

Rationale for combination therapy of chronic myelogenous leukaemia with imatinib and irradiation or alkylating agents: implications for pretransplant conditioning

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The tyrosine kinase activity of the BCR–ABL oncoprotein results in reduced apoptosis and thus prolongs survival of chronic myelogenous leukaemia cells. The tyrosine kinase inhibitor imatinib (formerly ST1571) was reported to selectively suppress the proliferation of BCR–ABL-positive cells. Assuming that imatinib could be included in pretransplantation conditioning therapies, we tested whether combinations of imatinib and γ -irradiation or alkylating agents such as busulfan or treosulfan would display synergistic activity in BCR–ABL-positive chronic myelogenous leukaemia BV173 and EM-3 cell lines. Further, primary cells of untreated chronic myelogenous leukaemia patients were assayed for colony forming ability under combination therapy with imatinib. Additionally, the cytotoxic effect of these combinations on BCR–ABL-negative cells was investigated. In the cell lines a tetrazolium based MTT assay was used to quantify growth inhibition after exposure to cytotoxic drugs alone or to combinations with imatinib. Irradiation was applied prior to exposure to imatinib. Interaction of drugs was analysed using the median-effect method of Chou and Talalay. The combination index was calculated according to the classic isobologram equation. The combination imatinib + γ -irradiation proved to be significantly synergistic over a broad range of cell growth inhibition levels in both BCR–ABL-positive cell lines and produced the strongest reduction in primary chronic myelogenous leukaemia colony-forming progenitor cells. Combinations of imatinib + busulfan and imatinib + treosulfan showed merely additive to antagonistic effects. Imatinib did not potentiate the effects of irradiation or cytotoxic agents in BCR–ABL-negative cells. Our data provide the basis to further develop imatinib-containing conditioning therapies for stem cell transplantation in chronic myelogenous leukaemia.

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Chronic myelogenous leukaemia (CML) is a clonal disorder of the pluripotent stem cell with involvement of all haemopoietic lineages and is characterised by preferential expansion of myeloid cells. On the cellular level, CML is distinguished by the Philadelphia chromosome (Nowell and Hungerford, 1960), an abnormally short chromosome 22, which is found in more than 90% of all cases of CML, arising from a reciprocal t(9;22) translocation (Rowley, 1973). The product of this translocation, a 210-kD BCR–ABL protein (p210^{BCR-ABL}), is characterised by a constitutively enhanced tyrosine kinase activity as compared with that of the normal c-ABL protein (Konopka *et al*, 1984) and is known to interfere with a variety of cytoplasmatic and cytoskeletal signalling proteins and cascades (Faderl *et al*, 1999; Deininger *et al*, 2000), which eventually leads to inhibition of apoptosis. The inherent resistance of residual BCR–ABL-positive cells to cytotoxic therapy is a major impediment to long-term management of CML.

After a median duration of 2–6 years under conventional therapy, the initial indolent chronic phase of the disease is followed by

the accelerated phase and terminal blast crisis, which results in the patient's death within 3–6 months (Kantarjian *et al*, 1998). Allogeneic stem cell transplantation (allo-SCT) still remains the only proven curative option and particularly younger patients can benefit from this procedure (Gratwohl *et al*, 1998).

For patients without a suitable donor, high-dose conditioning therapy followed by autologous SCT (auto-SCT) is an option and durable remissions can be achieved in patients transplanted in the early chronic phase (Carella *et al*, 1999). The value of auto-SCT as consolidative treatment in blast crisis patients or patients with BCR–ABL-positive leukaemia transplanted in remission is the subject of ongoing trials (Goekbuget *et al*, 2001). Detection of residual leukaemic cells after high-dose chemotherapy is a strong predictor for clinical relapse in BCR–ABL-positive disease (van Dongen *et al*, 1998). Strategies to improve the conditioning therapy may help to increase the remission rates.

One of the new and currently very promising therapeutic strategies for CML is the targeting of the BCR–ABL tyrosine kinase (Druker *et al*, 1996). Recently reported clinical trials with the Abl tyrosine kinase inhibitor imatinib in blast crisis (Druker *et al*, 2001a), interferon-refractory or -intolerant chronic phase (Druker *et al*, 2001b), and in accelerated phase CML patients (Talpa *et al*, 2000) showed encouraging results.

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Nevertheless, 60% of myeloid blast crisis and almost all lymphoid blast crisis patients eventually relapsed (Druker *et al*, 2001a). Relapses were also observed in the accelerated and even in the chronic phase of the disease (Gorre *et al*, 2001; Hochhaus *et al*, 2001). Combination of imatinib with established cytotoxic therapy especially for the control of minimal residual disease as can be achieved by marrow-ablative conditioning therapy may allow to further reduce the leukaemic cell pool.

MATERIALS AND METHODS

Cells

The BCR–ABL-positive BV173 (lymphatic blast crisis) and EM-3 (myeloid blast crisis) human cell lines and the BCR–ABL-negative KG1a and HL-60 human leukaemic cell lines were obtained from the German Collection of Microorganisms and Cell Cultures (Braunschweig, Germany). The BCR–ABL status of the cell lines was confirmed using reverse transcriptase-polymerase chain reaction (data not shown). Cells were grown in RPMI-1640 medium supplemented with 10% (BV173, HL-60 and EM-3) or with 20% (KG1a) heat inactivated foetal calf serum (FCS), 2 mM L-glutamine, and penicillin/streptomycin (Life Technologies GmbH, Eggenstein, Germany) at 37°C in a fully humidified atmosphere of 95% air and 5% CO₂. Primary cells were obtained from two

patients in chronic phase CML. Mononuclear cells were isolated by Ficoll gradient centrifugation (Biocoll separating solution, Biochrom KG, Berlin, Germany).

