MINI REVIEW



An update on adrenocortical cell lines of human origin

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Abstract

Adrenocortical carcinoma (ACC) is a rare, heterogenous and highly malignant disease. Management of ACC is dependent on disease stage with complete surgical resection as the only potentially curative option. However, advanced, un-resectable, metastatic stages and also recurrences often require systemic treatments, which are unfortunately nowadays still unsatisfactory. The scarcity of preclinical models reflecting patient heterogeneities and furthermore drug-resistant phenotypes, has hampered the progress and development of new therapies in recent years. In this review, we provide an overview on the classical models and substantial progress which has been made over the last years in context of this aggressive disease.

Adrenocortical carcinoma (ACC) is a rare and highly malignant disease with an estimated incidence of ~0.5–2 new cases per million people per year. Clinical treatment of choice is surgery, however, the diagnosis often highly delayed thereby leading to advanced, unresectable and metastatic stages with very limited prognosis and a 5-years overall survival being <15% [1, 2]. Etoposide, cisplatin, doxorubicin and mitotane (EDP-M) constitute the current gold standard for metastatic ACC, which, however is not

satisfactory and results frequently in clinical toxicity [1]. The scarcity of human adrenocortical cell lines hampered in recent years more detailed preclinical investigations and the development of strategies considering for example various genotypes, secretion profiles and drug resistant phenotypes. However, since 2016 we observe tremendous progress in this field with overall five new human adrenocortical cell lines developed. In this review, we provide an overview on the current state of the art.

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NCI-H295

Gazdar et al. reported in 1990 on the first human adrenocortical cell line, NCI-H295, which was obtained in 1980 from a 48-year-old woman diagnosed with a primary ACC (Fig. 1; [3]). After surgical resection and subsequent processing, the established cells underwent long-term culture over many years. Radioimmunoassays and mass spectrometry revealed then 7 to 9 years after cell line establishment that the cells show the ability to produce all major adrenal steroids including corticosteroids, mineralocorticoids, androgens and estrogens [3]. Since then until today NCI-H295 (including its substrains H295A, H295R, H295RA, HAC13, HAC15 and HAC50) is the most commonly implemented model in context of adrenal steroidogenesis [4–7]. The HACsubstrains result from the attempt to develop a new human adrenocortical carcinoma (HAC) cell line with ACTH responsiveness. Clonal populations of adrenocortical cells were isolated from a tumor of a 11-month-old female patient



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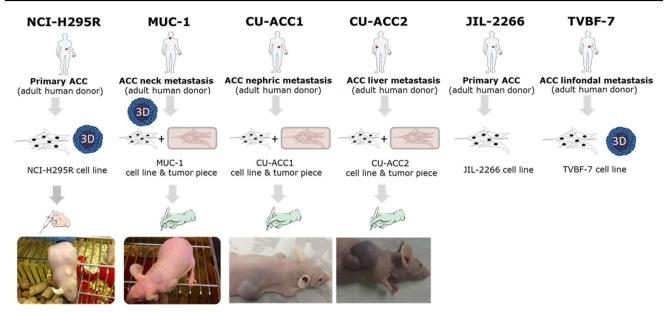


Fig. 1 Overview in the currently available human adrenocortical cell lines. Modified from [12]

Table 1 Known mutations of ACC cell lines

Cell line	Mutations	References
NCI-H295R	Mutation on TP53 gene - homozygous deletion of exons 8–9 Mutation on CTNNB1 gene - c.T133C:p.S45P, activating	[11, 13, 14, 54]
MUC-1	Mutation in TP53 gene - c.1024delC:p.R342fs.	[10, 12, 13].
CU-ACC1	Mutation on CTNNB1 gene - c.G100A:p.G34R, predicted as a gain-of-function	[41]
CU-ACC2	Mutation on TP53 gene - c.G337A:p.G245S, predicted as a loss of function. Mutation on MSH2 gene - homozygous deletion of exons 1–6	[41]
JIL-2266	Hemizygous mutation on TP53 gene - c.859G > T: p.E287X, stop-gain mutation Mutation on MUTYH gene - c.316C > T: p.R106W, loss of function	[48]
TVBF-7	Mutation on APC gene - c.739C > T:p.Q247*, non-sense	[13]

