# *CCNE1* amplification is associated with aggressive potential in endometrioid endometrial carcinomas

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Received October 6, 2015; Accepted November 8, 2015

DOI: 10.3892/ijo.2015.3268

Abstract. The clinicopathological significance of amplification was investigated of the gene encoding cyclin E (CCNE1) and we assessed whether CCNE1 was a potential target in endometrioid endometrial carcinomas. CCNE1 amplification and CCNE1 or F-box and WD repeat domain-containing 7 (FBXW7) expression in endometrial endometrioid carcinoma was assessed by immunohistochemistry and fluorescence in situ hybridization. CCNE1 knockdown by small interfering RNA (siRNA) was used to assess the CCNE1 function. The results showed that CCNE1 amplification was present in 9 (8.3%) of 108 endometrial carcinomas. CCNE1 amplification was correlated with high histological grade (Grade 3; P=0.0087) and lymphovascular space invasion (P=0.0258). No significant association was observed between CCNE1 amplification and FIGO stage (P=0.851), lymph node metastasis (P=0.078), body mass index (P=0.265), deep myometrial invasion (P=0.256), menopausal status (P=0.289) or patient age (P=0.0817). CCNE1 amplification was significantly correlated with shorter progression-free and overall survival (P=0.0081 and 0.0073, respectively). CCNE1 protein expression or loss of FBXW7 expression in endometrial endometrioid carcinoma tended to be correlated with shorter progression-free and overall survival; however, this difference was not statistically significant. Multivariate analysis showed that CCNE1 amplification was an independent prognostic factor for overall survival but not for progression-free survival (P=0.0454 and 0.2175, respectively). Profound growth inhibition was observed in siRNA-transfected cancer cells with endogenous CCNE1 overexpression compared with that in cancer cells having low

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CCNE1 expression. *CCNE1* amplification was independent of p53, HER2, MLH1 and ARID1A expression but dependent on PTEN expression in endometrial carcinomas. These findings indicated that *CCNE1* amplification was critical for the survival of endometrial endometrioid carcinomas. Furthermore, the effects of *CCNE1* knockdown were dependent on the CCNE1 expression status, suggesting that CCNE1-targeted therapy may be beneficial for patients with endometrial endometrioid carcinoma having *CCNE1* amplification.

#### Introduction

Endometrial carcinomas are the most common gynecological malignancies and the fifth most common cancer in women worldwide (1). Endometrial carcinomas can be subdivided into two main categories: type I, estrogen-dependent endometrioid carcinoma; and type II, estrogen-independent non-endometrioid carcinoma (2,3).

Studies have shown that exposure to high levels of estrogen is an important risk factor for endometrial carcinomas (2). Endometrioid carcinoma represents ~80-85% of all endometrial malignancies. Previous studies have identified various genetic alterations in endometrioid carcinoma, including silencing of the phosphatase and tensin homolog deleted on chromosome 10 (*PTEN*) gene, microsatellite instability (MSI), and mutations in *K-ras* and/or *PIK3CA* (3-6).

While the majority of endometrial carcinomas are diagnosed at an early stage, resulting in a favorable prognosis, women diagnosed with advanced or recurrent disease have much lower survival rates and limited adjuvant treatment options. Over the past few decades, survival rates in patients with advanced disease have not improved sufficiently.

Our recent genome-wide sequencing analyses of all exons and transcriptomes in uterine serous carcinomas (USCs) showed that approximately half of all USC cases harbor either somatic mutations in F-box and WD repeat domaincontaining 7 (*FBXW7*) or gene amplification in *CCNE1* (encoding cyclin E1) (7). The cyclin E-FBXW7 pathway is thought to be one of the most important pathways in USC development (7).

FBXW7 is the substrate recognition component of the Skp1-Cul1-F-box (SCF) ubiquitin-ligase and is located within 4q32, a chromosomal region that is commonly deleted in

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*Key words:* cyclin E1, gene amplification, F-box and WD repeat domain-containing 7, endometrial carcinoma

cancers (8-10). FBXW7 acts as a tumor suppressor by targeting several oncogenic regulators of proliferation, growth and apoptosis for proteasomal degradation. These include cyclin E, c-MYC, Notch and MCL1 (11-14). Furthermore, lower expression of FBXW7 contributes to lymph node metastasis, tumor size and poor prognosis in gastric cancer (15).

DNA copy number alterations, including amplification, deletion, and aneuploidy in chromosomes, are the hallmarks of neoplasia (16). Amplification of chromosomal regions plays a critical role in tumor development. Increases in the copy numbers of oncogenes promote initiation and progression of a variety of solid tumors, whereas amplification of genes that modify or detoxify chemotherapeutic agents can lead to drug resistance and is associated with tumor recurrence (17,18). Well-known amplified oncogenes include c-myc, ERBB2, EGFR, AKT2, CCND1 and CCNE1. Studies on these genes have not only provided new insights into the mechanisms of cancer development but also revealed significant translational implications. For example, ERBB2 and EGFR are molecular targets of the humanized antibodies trastuzumab (Herceptin) and matuzumab, respectively, which are used in the treatment of breast and lung cancer.

Although CCNE1 amplifications and FBXW7 mutations are frequently present in serous-type endometrial cancer, the clinicopathological and prognostic roles of these modifications in endometrioid-type endometrial cancer are still unclear. Inactivating mutations in tumor suppressors could participate not only in tumor initiation, but also in tumor progression and response to therapy. In the present study, we examined the prognostic and clinicopathological significance of CCNE1 amplification/expression and loss of FBXW7 expression in endometrial carcinoma by investigating the relationship between CCNE1 amplification/expression or FBXW7 expression and various clinicopathological variables in endometrial carcinoma. In addition, we compared phenotypes in cultured endometrial carcinoma cells with variations in CCNE1 expression levels after transfection with small interfering RNA (siRNA) targeting CCNE1.