Irradiation and alkylating agents

γ -irradiation, busulfan and treosulfan were chosen for combination treatment in this study. A ¹³⁸Cs source at a dose-rate of 0.72 Gy min⁻¹ was used for irradiation. The busulfan solution (Busulfex injection) was obtained from Orphan Medical, Inc., Minnetonka, MN, USA, in a 10 ml ampoule containing busulfan 60 mg, dimethylacetamide 3.333 ml and polyethylene glycole 6.667 ml. Treosulfan (Ovostat 1000) was provided by Medac, Hamburg, Germany. Imatinib was provided by Novartis Pharma AG, Basel, Switzerland. Serial dilutions of the drugs were prepared by diluting the stock solutions in sterile Dulbecco's phosphate buffered saline (Dulbecco's PBS, Life Technologies GmbH) freshly prior to beginning of incubation.

Simultaneous exposure to imatinib in combination with busulfan or treosulfan was chosen to ensure a continuous inhibition of the BCR–ABL tyrosine kinase. Irradiation was applied prior to incubation with imatinib. Cells were either treated with six increasing doses (doubling with each increment) of busulfan, treosulfan, irradiation or imatinib alone or with the respective combinations. The doses for BV173 cells are given in Table 1. For EM-3 cells following dose ranges were used: γ -irradiation

Table 1 Growth inhibition of BCR–ABL-positive BV173 cells by combinations of imatinib with γ -irradiation, busulfan or treosulfan (MTT assay)

	Imatinib (μM)						
	0	0.025	0.05	0.1	0.2	0.4	0.8
A							
Irradiation (Gy)							
0	0	0.047	0.078	0.173	0.459	0.710	0.816
0.375	0.143	0.256	0.312	0.484	0.672	0.833	0.903
0.75	0.265	0.374	0.465	0.583	0.778	0.888	0.937
1.5	0.520	0.596	0.666	0.786	0.884	0.955	0.973
3	0.743	0.768	0.808	0.863	0.941	0.977	0.985
6	0.910	0.957	0.964	0.980	0.990	0.992	0.992
12	0.956	0.986	0.990	0.991	0.994	0.996	0.994
B							
Busulfan (μM)							
0	0	0.007	0.035	0.146	0.431	0.665	0.736
10	0.010	0.012	0.037	0.200	0.496	0.674	0.768
20	0.011	0.020	0.059	0.240	0.513	0.705	0.778
40	0.092	0.103	0.143	0.352	0.616	0.752	0.811
80	0.376	0.432	0.449	0.577	0.726	0.865	0.917
160	0.853	0.888	0.858	0.908	0.972	0.992	0.997
320	0.995	0.997	0.998	0.998	0.996	0.997	0.996
C							
Treosulfan (μM)							
0	0	0.041	0.067	0.111	0.343	0.648	0.800
1	0.005	0.055	0.053	0.210	0.461	0.732	0.832
2	0.096	0.185	0.196	0.367	0.543	0.754	0.864
4	0.411	0.543	0.585	0.725	0.788	0.845	0.921
8	0.791	0.865	0.866	0.891	0.929	0.932	0.953
16	0.909	0.960	0.959	0.962	0.970	0.973	0.980
32	0.944	0.985	0.980	0.983	0.983	0.986	0.987

The tests with single agents and the respective combinations were carried out simultaneously. The effect of imatinib alone was determined separately for **A**, **B** and **C** explaining the slightly different values. Numbers represent F_a values (e.g. $F_a = 0.5$ corresponds to 50% growth inhibition). For each monotherapy six different doses and for each combination a total of 36 dose combinations were tested. All experiments were performed in triplicate. Here, mean values of triplicates are given. Bold numbers (representing values for equitoxic dose ratio combinations) were selected for further analysis on the basis of the ED₅₀ values of both single modality treatments: e.g. ED₅₀ of imatinib in **A** is approximately 0.2 μM , ED₅₀ of γ -irradiation is approximately 1.5 Gy; the respective approximately equitoxic dose ratio is 0.2 $\mu\text{M}/1.5$ Gy or 1:7.5. Accordingly, approximately equitoxic dose ratios of imatinib/busulfan and imatinib/treosulfan are 1:400 and 1:20 ($\mu\text{M}/\mu\text{M}$), respectively. Together with the respective F_a values for single modality treatments bold figures F_a values were used for combination index calculations using the CalcuSyn software.

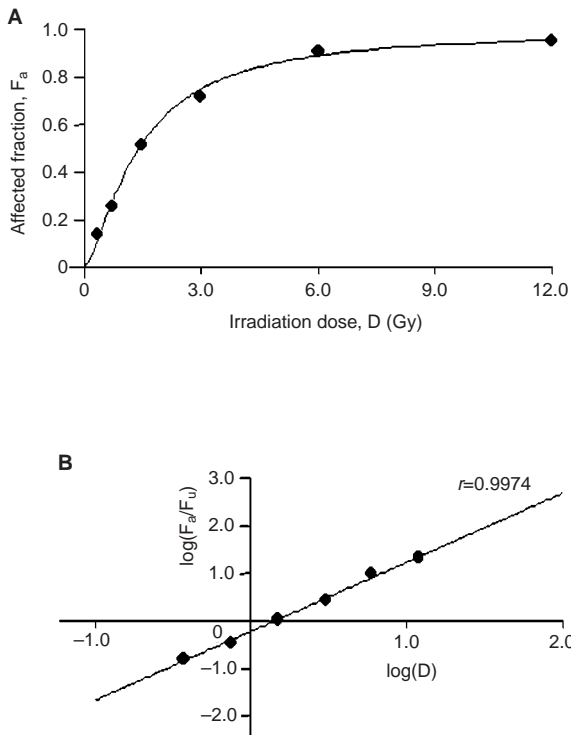


Figure 1 Example for the dose response curve of irradiation in BCR–ABL-positive BV173 cells (**A**) and the data transformation to allow calculation of the level of growth inhibition at a given dose according to the median effect equation of Chou and Talalay (**B**). The dose-response relationship is displayed as a linear regression line ($r=0.997$). The x-intercept of the median effect-plot (**B**) represents the median effect dose (D_m , ED_{50}) at which a 50% growth inhibition ($F_a=0.5$) occurs. The six data points in **A** and **B** correspond to the six original data points of irradiation alone in Table 1 (1st data column). F_a , affected fraction; F_u , unaffected fraction; D , dose.