with hypertension and hirsutism. Later on, single-nucleotide polymorphism analysis revealed that the clones were isolated from contaminated H295R cells. Accordingly, compared to the original NCI-H295 strain, isolated from a mixed population of tumor cells, the HAC subclones are monoclonal [8, 9]. Of note, the reported steroidogenic capacities differ considerably for NCI-H295 and its substrains. This applies also to not specifically selected subclones which naturally occur over time in various labs and are in part maybe also due to varying culture conditions which additionally influence the secretory output [6, 10].

Genetically, NCI-H295 cells are characterized by a large deletion in the TP53 locus [11–13] and are also known to carry an activating CTNNB1 mutation (Table 1, [12–14]). A comprehensive analysis of the mutational landscape of NCI-H295, basing on whole-exome sequencing, is also available since 2019 [15].

Gazdar et al. reported in the original study also on NCI-H295's tumorigenicity when inoculated subcutaneously into athymic nude mice ([3], Fig. 1). Until today NCI-H295 represents, thus, also the most commonly used subcutaneous xenograft model for human ACC and was later also applied as ACC hepatic metastasis model [16, 17]. Accordingly, various preclinical studies have been performed since 1990 in pathophysiological and therapeutic context. It could be for example demonstrated, that Wnt/ β-catenin pathway [12, 13] and SF-1 modulations are involved in pathogenesis and potential targets for the treatment of ACC [18, 19]. NCI-H295 were also repeatedly implemented in context of IGF-System and IGF1-receptor inhibiting approaches in ACC [20-23]. Using NCI-H295 as a tool, it could be furthermore demonstrated that mitotane is an inhibitor of sterol-O-acyl-transferase 1 (SOAT1) leading to accumulation of toxic lipids and subsequent ER stress



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[24]. Studies including NCI-H295 enabled also the investigation of preclinical experiments investigating the current clinical gold standard EDP-M (etoposide, doxorubicin, cisplatin, mitotane) and potential next-generation schemes involving nano-technological modified variants of the parental drugs [25, 26].

Nowadays, NCI-H295 is commonly implemented together with the next-generation models outlined below. Thus, further examples and most recent pre-clinical implementation of NCI-H295 will be context-dependent added in the following sections.

MUC-1

In 2016, we have reported on MUC-1, the second human adrenocortical cell line developed in this field. In contrast to the chemo-naïve and often pre-clinically good responding NCI-H295, MUC-1 represents a clinically systemically treated, but EDP-M-resistant model obtained from a distant ACC metastasis of a male patient. MUC-1 has been established as cell line for in vitro, but also as patientderived tissue xenograft for in vivo settings (Fig. 1, [10, 23]). The model shows the for metastases frequently observed increased cellular plasticity, which can be e.g. seen in highly elevated drug tolerance for a wide range of chemotherapeutic, phytochemical, molecular targeted drugs and also combinatory regimens which is far beyond resistance against the originally applied EDP-M scheme [10, 27– 36]. Recently, a broad comparative drug screen of clinically relevant chemotherapies and targeted therapies has been performed in NCI-H295R and MUC-1. Interestingly, these experiments identified the very classical combination gemcitabine and cisplatin as highly promising treatment, but indicated in parallel also strong molecular signs of acquiring gemcitabine resistance as represented by treatment-dependent ribonucleotide reductase-upregulation in both tumor models. Of note, a combination of gemcitabine, cisplatin and the dual ribonucleotide-reductaseinhibitor COH-29 led to previously total cell killing for both tumor models [33].

