SEM and analyzed by one-way ANOVA followed by Tukey posttest for multiple comparisons. *P*<0.05 was considered statistically significant.

Results

Terlipressin or norepinephrine improved hemodynamics after acute MI

As shown in Fig. 1, acute MI led to significantly decreased MAP (all P<0.05, MI vs. Sham; Fig. 1A) and slightly increased heart rate (all P>0.05; Fig.1B) throughout the whole reperfusion phase. The current dosage of norepinephrine or terlipressin effectively elevated MAP (all P<0.05, vs. MI; Fig. 1A), and there was no significant difference in MAP between TP group and NE group after MI (all P>0.05).

Terlipressin or norepinephrine alleviated intestinal and renal injuries after acute MI

As shown in Figure 2A, the Chiu's score and DAO level in the MI group were significantly higher than that in the Sham group (both P<0.05). Terlipressin or norepinephrine significantly attenuated Chiu's score and DAO level (all P<0.05 vs. MI; Fig. 2A), and there was no statistical difference in Chiu's score and DAO level between group TP and NE (all P>0.05).

Meanwhile, the serum BUN and creatinine levels

and the injury score of the kidney in the MI group were higher than that in the Sham group (all P<0.05; Fig. 2B). Either terlipressin or norepinephrine infusion significantly attenuated renal injuries (all P<0.05 vs. MI; Fig. 2B), and there was no statistical difference in these parameters between group TP and NE (all P>0.05; Fig. 2B).

The survival time and mortality rate of the rats in the MI group were 3 hours (2-24 hours) and 85.7%, respectively (Fig. 2C). Terlipressin or norepinephrine significantly increased survival time and survivors (both P<0.05 vs. MI; Fig. 2C).

Terlipressin or norepinephrine reduced intestinal epithelial apoptosis after acute MI

As shown in Fig. 3A, the apoptotic index in TUNEL analysis of the intestinal epithelia was significantly higher in the MI group than in the Sham group (P<0.05), while the apoptosis of intestinal epithelial cells in the TP and NE groups were markedly reduced when compared with the MI group (both P<0.05). Moreover, cleaved capsase-3 expression notably increased in intestinal mucosal tissues after MI (P<0.05 vs. Sham; Fig. 4B) and was significantly reduced by the use of terlipressin or norepinephrine (both P<0.05 vs. MI; Fig. 3B).

The apoptotic index and cleaved caspase-3 expression in the renal tissues were also increased in the MI group (both P<0.05 vs. Sham; Fig. S1). However, neither terlipressin nor norepinephrine reduced renal apoptosis (all P>0.05 vs. MI; Fig. S1). Moreover, neither terlipressin nor norepinephrine significantly inhibited the 8-isoprostane upregulation induced by acute MI (both P>0.05 vs. MI; Fig. S2).



Figure 1. Effects of terlipressin or norepinephrine infusion on mean arterial pressure (A) and heart rate (B) during the study period. Animals underwent various interventions based on the experimental protocol. Data are expressed as mean \pm SEM, n=4 to 5. Results were compared by two-way ANOVA with repeated measures followed by Bonferroni posttest. * *P*<0.05 vs. Sham group; % *P*<0.05 vs. MI group, WT group or T+W group. Timepoint 1: baseline, 2: clamping the SMA, 3: immediately after unclamping the SMA, 4: 3 minutes after unclamping, 6: 60 minutes after unclamping, 7: 120 minutes after unclamping. Sham group: the SMA of rat was exposed but not occluded; MI group: acute mesenteric ischemia model was produced by clamping the SMA; TP group: terlipressin was infused after unclamping the SMA; NE group: norepinephrine was infused after unclamping the SMA; WT group: wortmannin, a specific PI3K inhibitor, was used after mesenteric ischemia, T+W group: terlipressin and wortmannin were both administered after ischemia. SMA: superior mesenteric artery; PI3K: phosphoinositide 3-kinase.