#### Materials and methods

Tissue samples. Formalin-fixed, paraffin-embedded tissue samples from 108 endometrioid-type endometrial carcinomas were used in this study. Samples were obtained from the Department of Obstetrics and Gynecology at the Shimane University Hospital. Diagnosis was based on conventional morphological examination of hematoxylin and eosin (H&E)stained sections. Cancer patients had received appropriate therapy at Shimane University Hospital between January 1998 and August 2010. Acquisition of tissue specimens and clinical information was approved by the institutional review board of Shimane University. Endometrial carcinomas were classified according to the surgical staging system of the International Federation of Gynecology and Obstetrics (FIGO). Invasive carcinomas were divided by stage, with 76 patients having stage I disease, 9 patients having stage II disease, 17 patients having stage III disease, and 6 patients having stage IV disease. All tumors were classified histologically according to the World Health Organization criteria. The median patient age was 60 years (range, 26-85 years). Primary surgeries included total hysterectomy in 39 patients, modified radical hysterectomy in 60 patients and radical hysterectomy in 9 patients. Systemic retroperitoneal lymphadenectomy was performed in ~90% of patients. Patients with risk factors for recurrence (e.g., deep myometrial invasion, cervical invasion, lymph node metastasis, lymphovascular space invasion and positive peritoneal cytology) underwent postoperative adjuvant external radiotherapy and/or 4-6 cycles of CAP therapy (cisplatin 100 mg/m<sup>2</sup>, doxorubicin 40 mg/m<sup>2</sup>, cyclophosphamide 500 mg/m<sup>2</sup>) or TC therapy (paclitaxel 175 mg/m<sup>2</sup>, carboplatin according to Chatelut's formula - area under the curve = 5 mg/m<sup>2</sup>). All 108 patients were included in the survival analysis. The follow-up period ranged from 5 to 139 months, with a median of 52 months.

*Fluorescence in situ hybridization (FISH).* BAC clones (RP11-345J21 and CTD-3005A16) containing the genomic sequences of the 19q12 amplicon at 15.00-15.25 Mb were purchased from Bacpac Resources Center (Children's Hospital, Oakland, CA, USA) and Invitrogen (Carlsbad, CA, USA). Bac clones located at Chr2q11.2 (e.g., RP11-127K18 and RP11-629A22) or at Ch19P12 (CTD-2518O18) were used to generate reference probes.

RP11-127K18, RP11-629A22 and CTD-2518O18 were labeled by nick translation with biotin-dUTP; RP11-345J21 and CTD-3005A16 were labeled similarly with digoxigenindUTP. To detect biotin-labeled and digoxigenin-labeled signals, slides were first incubated with FITC-avidin (Vector Laboratories, Burlingame, CA, USA) and an anti-digoxigenin mouse antibody (Roche Molecular Biochemicals, Mannheim, Germany). Slides were subsequently incubated with a biotinylated anti-avidin antibody (Vector Laboratories) and tetramethylrhodamine B isothiocyanate (TRITC)-conjugated rabbit anti-mouse antibodies (Sigma, St. Louis, MO, USA). The final incubation was with FITC-avidin and TRITC-conjugated goat anti-rabbit antibodies (Sigma). Slides were counterstained with 4',6'-diamidino-2-phenylindole stain (Sigma).

Fluorescence *in situ* hybridization (FISH) signals were evaluated with an Olympus fluorescence microscope BX41 (Olympus, Tokyo, Japan) by two individuals who were blinded to the treatment history of each patient. Separate narrow band pass filters were used for detection of tetramethylrhodamine B isothiocyanate, FITC, and 4',6'-diamidino-2-phenylindole staining signals. Using a x60 objective lens, ~100 tumor cells were examined for each specimen, and the numbers of fluorescent signals within tumor cells from the *CCNE1* gene BAC probe and chromosome 2q11.2 or 19p12 reference BAC probe were recorded. Amplification of *CCNE1* was defined as the ratio of *CCNE1* BAC probe signals to chromosome 2 or chromosome 19 centromeric reference BAC probe signals of 2:1 or more.

*Immunohistochemistry*. Paraffin-embedded tissues were organized into tissue microarrays, which were made by removing tumor cores (3 mm in diameter) from each block. Selection of the area to core was made by a gynecologic oncologist (K.N.) and pathology technician (K.I.) and was based on a review of the H&E slides.

Expression of CCNE1 and FBXW7 was assessed by immunohistochemistry and/or western blot analysis. The antibodies used in the present study were mouse monoclonal antibodies targeting CCNE1 (Zymed Laboratories-Invitrogen, Carlsbad, CA, USA) and mouse monoclonal antibodies targeting FBXW7 (Abcam, Cambridge, MA, USA). Immunohistochemistry studies for CCNE1 and FBXW7 were performed on tissue microarrays at a dilution of 1:250 or 1:50 followed by detection with the EnVision+ system using the peroxidase method (Dako, Carpinteria, CA, USA). After antigen retrieval in sodium citrate buffer, slides were incubated with antibodies overnight at 4°C. Slides for all samples were evaluated with a light microscope by two researchers; the researchers were blinded to the clinicopathological factors. The antibody staining intensity was then analyzed in glands using the HSCORE (19). The modified HSCORE was calculated as follows: HSCORE =  $\Sigma P_i(i)$ , where *i* is the intensity of staining (0, undetectable, 1, weakly positive, 2, moderately positive, 3, intensely positive) and  $P_i$  is a score based on the percentage of stained cells for each intensity, varying from 0 to 100%.