0.75–24 Gy, busulfan 5–160 μM , treosulfan 2–64 μM , and imatinib 0.025–0.8 μM .

MTT assay

The MTT assay is based on the cleavage of the yellow tetrazolium salt 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide

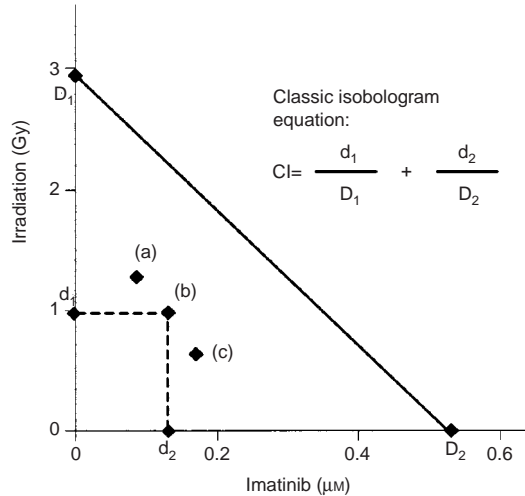


Figure 2 Example of a classic isobologram at ED_{75} . Imatinib is combined with γ -irradiation in BV173 cells. The ED_{75} values were calculated from a series of primary data such as presented in Table 1 using CalcuSyn software. Means of three experiments were calculated and an isobologram was drawn by MS Excel. The three data points shown (**a**, **b**, and **c**) correspond to imatinib [μM]/ γ -irradiation [Gy] ratios of 1:15, 1:7.5 (approximately equitoxic combination), and 1:3.75, respectively. The corresponding CI values as calculated following the classic isobologram equation are 0.572, 0.583, and 0.593. As CI values of <1 express synergism, imatinib + γ -irradiation represents an example of a strongly synergistic combination.

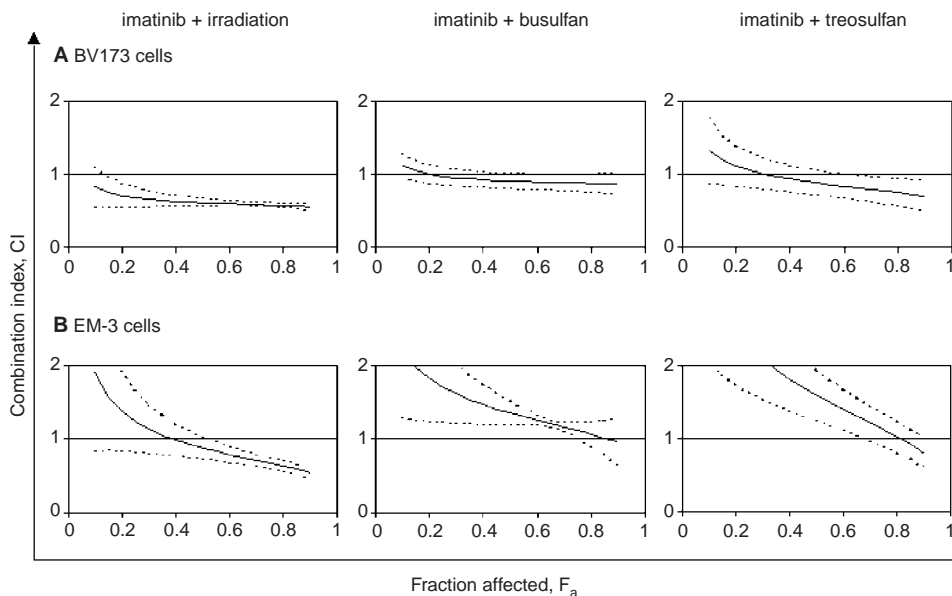


Figure 3 Combination index (CI) as a function of affected fraction (F_a) in BCR–ABL-positive BV173 (**A**) and EM-3 (**B**) cells. F_a values of e.g. 0.25, 0.5 or 0.75 correspond to 25, 50 or 75% growth inhibition. CI values were calculated for distinct F_a values (e.g. F_a 0.1, 0.2, etc.) on the basis of experimentally determined F_a data such as shown in Table 1. By this method, CI values for different levels of growth inhibition and different drug combinations can easily be compared. The synergistic activity increases with higher levels of growth inhibition in all combinations (most apparently in the combination γ -irradiation + imatinib) suggesting optimal (maximal) activity of the pro-apoptotic treatment (γ -irradiation, busulfan, treosulfan) when imatinib is efficiently blocking BCR–ABL tyrosine kinase activity and the associated anti-apoptotic mechanisms. *Solid lines*, means; *dotted lines*, 1 s.d.-range.

(MTT; SIGMA) to dark blue formazan crystals by viable cells (Mosmann, 1983). A high correlation between the viable cell number and formazan production has been reported (Finlay et al, 1986; Pieters et al, 1988). *In vitro* blast cell survival measured by the MTT assay correlates highly with blast cell proliferation measured by the classical ³H-thymidine incorporation assay (Norgaard et al, 1996). The MTT assay is widely used for assessment of combination therapies employing imatinib (Thiesing et al, 2000; Kano et al, 2001) and was performed as previously described (Topaly et al, 2001).

