

Research Paper

## Cloning and Characterization of a 2-Cys Peroxiredoxin in the Pine Wood Nematode, *Bursaphelenchus xylophilus*, a Putative Genetic Factor Facilitating the Infestation

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### Abstract

The pine wood nematode, *Bursaphelenchus xylophilus*, is an invasive plant parasitic nematode and a worldwide quarantine pest. An indigenous species in North America and the causal agent of pine wilt disease, *B. xylophilus* has devastated pine production in South-eastern Asia including Japan, China, and Korea since its initial introduction in the early 1900s. The reactive oxygen species (ROS) is the first line of defense utilized by host plants against parasites, while nematodes, counteractively, employ antioxidants to facilitate their infection. Peroxiredoxins (Prxs) are a large class of antioxidants recently found in a wide variety of organisms. In this report, a gene encoding a novel 2-cysteine peroxiredoxin protein in *B. xylophilus* was cloned and characterized. The 2-cysteine peroxiredoxin in *B. xylophilus* (herein refers to as "BxPrx") is highly conserved in comparison to 2-cysteine peroxiredoxins (Prx2s) in other nematodes, which have two conserved cysteine amino acids (Cp and Cr), a threonine-cysteine-arginine catalytic triad, and two signature motifs (GGLG and YF) sensitive to hydrogen peroxide. *In silico* assembly of BxPrx tertiary structure reveals the spatial configuration of these conserved domains and the simulated BxPrx 3-dimensional structure is congruent with its presumed redox functions. Although no signal peptide was identified, BxPrx was abundantly expressed and secreted under the *B. xylophilus* cuticle. Upon further analysis of this leader-less peptide, a single transmembrane  $\alpha$ -helix composed of 23 consecutive hydrophobic amino acids was found in the primary structure of BxPrx. This transmembrane region and/or readily available ATP binding cassette transporters may facilitate the transport of non-classical BxPrx across the cell membrane. Recombinant BxPrx showed peroxidase activity *in vitro* reducing hydrogen peroxide using glutathione as the electron donor. The combined results from gene discovery, protein expression and distribution profiling (especially the "surprising" presence under the nematode cuticle), and recombinant antioxidant activity suggest that BxPrx plays a key role in combating the oxidative burst engineered by the ROS defense system in host plants during the infection process. In summary, BxPrx is a genetic factor potentially facilitating *B. xylophilus* infestation.

Key words: *Bursaphelenchus xylophilus*, pine wilt disease, reactive oxygen species, Peroxiredoxin, hydrogen peroxide.

## Introduction

The pinewood nematode, *Bursaphelenchus xylophilus* (Steiner and Buhrer) Nickle, is the pathogenic agent of pine wilt disease, which has caused serious damage to pine forests in Japan, Korea, and China [1]. *Bursaphelenchus xylophilus* is native to North America [2] and was introduced to Japan in the 1900s, South Korea in the 1980s, China in 1982 [3], and later to Portugal, Canada and 40 other countries [4, 5]. In China alone, the combined damage and management costs exceed 4 billion US dollars annually [6].

### Biotic and abiotic factors affecting *B. xylophilus* infection

*Bursaphelenchus xylophilus* completes its life cycle within a temperature range of 25-30 °C; therefore, the population density of *B. xylophilus* normally peaks after summer, decreases from the beginning of the winter and reaches its lowest point the following spring [7]. Water status in pines also plays an important role in the pine-nematode relationship, and a reduced transpiration rate in pine can stimulate population growth of *B. xylophilus* [8]. In addition to abiotic factors, such as temperature and water stress, biotic factors also play a prominent role in the infection and distribution of this plant pathogenic nematode. A better understanding of these biotic factors will provide essential information to control the spread of this devastating disease in pine trees. The innate resistance of pine is one of the most important biotic factors limiting the occurrence of pine wilt disease. *Bursaphelenchus xylophilus* can rapidly reproduce and spread in susceptible pine trees, but its movement is restricted among resistant pine trees [9]. It has been reported that resistant pine species produce additional secondary metabolites to recognize and suppress the invasion of pathogens [10].

After millions of years of co-evolution with parasites, host plants have evolved various resistant mechanisms against the parasite infection. In many plants, the first line of defense involves an oxidative burst which generates toxic reactive oxygen species (ROS), including superoxide anion ( $O_2^-$ ), hydrogen peroxide ( $H_2O_2$ ) and the hydroxyl radical ( $\cdot OH$ ) [11]. ROS production in plants is particularly important in the defense and recognition of novel pathogens [12], because it can affect many key events in plant-pathogen interactions, including signal transduction, antimicrobial effect, membrane lipoxidation, cell wall modification, induced resistance, and hypersensitive cell death [11]. Counteractively, parasites produce antioxidants to protect them from the host plant's ROS defenses [13]. Therefore, removing ROS in host plants is critical for the survival of parasites.

Given the fact that *B. xylophilus* can successfully infest and spread in pine trees, antioxidant proteins likely play a key role in the host-parasite interaction.

### Antioxidants and peroxiredoxins

The major antioxidants in eukaryotic organisms include superoxide dismutase which detoxifies the superoxide anion, catalase, glutathione peroxidase which is involved in the breakdown of cellular peroxides, and peroxiredoxin which functions to inactivate hydrogen peroxide [13, 14 and 15]. Based on the catalytic mechanisms and the presence of either one or two highly conserved cysteine residues, peroxiredoxins (Prxs) are classified into 1-Cys, typical 2-Cys, and atypical 2-Cys Prxs. Catalytic reactions of the typical 2-Cys, atypical 2-Cys, and 1-Cys Prx are through the formation of intermolecular disulfide, inner-molecular disulfide [16], and the participation of small chemical thiols as reduced agents, respectively [17]. Prxs are multifunctional proteins, for example they function as antioxidants, by reducing alkyl peroxide and hydrogen peroxide to alcohol or water, respectively [18, 19]. They can also protect against phospholipid peroxidation [20] and serve as a peroxynitrite reductase for the detoxification of the reactive nitrogen species (RNS) [21] to protect cells from oxidant-induced membrane damage and prevent cell death [22]. In addition, Prxs are also considered to be signal transmitters in phosphorylation and transcriptional regulation, presumably through regulation of hydrogen peroxide [23]. Differing from other antioxidants, Prxs have no cofactors, such as metals or prosthetic groups for its antioxidant activity. The highly conserved cysteine residues are their active sites [16]. The cysteine residue near the N-terminus is referred to as Cp, which exists in all Prxs; whereas the cysteine residue near the C-terminus is referred to as Cr, and it only exists in the 2-Cys Prxs [16].