Immunohistochemistry methods for ARID1A, PTEN, MLH1, p53, HER2 ER and progesterone receptor (PR), as well as evaluation criteria, have been previously described (20,21).

*Cell culture and cell lines*. The human endometrial carcinoma cell lines HEC251 (endometrioid carcinoma), HEC50B (endometrioid carcinoma G3), and HEC108 (endometrioid carcinoma) were obtained from the Japanese Health Science Research Resources Bank (Osaka, Japan). HHUA (endometrioid carcinoma) and JHUC-1 (endometrial carcinosarcoma) cells were obtained from Riken Bioresource Center (Ibaragi, Japan).

Western blot analysis. Cell lysates were prepared by dissolving cell pellets in Laemmli sample buffer (Bio-Rad Laboratories, Hercules, CA, USA) supplemented with 5% beta-mercaptoethanol (Sigma). Western blot analysis was performed on endometrial carcinoma cell lines, including HEC251, HEC50B, HHUA, JHUC-1 and HEC108 cells. Similar amounts of total protein from each lysate were loaded and separated on 10% Tris-Glycine-SDS polyacrylamide gels (Novex, San Diego, CA, USA) and electroblotted to Millipore Immobilon-P polyvinylidene difluoride membranes (Millipore, Bedford, MA, USA). Membranes were probed with anti-CCNE1 antibodies (1:1,000 dilution; Zymed Laboratories-Invitrogen) followed by peroxidase-conjugated anti-mouse or anti-rabbit immunoglobulin (1:20,000 dilution). The same membrane was probed with antibodies targeting GAPDH (1:10,000 dilution; Cell Signaling Technology, Beverly, MA, USA) as a loading control. Western blots were developed by chemiluminescence (Pierce, Rockford, IL, USA).

siRNA-mediated knockdown of CCNE1 gene expression. Two siRNAs that targeted CCNE1 were designed with the following sense sequences: UCAGUUGACAGUGUACAAUGCCUTT and UGACUUACAUGAAGUGCUACUGCCG. Control siRNA (luciferase siRNA) was purchased from Integrated DNA Technologies (Coralville, IA, USA). Cells were seeded onto 96-well plates and transfected with siRNAs using Oligofectamine (Invitrogen). Cell numbers were determined indirectly using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay 72 h after transfection (22). *Cell proliferation assay.* Cells were seeded in 96-well plates at a density of 3,000 cells/well. Cell numbers were determined indirectly using MTT assays (22). Data were expressed as the mean  $\pm$  standard deviation (SD) from triplicate determinations.

Statistical methods for clinical correlations. Overall survival was calculated from the date of diagnosis to the date of death or last follow-up. Patients with and without CCNE1 or FBXW7 expression or *CCNE1*-amplified tumors had similar ages and performance status distributions. Statistical significance was determined by the log-rank test. Data were censored when patients were lost to follow-up.

#### Results

Relationships among CCNE1 amplification, CCNE1 protein expression, FBXW7 protein expression and clinicopathological factors. As shown in Table I, the frequency of CCNE1 amplification in high-grade carcinomas (G3; 20.6%: 6/29) was significantly higher than that in low-grade tumors (G1, G2; 3.8%: 3/79; P=0.0087). CCNE1 amplification was also correlated with lymphovascular space invasion (P=0.0258). There were no significant correlations between CCNE1 amplification and FIGO stage (P=0.851), lymph node metastasis (P=0.078), patient age (P=0.0817), body mass index (P=0.265), deep myometrial invasion (P=0.256) or menopausal status (P=0.289; Table I).

Patients were stratified into one of two groups depending on the median CCNE1 immunohistochemical HSCORE. The relationships between CCNE1 protein expression and clinicopathological factors are shown in Table I. CCNE1 protein expression was significantly correlated with highgrade carcinoma (P=0.0169), lymphovascular space invasion (P=0.0195), postmenopausal status (P=0.0281), and patient age (P=0.0209). There was no significant correlation between CCNE1 protein expression and FIGO stage (P=0.0601), body mass index (P=0.5148), lymph node metastasis (P=0.7808), or deep myometrial invasion (P=0.1651).

Negative expression of FBXW7 (FBXW7 immunohistochemical HSCORE = 0) was observed in 62% (63/101) of the analyzed tumors. Patients were stratified into one of two groups depending on the status of the negative FBXW7 immunohistochemical HSCORE. The relationships between FBXW7 protein expression and clinicopathological factors are shown in Table I. Negative FBXW7 protein expression was significantly correlated with deep myometrial invasion (P=0.0144), lymphovascular space invasion (P=0.0414) and patient age (P=0.038). There was no significant correlation between negative FBXW7 protein expression and FIGO stage (P=0.2439), tumor grade (P=0.228), lymph node metastasis (P=0.4805), postmenopausal status (P=0.882) or body mass index (P=0.162).