Determination of Combination Index (CI)

Dose-effect relationships were analysed using the median-effect method (Chou and Talalay, 1984; Topaly et al, 2001), an example is given in Figure 1. The combination index (CI) was used to express synergism (CI<1), additivity (CI=1) or antagonism (CI>1) and was calculated according to the classic isobologram equation:

$$CI = d_1/D_1 + d_2/D_2 \quad (1)$$

In this equation, *D*₁ and *D*₂ represent the doses of drug 1 and drug 2 alone, required to produce x% effect, and *d*₁ and *d*₂ are the doses of drugs 1 and 2 in combination required to produce the same effect. Figure 2 illustrates this equation using three different dose ratio combinations of the two treatment modalities, each effecting 75% growth inhibition (ED₇₅). Since different CI values can be observed at different levels of growth inhibition (fraction affected, *F*_a), presentation of data in CI vs affected fraction (*F*_a) plots is reasonable (Figure 3). CI values were initially calculated for three separate experiments using CalcuSyn Software and then transferred to MS Excel for calculation of mean values and standard deviation (s.d.); *F*_a-CI plots were drawn using MS Excel.

CFC Assay

4 × 10⁴ (patient 1) or 2 × 10⁵ cells (patient 2), cell numbers producing 50–70 colony-forming units granulocyte/macrophage (CFU-GM) per plate in untreated controls, were plated in duplicate in

MethoCult GF H4434 methylcellulose (MC; Stem Cell Technologies, Vancouver, Canada) containing SCF (50 ng ml⁻¹), GM-CSF (10 ng ml⁻¹), IL-3 (10 ng ml⁻¹), and erythropoietin (3 units ml⁻¹) and incubated for 14 days at 37°C in humidified atmosphere containing 5% CO₂ as previously described (Kasper et al, 1999). Individual CFU-GMs were counted. Mean values of

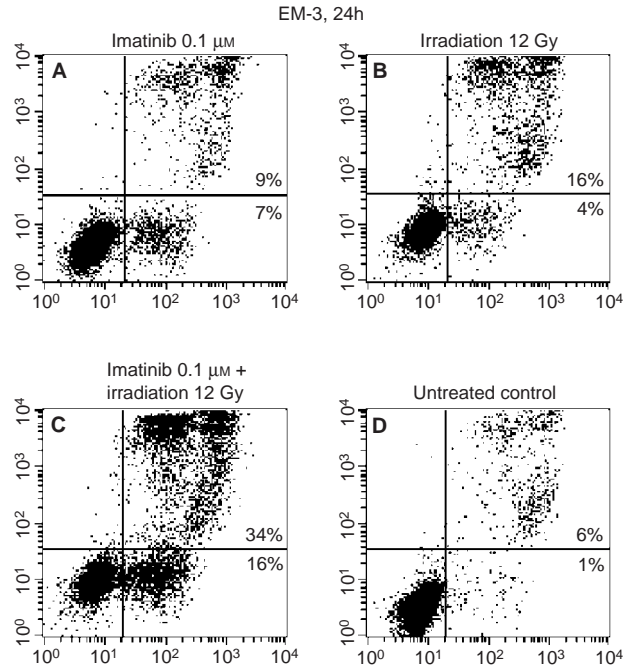


Figure 4 Apoptosis in EM-3 cells 24 h after start of incubation as detected by the annexin V/propidium iodide assay. Compared to treatment with imatinib alone or γ-irradiation alone (A, imatinib; B, γ-irradiation) a striking increase in the early apoptotic (lower right quadrant: annexin V+/PI-) and dead cell (upper right quadrant: annexin V+/PI+) fractions was observed (C) when the cells were treated with the combination. D, untreated cells.

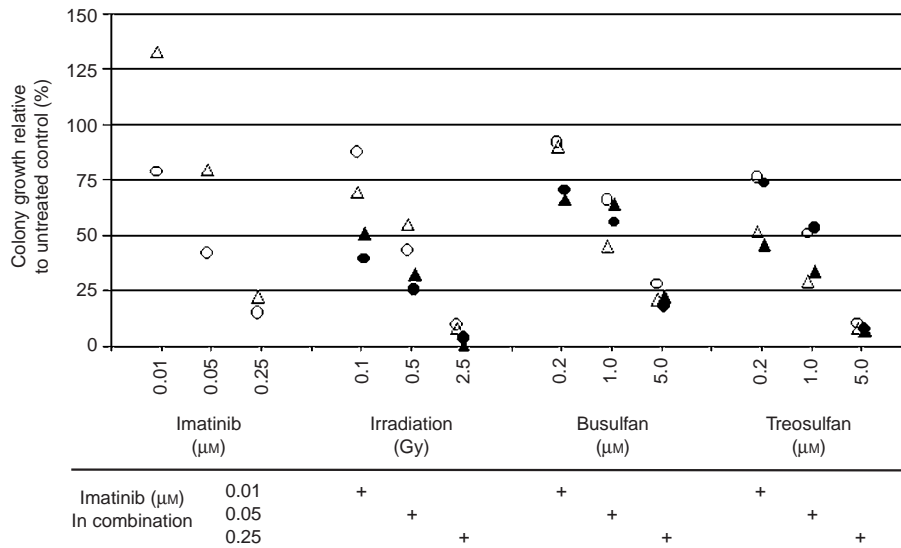


Figure 5 Activity of imatinib combinations in primary CML progenitor cells obtained from two patients. Imatinib + γ-irradiation produced a stronger growth inhibition than either treatment alone over the whole dose range tested as opposed to imatinib + busulfan and imatinib + treosulfan. Triangles, patient 1; circles, patient 2; open symbols, monotherapy; filled symbols, combination with imatinib.

duplicates for each patient were used for further analysis. Doses of the drugs and γ -irradiation used in CFC assay are given in Figure 5.