Despite recent progresses in the genomic [24] and epigenomic [25] understanding of this world-wide quarantine pest, there is virtually no information available regarding the *Prx* gene in plant pathogenic *B. xylophilus* and its potential role in engineering the antioxidative counterattack against pine hosts. In this study, we pursue the following objectives: (1) Clone and sequence *peroxiredoxin* in the plant pathogenic *B. xylophilus* (*BxPrx*) using RACE-PCR; (2) Gain a better understanding of the transmembrane pathway of leader-less *BxPrx* based on the *in silico* structural analysis of *BxPrx*; and (3) Express recombinant *BxPrx* heterogeneously in a bacterial system and investigate the tissue localization of *BxPrx* by

immunohistochemistry using a polyclonal antibody raised against recombinant BxPrx.

## Materials and Methods

### Bursaphelenchus xylophilus culture and maintenance

The *B. xylophilus* isolate JSZJ1-6 [3] was kindly provided by Dr. Bingyan Xie (Institute of Vegetables and Flowers, Chinese Academy of Agricultural Sciences). *B. xylophilus* was maintained at 25 °C for one week on a necrotrophic fungus, *Botrytis cinerea*, grown on a PDA (Potato Dextrose Agar) culture dish. Mixed stages of *B. xylophilus* were then harvested with a Baermann funnel, and washed with M9 buffer (KH<sub>2</sub>PO<sub>4</sub> 30 g/L, K<sub>2</sub>HPO<sub>4</sub> 60 g/L, NaCl 50 g/L) before subjecting the isolate to subsequent experiments [26].

### BxPrx gene discovery and sequence analysis

A thorough search of available sequences at a web-based nematode genomics database, Nematode.Net v3.0 (<http://www.nematode.net>) showed that Prx2s are evolutionarily conserved in all extant nematode species. After alignment of the deduced protein sequences of Prx2 from EST libraries obtained at this website, the degenerate primers (Table S1) were designed based on several highly conserved domains for the cloning of BxPrx. Total RNA was extracted from mixed stages of *B. xylophilus* using RNeasy Mini kit (Qiagen, Hamburg, Germany) according to the manufacturer's protocols. The first strand cDNA used in 3' RACE and 5' RACE were synthesized with 3' full RACE core set (Takara, Dalian, China) and Firstchoice® RLM RACE Kit (Ambion, Foster City, CA, USA), respectively. The 3' end product of the BxPrx gene was obtained with oligodT-3 site adaptor primer (Takara) and the degenerate primer BxPrx3S. 5' end sequence was obtained with 5' RACE outer primer (Ambion) and BxPrx5R. The entire sequence was amplified by the primer set BxPrxSB and BxPrxRS (Table S1) which include restriction sites BamH I and Sal I to the 5' and 3' ends, respectively, and the resulting PCR products were validated by direct sequencing.

The deduced protein sequence, pI (Iso-electric Point) and MW (Molecular Weight) of BxPrx were predicted by DNAMAN v.5.2.2 (Lynnon Biosoft, Quebec, Canada). The catalytic triad of BxPrx was determined by a Conserved Domain search on the NCBI website (<http://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi>) [27]. Protein tertiary structure was predicted by an online protein structure homology-modeling server SWISS-MODEL using the automated mode and modified by PyMOL-v1.3r1 software. All published EST sequences of Prx2 in nema-

todes were obtained from Nematode.Net (<http://www.nematode.net>). The signal peptide of BxPrx was determined using SignalP (<http://www.cbs.dtu.dk/services/SignalP/>) [28], and the transmembrane region of BxPrx was predicted by TMHMM (<http://www.cbs.dtu.dk/services/TMHMM-2.0/>). The hydrophobicity profile of BxPrx was analyzed by DNAMAN v.5.2.2. Secretome 2.0 Server (<http://www.cbs.dtu.dk/services/SecretomeP/>) was used to characterize the non-classical secreted proteins.

### Phylogenetic relationship of BxPrx with other Prx2s

All Prx2 sequences in animals which have the full open reading frames (ORFs) were extracted from GenBank, loaded in MEGA 4, and aligned. The phylogenetic relationship of BxPrx with other Prx2s was revealed using two different methods: (1) the neighbor-joining (NJ) analysis with an Amino: Poisson correction model; (2) the maximum parsimony (MP) analysis. The sites containing missing data or alignment gaps were removed in a pair-wise fashion. A total of 1,000 bootstrap replications were used to test topology. Bootstrap values (>50) were reported with each branch and the taxonomic origins of the sequences were highlighted.

### Nuclear localization of BxPrx

Intracellular localization of BxPrx was investigated through transient expression of the Cam35S::GFP-BxPrx fusion construct which was bombarded into onion epidermal cells [29] using Model PDS-1000/He Biolistic Particle Delivery System (Baumbusch, LO.). Plasmid Cam35S::GFP-BxPrx was constructed by inserting the entire coding sequence of BxPrx into the Cam35S-GFP plasmid behind the Cam35S promoter (Cauliflower Mosaic virus 35S promoter) and GFP (Green Fluorescent Protein) reporter gene after digestion with BamH I and Sal I (Fig. S1A). The resulting in-frame fusion was confirmed by restriction mapping and DNA sequencing. Expression was detected against a blank Cam35S-GFP control with a Bx60 Olympus microscope using a blue wide band cube for fluorescence observation.

### Recombinant protein expression, purification and validation

A recombinant expression plasmid was constructed by ligating the entire open reading frame of BxPrx into a bacterial expression vector pET-28a (Fig. S1B). The resulting recombinant construct was validated by the direct sequencing of PCR products and then transformed into an *Escherichia coli* BL21 (DE3) competent cell. *Escherichia coli* was cultured at 37 °C

for 16 hrs with shaking at 250 rpm. It was then diluted 1:50 (V:V) into liquid LB (Lysogeny Broth), and the liquid medium was shaken continuously until the OD<sub>600</sub> value reached 0.4-0.6. The induction conditions, such as isopropyl β-D-1-thiogalactopyranoside (IPTG) concentration and incubation temperature, were optimized to maximize the yield of soluble recombinant BxPrx. A series of IPTG concentrations ranging from 0, 1, 2, and 5 mM was tested. The induction was then carried out at two different temperatures: 37 °C, 250 rpm shaking for three hours, and 20 °C, 150 rpm shaking for 16 hrs. 100 ml of bacterial culture from each treatment was precipitated by centrifugation at 4,000 g for 15 min. Then the pellets were dissolved in the lysis buffer (50 mM NaH<sub>2</sub>PO<sub>4</sub>, 300 mM NaCl, 10 mM Imidazole, pH 7.0), and sonicated (250 W, 10 s × 10 times, 30 s interval, in ice) to break up the bacteria. After centrifugation at 4 °C, 12,000 rpm for 20 min, the supernatant of sonicated fraction was mixed with Ni-NTA agarose (Qiagen, Hamburg Germany). To purify recombinant BxPrx, the resulting protein was washed (50 mM NaH<sub>2</sub>PO<sub>4</sub>, 300 mM NaCl, 20 mM imidazole, pH 7.0) and then eluted (50 mM NaH<sub>2</sub>PO<sub>4</sub>, 300 mM NaCl, 250 mM Imidazole, pH 7.0). A total of 10 elution fractions were collected with 500 µl in each fraction.