*Effects of CCNE1 amplification or CCNE1 protein expression on progression-free survival.* Next, we evaluated the prognostic relevance of CCNE1 amplification or *CCNE1* protein expression in terms of progression-free survival. Kaplan-Meier estimates of progression-free/overall survival are plotted in Fig. 2. Amplification of *CCNE1* correlated with shorter progression-free survival in patients with endometrial

|                               |          | CCNE1 gene amplification |          |         | CCNE1<br>protein<br>expression |      |         | FBXW7 protein expression |          |         |
|-------------------------------|----------|--------------------------|----------|---------|--------------------------------|------|---------|--------------------------|----------|---------|
| Factors                       | Patients | Negative                 | Positive | P-value | Low                            | High | P-value | Negative                 | Positive | P-value |
| FIGO stage                    |          |                          |          |         |                                |      |         |                          |          |         |
| Ι                             | 75       | 69                       | 6        | 0.851   | 42                             | 33   | 0.0601  | 41                       | 34       | 0.2439  |
| II, III, IV                   | 33       | 30                       | 3        |         | 12                             | 21   |         | 22                       | 11       |         |
| Grade                         |          |                          |          |         |                                |      |         |                          |          |         |
| G1, G2                        | 79       | 76                       | 3        | 0.0087  | 45                             | 34   | 0.0169  | 45                       | 34       | 0.228   |
| G3                            | 29       | 23                       | 6        |         | 9                              | 20   |         | 18                       | 11       |         |
| Lymph node metastasis         |          |                          |          |         |                                |      |         |                          |          |         |
| Negative                      | 93       | 87                       | 6        | 0.078   | 47                             | 46   | 0.7808  | 53                       | 40       | 0.4805  |
| Positive                      | 15       | 12                       | 3        |         | 7                              | 8    |         | 10                       | 5        |         |
| Depth (myometrial invasion)   |          |                          |          |         |                                |      |         |                          |          |         |
| a, b                          | 67       | 63                       | 4        | 0.256   | 37                             | 30   | 0.1651  | 33                       | 34       | 0.0144  |
| c                             | 41       | 36                       | 5        |         | 17                             | 24   |         | 30                       | 11       |         |
| Lymphovascular space invasion |          |                          |          |         |                                |      |         |                          |          |         |
| Negative                      | 62       | 60                       | 7        | 0.0258  | 37                             | 25   | 0.0195  | 31                       | 31       | 0.0414  |
| Positive                      | 46       | 39                       | 7        |         | 17                             | 29   |         | 32                       | 14       |         |
| Menopause                     |          |                          |          |         |                                |      |         |                          |          |         |
| Peri, pre                     | 28       | 27                       | 1        | 0.289   | 19                             | 9    | 0.0281  | 16                       | 12       | 0.882   |
| Post                          | 80       | 72                       | 8        |         | 35                             | 45   |         | 47                       | 33       |         |
| Body mass index               |          |                          |          |         |                                |      |         |                          |          |         |
| <25                           | 79       | 71                       | 8        | 0.265   | 41                             | 38   | 0.5148  | 47                       | 32       | 0.162   |
| ≥25                           | 29       | 28                       | 1        |         | 13                             | 16   |         | 16                       | 13       |         |
| Age (years)                   |          |                          |          |         |                                |      |         |                          |          |         |
| <60                           | 54       | 52                       | 2        | 0.0817  | 33                             | 21   | 0.0209  | 31                       | 23       | 0.038   |
| ≥60                           | 54       | 47                       | 7        |         | 21                             | 33   |         | 32                       | 22       |         |

| Table I. | Association bet | ween CCNE1 g | gene amplification | and clinocopathe | ological factor | rs in patien | ts with end | ometrial can | cer |
|----------|-----------------|--------------|--------------------|------------------|-----------------|--------------|-------------|--------------|-----|
|          |                 | 4            | , .                |                  |                 |              |             |              |     |

endometrioid carcinomas. A total of 108 patients were diagnosed at stages I-IV. Among them, the 9 patients with CCNE1 amplifications had shorter progression-free survival than those without gene amplification (P=0.0081; log-rank test; Fig. 2A). Univariate analysis demonstrated that deep myometrial invasion (P=0.0008; log-rank test), high-grade histological subtype (P<0.0001; log-rank test), lymph node metastasis (P<0.0001; log-rank test), lymphovascular space invasion (P=0.001; logrank test) and *CCNE1* amplification (P=0.0081; log-rank test) correlated with shorter progression-free survival. CCNE1 HSCORE tended to influence progression-free survival, but this relationship was not statistically significant (P=0.3148; Fig. 2B). We excluded lymph node metastasis and lymphovascular space invasion from multivariate analysis because these two factors are similar events in the clinical setting and to prevent any issues with statistical analyses. Therefore, for multivariate analysis, we chose FIGO stage, tumor grade, depth of myometrial invasion and CCNE1 amplification. When the data were stratified for multivariate analysis, high-grade

histological subtype and deep myometrial invasion remained significantly associated with shorter progression-free survival (P=0.0003 and P=0.0142, respectively; Table II).

*Effects of CCNE1 amplification or CCNE1 protein expression on overall survival.* Next, we examined the prognostic relevance of *CCNE1* amplification or CCNE1 protein expression in terms of overall survival. Kaplan-Meier estimates of progression-free/overall survival are plotted in Fig. 3. Amplification of *CCNE1* correlated with shorter overall survival in patients with endometrial endometrioid carcinomas treated. A total of 108 patients were diagnosed at stages I-IV. Among them, the 9 patients with *CCNE1* amplification had shorter overall survival than those without CCNE1 amplification (P=0.0073; log-rank test; Fig. 2C). Univariate analysis demonstrated that FIGO stage III or IV (P=0.035; log-rank test), highgrade histological subtype (P<0.0001; log-rank test), lymph node metastasis (P<0.0001; log-rank test), lymph node