Also regarding metabolic re-programming MUC-1 provides new insights and opportunities. Warde et al. recently initiated comprehensive analyses in terms of lipid metabolism and found significant differences in cholesterol storage and lipid droplet re-modelling between mitotane sensitive (NCI-H295R) and resistant (MUC-1) ACC cells [31, 37]. Fittingly, we found recently also signs of differential HSD17B4 regulation in MUC-1, a gene with a dual role in steroidogenesis and fatty acid oxidation [13]. HSD17B4 expression is also known to be involved in the development of castration resistant prostate cancer, leading to metabolic re-programming and poor prognosis [38, 39]. In this context

we have lately also characterized steroidogenic signaling in a panel of ACC cells [13]. In accordance with the original patient record of a diffuse steroidogenic phenotype, MUC-1 demonstrates here basally a profile with comparably low steroidogenic activity and secretion. However, e.g. upon forskolin stimulation MUC-1 shows tremendous steroidogenic activity for all steroidogenic precursors, strong electrophysiological response and specific focus on the androgen axis in the resulting functional secretion profile. While MUC-1 demonstrates under these conditions furthermore specific upregulation of the androgen receptor gene, converse regulations are detectable for NCI-H295R. While even a role of adrenal derived androgens is under discussion in castration resistant prostate cancer [40], comprehensive preclinical investigations in terms of androgenic signaling in ACC are currently widely lacking. Basing on our newest findings, MUC-1 represents in this context the most promising cell line.

Moreover, as recently reported for the first time at the 8th International ACC Symposium in Brescia, Italy 2021, further experiments indicate for MUC-1 spontaneous metastatic potential which could be not reported for any other human ACC cell line so far (https://www.acc2021brescia.com/). Recently published experiments on the chemokine axis and related tumor aggressiveness confirm differential regulations for CXCR4 and CXCL12 in NCI-H295R and MUC-1, which also suggest varying potential in terms of metastasization [34].

Genetic analyses demonstrate for MUC-1 a somatic deletion/frameshift mutation in TP53 (Table 1), but no mutations for example in MEN1, PRKAR1A, CTNNB1, APC, ZNRF-3, IGF-2, EGFR, RB1, BRCA1, BRCA2, RET, GNAS and PTEN [10, 13]. Whole genome and whole-bisulfite sequencings have been performed for the panel of NCI-H295, MUC-1 and a new ACC cell line named TVBF-7 (please see below). These overall data sets will be available soon.

CU-ACC1 and **CU-ACC2**

In 2018, two further cell lines were established from metastatic ACC in female patients, namely CU-ACC1 and CU-ACC2 (Fig. 1, [41]). The CU-ACC1 cell line derives from a perinephric ACC metastasis [41] and secretes higher levels of cortisol compared to NCI-H295R cells, while no aldosterone secretion was reported. Genetic analysis reveals an activating point mutation of CTNNB1 gene [41], a well known ACC driver gene [42]. The CU-ACC2 cell line was established from a liver metastasis in a Lynch syndrome patient with non-secreting primary ACC [41]. Despite high abundance of CYP11A1 [43], CU-ACC2 cells secrete only minute amount of cortisol and are unresponsive to hormone



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stimulation [41]. CU-ACC2 cells represent wild type for CTNNB1, while a mutation in TP53 is reported (Table 1, [7, 41]). In addition, in line with the Lynch syndrome, a heterozygous deletion of 1-6 exons of MSH2 gene was detected. The mutation on the CTNNB1 gene found in CU-ACC1 is within the ubiquitination recognition motif. Currently, this mutation is not fully characterized but it is predicted to be activating [41]. These ACC experimental cell models, together with the NCI-H295R cells, have been recently used to demonstrate sensitivity of ACC to ferroptosis and its dependence on the active steroid synthetic pathways [43]. Moreover, these models were useful to identify the maternal embryonic leucine zipper kinase (MELK) [44], the mitotic PDZ-binding kinase (PBK) [45] and inhibition of aurora kinase and beta-catenin pathway as potentially druggable targets for ACC [46]. Furthermore, the CU-ACC2 cell line allowed to set up the first humanized ACC patient-derived xenograft mouse [47], elucidating the effects of immunotherapy and its mechanism of action. Authors suggested that the effects of an anti-PD1 on tumor microenvironment would recapitulate effects of the immune milieu of the tumor in the matching patient.