After SDS-PAGE and staining by coomassie blue, the band of the purified BxPrx protein was cut out from the gel. After being dipped in a 100 µl discoloration solution (H<sub>2</sub>O: methonal: acetic acid = 5: 4: 1) for 20 min, dehydrated with 100 µl acetonitrile for 10 min, and alkylated with 50 µl indoacetamide for 30 min in dark, the product was treated with 15 µl 0.01 µg/µl trypsin for 16 h, and then was extracted with 100 µl 5 % TFA (trifluoroacetic acid) at 40 °C for 1h. After drying under nitrogen and mixed with 5 µl 0.1 % TFA, the sample was loaded into an ABI-4800 MALDI-TOF-TOF for a confirmation test of the induced recombinant protein.

### BxPrx activity assay

The activity of recombinant BxPrx on the reduction of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and cumene hydroperoxide (C<sub>9</sub>H<sub>12</sub>O<sub>2</sub>) was measured at OD<sub>340</sub> according to Kwatia [30]. The reaction mixture contained 5 mM potassium phosphate (pH 7.0), 1 mM EDTA, 0.1 mM NADPH, 1mM L-glutathione, and 0.1 unit/ml glutathione reductase (Sigma). One unit of activity is defined as the amount of enzyme required to cause the oxidation of 1 nmol of NADPH per minute.

### BxPrx antiserum production and immunoblot analysis

A polyclonal antiserum against recombinant BxPrx was obtained by consecutive 4-week injections of the purified protein into a rabbit. For the first injection, 1 mg of BxPrx protein was injected concurrently with 1 mg Freund's Complete Adjuvant (Sigma, St. Louis, MO). In the following 3 weeks, 1 mg of protein was injected mixed with 1 mg of Freund's Incomplete Adjuvant (Sigma, St. Louis, MO). The serum supernatant were obtained after centrifugation of collected blood samples subjected to immunoblot analysis at 4 °C, 12,000 rpm for 15 min.

To test the quality of resulting anti-BxPrx serum, 30 µg of total protein extract from *B. xylophilus* and *B. mucronatus* were separately loaded in 12 % SDS-PAGE for the immunoblot analysis. The protein bands were transferred from SDS-PAGE to a nitrocellulose (NC) membrane. Goat anti-rabbit IgG-alkaline phosphatase (Fermentas, MD) (diluted by 1: 10000) was used as the secondary antibody. Immunoreactions were visualized by adding 5-bromo-4-chloro-3-indolyl-phosphate and p-nitroblue tetrazolium chloride (Promega, WI).

### Immunohistocalization

Mixed stages of *B. xylophilus* were fixed in the solution of paraformaldehyde (4 %) and glutaraldehyde (2 %) in 0.1 M phosphate buffered saline (PBS) (pH 7.4), dehydrated in an ethanol series and embedded in LR (London Resin Company) White resin with polymerization at 60 °C. Ultrathin sections (60-80 nm) were cut with a diamond knife on a Reichert Ultracut Ultramicrotome (Reichert Company, Vienna, Austria). For immunohistochemistry, the grids were subsequently floated on droplets of the following solutions: PBS (containing 50 mM glycine), PBGT (PBS containing 0.2 % gelatine, 1 % bovine serum albumin and 0.02 % Tween-20), the primary antibody (Sigma) diluted with PBGT, and the secondary antibody diluted with PBGT. Optional silver intensification increased the size of the gold granules from 10 to about 40 nm, and 2 % of uranyl acetate increased the tissue contrast for observation in the transmission electron microscope (HITACHI H-7500, Hitachi, Tokyo, Japan). The primary antibody was diluted 1: 2000 and incubated overnight at 4 °C. The secondary antibody, an anti-mouse IgG which was coupled to 10 nm colloidal gold (AuroProbe EM, GAR G10; Amersham Biosciences, Little Chalfont, Buckinghamshire, UK), was diluted 20-fold and incubated at room temperature for 60-90 min.

## Results

### Cloning and sequence analysis of BxPrx

The full length cDNA of *BxPrx* was determined through RACE-PCR and was deposited in GenBank under accession number EU095848. The cDNA of *BxPrx* contains 588 bp ORF encoding 195 amino acids. The predicted isoelectric point (pI) and molecular weight (MW) are 6.54 and 21,854 Da, respectively. *BxPrx* contained two conserved functional cysteine regions flanked by valine and proline residues (VCP). The cysteines, located at the peptide positions 49 and 170, respectively, are generally referred to as Cp and Cr (Fig. 1A and 1B). *BxPrx* is, therefore, classified as a 2-cys peroxiredoxin [17]. Moreover, Cp is a part of the catalytic triad. Two conserved motifs GGLG and YF are also identified in *BxPrx*, which are critical as a signal transmitter through sensitive recognition of H<sub>2</sub>O<sub>2</sub> changes. Prx2 have been found in human, animal, plant-parasitic, and free living nematodes, and *BxPrx* shares a high degree of similarity with Prx2 from other nematodes, such as *Ascaris suum* [159/195 (81 %)], *Caenorhabditis elegans* [157/195 (80 %)], *Globodera rostochiensis* [153/193 (79 %)].

A catalytic triad consisting of threonine, cysteine, and arginine are found to be in close spatial proximity and forms a triangle shape in their 3-D tertiary structure to facilitate the redox catalytic reaction (Fig. 1B). The two motifs GGLG and YF are located at the opposite ends in the tertiary configuration, and Cp is closer to the ATP binding motif GGLG to initiate the catalytic reaction of *BxPrx* with the supply of ATP. Motif YF helix is located above the Cr in their spatial configuration, and this arrangement could potentially block the reduction reaction of *BxPrx* from the oxidized status and it is important for Prx2's function as a signal transmitter.