Figure 3. Effects of terlipressin or norepinephrine infusion on intestinal epithelial apoptosis after acute MI. (A) The ileal sections were stained by using TUNEL and evaluated with apoptotic epithelial cells under light microscopy (×200). Representative microscopic images from various groups were presented in the upper panel, and apoptotic nuclei were stained dark brown (red arrows). Analysis of apoptotic index was presented in the lower left-hand. (B) The cleaved caspase-3 expression in intestinal epithelial tissues was detected by western blotting assay. Representative bands of densitometry analysis from each group were shown in the upper side, and analysis of quantitative changes in caspase-3 expressions was shown in the bottom. Data were expressed as mean 4 SEM, n=4 to 7. Results were compared by ANOVA with Tukey posttest. * P < 0.05 vs. Sham group; # P < 0.05 vs. MI group; \$ P < 0.05 vs. TP group. Sham group: the SMA of rat was exposed but not occluded; MI group: acute mesenteric ischemia model was produced by clamping the SMA; TP group: terlipressin was infused after unclamping the SMA; NE group: norepinephrine was infused after unclamping the SMA; WT group: wortmannin, a specific PI3K inhibitor, was used after mesenteric ischemia; T+W group: terlipressin and wortmannin were both administered after ischemia. MI: mesenteric ischemia; SMA: superior mesenteric artery; PI3K: phosphoinositid 3-kinase; TUNEL: terminal deoxynucleotidyl transferase biotin-dUTP nick end-labeling; c-caspase-3: cleaved caspase.

Terlipressin inhibited macrophage M1 polarization and excessive inflammation after acute MI

As shown in Fig. 4A, *in vitro*, M1 markers (iNOS and IL-1 β) mRNA expressions were significantly increased after LPS treatment (both *P*<0.05 vs. Control), whereas M2 markers (Arg1 and FIZZ1) did not significantly change (both *P*>0.05 vs. Control). Terlipressin inhibited iNOS and IL-1 β mRNA expressions in RAW264.7 cells (both *P*<0.05 vs. LPS; Fig. 4A). Meanwhile, the increased TNF- α and IL-1 β levels in culture medium after LPS challenge were significantly reduced by terlipressin (both *P*<0.05 vs. LPS; Fig. 4B).

Furthermore, the number of iNOS-positive cells were significantly increased in intestinal and renal tissues in group MI (both P<0.05 vs. Sham; Fig. 4C),

but the number of CD163-positive cells did not significantly change (both *P*>0.05 vs. Sham; Fig. 4D). Terlipressin, but not norepinephrine, reduced the number of iNOS-positive cells after MI (both *P*<0.05 vs. MI; Fig. 4C), whereas CD163-positive cells were not significantly changed by terlipressin (both *P*>0.05 vs. MI; Fig. 4D). Similarly, the increased concentration of TNF- α and IL-1 β in the intestinal and renal tissues caused by acute MI were reduced in the group TP (all P<0.05 vs. MI; Fig. 5E) but not in the group NE (all *P*>0.05 vs. MI; Fig. 4E).

In rat's serum, the concentration of TNF- α , IL-1 β and IL-6 were higher in the group MI (all *P*<0.05 vs. Sham; Fig. 5). Terlipressin significantly decreased the levels of these cytokines in the serum (both *P*<0.05 vs. MI), whereas norepinephrine failed to decrease the concentration of TNF- α , IL-1 β and IL-6 (all *P*>0.05 vs. MI; Fig. 5).