JIL-2266

JIL-2266 is a newly established ACC cell line presenting high tumor mutational burden (TMB) and loss of heterozygosity (LOH) [48]. Similar to NCI-H295R cells, JIL-2266 is derived from a female patient with a primary ACC (Fig. 1). The subsequent characterization of JIL-2266 revealed intermediate-to-low expression of SF-1, while the hormonal production and steroidogenic enzyme expression is reported to depend on culture media composition [48]. Exome sequencing demonstrates the presence of a pathogenic germline mutation in the MUTYH gene, which is linked to base excision repair inactivation, leading to high TMB. Further genetic analysis showed a hemizygous stop-gain mutation in the TP53 gene (Table 1), which presented high expression, and somatic nucleotide variant in the ZNRF3 gene. Similar to MUC-1 and CU-ACC2, JIL-2266 holds wild type CTNNB1.

TVBF-7

Recently, the new ACC cell line TVBF-7 has been proposed by our group [13]. These cells represent the most recently developed ACC cell line, formerly known as primary cell culture ACC115m [35, 36]. Cells were established from a perirenal lymph-node metastasis of a male ACC patient which, similar to MUC-1, underwent progression upon clinical EDP-M treatment [13]. After

establishment of a primary cell culture, the cells were found to be continuously passageable. Moreover, STR-analysis revealed a unique STR-profile, which could be furthermore linked to the original patient tumor [13]. Interestingly, although the patient did not report signs of hormone excess, TVBF-7 cells demonstrate basally high levels of cortisol and related candidates of the glucocorticoid axis. Moreover, they are unresponsive to a selection of known physiological stimuli, thereby indicating autonomous cortisol secretion [13, 49]. Fittingly, further genetic investigations indicate an altered Wnt/β-Catenin pathway due to the detection of a non-sense APC mutation (Table 1). Except for the ACTH receptor (MC2R), these cells express, compared to NCI-H295R, low level of hormonal receptor at gene level [13, 35]. The expression of estrogen receptor α is comparable to that of NCI-H295R, while the expression of estrogen receptor β is lower [35]. TVBF-7 cells have already been used as an experimental cell model to test the efficacy of drugs proven effective in ACC, in addition to other ACC cell lines. Indeed, we demonstrated that progesterone receptors may be a druggable target in these ACC cells [35, 36], thus confirming the correlation between the level of the progesterone receptor expression and the sensitivity of ACC cells to the cytotoxic effect elicited by progesterone [35, 36, 50, 51]. Finally, the Cdk4/6 inhibitor ribociclib induces cytotoxicity and reduction of the cell proliferation rate in these cells, although the cell cycle perturbation appears to differ compared to other ACC cell lines. TVBF-7 cells accumulate, as expected, in G1 phase, while NCI-H295R cells and MUC-1 cells accumulate in G2 phase [36], thus supporting the experimental advantage to have different cell models that could reflect the different ACC behavior, both in vitro and in vivo. Interestingly, in line with the shared metastatic origin from patients in progression after EDP-M, the TVBF-7 cells reflect some, but not all, characteristics with the cell line MUC-1. In a recent published work, the exposure of these both cell models to mitotane, to progesterone and to the CDK4/6 inhibitor ribociclib alone or in combined setting, revealed a lower sensitivity compared to the NCI-H295R cells [36]. The different responses to pharmacological treatments underline the importance of their different phenotype.

Finally, a detailed report on the establishment of ACC tumor spheroids and organoids including NCI-H295R, MUC-1 and TVBF-7 in a high-throughput format is currently under review. A pre-print of the manuscript can be found here (Fig. 1, [52]).