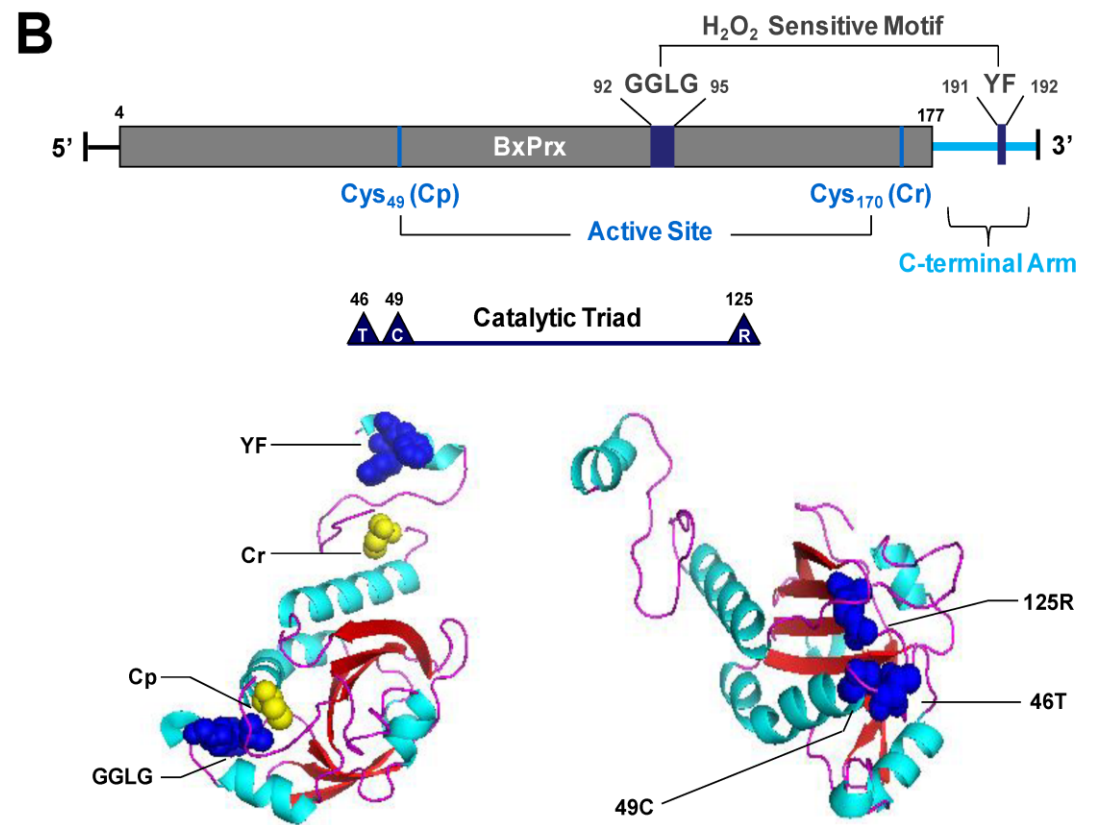
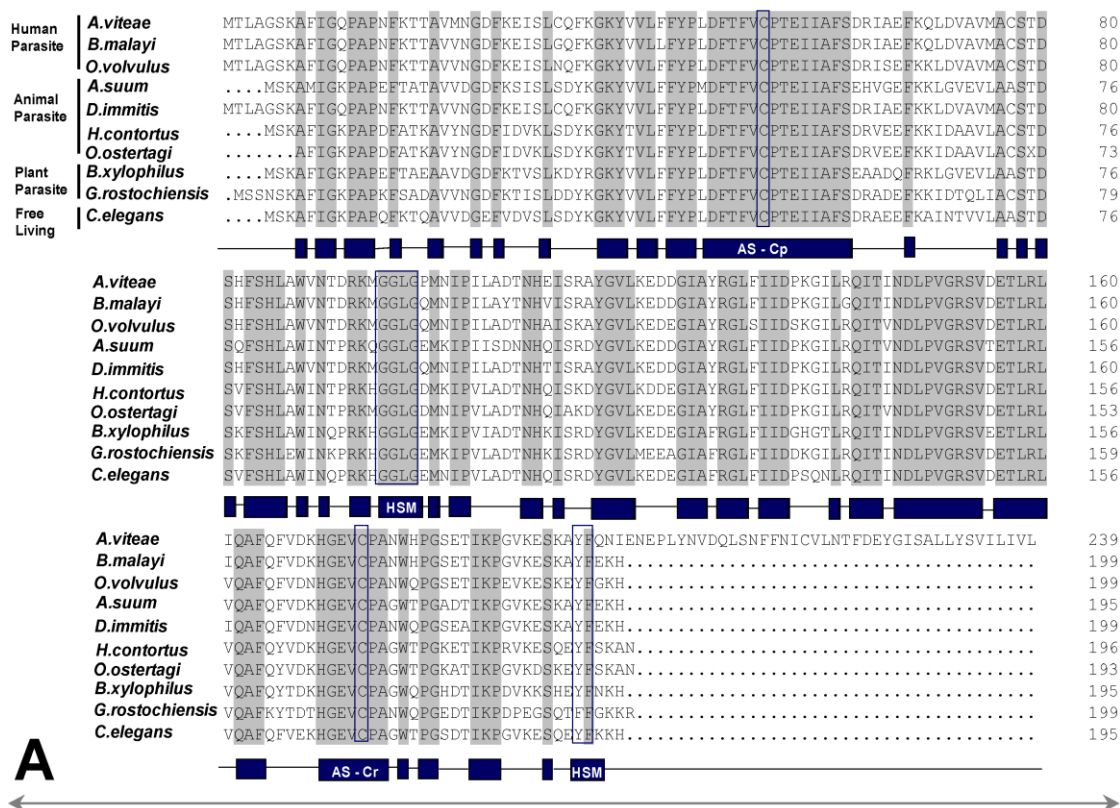
### Phylogenetic analysis

The phylogenetic relationship of *BxPrx* with other Prx2s from the animal kingdom is depicted in Fig. 2. Phylogenies derived from both neighbor joining (NJ) and maximum parsimony (MP) methods were generally congruent, and the results of the two phylogenetic trees and associated bootstrap supports are displayed in Fig. 2. Prx2 sequences came from five clades of nematodes with different life histories (Table 1).