Figure 5. Effects of terlipressin or norepinephrine infusion on proinflammatory cytokines in serum after acute MI. The concentration of proinflammatory cytokines in rat's serum was detected by using ELISA assay, respectively. Analyses of TNF- α , IL-1 β an IL-6 concentration in serum were presented. Data were expressed as mean±SEM, n=4 to 6. Results were compared by ANOVA with Tukey posttest. * P < 0.05 vs. Sham group; # P < 0.05 vs. MI group; \$ P < 0.05 vs. TP group. Sham group: the SMA of rat was exposed but not occluded; MI group: acute mesenteric ischemia model was produced by clamping the SMA; TP group: acute mesenteric ischemia model was produced by clamping the SMA; TP group: terlipressin was infused after unclamping the SMA; WT group: terlipressin and wortmannin were both administered after ischemia. MI: mesenteric ischemia; SMA: superior mesenteric artery; Pl3K: phosphoinositide 3-kinase; TNF: tumor necrosis factor; IL: interleukin; ELISA: enzyme-linked immunosorbent assay.

Terlipressin relieved organ injuries after acute MI via PI3K/Akt pathway

As shown in Fig. 6A and 6B, *in vivo*, the PI3K and p-Akt protein expressions in the intestinal mucosa and renal tissues were elevated after acute MI (all P<0.05 vs. Sham). The use of terlipressin moderately, but not statistically, enhanced PI3K and p-Akt expressions in the intestinal epithelia and renal tissues (all P>0.05 vs. MI; Fig. 6A and 6B). Similarly, the PI3K

and p-Akt expressions in the cultured RAW264.7 cells were also elevated after LPS challenge (both P<0.05 vs. Control; Fig. 6C). And terlipressin slightly increased PI3K and p-Akt expressions in the RAW264.7 cells with LPS treatment (both P>0.05 vs. LPS; Fig. 6C).

In vivo, wortmannin alone (WT group) produced no active effects on the changes caused by MI (all *P*>0.05 vs. MI; Fig. 1-5) but decreased the PI3K and p-Akt expressions in the cells and tissues (all *P*<0.05 vs. MI; Fig. 6). On the other hand, wortmannin totally diminished the active effects of terlipressin on the PI3K/Akt expressions (Fig. 6), the intestinal epithelial apoptosis, macrophages M1 polarization and inflammatory activation (Figs. 3-5), and the MAP, organ damages and animal survival (Fig. 1 and 2) in the T+W group, when compared with the TP group (all *P*<0.05).

Discussion

In the current study, occluding and unclamping of SMA successfully induced vasodilatory shock and caused intestinal and renal injury (Figs. 1 and 2). Our dosing regimens of terlipressin and norepinephrine mimicked the applications of vasoactive drugs in the clinical settings, and the current infusing doses in rats are approximately equal to usual dosage of terlipressin and norepinephrine for shock treatment in humans according to a classical conversion of drug dose from animal to human [19]. Several clinical evidences suggested that terlipressin, as compared with norepinephrine, did not increased digestive complications in the treatment of vasodilatory shock and hepatorenal syndrome [20-22]. However, to the best of our knowledge, no study has compared the efficacy of vasopressin receptors agonist versus norepinephrine on the organ functions and outcome following acute MI. The current data showed that either terlipressin or norepinephrine application could effectively increase MAP (Fig. 1A), improve the organ damages and reduce mortality (Fig. 2) after severe MI insult in rats.

In the present study, acute MI and following reperfusion induced sustaining and severe hypotension, whose hemodynamic data were line with previous findings [15, 23]. For shock treatment, it is essential to restore MAP timely to guarantee adequate tissue perfusion and oxygen delivery. The either administration of terlipressin or norepinephrine rapidly and effectively improved MAP (Fig. 1A) and subsequently relieved organ damages following acute MI insult (Fig. 2). This suggests that the timely restoration of MAP and organ perfusion pressure delivered by these two vasoactive drugs plays an important role in the intestinal and



unclamping the SMA; WT group: wortmannin, a specific PI3K inhibitor, was used after mesenteric ischemia; T+W group: terlipressin and wortmannin were both administered after ischemia. MI: mesenteric ischemia; SMA: superior mesenteric artery; PI3K: phosphoinositide 3-kinase; p-Akt: phospho-protein kinase B; GAPDH:

renal protection. Besides impaired organ perfusion, intestinal epithelial excessive apoptosis and inflammation, well macrophage M1 as as polarization, were also involved in the pathogenesis of MI-induced organ injury [15, 18, 24]. Apoptosis is a major mode of intestinal epithelial cell death after acute MI attack, and inhibition of apoptosis can prevent the destruction of mucosal barrier in such pathophysiological events [25-27]. Two previous studies have reported that terlipressin could decrease cerebral and hepatic apoptosis in ischemic conditions [28, 29]. Similarly, the results of the apoptotic index and cleaved caspase-3 expression explicitly showed that terlipressin management could significantly inhibit intestinal mucosal apoptosis (Fig. 3) in intestinal ischemia. Interestingly, neither terlipressin

glyceraldehyde-3-phosphate dehydrogenase; DMSO: dimethyl sulfoxide; LPS: lipopolysaccharide.

nor norepinephrine inhibited renal apoptosis following MI (Fig. S1). Apoptosis in distant organs caused by acute MI needs further investigations.

Our previous study suggested that M1 macrophages polarization was promoted after acute MI, and that macrophage depletion or enhancing switch from proinflammatory M1 to antiinflammatory M2 macrophages phenotypes could ameliorate intestinal injury and improved survival [18, 30]. Chang and Jan found that, vasopressin inhibited the activation of cultured macrophages and the release of proinflammatory cytokines following endotoxin challenge [31, 32]. To the best of our knowledge, the current study for the first time showed that, both in vitro and in vivo, V1 receptor markedly blocked macrophages agonist M1

polarization after acute MI challenge whereas it produced no distinct effect on M2 polarization (Fig. 4). We speculate that LPS migrated from the gut lumen induced macrophages M1 polarization in various organs after MI, and that increased M1 macrophages and subsequently produced excessive inflammatory responses led to tissue injury and organ failure. Because V1 receptor agonist possesses anti-inflammatory effects [31, 32], terlipressin, as compared with norepinephrine, potently restrains the activated M1 macrophages in various organs and then reduces the release of inflammatory cytokines after acute MI (Figs. 4 and 5). The present data strongly indicated that terlipressin offered reliable and efficient protections for acute MI-induced multiple organ dysfunction when compared with traditional vasopressor. These novel findings may help laboratory investigators and clinicians to deeply understand mechanism of distant organ injury caused by MI and to choose the optimal vasoactive drugs in MI-relevant diseases.

In cardiovascular system, PI3K inhibitors in particular have the potential to reduce blood pressure [33], and PI3K inhibitors could block the phenylephrine-induced arterial contraction and reduce MAP under a volatile anesthetic sevoflurane conditioning [34]. Vasopressin receptors are G protein-coupled receptors and PI3K has been implicated in a signal transducer of G proteinassociated signaling. It has been demonstrated that the vasoconstricting effect of vasopressin through V1 receptor is mediated by phosphotidylinositol pathway [35]. Interestingly, by activating PI3K/Akt signaling, vasopressin also exerts its ability to inhibit the activation of downstream NF-KB and the upregulation of inflammatory mediators in macrophage induced by endotoxin [32]. It has been widely proven that the activation of PI3K/Akt signaling could reduce the M1 polarization by downregulating the TLR signaling and activating mTOR which then decreases the NF-кВ activation [36, 37]. Previous studies have shown that Akt isoforms hold the key role in the regulation of macrophage activation [36, 38]. Specifically, Akt1 kinase contributes to M2 polarization while Akt2 kinase contributes to M1 polarization due to their opposite roles in regulating miR-155 and its target, C/EBPb, a master regulator of macrophage differentiation [38]. We speculate a dominant activation of Akt2 kinase but not Akt1 in current MI model and thus only M1 marker changed notably. Also, our previous study and Miller's study have demonstrated the antiapoptotic effect of vasopressin and terlipressin via activating PI3K/Akt signaling [13, 39]. The current data demonstrated that wortmannin could diminish