Conclusions

In sum, a lot of progress has been made over the last years and changed the preclinical landscape for ACC. The new



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ACC experimental cell models, together with the worldwide-used NCI-H295R cell line, give to researchers instruments that are in line with the well known heterogeneity of this disease and have the potential to reveal so far unknown patient sub-type characteristics. Furthermore, ACC cell lines of metastatic origin can offer useful models to investigate the pharmacological effect of drugs and the pathological molecular alteration in the context of metastatic and EDP-M-progressed disease, which is specifically challenging in the clinic management and closely related with poor prognosis. Finally, we would like to underline a key point in using cancer cell lines in research and drug development, that is the risk of cell line misidentification [53]. A periodically authentication by STR analysis as well as the expression of ACC markers, such as Steroidogenic Factor 1 could help to avoid this.

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Compliance with ethical standards

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References

- B. Kiesewetter et al. Management of adrenocortical carcinoma: are we making progress? Ther. Adv. Med. Oncol. 13, 17588359211038409 (2021)
- M. Fassnacht et al. Adrenocortical carcinomas and malignant phaeochromocytomas: ESMO-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann. Oncol. 31 (11), 1476–1490 (2020)
- A.F. Gazdar et al. Establishment and characterization of a human adrenocortical carcinoma cell line that expresses multiple pathways of steroid biosynthesis. Cancer Res. 50(17), 5488–5496 (1990)
- A. Dierks et al. Identification of adrenal genes regulated in a potassium-dependent manner. J. Mol. Endocrinol. 45(4), 193–206 (2010)
- K.E.M. Ahmed et al. LC-MS/MS based profiling and dynamic modelling of the steroidogenesis pathway in adrenocarcinoma H295R cells. Toxicol. Vitr. 52, 332–341 (2018)

 M. Kurlbaum et al. Steroidogenesis in the NCI-H295 Cell Line Model is strongly affected by culture conditions and substrain. Exp. Clin. Endocrinol. Diabetes 128(10), 672–680 (2020)