**Table 1.** *BxPrx* homologues in nematodes

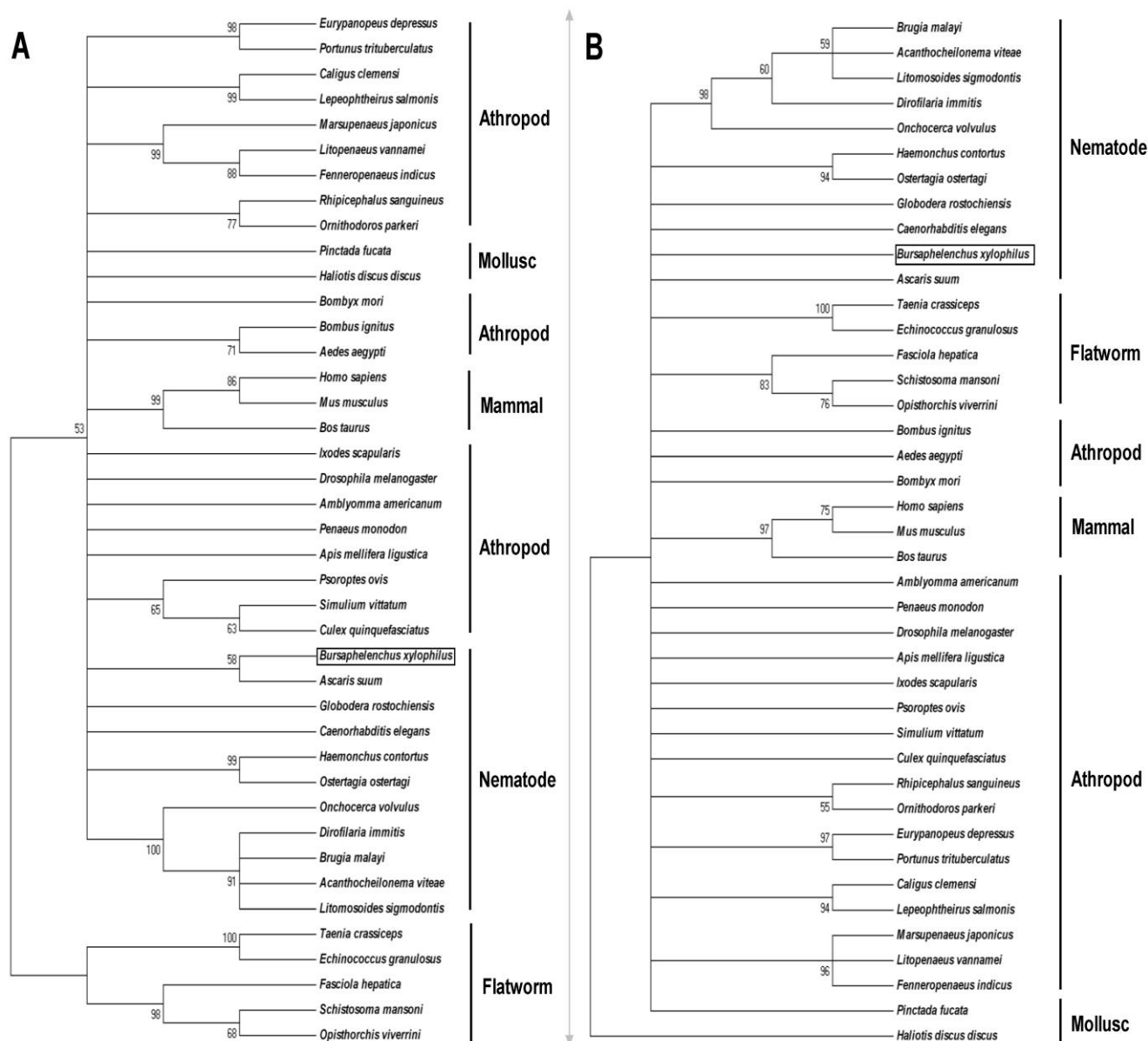
Clade	Species	Host	Contig ID	RF <sup>a</sup>	H-Score <sup>s</sup>	e-Value <sup>e</sup>
Clade I	<i>Xiphinema index</i>	Plants	XI01176	+2	806	6.0e <sup>-81</sup>
	<i>Trichinella spiralis</i>	Human	TS00204	+3	573	4.7e <sup>-56</sup>
Clade III	<i>Ascaris suum</i>	Human	AS00117	+1	<b>849</b>	5.1e <sup>-85</sup>
	<i>Toxocara canis</i>	Dogs, cats, and human	TX02017	+2	786	2.2e <sup>-78</sup>
	<i>Dirofilaria immitis</i>	Dogs and cats	DI00483	+2	758	2.0e <sup>-75</sup>
	<i>Brugia malayi</i>	Human	BM01405	+2	438	2.1e <sup>-41</sup>
Clade IVa	<i>Strongyloides stercoralis</i>	Human	SS01309	+2	805	5.5e <sup>-81</sup>
	<i>Parastrongyloides trichosuri</i>	Possums	PT00761	+3	804	7.6e <sup>-81</sup>
	<i>Strongyloides ratti</i>	Mice	SR00321	+1	650	1.5e <sup>-64</sup>
Clade IVb	<i>Zeldia punctata</i>	Free living (bacterivore)	ZP00073	+1	830	6.9e <sup>-83</sup>
	<i>Heterodera glycines</i>	Plant (soybean)	HG03313	+2	817	1.7e <sup>-81</sup>
	<i>Meloidogyne hapla</i>	Plants (root knot)	MH01325	+2	810	1.1e <sup>-80</sup>
	<i>Meloidogyne paranaensis</i>	Plant (coffee)	MP00535	+2	807	2.0e <sup>-80</sup>
	<i>Meloidogyne incognita</i>	Plants (root knot)	MI02462	+3	807	2.1e <sup>-80</sup>
	<i>Pratylenchus penetrans</i>	Plant (meadow)	PE00136	+3	718	5.6e <sup>-71</sup>
	<i>Meloidogyne arenaria</i>	Plants (root knot)	MA03144	+1	680	6.1e <sup>-67</sup>
	<i>Meloidogyne javanica</i>	Plants (root knot)	MJ00986	+2	611	1.1e <sup>-59</sup>
	<i>Globodera pallida</i>	Plant (potato cyst)	GP03375	+2	560	2.7e <sup>-54</sup>
	<i>Ditylenchus africanus</i>	Plant (peanut)	DA00204	+3	450	1.3e <sup>-53</sup>
	<i>Globodera rostochiensis</i>	Plant (potato cyst)	GR13953	+2	282	4.8e <sup>-43</sup>
	<i>Meloidogyne chitwoodi</i>	Plants (root knot)	MC03207	+1	239	6.4e <sup>-20</sup>
	Clade V	<i>Caenorhabditis elegans</i>	Free living (bacterivore)	F09E5.15	+1	<b>843</b>
<i>Nippostrongylus brasiliensis</i>		Rats	NB00963	+2	<b>835</b>	3.3e <sup>-83</sup>
<i>Ancylostoma ceylanicum</i>		Human and hamster	AE00970	+2	833	5.4e <sup>-83</sup>
<i>Caenorhabditis remanei</i>		Free living (bacterivore)	CR02463	+1	831	8.1e <sup>-83</sup>
<i>Pristionchus pacificus</i>		Free living (bacterivore)	PP00536	+3	831	8.4e <sup>-83</sup>
<i>Haemonchus contortus</i>		Sheep and goats	HC02224	+2	792	1.2e <sup>-78</sup>
<i>Ancylostoma caninum</i>		Dogs	AC03236	+2	761	2.5e <sup>-75</sup>
<i>Ostertagia ostertagi</i>		Cattles	OS00892	+3	367	1.2e <sup>-67</sup>

"a": Reading Frame. "s": High Score, species of the first three highest score were highlighted. "E": Smallest sum probability P (N)



**Figure 1. Sequence and structural analysis of BxPrx.** (A) Alignment of BxPrx with orthologues in other nematode species. Grey blocks represent conserved amino acids across all nematodes. The schematic drawing of conserved domains of BxPrx is shown in blue blocks. Two conserved functional active cysteine sites (AS-Cp and AS-Cr) and two H<sub>2</sub>O<sub>2</sub> sensitive (HS) motifs (GGLG and YF) are highlighted in blue boxes. Accession numbers of Prx2s used in this study are *Acanthocheilonema viteae*

[AAL91102], *Brugia malayi* [Q17172], *Onchocerca volvulus* [AAC32810], *Ascaris suum* [Q9NL98], *Dirofilaria immitis* [AAC38831], *Haemonchus contortus* [AAT28331], *Ostertagia ostertagi* [CAD20737], *Bursaphelenchus xylophilus* [ABW81468], *Globodera rostochiensis* [CAB48391], and *Caenorhabditis elegans* [NP\_872052]. (B) Schematic drawing of BxPrx primary and tertiary structures. The grey bar shows the conserved typical 2-Cys Prx domain. The two conserved active sites (Cp and Cr), H<sub>2</sub>O<sub>2</sub> sensitive motifs (GGLG and YF), and catalytic triad (Thr-Cys-Arg) are highlighted in BxPrx tertiary structure as colored spheres. One sphere represents one of the atoms in the corresponding amino acids.