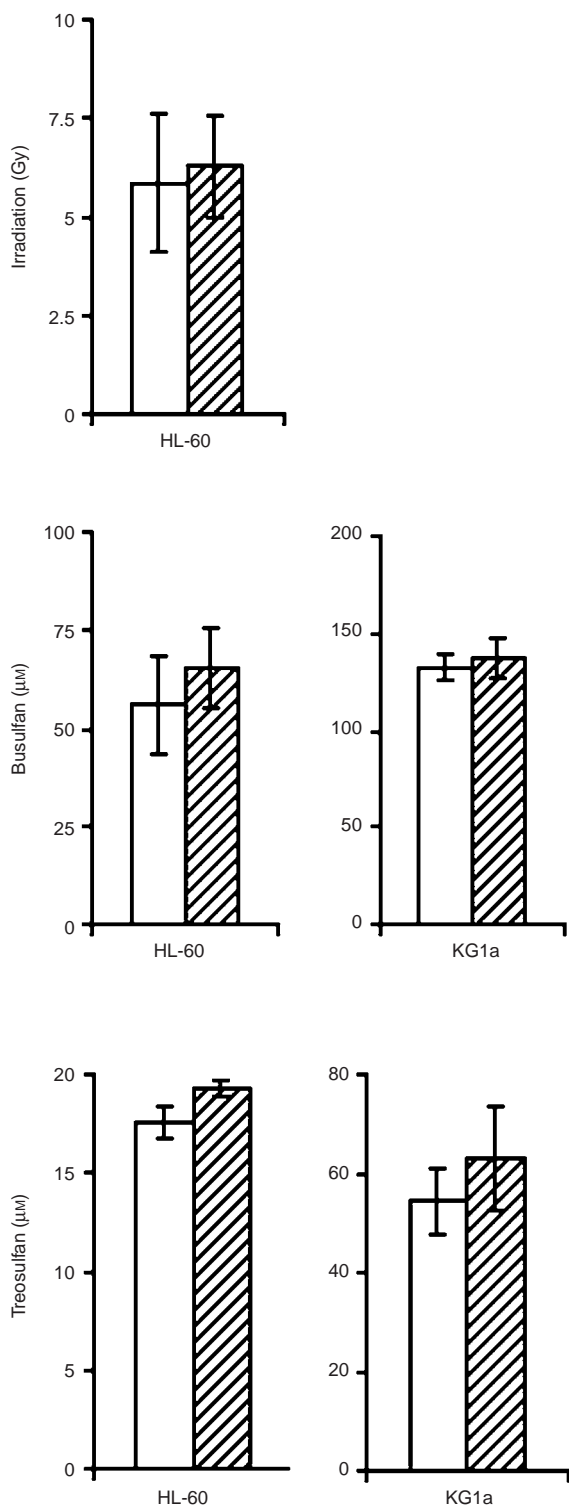


Figure 6 Effects of γ -irradiation, busulfan or treosulfan (each $\pm 1.0 \mu\text{M}$ imatinib) on BCR-ABL-negative cells: ED₅₀ values of γ -irradiation, busulfan or treosulfan alone and the respective ED₅₀ values in combination with imatinib do not differ significantly. Addition of imatinib does not increase the cytotoxicity on BCR-ABL-negative cells. White columns, cytotoxic treatment alone; hatched columns, respective combination with imatinib; bars, 1 s.d. Means of three paired experiments are presented ($P > 0.05$).

Apoptosis assay

Annexin V/propidium iodide apoptosis assay was performed using the Annexin V-FITC Kit (Immunotech, Marseille, France) 24 h after beginning of incubation of EM-3 cells according to the manufacturer's protocol. The data were collected and analysed with a FACScalibur flow cytometer (Becton Dickinson, Heidelberg, Germany) using the CellQuest Software (Becton Dickinson).

Statistics

Results are shown as mean values \pm standard deviation (s.d.) of three separate experiments. Statistical significance of the data was calculated by Student *t*-test. A significance level of $P < 0.05$ was chosen.

RESULTS

Single agent therapy

ED₅₀ values of imatinib alone were assessed following 48 h of incubation for all cell lines. The BCR-ABL-positive cell lines BV173 and EM3 were highly susceptible to imatinib with ED₅₀ values of $0.30 \pm 0.08 \mu\text{M}$ (mean \pm s.d., $n=9$) and $0.11 \pm 0.03 \mu\text{M}$ ($n=3$), respectively. The ED₅₀ values in BCR-ABL-negative cell lines HL-60 and KG1a were $17.1 \pm 0.7 \mu\text{M}$ and $26.7 \pm 3.3 \mu\text{M}$, respectively. The therapeutic index of imatinib *in vitro*, defined as the ratio of ED₅₀ in BCR-ABL-negative cells (dose-limiting side effect) to ED₅₀ in BCR-ABL-positive cells (therapeutic effect), ranged from 57 to 243. For assessment of combination effects in BCR-ABL-negative cells a dose of $1.0 \mu\text{M}$ imatinib was used since this concentration failed to measurably affect their growth while causing more than 80% of growth inhibition in BV173 and EM-3 cells.