hemodynamic improvement induced bv the terlipressin after MI insult whereas wortmannin alone produced no active effect (Fig. 1A), suggesting that PI3K inhibitor may abolish V1 receptor-mediated vasoconstricting effect. Wortmannin also significantly eliminated the protective effects of terlipressin on macrophage M1 polarization reducing and attenuating organ damages after MI challenge by inhibiting PI3K activation (Fig. S3). Our clinically meaningful results suggest that vasopressin V1 receptor agonists are not suitable for patients with PI3K inhibitors therapy (e.g. treatment and prevention of cancer).

There were some possible limitations in this study. First, the present results indicated that terlipressin provides organ protection by elevating blood pressure and inhibiting apoptosis and M1 polarization. However, it is very difficult to distinguish whether the main effect of terlipressin on MI insult is attributed to the improved organ perfusion or the repression of apoptosis and M1 polarization. Second, in this experimental study, severe hypotension was induced by intestinal ischemia and reperfusion. Whereas, in the clinical settings, hypotensive shock in the patients with acute MI was usually induced by other critical conditions (e.g. sepsis and hemorrhage). Final, in this study, we macrophages that decreased M1 speculated generation (Fig. 4) caused by terlipressin may play a key role in the suppression of systemic inflammation (Fig. 5). Further verification is required to confirm this hypothesis.

Taken together, MI leads to vasodilatory hypotension and notable intestinal and renal injury. The use of terlipressin or norepinephrine effectively improves hemodynamics and organ damages, and subsequently reduces mortality. Terlipressin delivers its protective effects by elevating blood pressure and inhibiting intestinal epithelial apoptosis and macrophage M1 polarization via PI3K/Akt pathway. Our study exhibits that terlipressin may be a promising vasoactive alternative for MI relevant conditions in clinical practices.

Abbreviations

MI: Mesenteric ischemia; ICU: Intensive care unit; SMA: superior mesenteric artery; NS: normal saline; TNF-a: tumor necrosis factor-a; IL: interleukin; PI3K/Akt pathway: phosphoinositide 3-kinase/ protein kinase B pathway; ELISA: Enzyme Linked Immunosorbent Assay; DAO: Diamine oxidase; BUN: blood urea nitrogen; GAPDH: glyceraldehyde-3phosphate dehydrogenase; TUNEL: terminal deoxynucleotidyl transferase biotin-dUTP nick end-labeling assay.

Supplementary Material

Supplementary figures and tables. http://www.medsci.org/v17p2751s1.pdf

Acknowledgements

We would like to thank Ms. Jie Li for her help in animal experiments. And we thank Ms. Brooke Wang for her help with the manuscript. This work was supported by grants from the National Natural Science Foundation of China (Grant 81701874 to Zi-Meng Liu), the Natural Science Foundation of Guangdong Province, China (Grant 2017A030313572 to Xu-Yu Zhang), and the Major Science and Technology Projects of Guangdong province, China (Grant 2012A080204018 to Xiang-Dong Guan).

Competing Interests

The authors have declared that no competing interest exists.