| Table II. Univariate analysis of progression-free prognostic factors in patients with endometrial carcinoma. | <b>T</b> 11 <b>T T T T T T T T T T</b> | • •              |                    | • .• .      | • . 1   | 1 . 1 1 1           |
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|  | rable II. Ollivariate analysis of      | progression nee  | prognostic ractors | in patients | with on | uomennai caremonna. |

| Factors                       | Patients | Univariate<br>hazard ratio | 95% CI    | P-value | Multivariate<br>hazard ratio | 95% CI   | P-value |
|-------------------------------|----------|----------------------------|-----------|---------|------------------------------|----------|---------|
| FIGO stage                    |          |                            |           |         |                              |          |         |
| I                             | 75       |                            |           |         |                              |          |         |
| II, III, IV                   | 33       | 2.8                        | 1.0-7.7   | 0.0532  | NA                           | NA       | NA      |
| Grade                         |          |                            |           |         |                              |          |         |
| G1, G2                        | 79       |                            |           |         |                              |          |         |
| G3                            | 29       | 27.1                       | 6.0-122.1 | <0.0001 | 18.2                         | 3.8-86.8 | 0.0003  |
| Lymph node metastasis         |          |                            |           |         |                              |          |         |
| Negative                      | 93       |                            |           |         |                              |          |         |
| Positive                      | 15       | 9.7                        | 3.4-27.7  | <0.0001 | NA                           | NA       | NA      |
| Depth (myometrial invasion)   |          |                            |           |         |                              |          |         |
| a, b                          | 67       |                            |           |         |                              |          |         |
| c                             | 41       | 8.824                      | 2.5-31.4  | 0.0008  | 5.3                          | 1.4-19.8 | 0.0142  |
| Lymphovascular space invasion |          |                            |           |         |                              |          |         |
| Negative                      | 62       |                            |           |         |                              |          |         |
| Positive                      | 46       | 30.5                       | 4.0-233.6 | 0.001   | NA                           | NA       | NA      |
| Menopause                     |          |                            |           |         |                              |          |         |
| Peri, pre                     | 28       |                            |           |         |                              |          |         |
| Post                          | 80       | 5.5                        | 0.7-42.0  | 0.0991  | NA                           | NA       | NA      |
| Body mass index               |          |                            |           |         |                              |          |         |
| <25                           | 79       |                            |           |         |                              |          |         |
| ≥25                           | 29       | 1.1                        | 0.3-3.5   | 0.8597  | NA                           | NA       | NA      |
| Age (years)                   |          |                            |           |         |                              |          |         |
| <60                           | 54       |                            |           |         |                              |          |         |
| ≥60                           | 54       | 2.9                        | 0.9-9.1   | 0.069   | NA                           | NA       | NA      |
| CCNE1 FISH                    |          |                            |           |         |                              |          |         |
| Normal                        | 9        |                            |           |         |                              |          |         |
| Amp                           | 99       | 4.7                        | 1.5-14.8  | 0.0081  | 2.2                          | 0.6-7.4  | 0.2175  |
| CCNE1 immunostaing            |          |                            |           |         |                              |          |         |
| Low                           | 54       |                            |           |         |                              |          |         |
| High                          | 54       | 1.7                        | 0.6-4.8   | 0.3148  | NA                           | NA       | NA      |
| FBXW7 immunostaing            |          |                            |           |         |                              |          |         |
| Positive                      | 45       |                            |           |         |                              |          |         |
| Negative                      | 63       | 1.9                        | 0.6-5.5   | 0.2637  | NA                           | NA       | NA      |

invasion (P=0.0011; log-rank test), and *CCNE1* amplification (P=0.0073; log-rank test) were correlated with shorter overall survival. High CCNE1 HSCOREs tended to be associated with poor overall survival; however, this relationship was not significant (P=0.2205; Fig. 2D). As described in the previous section, lymph node metastasis and lymphovascular space invasion were excluded from multivariate analysis. Therefore, for multivariate analysis, we chose FIGO stage, tumor grade, depth of myometrial invasion and *CCNE1* amplification. When data were stratified for multivariate analysis, high-grade subtype, deep myometrial invasion and *CCNE1* amplification

remained significantly associated with shorter overall survival (P=0.0006, P=0.0252 and P=0.0454, respectively; Table III).

*Effects of FBXW7 protein expression on disease-free/overall survival.* Because FBXW7 is the substrate recognition component of the Skp1-Cul1-F-box (SCF) ubiquitin-ligase, which targets CCNE1, we next analyzed whether loss of FBXW7 immunostaining was related to poor survival and whether there was a positive relationship between loss of FBXW7 protein expression and CCNE1 protein expression. Immunoreactivity for FBXW7 was detected in tumor cell