The sensitivity of cells to busulfan was relatively uniform ranging from an ED₅₀ of $56.2 \pm 12.4 \mu\text{M}$ in HL-60 to $90.8 \pm 11.3 \mu\text{M}$ in BV173 cells. On the contrary, the sensitivity to γ -irradiation varied widely. BV173 cells proved to be most sensitive (ED₅₀ = $1.70 \pm 0.95 \text{ Gy}$) while in EM-3 cells the ED₅₀ was $6.39 \pm 0.69 \text{ Gy}$; in KG1a cells 10 Gy produced only minor growth inhibition of less than 20%. The ED₅₀ of treosulfan ranged from $5.03 \pm 0.24 \mu\text{M}$ in BV173 cells to $54.4 \pm 3.7 \mu\text{M}$ in KG1a cells.

In all experiments the linear correlation coefficient of the median-effect plot (*r*) was > 0.96 providing a reliable basis for further calculations.

Combination treatment of BCR-ABL-positive cells

Therapeutic effects of approximately equitoxic molar ratios were chosen for the assessment of the combination index (CI). Since each cell line displayed different sensitivity to single modality treatments tested, the equitoxic molar ratios were adjusted individually for each BCR-ABL-positive cell line (bold numbers in Table 1). An example of the experimental design is given in Table 1.

Both in BV173 and in EM-3 cells significant synergistic effects ($P < 0.05$) of imatinib + γ -irradiation were observed. In BV173 cells, the synergistic effect became apparent already at inhibition levels above 30% ($F_a=0.3$) and still increased at higher inhibition levels. In combinations with busulfan or treosulfan, only additive effects were observed (Figure 3A).

In EM-3 cells, higher CI values were generally observed with all combinations tested. A synergism of imatinib + γ -irradiation became apparent at higher levels of growth inhibition than in BV173 cells (Figure 3B). Combinations with busulfan and treosulfan were again additive or even slightly antagonistic (Figure 3B).

The annexin V/propidium iodide apoptosis assay showed a considerable increase of the apoptotic cell fraction in EM-3 cells treated with imatinib + γ -irradiation as compared to treatment with imatinib or γ -irradiation alone (Figure 4). This finding confirms that the observed synergism is based on a pro-apoptotic mechanism of action.

In CFC assays with primary CML progenitor cells combination of imatinib and γ -irradiation produced a stronger growth inhibition than either treatment alone over the whole dose range tested (Figure 5). Combinations of imatinib + busulfan and imatinib + treosulfan were inferior.

Combination treatment of BCR–ABL-negative cells

Since imatinib itself had no measurable effect on BCR–ABL-negative cells at concentrations up to 1.0 μ M, the median-effect method could not be applied to assess combination effects. Therefore, ED₅₀ values of γ -irradiation, busulfan or treosulfan alone were compared with the respective ED₅₀ values in the presence of 1.0 μ M imatinib. As shown in Figure 6, no increase of γ -irradiation-, busulfan- or treosulfan-induced cytotoxicity by imatinib was observed; the respective ED₅₀ values were even slightly higher when combined with imatinib (not significant).

DISCUSSION

This study shows a strong synergism of the combination of imatinib + γ -irradiation in BCR–ABL-positive lymphoid and myeloid blast crisis cells and its superiority to combinations of imatinib with the alkylating agents busulfan or treosulfan which were merely additive. Imatinib + γ -irradiation was also the most active combination against primary colony-forming CML cells. In BCR–ABL+ cell lines, doses chosen (with the exception of busulfan) came close to the clinically relevant dose range; in primary CML progenitors we chose a dose range of 0.1–2.5 Gy γ -irradiation and 0.2–5.0 μ M busulfan or treosulfan (Figure 5) which would be feasible to be used *in vivo* (Körbling *et al*, 1986; Jacobson *et al*, 2001; Scheulen *et al*, 2000). Synergistic effects of imatinib with γ -irradiation and chemotherapeutic agents in leukaemic cell lines correlated well with the effects in primary CML cells.

In-vitro combination treatment data have already contributed to the design of current clinical trials with imatinib (Fruehauf and Hochhaus, 2001; Druker *et al*, 2001c). Consideration of the present results for clinical pretransplant conditioning therapy of CML patients should be discussed.

Since imatinib alone has little or no measurable effect and does not potentiate the effects of established cytotoxic treatment in BCR–ABL-negative cells at therapeutically relevant concentrations, even merely additively acting imatinib-containing combinations as shown in CML cells should exert more selective toxicity on the leukaemic cell clone as compared with either chemo- or radio-

therapy alone. Synergistic combinations exert this effect even stronger.

Total body irradiation (TBI) or busulfan are contained in marrow ablative conditioning regimens and treosulfan (Loeb, 1964) is being tested in clinical trials in toxicity-reduced conditioning (Casper *et al*, 2000; Ploemacher *et al*, 2000). TBI/cyclophosphamide and busulfan/cyclophosphamide regimens are used alternatively for pretransplant conditioning therapy. A synergism between imatinib and mafosfamide, an active form of cyclophosphamide representing an essential part of the conditioning therapy, has been shown previously (Topaly *et al*, 2001). It is controversial which conditioning regimen, TBI/cyclophosphamide or busulfan/cyclophosphamide, should be preferred for conditioning in the clinical setting. For treatment of BCR–ABL-positive acute lymphoblastic leukaemia or lymphatic blast crisis of CML, TBI-containing regimens are generally preferred due to the higher irradiation sensitivity of lymphoid blasts and due to the risk of relapse in the central nervous system (Grandos *et al*, 2000). In the myeloid blast crisis the choice of regimen is less uniform. Although TBI-containing conditioning seems to be more toxic as compared to its busulfan-containing counterpart, the speed of engraftment, event-free survival and relapse rate are not significantly different (Clift *et al*, 1994). We hypothesise that in the autologous (Pigneux *et al*, 1999) and possibly in the allogeneic transplantation setting TBI/cyclophosphamide + imatinib based on their synergistic effects could produce superior disease-free survival, even though risks and benefits should be thoroughly evaluated in future clinical trials. Treosulfan is employed alternatively to busulfan in conditioning regimens (Casper *et al*, 2000) and could also be supplemented with imatinib in CML patients. As a next step of preclinical testing, these combinations can be assessed in a 'mock-conditioning' scenario, i.e. in a mixed population of BCR–ABL-positive and -negative cells.