References

- Leone M, Bechis C, Baumstarck K, et al. Outcome of acute mesenteric ischemia in the intensive care unit: a retrospective, multicenter study of 780 cases. Intensive Care Med. 2015; 41: 667-76.
- Yasuhara H. Acute mesenteric ischemia: the challenge of gastroenterology. Surg. Today. 2005; 35: 185-95.
- Ozban M, Aydin C, Cevahir N, et al. The effect of melatonin on bacterial translocation following ischemia/reperfusion injury in a rat model of superior mesenteric artery occlusion. BMC Surg. 2015; 15: 18.
- Biffl WL and Moore EE. Splanchnic ischaemia/reperfusion and multiple organ failure. Br J Anaesth. 1996; 77: 59-70.
- Caluwaerts M, Castanares-Zapatero D, Laterre PF, et al. Prognostic factors of acute mesenteric ischemia in ICU patients. BMC Gastroenterol. 2019; 19: 80.
- Daviaud F, Grimaldi D, Dechartres A, et al. Timing and causes of death in septic shock. Ann Intensive Care. 2015; 5: 16.
- Anderson JE, Brown IE, Olson KA, et al. Nonocclusive mesenteric ischemia in patients with methamphetamine use. J Trauma Acute Care Surg. 2018; 84: 885-892.
- Liu ZM, Chen J, Kou Q, et al. Terlipressin versus norepinephrine as infusion in patients with septic shock: a multicentre, randomised, double-blinded trial. Intensive Care Med. 2018; 44: 1816-1825.
- Schmitt W, Wagner-Thiessen E and Lux G. Ischaemic colitis in a patient treated with glypressin for bleeding oesophageal varices. Hepatogastroenterology. 1987; 34: 134-6.
- Kim HR, Lee YS, Yim HJ, et al. Severe ischemic bowel necrosis caused by terlipressin during treatment of hepatorenal syndrome. Clin Mol Hepatol. 2013; 19: 417-20.
- 11. Borrego R, López-Herce J, Mencía S, et al. Severe ischemia of the lower limb and of the intestine associated with systemic vasoconstrictor therapy and femoral arterial catheterization. Pediatr Crit Care Med. 2006; 7: 267-9.
- Asfar P, Bracht H and Radermacher P. Impact of vasopressin analogues on the gut mucosal microcirculation. Best Pract Res Clin Anaesthesiol. 2008; 22: 351-8.
- Liu ZM, Zhang XY, Chen J, et al. Terlipressin protects intestinal epithelial cells against oxygen-glucose deprivation/re-oxygenation injury via the phosphatidylinositol 3-kinase pathway. Exp Ther Med. 2017; 14: 260-266.
- Qiu X, Huang Y, Xu J, et al. Effects of terlipressin on microcirculation of small bowel mesentery in rats with endotoxic shock. J Surg Res. 2014; 188: 503-9.
- Zhang XY, Liu ZM, Wen SH, et al. Dexmedetomidine administration before, but not after, ischemia attenuates intestinal injury induced by intestinal ischemia-reperfusion in rats. Anesthesiology. 2012; 116: 1035-46.
- Luk GD, Bayless TM and Baylin SB. Diamine oxidase (histaminase). A circulating marker for rat intestinal mucosal maturation and integrity. J Clin Invest. 1980; 66: 66-70.
- Gao S, Wang Y, Li D, et al. TanshinoneIIA Alleviates Inflammatory Response and Directs Macrophage Polarization in Lipopolysaccharide-Stimulated RAW264.7 Cells. Inflammation. 2019; 42: 264-275.
- Liu WF, Wen SH, Zhan JH, et al. Treatment with Recombinant Trichinella spiralis Cathepsin B-like Protein Ameliorates Intestinal Ischemia/Reperfusion Injury in Mice by Promoting a Switch from M1 to M2 Macrophages. J. Immunol. 2015; 195: 317-28.
- Reagan-Shaw S, Nihal M and Ahmad N. Dose translation from animal to human studies revisited. Faseb j. 2008; 22: 659-61.