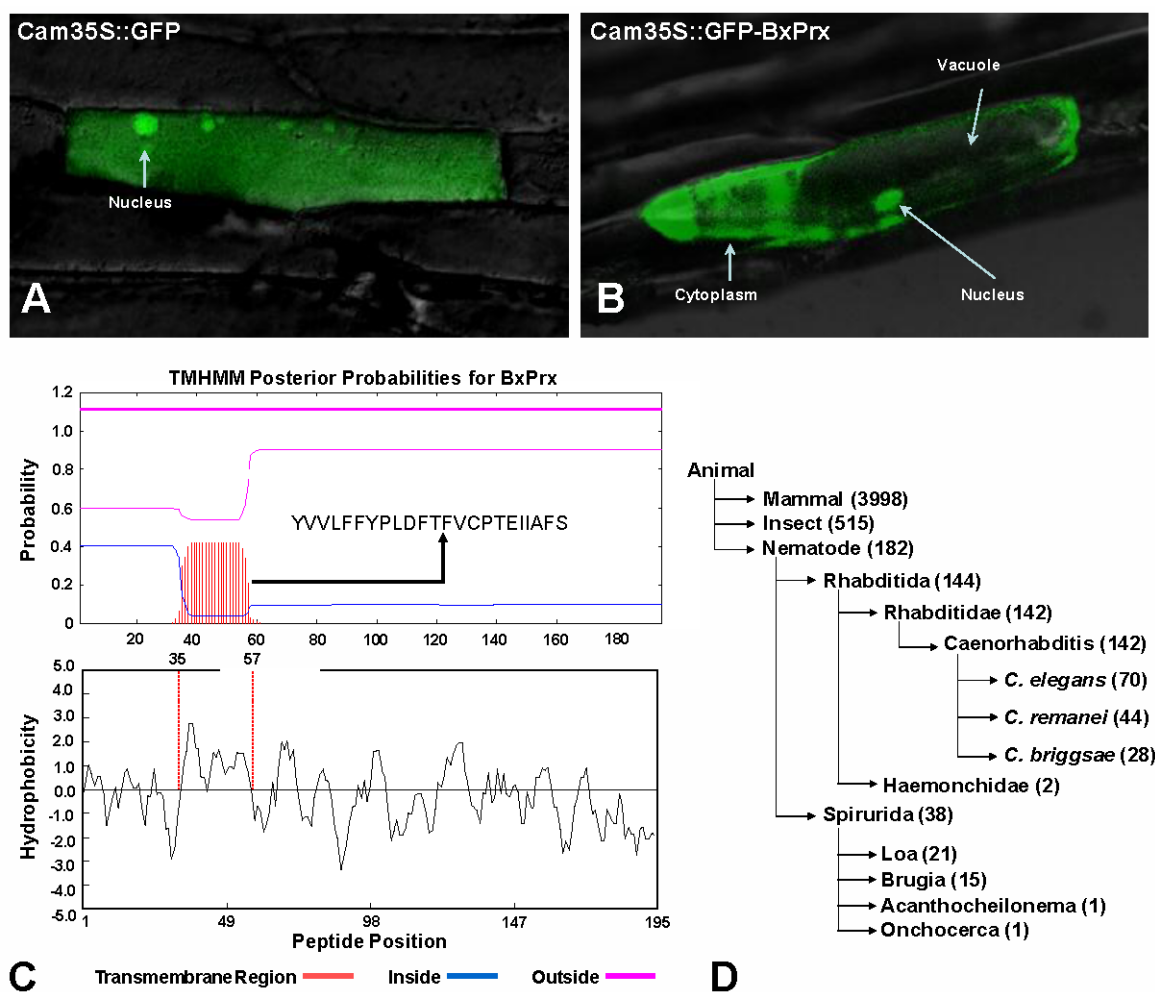


**Figure 2. Phylogenetic relationship of BxPrx with Prx2s from other organisms. (A)** Neighbor-joining (NJ) consensus tree. **(B)** Maximum parsimony (MP) consensus tree. The branch length of the NJ and MP tree indicates evolutionary distance. Accession numbers of the Prx2s used in this study are *Xenopus tropicalis* [NP\_989001], *Ascaris suum* [Q9NL98], *Onchocerca volvulus* [AAC32810], *Onchocerca ochengi* [AAC77922], *Dirofilaria immitis* [AAC38831], *Brugia malayi* [Q17172], *Acanthocheilonema viteae* [AAL91102], *Litomosoides sigmodontis* [AAG10102], *Ostertagia ostertagi* [CAD20737], *Haemonchus contortus* [AAT28331], *Caenorhabditis elegans* [NP\_872052], *Globodera rostochiensis* [CAB48391], *Bursaphelenchus xylophilus* [ABW81468], *Fenneropenaeus indicus* [ACS91344], *Bombus ignitus* [ACP44066], *Helicoverpa armigera* [ABW96360], *Bombus mori* [NP\_001037083], *Osmerus mordax* [ACO09982], *Thunnus maccoyii* [ABW88997], *Danio rerio* [NP\_001002468], *Mus musculus* [NP\_035693], *Rattus norvegicus* [AAH58481], and *Homo sapiens* [NP\_005800]. Calculated bootstrap value is attached to each branch and the value is a confidence level supporting this arrangement. BxPrx is highlighted in a box.

The deduced ORFs have the two conserved functional cysteine sites which suggest a similar antioxidant function through the same catalytic mechanism. Among all the sequences, Prx2 from *A. suum* has the highest identity with BxPrx which is consistent with the results of phylogenetic analysis, in which BxPrx has the closest evolutionary relationship with a Prx2 in *A. suum*. Additionally, two main clades are resolved from the NJ tree, flatworms and a cluster of nematodes, arthropods and mammals; In MP tree, all the Prx2s originated from a same evolutionary root, only with an isolated branch of *Haliotis discus discus* from taxonomic level of mollusc.

### Cellular distribution of BxPrx

Based on the primary structure analysis, there is no signal peptide at the N-terminal of BxPrx, which is consistent with other known Prx2s. However, the cellular distribution study using a GFP reporter system demonstrated that BxPrx is located within both the cytoplasm and nucleus of the onion epidermis cell (Fig. 3A and 3B). The green fluorescent signals emitted by the blank Cam35S-GFP construct were detected in all portions of the onion cell, including the nucleus, cytoplasm and vacuole (Fig. 3A); whereas signals from the Cam35S::GFP-BxPrx construct were predominately located in the nucleus and cytoplasm (Fig. 3B).