| Factors                       | Patients | Univariate<br>hazard ratio | 95% CI    | P-value  | Multivariate<br>hazard ratio | 95% CI   | P-value |
|-------------------------------|----------|----------------------------|-----------|----------|------------------------------|----------|---------|
| FIGO stage                    |          |                            |           |          |                              |          |         |
| I                             | 75       |                            |           |          |                              |          |         |
| II, III, IV                   | 33       | 3                          | 1.1-8.4   | 0.035    | 1.4                          | 0.4-4.2  | 0.5957  |
| Grade                         |          |                            |           |          |                              |          |         |
| G1, G2                        | 79       |                            |           |          |                              |          |         |
| G3                            | 29       | 26.7                       | 5.9-120.3 | <0.0001  | 15.7                         | 3.2-76.1 | 0.0006  |
| Lymph-node metastasis         |          |                            |           |          |                              |          |         |
| Negative                      | 93       |                            |           |          |                              |          |         |
| Positive                      | 15       | 11.7                       | 4.0-33.8  | < 0.0001 | NA                           | NA       | NA      |
| Depth (myometrial invasion)   |          |                            |           |          |                              |          |         |
| a, b                          | 67       |                            |           |          |                              |          |         |
| c                             | 41       | 8.8                        | 2.5-31.4  | 0.0008   | 4.6                          | 1.2-17.4 | 0.0252  |
| Lymphovascular space invasion |          |                            |           |          |                              |          |         |
| Negative                      | 62       |                            |           |          |                              |          |         |
| Positive                      | 46       | 29.4                       | 3.9-224.3 | 0.0011   | NA                           | NA       | NA      |
| Menopause                     |          |                            |           |          |                              |          |         |
| Peri, pre                     | 28       |                            |           |          |                              |          |         |
| Post                          | 80       | 5.3                        | 0.7-40.2  | 0.1081   | NA                           | NA       | NA      |
| Body mass index               |          |                            |           |          |                              |          |         |
| <25                           | 79       |                            |           |          |                              |          |         |
| ≥25                           | 29       | 1.1                        | 0.4-3.6   | 0.883    | NA                           | NA       | NA      |
| Age (years)                   |          |                            |           |          |                              |          |         |
| <60                           | 54       |                            |           |          |                              |          |         |
| ≥60                           | 54       | 2.9                        | 0.9-9.1   | 0.067    | NA                           | NA       | NA      |
| CCNE1 FISH                    |          |                            |           |          |                              |          |         |
| Normal                        | 9        |                            |           |          |                              |          |         |
| Amp                           | 99       | 4.8                        | 1.5-15.2  | 0.0073   | 3.8                          | 1.0-14.0 | 0.0454  |
| CCNE1 immunostaing            |          |                            |           |          |                              |          |         |
| Low                           | 54       |                            |           |          |                              |          |         |
| High                          | 54       | 1.9                        | 0.7-5.4   | 0.2205   | NA                           | NA       | NA      |
| FBXW7 immunostaing            |          |                            |           |          |                              |          |         |
| Positive                      | 45       |                            |           |          |                              |          |         |
| Negative                      | 63       | 1.9                        | 0.7-5.7   | 0.2307   | NA                           | NA       | NA      |

| Table III. Univariate analy | ysis of overall progno | stic factors in patients | with endometrial carcinoma. |
|-----------------------------|------------------------|--------------------------|-----------------------------|

nuclei (Fig. 1). The mean HSCORE of FBXW7 was 46 (range, 0-300). The CCNE1 immunohistochemical HSCORE was not correlated with the FBXW7 immunohistochemical HSCORE (data not shown). Negative expression of FBXW7 (FBXW7 immunohistochemical HSCORE = 0) was observed in 58.3% (63/108) of the analyzed tumors. Patients were stratified into one of two groups depending on the status of negative FBXW7 immunohistochemical HSCORE. Kaplan-Meier estimates of progression-free/overall survival are plotted in Fig. 2. Negative FBXW7 expression tended to correlate with shorter overall/progression-free survival in patients with endometrial

endometrioid carcinoma; however, this relationship was not statistically significant (Fig. 2E and F).

Relationship between CCNE1 amplification and PTEN, p53, HER2, MLH1 or ARID1A expression in endometrial endometrioid carcinoma. We previously performed immunohistochemical analysis to evaluate the status of PTEN, p53, HER2, MLH1 and ARID1A expression among analyzed tumor samples (20). We then investigated the correlations between the expression of these molecules and CCNE1 amplification (Table IV). No significant correlations were observed between



Figure 1. Dual-color fluorescence *in situ* hybridization (FISH) was used to detect amplification of the *CCNE1* gene in endometrial endometrioid carcinomas. (A) FISH analysis revealed a homogeneously stained region in the carcinoma portion with *CCNE1* amplification. (B and D) A case of endometrial endometrioid carcinoma with positive immunoreactivity for CCNE1 and FBXW7 in endometrial carcinoma tissues. (C and E) A case with negative CCNE1 and FBXW7 staining in endometrial carcinoma tissues.

Table IV. Association between *CCNE1* gene amplification and status of p53, hMLH1, Her2, PTEN and ARID1A in patients with endometrial cancer.

|          |          | CCI<br>amp |               |         |
|----------|----------|------------|---------------|---------|
| Factors  | Patients | Normal     | Amplification | P-value |
| p53      |          |            |               |         |
| Negative | 58       | 55         | 3             | 0.2005  |
| Positive | 50       | 44         | 6             |         |
| hMLH1    |          |            |               |         |
| Negative | 53       | 48         | 5             | 0.6846  |
| Positive | 55       | 51         | 4             |         |
| Her2     |          |            |               |         |
| Low      | 40       | 36         | 4             | 0.6308  |
| High     | 68       | 63         | 5             |         |
| PTEN     |          |            |               |         |
| Negative | 27       | 25         | 2             | 0.041   |
| Positive | 81       | 74         | 7             |         |
| ARID1A   |          |            |               |         |
| Negative | 27       | 26         | 1             | 0.3149  |
| Positive | 81       | 73         | 8             |         |

*CCNE1* amplification and the expression of these proteins, except for positive PTEN expression (P=0.041).