- K. Nanba, A.R. Blinder, W.E. Rainey, Primary cultures and cell lines for in vitro modeling of the human adrenal cortex. Tohoku J. Exp. Med. 253(4), 217–232 (2021)
- J. Parmar, R.E. Key, W.E. Rainey, Development of an adrenocorticotropin-responsive human adrenocortical carcinoma cell line. J. Clin. Endocrinol. Metab. 93(11), 4542–4546 (2008)
- T. Wang, W.E. Rainey, Human adrenocortical carcinoma cell lines. Mol. Cell Endocrinol. 351(1), 58–65 (2012)
- C. Hantel et al. Targeting heterogeneity of adrenocortical carcinoma: evaluation and extension of preclinical tumor models to improve clinical translation. Oncotarget 7(48), 79292–79304 (2016)
- M. Reincke et al. p53 mutations in human adrenocortical neoplasms: immunohistochemical and molecular studies. J. Clin. Endocrinol. Metab. 78(3), 790–794 (1994)
- E.M. Pinto, K. Kiseljak-Vassiliades, C. Hantel, Contemporary preclinical human models of adrenocortical carcinoma. Curr. Opin. Endocr. Metab. Res. 8, 139–144 (2019)
- S.B.C. Sigala, D. Penton, A. Abate, M. Peitzsch, D. Cosentini, G. A.M. Tiberio, S.R. Bornstein, A. Berruti, C. Hantel, A comprehensive investigation of steroidogenic signaling in classical and new experimental cell models of adrenocortical carcinoma. Cells, 2022. 11
- F. Tissier et al. Mutations of beta-catenin in adrenocortical tumors: activation of the Wnt signaling pathway is a frequent event in both benign and malignant adrenocortical tumors. Cancer Res. 65(17), 7622–7627 (2005)
- N.G. Nicolson, R. Korah, T. Carling, Adrenocortical cancer cell line mutational profile reveals aggressive genetic background. J. Mol. Endocrinol. 62(4), 179–186 (2019)
- A. Morin et al. Establishment of a mouse xenograft model of metastatic adrenocortical carcinoma. Oncotarget 8(31), 51050–51057 (2017)
- C. Hantel, F. Beuschlein, Xenograft models for adrenocortical carcinoma. Mol. Cell Endocrinol. 421, 28–33 (2016)
- M. Doghman et al. Integrative analysis of SF-1 transcription factor dosage impact on genome-wide binding and gene expression regulation. Nucleic Acids Res. 41(19), 8896–8907 (2013)
- M. Doghman et al. Increased steroidogenic factor-1 dosage triggers adrenocortical cell proliferation and cancer. Mol. Endocrinol. 21(12), 2968–2987 (2007)
- A. Logie et al. Establishment and characterization of a human adrenocortical carcinoma xenograft model. Endocrinology 141(9), 3165–3171 (2000)
- T.C. Brown et al. Insulin-like growth factor and SLC12A7 dysregulation: a novel signaling hallmark of non-functional adrenocortical carcinoma. J. Am. Coll. Surg. 229(3), 305–315 (2019)
- C. Hantel et al. Liposomal doxorubicin-based treatment in a preclinical model of adrenocortical carcinoma. J. Endocrinol. 213(2), 155–161 (2012)
- F. Beuschlein et al. IGF1-R inhibition and liposomal doxorubicin: progress in preclinical evaluation for the treatment of adrenocortical carcinoma. Mol. Cell Endocrinol. 428, 82–88 (2016)
- S. Sbiera et al. Mitotane Inhibits Sterol-O-Acyl transferase 1 triggering lipid-mediated endoplasmic reticulum stress and apoptosis in adrenocortical carcinoma cells. Endocrinology. 156(11), 3895–3908 (2015)
- C. Hantel et al. Liposomal polychemotherapy improves adrenocortical carcinoma treatment in a preclinical rodent model. Endocr. Relat. Cancer 21(3), 383–394 (2014)
- S. Jung et al. Preclinical progress and first translational steps for a liposomal chemotherapy protocol against adrenocortical carcinoma. Endocr. Relat. Cancer 23(10), 825–837 (2016)



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 A. Hasanovic et al. Targeting the multidrug transporter Patched potentiates chemotherapy efficiency on adrenocortical carcinoma in vitro and in vivo. Int J. Cancer 143(1), 199–211 (2018)

- C. Siebert et al. Heat shock protein 90 as a prognostic marker and therapeutic target for adrenocortical carcinoma. Front Endocrinol. (Lausanne) 10, 487 (2019)
- R. Liang et al. Targeted gene expression profile reveals CDK4 as therapeutic target for selected patients with adrenocortical carcinoma. Front Endocrinol. (Lausanne) 11, 219 (2020)
- M. Fragni et al. In vitro cytotoxicity of cabazitaxel in adrenocortical carcinoma cell lines and human adrenocortical carcinoma primary cell cultures(). Mol. Cell Endocrinol. 498, 110585 (2019)
- 31. K.M. Warde et al. Liver X receptor inhibition potentiates mitotane-induced adrenotoxicity in ACC. Endocr. Relat. Cancer 27(6), 361–373 (2020)
- A. Abate et al. Cytotoxic effect of trabectedin in human adrenocortical carcinoma cell lines and primary cells. Cancers (Basel) 12 (4), (2020)
- C. Bothou et al. Novel insights into the molecular regulation of ribonucleotide reductase in adrenocortical carcinoma treatment. Cancers (Basel) 13(16), (2021)
- 34. G. Cantini et al. Stimulated expression of CXCL12 in adrenocortical carcinoma by the PPARgamma ligand rosiglitazone impairs cancer progression. J Pers. Med. **11**(11), (2021)
- E. Rossini et al. Cytotoxic effect of progesterone, tamoxifen and their combination in experimental cell models of human adrenocortical cancer. Front Endocrinol. (Lausanne) 12, 669426 (2021)
- A. Abate et al. Ribociclib cytotoxicity alone or combined with progesterone and/or mitotane in in vitro adrenocortical carcinoma cells. Endocrinology (2021)
- K. Warde et al. Investigating the role of cholesterol and lipid trafficking in mitotane resistance in adrenocortical carcinoma. J. Endocr. Soc. 5(Supplement_1), A70–A70 (2021)
- 38. K.K. Rasiah et al. HSD17B4 overexpression, an independent biomarker of poor patient outcome in prostate cancer. Mol. Cell Endocrinol. **301**(1–2), 89–96 (2009)
- H.K. Ko et al. Loss of an androgen-inactivating and isoformspecific HSD17B4 splice form enables emergence of castrationresistant prostate cancer. Cell Rep. 22(3), 809–819 (2018)
- M. Barnard et al. The role of adrenal derived androgens in castration resistant prostate cancer. J. Steroid Biochem Mol. Biol. 197, 105506 (2020)