In conclusion, this is the first report on synergistic interaction between imatinib and γ -irradiation. It highlights the potential of combination treatment of CML employing specific BCR–ABL tyrosine kinase targeting and provides a rationale for the design of future conditioning protocols for stem cell transplantation in CML.

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REFERENCES

- Carella AM, Lerma E, Corsetti MT, Dejana A, Basta P, Vassallo F, Abate M, Soracco M, Benvenuto F, Figari O, Podesta M, Piaggio G, Ferrara R, Sessarego M, Parodi C, Pizzuti M, Rubagotti A, Occhini D, Frassoni F (1999) Autografting with philadelphia chromosome-negative mobilized hematopoietic progenitor cells in chronic myelogenous leukaemia. *Blood* **93**: 1534–1539
- Casper J, Wilhelm S, Steiner B, Wolff D, Hammer U, Wegener R, Kleine H-D, Freund M (2000) Toxicity-reduced conditioning with treosulfan and fludarabine prior to allogeneic blood stem cell transplantation. *Blood* **96**:298b
- Chou TC, Talalay P (1984) Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. *Adv Enzyme Regul* **22**: 27–55
- Clift RA, Buckner CD, Thomas ED, Bensinger WI, Bowden R, Bryant E, Deeg HJ, Doney KC, Fisher LD, Hansen JA (1994) Marrow transplantation for chronic myeloid leukaemia: a randomized study comparing cyclophosphamide and total body irradiation with busulfan and cyclophosphamide. *Blood* **84**: 2036–2043
- Deininger MW, Vieira S, Mendiola R, Schultheis B, Goldman JM, Melo JV (2000) BCR–ABL tyrosine kinase activity regulates the expression of multiple genes implicated in the pathogenesis of chronic myeloid leukaemia. *Cancer Res* **60**: 2049–2055
- Druker BJ, Tamura S, Buchdunger E, Ohno S, Segal GM, Fanning S, Zimmermann J, Lydon NB (1996) Effects of a selective inhibitor of the Abl tyrosine kinase on the growth of bcr/abl positive cells. *Nat Med* **2**: 561–566