- Serpa Neto A, Nassar AP, Cardoso SO, et al. Vasopressin and terlipressin in adult vasodilatory shock: a systematic review and meta-analysis of nine randomized controlled trials. Crit Care. 2012; 16: R154.
- Sharma P, Kumar A, Shrama BC, et al. An open label, pilot, randomized controlled trial of noradrenaline versus terlipressin in the treatment of type 1 hepatorenal syndrome and predictors of response. Am. J. Gastroenterol. 2008; 103: 1689-97.
- Alessandria C, Ottobrelli A, Debernardi-Venon W, et al. Noradrenalin vs terlipressin in patients with hepatorenal syndrome: a prospective, randomized, unblinded, pilot study. J. Hepatol. 2007; 47: 499-505.
- Liu C, Shen Z, Liu Y, et al. Sevoflurane protects against intestinal ischemiareperfusion injury partly by phosphatidylinositol 3 kinases/Akt pathway in rats. Surgery. 2015; 157: 924-33.
- Wang Z, Li Z, Feng D, et al. Autophagy Induction Ameliorates Inflammatory Responses in Intestinal Ischemia-Reperfusion Through Inhibiting NLRP3 Inflammasome Activation. Shock. 2019; 52: 387-395.
- Li Y, Wen S, Yao X, et al. MicroRNA-378 protects against intestinal ischemia/reperfusion injury via a mechanism involving the inhibition of intestinal mucosal cell apoptosis. Cell Death Dis. 2017; 8: e3127.
- Wen S, Ling Y, Yang W, et al. Necroptosis is a key mediator of enterocytes loss in intestinal ischaemia/reperfusion injury. J Cell Mol Med. 2017; 21: 432-443.
- Wen SH, Li Y, Li C, et al. Ischemic postconditioning during reperfusion attenuates intestinal injury and mucosal cell apoptosis by inhibiting JAK/STAT signaling activation. Shock. 2012; 38: 411-9.
- Liu X, Luo G, Jiang J, et al. Signaling through hepatocyte vasopressin receptor 1 protects mouse liver from ischemia-reperfusion injury. Oncotarget. 2016; 7: 69276-69290.
- Ida KK, Otsuki DA, Sasaki AT, et al. Effects of terlipressin as early treatment for protection of brain in a model of haemorrhagic shock. Crit Care. 2015; 19: 107.
- Wen S, Li X, Ling Y, et al. HMGB1-associated necroptosis and Kupffer cells M1 polarization underlies remote liver injury induced by intestinal ischemia/ reperfusion in rats. FASEB J. 2020.
- Chang YY, Yang CH, Wang SC, et al. Vasopressin inhibits endotoxin binding in activated macrophages. J. Surg. Res. 2015; 197: 412-8.
- Jan WC, Kao MC, Yang CH, et al. Phosphoinositide 3-Kinase Is Involved in Mediating the Anti-inflammation Effects of Vasopressin. Inflammation. 2017; 40: 435-441.
- Ghigo A, Morello F, Perino A, et al. Therapeutic applications of PI3K inhibitors in cardiovascular diseases. Future Med Chem. 2013; 5: 479-92.
 Miyamoto Y, Feng GG, Satomi S, et al. Phosphatidylinositol 3-kinase
- Miyamoto Y, Feng GG, Satomi S, et al. Phosphatidylinositol 3-kinase inhibition induces vasodilator effect of sevoflurane via reduction of Rho kinase activity. Life Sci. 2017; 177: 20-26.
- den Ouden DT and Meinders AE. Vasopressin: physiology and clinical use in patients with vasodilatory shock: a review. Neth J Med. 2005; 63: 4-13.
- Vergadi E, Ieronymaki E, Lyroni K, et al. Akt Signaling Pathway in Macrophage Activation and M1/M2 Polarization. J Immunol. 2017; 198: 1006-1014.
- Yang QC, Wu CC, Cao LY, et al. Increased Expression of LAMTOR5 Predicts Poor Prognosis and Is Associated with Lymph Node Metastasis of Head and Neck Squamous Cell Carcinoma. Int J Med Sci. 2019; 16: 783-792.
- Arranz A, Doxaki C, Vergadi E, et al. Akt1 and Akt2 protein kinases differentially contribute to macrophage polarization. Proc Natl Acad Sci U S A. 2012; 109: 9517-22.
- Miller RL, Sandoval PC, Pisitkun T, et al. Vasopressin inhibits apoptosis in renal collecting duct cells. Am J Physiol Renal Physiol. 2013; 304: F177-88.