**Figure 3. Intracellular localization of BxPrx.** (A) 35S::GFP, GFP fluorescence signal was ubiquitously distributed in the whole cell. (B) 35S::GFP-BxPrx, GFP signal was strictly located in the cytoplasm and nucleus. The photo was created by overlaying the differential interference contrast light micrographs with the fluorescence micrographs. C & D presents two plausible transmembrane mechanisms for BxPrx to reach the cytoplasm. (C) Transmembrane region within BxPrx. A 23-amino acid transmembrane region was predicted by TMHMM. The sequence of the predicted transmembrane region is provided, and the transmembrane helix is highlighted in red. Hydrophobicity profile of BxPrx was analyzed by DNAMAN. The 23 consecutive hydrophobic amino acids coincide with the single transmembrane region in BxPrx. (D) A survey of the available ATP-binding cassette transporter genes in animals including nematodes.

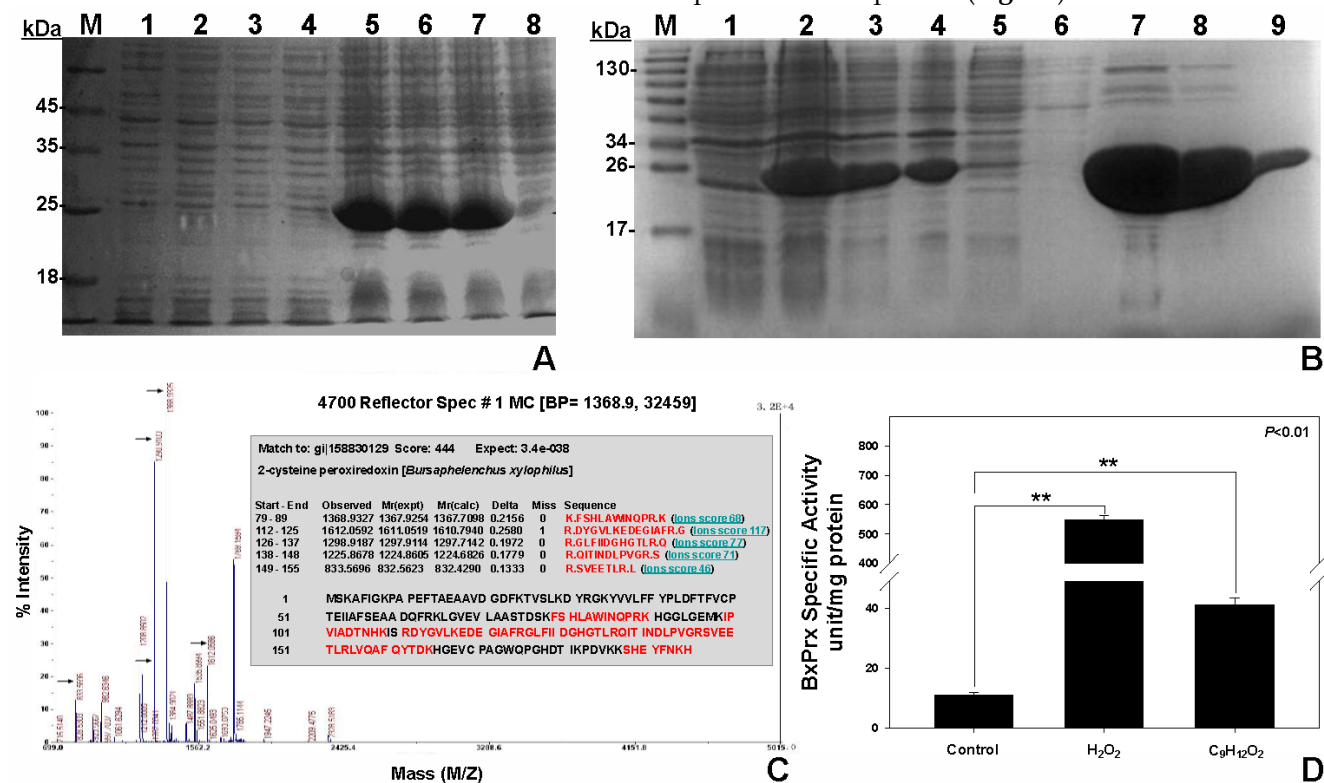


The *in silico* secretome study supports the cellular localization of BxPrx in the onion epidermal cells. The estimated NN-score of BxPrx as a secretome is 0.597 (above the default threshold of 0.5), which means that BxPrx is a non-classical secretable protein. These results indicate that BxPrx, a leader-less peptide, can be secreted and enters the cytoplasm without a leading signal peptide. A transmembrane helix composed of 23 hydrophobic amino acids provided a plausible transmembrane mechanism for the leader-less BxPrx (Fig. 3C). In addition, surveying GenBank shows that ATP-binding cassette transporters (ABC-transporter) are commonly found in animals including all nematodes for which sequence information is available (Fig. 3D). ABC transporters typically share a highly conserved ATPase domain and provide energy for transportation and other biological processes. Therefore, it is germane to speculate that transmembrane helix and ABC transporters could

potentially facilitate the transmembrane activity of the leader-less BxPrx in *B. xylophilus*.

### Recombinant BxPrx: optimization and validation

Three concentrations of IPTG (1, 2, and 5 mM) and two different incubation temperatures (20 and 37 °C) were examined to maximize the cytosolic induction of recombinant pET-28a-BxPrx (Fig. 4). The optimal induction conditions were determined to be 1 mM IPTG and 20 °C for 16 hrs because 1) the induction of recombinant BxPrx by IPTG showed no significant improvement beyond 1 mM concentration (Fig. 4A), and 2) the yield of soluble BxPrx recombinant protein was greatly reduced at 37 °C, although the overall production of BxPrx recombinant protein was significantly increased at a higher incubation temperature. Under optimal conditions, 100 ml of bacterial culture produced 0.949±0.025 g of precipitate. After a single step purification procedure (Ni-NTA agarose), it generated 1.233±0.067 mg of purified BxPrx protein (Fig. 4B).