- K. Kiseljak-Vassiliades et al. Development of new preclinical models to advance adrenocortical carcinoma research. Endocr. Relat. Cancer 25(4), 437–451 (2018)
- S. Grisanti et al. Molecular genotyping of adrenocortical carcinoma: a systematic analysis of published literature 2019-2021. Curr. Opin. Oncol. 34(1), 19–28 (2022)
- I. Weigand et al. Active steroid hormone synthesis renders adrenocortical cells highly susceptible to type II ferroptosis induction. Cell Death Dis. 11(3), 192 (2020)
- K. Kiseljak-Vassiliades et al. Elucidating the role of the maternal Embryonic Leucine Zipper Kinase in Adrenocortical Carcinoma. Endocrinology 159(7), 2532–2544 (2018)
- A. Kar et al. Targeting PDZ-binding kinase is anti-tumorigenic in novel preclinical models of ACC. Endocr. Relat. Cancer 26(10), 765–778 (2019)
- A.G. Maria et al. Inhibition of Aurora kinase A activity enhances the antitumor response of beta-catenin blockade in human adrenocortical cancer cells. Mol. Cell Endocrinol. 528, 111243 (2021)
- 47. J. Lang et al. Development of an Adrenocortical Cancer Humanized Mouse Model to characterize anti-PD1 effects on tumor microenvironment. J. Clin. Endocrinol. Metab. **105**(1), (2020)
- L.S. Landwehr et al. A novel patient-derived cell line of adrenocortical carcinoma shows a pathogenic role of germline MUTYH mutation and high tumour mutational burden. Eur. J. Endocrinol. 184(6), 823–835 (2021)
- G.A. Ueland et al. Diagnostic testing of autonomous cortisol secretion in adrenal incidentalomas. Endocr. Connect 9(10), 963–970 (2020)
- M. Fragni et al. In vitro antitumor activity of progesterone in human adrenocortical carcinoma. Endocrine 63(3), 592–601 (2019)
- C. Fiorentini et al. Antisecretive and antitumor activity of abiraterone acetate in human adrenocortical cancer: a preclinical study. J. Clin. Endocrinol. Metab. 101(12), 4594

 –4602 (2016)
- S. Bornstein et al. Innovative multidimensional models in a highthroughput-format for different cell types of endocrine origin. p. 2022.01.10.475686 (2022)
- American Type Culture Collection Standards Development Organization Workgroup ASN-0002, Cell line misidentification: the beginning of the end. Nat. Rev. Cancer 10(6), 441–448 (2010)
- L. Cerquetti et al. Mitotane increases the radiotherapy inhibitory effect and induces G2-arrest in combined treatment on both H295R and SW13 adrenocortical cell lines. Endocr. Relat. Cancer 15(2), 623–634 (2008)