- Druker BJ, Sawyers CL, Kantarjian H, Resta DJ, Reese SF, Ford JM, Capdeville R, Talpaz M (2001a) Activity of a Specific Inhibitor of the BCR-ABL Tyrosine Kinase in the Blast Crisis of Chronic Myeloid Leukemia and Acute Lymphoblastic Leukemia with the Philadelphia Chromosome. *N Engl J Med* **344**: 1038–1042
- Druker BJ, Talpaz M, Resta DJ, Peng B, Buchdunger E, Ford JM, Lydon NB, Kantarjian H, Capdeville R, Ohno-Jones S, Sawyers CL (2001b) Efficacy and safety of a specific inhibitor of the BCR-ABL tyrosine kinase in chronic myeloid leukaemia. *N Engl J Med* **344**: 1031–1037
- Druker BJ, Kantarjian HM, Talpaz M, Paquette R, Mauro MJ, Rosamilia M, Zoellner U, Resta D, Capdeville R, Sawyers CL (2001c) A Phase I Study of Gleevec (Imatinib Mesylate) Administered Concomitantly with Cytosine Arabinoside (Ara-C) in Patients with Philadelphia Positive Chronic Myeloid Leukemia (CML). *Blood* **98**: 845a–846a
- Faderl S, Talpaz M, Estrov Z, O'Brien S, Kurzrock R, Kantarjian HM (1999) The biology of chronic myeloid leukaemia. *N Engl J Med* **341**: 164–172
- Finlay GJ, Wilson WR, Baguley BC (1986) Comparison of in vitro activity of cytotoxic drugs towards human carcinoma and leukaemia cell lines. *Eur J Cancer Clin Oncol* **22**: 655–662
- Fruehauf S, Hochhaus A (2001) A multicenter study to determine the efficacy and safety of the combination of STI571 (Glivec[®]) with mitoxantrone/etoposide and cytarabine in patients with chronic myeloid leukemia in myeloid blast crisis (CSTI571A-DE-01). Website: www.kompetenznetz-leukaemie.de/kn_home/Studien/Dokumente/114.pdf
- Goekbuget N, Arnold R, Buechner T, Ganser A, Ludwig WD, Maschmeyer G, Rieder H, Thiel E, Messerer D, Hoelzer D for the GMALL Study Group (2001) Intensification of induction and Consolidation Improves Only Subgroups of Adult ALL: Analysis of 1200 Patients in GMALL Study 05/93. *Blood* **98**: 802a
- Granados E, de La Camara R, Madero L, Diaz MA, Martin-Regueira P, Steegmann JL, Arranz R, Figuera A, Fernandez-Ranada JM (2000) Hematopoietic cell transplantation in acute lymphoblastic leukaemia: better long term event-free survival with conditioning regimens containing total body irradiation. *Haematologica* **85**: 1060–1067
- Gratwohl A, Hermans J, Goldman JM, Arcese W, Carreras E, Devergie A, Frassoni F, Gahrton G, Kolb HJ, Niederwieser D, Ruutu T, Vernant JP, de Witte T, Apperley J (1998) Risk assessment for patients with chronic myeloid leukaemia before allogeneic blood or marrow transplantation. Chronic Leukemia Working Party of the European Group for Blood and Marrow Transplantation. *Lancet* **352**: 1087–1092
- Gorre ME, Mohammed M, Ellwood K, Hsu N, Paquette R, Rao PN, Sawyers CL (2001) Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. *Science* **293**: 876–880
- Hochhaus A, Kreil S, Corbin A, La Rosee P, Lahaye T, Berger U, Cross NC, Linkesch W, Druker BJ, Hehlmann R, Gambacorti-Passerini C, Corneo G, D'Incali M (2001) Roots of clinical resistance to STI-571 cancer therapy. *Science* **293**: 2163
- Jacobson P, Park JJ, DeFor TE, Thrall M, Abel S, Krivit W, Peters C (2001) Oral busulfan pharmacokinetics and engraftment in children with Hurler syndrome and other inherited metabolic storage diseases undergoing hematopoietic cell transplantation. *Bone Marrow Transplant* **27**: 855–861
- Kano Y, Akutsu M, Tsunoda S, Mano H, Sato Y, Honma Y, Furukawa Y (2001) In vitro cytotoxic effects of a tyrosine kinase inhibitor STI571 in combination with commonly used antileukemic agents. *Blood* **97**: 1999–2007
- Kantarjian HM, Giles FJ, O'Brien SM, Talpaz M (1998) Clinical course and therapy of chronic myelogenous leukaemia with interferon-alpha and chemotherapy. *Hematol Oncol Clin North Am* **12**: 31–80
- Kasper B, Fruehauf S, Schiedmeier B, Buchdunger E, Ho AD, Zeller WJ (1999) Favorable therapeutic index of a p210(BCR-ABL)-specific tyrosine kinase inhibitor; activity on lineage-committed and primitive chronic myelogenous leukaemia progenitors. *Cancer Chemother Pharmacol* **44**: 433–438
- Körbling M, Dörken B, Ho AD, Pezzutto A, Hunstein W, Fliedner TM (1986) Autologous transplantation of blood-derived hemopoietic stem cells after myeloablative therapy in a patient with Burkitt's lymphoma. *Blood* **67**: 529–532
- Konopka JB, Watanabe SM, Witte ON (1984) An alteration of the human c-abl protein in K562 leukaemia cells unmasks associated tyrosine kinase activity. *Cell* **37**: 1035–1042
- Loeb V (1964) Dihydroxybusulfan (NSC-39069) in chronic myelocytic leukaemia and miscellaneous malignant neoplasms. *Cancer Chemother Rep* **42**: 39–43
- Mosmann T (1983) Rapid Colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assay. *J Immunol Methods* **65**: 55–63
- Norgaard JM, Langkjer ST, Palshof T, Clausen N, Pedersen B, Hokland P (1996) Relation of blast cell survival and proliferation to chemotherapy resistance in AML. *Br J Haematol* **93**: 888–897
- Nowell PC, Hungerford DA (1960) A minute chromosome in chronic granulocytic leukaemia. *Science* **132**: 1497–1501
- Pieters B, Huismans DR, Leyva A, Veerman AJP (1988) Adaptation of the rapid automated tetrazolium dye based (MTT) assay for chemosensitivity testing in childhood leukaemia. *Cancer Letters* **41**: 323–332
- Pigneux A, Faberes C, Boiron JM, Mahon FX, Cony-Makhoul P, Agape P, Lounici A, Bernard P, Bilhou-Nabera C, Bouzgarrou R, Marit G, Reiffers J (1999) Autologous stem cell transplantation in chronic myeloid leukaemia: a single center experience. *Bone Marrow Transplant* **24**: 265–270
- Ploemacher RE, Westerhof GR, Blokland I, Baumgart J, Down JN (2000) Treosulfan as an alternative conditioning agent in bone marrow transplantation. *Bone Marrow Transplant* **25**(Suppl 1): 421
- Rowley JD (1973) A new consistent chromosomal abnormality in chronic myelogenous leukaemia identified by quinacrine fluorescence and Giemsa staining. *Nature* **243**: 290–293
- Scheulen ME, Hilger RA, Oberhoff C, Casper J, Freund M, Josten KM, Bornhauser M, Ehniger G, Berdel WE, Baumgart J, Harstrick A, Bojko P, Wolf HH, Schindler AE, Seeber S (2000) Clinical phase I dose escalation and pharmacokinetic study of high-dose chemotherapy with treosulfan and autologous peripheral blood stem cell transplantation in patients with advanced malignancies. *Clin Cancer Res* **6**: 4209–4216
- Talpaz M, Silver RT, Druker B, Paquette R, Goldman JM, Reese SF, Capdeville R (2000) A phase II study of STI571 in adult patients with Philadelphia chromosome positive chronic myeloid leukaemia in accelerated phase. *Blood* **96**: 469a
- Thiesing JT, Ohno-Jones S, Kolibaba KS, Druker BJ (2000) Efficacy of STI571 an abl tyrosine kinase inhibitor, in conjunction with other antileukemic agents against BCR-ABL-positive cells. *Blood* **96**: 3195–3199
- Topaly J, Zeller WJ, Fruehauf S (2001) Synergistic activity of the new ABL-specific tyrosine kinase inhibitor STI571 and chemotherapeutic drugs on BCR-ABL-positive chronic myelogenous leukaemia cells. *Leukemia* **15**: 342–347
- van Dongen JJ, Seriu T, Panzer-Grumayer ER, Biondi A, Pongers-Willemsse MJ, Corral L, Stolz F, Schrappe M, Masera G, Kamps WA, Gadner H, van Wering ER, Ludwig WD, Basso G, de Bruijn MA, Cazzaniga G, Hettlinger K, van der Does-van den Berg A, Hop WC, Riehm H, Bartram CR (1998) Prognostic value of minimal residual disease in acute lymphoblastic leukaemia in childhood. *Lancet* **352**: 1731–1738