Effects of CCNE1 knockdown on endometrial endometrioid carcinoma in vitro. A panel of endometrial carcinoma cell lines was analyzed for CCNE1 amplification and protein expression status (Fig. 3A). However, there were no CCNE1 amplification cell lines in the tested cells, as measured by FISH analysis (data not shown). Transfection with CCNE1 siRNA significantly reduced CCNE1 protein expression compared with control siRNA transfection (Fig. 3B). However, CCNE1 expression status correlated with the growth inhibition induced by CCNE1 siRNA (Fig. 3C). Reduction of CCNE1 expression significantly inhibited cell growth in CCNE1-overexpressing cells, including JHUC-1 and HEC108 cells. In contrast, transfection with CCNE1 siRNA did not affect cell growth in HEC251, HEC50B and HHUA cells, which exhibit moderate or minimal CCNE1 expression, suggesting that endometrial carcinomas with CCNE1 overexpression are more dependent on activation of the CCNE1-related pathway for cell proliferation and survival than those without CCNE1 expression.

## Discussion

Notably, we found that there was a significantly higher frequency of *CCNE1* amplification in high-grade endometrial



Figure 2. *CCNE1* amplification is correlated with shorter progression-free/overall survival in patients with endometrial endometrioid carcinoma. (A) Kaplan-Meier survival analysis showed that *CCNE1* amplification (solid line, n=9) was associated with shorter progression-free survival than absence of *CCNE1* amplification (dashed line, n=99) (P=0.0081, log-rank test). CCNE1 overexpression was not significantly associated with progression-free/overall survival in patients with endometrial endometrioid carcinoma. (B) Kaplan-Meier survival analysis showed that a high CCNE1 expression (solid line, n=44) was associated with shorter relapse-free survival than low *CCNE1* expression; however, the difference was not statistically significant (dashed line, n=54; P=0.3148, log-rank test). (C) Kaplan-Meier survival analysis showed that *CCNE1* amplification (solid line, n=9) was associated with shorter overall survival than absence of *CCNE1* amplification (dashed line, n=99; P=0.0073, log-rank test). (D) Kaplan-Meier survival analysis showed that *CNE1* expression (solid line, n=54) was associated with shorter overall survival than low *CCNE1* expression; however, this difference was not statistically significant (dashed line, n=54; P=0.2105, log-rank test). Negative FBXW7 expression was not significantly associated with progression-free/overall survival than low *CCNE1* expression; however, this difference was not statistically significant (dashed line, n=54; P=0.2205, log-rank test). Negative FBXW7 expression was not significantly associated with progression-free/overall survival analysis showed that negative FBXW7 expression; however, this difference was not statistically significant (dashed line, n=54; P=0.3148, log-rank test). (F) Kaplan-Meier survival analysis showed that negative FBXW7 expression (solid line, n=63) was associated with shorter progression-free survival analysis showed that negative FBXW7 expression; however, this difference was not statistically significant (dashed line, n=54; P=0.3148, log-rank t

endometrioid carcinomas compared with that in low-grade endometrial endometrioid carcinomas. This suggested that high-grade and low-grade tumors may be distinguished based on characteristic genetic alterations. Additionally, this observation further supports the hypothesis that endometrial carcinoma arises from multiple pathways (7,23). In this model, high-grade and low-grade carcinomas develop independently from one another and are characterized by different molecular genetic changes and gene expression profiles (7,23).



Figure 3. Western blot analysis. (A) Western blot analysis showed that CCNE1 protein expression was higher in JHUC-1 and HEC108 cells than in other cell lines. (B) Western blot analysis showed that CCNE1 protein expression was significantly decreased in *CCNE1* siRNA-transfected cells compared with control siRNA-transfected cells. (C) Effects of *CCNE1* siRNA on cell proliferation. Cell numbers were determined at 72 h after transfection with *CCNE1* siRNA or control siRNA. Endometrial carcinoma cells with CCNE1 overexpression were more sensitive to growth inhibition by *CCNE1* siRNA than cells without CCNE1 overexpression. The mean and SD were obtained from three experiments.

Our findings were consistent with those of a previous report showing a positive correlation between CCNE1 immunostaining and tumor grade (24). Taken together, these data, including our current FISH and immunostaining analyses, suggested that alterations in the *CCNE1* gene could be a specific event in high-grade endometrial endometrioid carcinoma.

Based on our combined FISH and immunohistochemical analysis, we found that *CCNE1* was consistently overexpressed in all tumors exhibiting *CCNE1* gene amplification (data not shown). However, some tumors overexpressed CCNE1 protein in the absence of *CCNE1* amplification. Therefore, we concluded that overexpression did not require gene amplification. Our findings suggested that the mechanism of CCNE1 overexpression may be dualistic, owing in part to gene amplification and in part to activation at the transcriptional level.

Bio markers that can predict clinical prognosis, including treatment response and overall survival, have substantial clinical impact on the management of patients with cancer (25). To further explore the clinical relevance of *CCNE1* alterations in endometrial endometrioid carcinomas, we evaluated the correlation between *CCNE1* amplification or protein expression and length of progression-free/overall survival in our population of patients with endometrial endometrial endometrial endometrial endometrial.

Interestingly, we found a strong correlation between poor prognosis and *CCNE1* amplification in patients with endometrial endometrioid carcinoma. However, CCNE1 protein overexpression was associated with an insignificant trend toward poor overall survival. CCNE1 functions as a regulatory subunit of CDK2, which is essential for both the G1 to S-phase transition and DNA replication. Moreover, CCNE1 plays a role in apoptosis and chromosomal instability in tumor cells (26). Cyclin E-CDK2 acts through phosphorylation of substrates involved in G<sub>1</sub> progression, S-phase entry and centrosome duplication. This complex also has kinase-independent functions involving the loading of mini-chromosome maintenance proteins onto origins of replication as quiescent cells enter the cell cycle (27).

CCNE1 amplification has been identified as a mechanism for overexpression in endometrial carcinomas (24,28). However, in the present study, we found a direct correlation between CCNE1 amplification and poor outcomes, consistent with the results of another report (29). To date, CCNE1 amplification has not been shown to be explicitly associated with primary treatment resistance. The mechanism underlying the association between CCNE1 amplification and shorter survival is not known; however, because mortality in patients with endometrial cancer is directly related to recurrence of disease after chemotherapy, it is conceivable that CCNE1 amplification may confer resistance to chemotherapy and/or enhance cell proliferation in chemoresistant recurrent tumors. Because the etiology of tumor recurrence is multifactorial, it is likely that *CCNE1* expression promotes the growth of tumor cells by providing them with higher proliferative capacity and lower apoptotic activity, as was shown in our previous study (30). Because the present study identified a previously undescribed pattern of CCNE1 amplification/expression in endometrial endometrioid carcinoma cells, further studies are required to elucidate the relationship between CCNE1 gene amplification/ expression and tumor sensitivity to chemotherapy

In the present study, the absence of FBXW7 expression in endometrial endometrioid carcinoma tissue was associated with deep myometrial invasion and lymphovascular space invasion, thereby potentially affecting overall survival and progression-free survival, with marginal statistical significance. Studies have demonstrated that FBXW7 regulates a wide range of cancer-associated substrates, and FBXW7 mutation leads to high expression of CCNE1, c-MYC, Notch and MCL1. c-MYC and CCNE1 are important components of the cell cycle machinery that are frequently deregulated in cancer and possess strong oncogenic activity. FBXW7 loss and c-MYC upregulation may play roles in the aggressive biological behaviors of cancer, which may in turn induce cell proliferation (31). Cells with increased c-MYC and reduced FBXW7 expression have been shown to be more invasive and to be associated with poor prognosis due to the high levels of matrix metalloproteinases (MMP2 and MMP9) (32,33). Indeed, the tendency to undergo lymph node and intrahepatic metastases may be correlated with these substrates in intrahepatic cholangiocarcinoma. Furthermore, mammalian target of rapamycin (mTOR) has been shown to function as an oncoprotein, controlling many cellular processes, such as cell proliferation and division. Interestingly, mTOR was also found to be degraded through an FBXW7-dependent

mechanism (34). As a major regulator of a set of oncoproteins, FBXW7 has been shown to be lost and mutated in various human cancers, such as human hepatocellular carcinoma and breast carcinoma (35,36). Recently, Mitsuhashi *et al* (37) reported that the Notch1-JAG1 axis is related to invasive properties and a poor prognosis in patients with endometrial cancer. Peiró *et al* (38) reported that mTOR expression is also related to poor outcomes in endometrial cancer. Taken together, the above findings provided evidence for the mechanisms through which loss of FBXW7 expression was related to poor outcomes in endometrioid carcinoma in the present study.

Molecular genetic evidence indicates that endometrial carcinomas are likely to develop as a result of a multistep process of oncogenic activation and tumor suppressor inactivation (39,40). In this study, we focused on some important genetic events of endometrial endometrioid carcinoma, i.e., inactivation of the tumor-suppressor genes PTEN, p53 and ARID1A; overexpression of the oncogene HER2; and MSI (5,40-42). In addition, we also analyzed the association between PTEN, HER2, p53, MLH1 and ARID1A expression and CCNE1 amplification. No significant relationships were observed between the expression of these proteins and CCNE1 amplification, except for positive PTEN expression; these data indicated that CCNE1 amplification was independent of p53, HER2, MLH1 and ARID1A expression in endometrial endometrioid carcinoma. Notably, CCNE1 amplification and PTEN loss may occur in a mutually exclusive manner. However, assessment of mutational status via direct sequencing was not performed in this study; only immunohistochemical analysis was carried out to compare different molecular events with CCNE1 amplification status. Thus, further studies are required to fully explore the relationship between CCNE1 amplification and other genetic events occurring in endometrial endometrioid carcinomas.

Cancer cells contain multiple genetic and epigenetic abnormalities. Despite this complexity, their growth and survival can often be impaired by inactivation of a single oncogene. This phenomenon, called 'oncogene addiction', provides a rationale for molecular targeted therapy (43). Based on this theory and our in vivo and in vitro findings, we propose that patients with endometrial endometrioid carcinoma having CCNE1 amplification may benefit from CDK inhibitor therapy if the disease recurs after conventional platinum and taxane or doxorubicin chemotherapy or from CDK inhibitor therapy as the primary therapy with the conventional combination of platinum and taxane or doxorubicin. To date, the CDK inhibitor E7070 has fared poorly in clinical trials for melanoma and lung cancer (44,45). However, its favorable therapeutic index and high selectivity may outweigh its shortcomings in CCNE1amplified endometrial endometrioid carcinoma. Therefore, we recommend that patients participating in future clinical trials evaluating the efficacy of CDK inhibitors for endometrial endometrioid carcinoma should be stratified based on CCNE1 amplification status.

In summary, we have demonstrated that phenotypic changes in endometrial endometrioid carcinomas in response to *CCNE1* inactivation depend on the expression status of *CCNE1*. The findings in the present study provide important insights into the biological roles of the CCNE1 signaling pathway in endometrial endometrioid carcinomas. Additionally, our observations have important therapeutic implications for patients with endometrial endometrioid carcinoma having *CCNE1* amplification. Endometrial endometrioid carcinomas with *CCNE1* amplification are clinically high-grade carcinomas with aggressive behavior (46,47). Therefore, detection of *CCNE1* amplification in endometrial endometrioid carcinomas may identify patients who will benefit from a CCNE1-specific inhibitor.

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