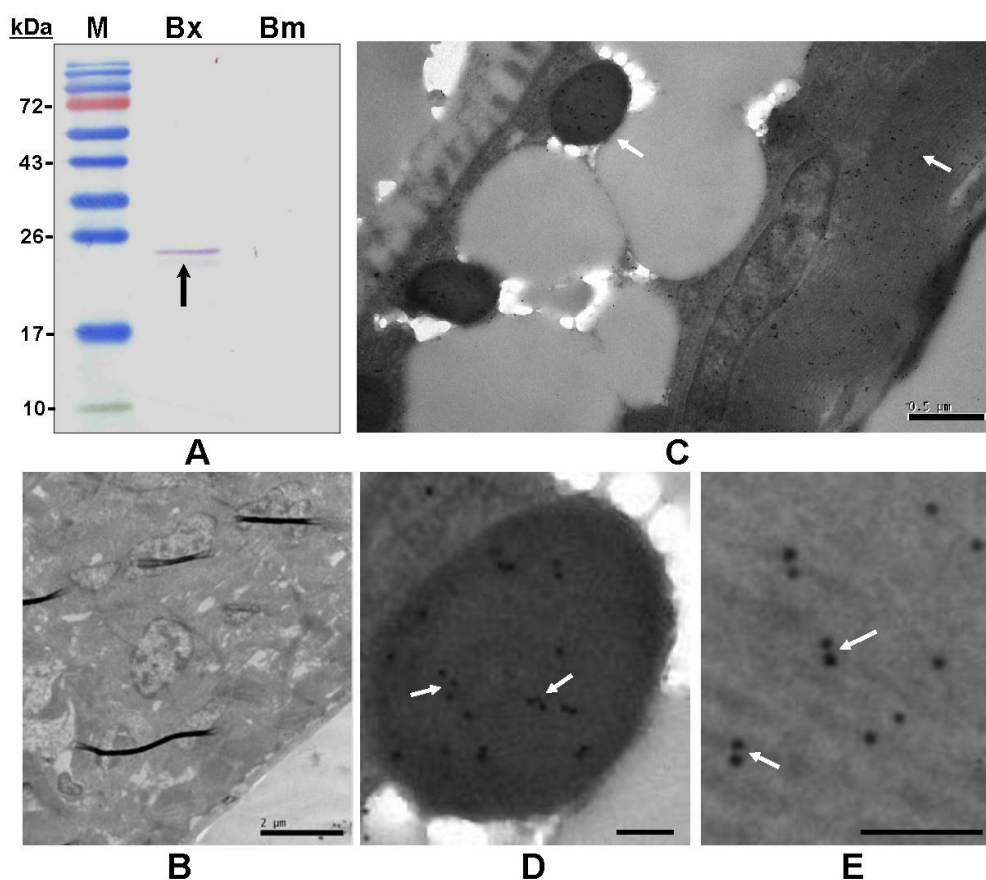
**Figure 4. Recombinant protein expression, purification and validation.** (A) Optimal concentration of IPTG for the induction of recombinant BxPrx. 1, 2, 3, and 4 are the protein expressions with blank pET-28a plasmid induced by 5, 2, 1, and 0 mM of IPTG, separately; 5, 6, 7, and 8 are the protein expressions of recombinant BxPrx induced by 5, 2, 1, and 0 mM of IPTG, respectively. (B) Recombinant BxPrx expression after induced by 1mM IPTG at 20 °C for 16 hrs. (1) without IPTG, (2) crude, (3) supernatant after sonication, (4) resuspended pellet after sonication, (5) 1<sup>st</sup> flow-through after initial protein loading (6) flow-through after washing solution, and (7-9) the 2<sup>nd</sup>, 3<sup>rd</sup>, 10<sup>th</sup> fraction of the eluted BxPrx. (C) Mass spectrometry test of the purified BxPrx. 5 peaks (highlighted with arrows) in the mass spectrum matched with 5 peptide segments (inset, highlighted in red) of the BxPrx protein in *B. xylophilus*. (D) Specific activity assay of recombinant BxPrx. Control, without BxPrx; Two different substrates, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and cumene hydroperoxide (C<sub>9</sub>H<sub>12</sub>O<sub>2</sub>), respectively, were used for the activity assay of recombinant BxPrx.

The identity of heterogeneously expressed protein was validated by the mass spectrometry analysis (Fig. 4C). The purified recombinant protein exhibited high sequence homology with 5 peptide fragments from BxPrx. The specific activity of BxPrx was measured at  $547.2 \pm 15.2$  units/mg protein and  $41.1 \pm 2.3$  units/mg protein to reduce hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) and cumene hydroperoxide ( $\text{C}_9\text{H}_{12}\text{O}_2$ ), respectively, with a negative control of  $11.1 \pm 0.8$  units/mg protein (Fig. 4D). Recombinant BxPrx showed highly specific activity toward hydrogen peroxide.

#### Immunoblot analysis and immunohistocalization study

The polyclonal antiserum raised against the recombinant BxPrx recognized the target BxPrx as a single band, but only detected the protein extracted from *B. xylophilus* not *B. mucronatus* (Fig. 5A). This

result suggested that the Prx2 in *B. mucronatus* either has a low homology with BxRrx or is expressed at significantly lower concentrations. *Bursaphelenchus mucronatus* is closely related to *B. xylophilus*, but only the latter one causes pine wilt disease. The differences in Prx2 structure and expression profile may lead to the distinctive differences in pathogenicity between the two species. Immunohistochemistry was carried out to determine the distribution of BxPrx in tissues (Fig. 5B-E). The results showed that BxPrx was abundantly expressed in *B. xylophilus* and the protein was predominantly localized under the cuticle surface (Fig. 5C), i.e., in the particles inside the body (Fig. 5D) and the muscle cells under the body surface (Fig. 5E). The negative control with preimmune serum showed no detectable binding in *B. xylophilus* samples (Fig. 5B).



**Figure 5.** Immunoblot analysis and immunohistocalization of BxPrx in *B. xylophilus*. (A) Immunoblot analysis of a polyclonal antiserum raised against recombinant BxPrx to total proteins extracted from *B. xylophilus* and *B. mucronatus*, respectively. M, Molecular marker; Bx, *B. xylophilus*; Bm, *B. mucronatus*. A single band was detected in *B. xylophilus*, but not in *B. mucronatus*. (B-E) Immunohistocalization of BxPrx in *B. xylophilus* using preimmune serum from a healthy rabbit as a negative control (B). BxPrx antiserum is found to bind to particles (D) and muscle fiber cells (E) under the cuticle of *B. xylophilus* (C). D and E are the close-up images of C. The scale bars of B-E are 2  $\mu\text{m}$ , 0.5  $\mu\text{m}$ , 0.1  $\mu\text{m}$  and 0.1  $\mu\text{m}$  respectively.

## Discussion

### BxPrx gene discovery

*Prx2* is a common theme in parasitic and free-living nematodes, and its primary structure is highly conserved. The alignment of BxPrx with Prx2 from other species demonstrates that BxPrx has two highly conserved cysteine functional sites for antioxidant activity. The N-terminal cysteine is referred to as peroxidatic cysteine (Cp), and the resolving cysteine (Cr) is located at the C-terminus. Cp is the primary site of enzyme oxidation which can attack peroxides that is subsequently oxidized to a cysteine sulfenic acid (Cp-SOH) [18]. Then, the reduction of the enzyme to its original active reduced state involves thioredoxin and Cr [31, 32]. The antioxidant catalytic reaction of BxPrx is assisted by the catalytic triad composed of threonine, cysteine, and arginine. In the BxPrx primary structure, arginine is about 80 amino acids away from threonine and cysteine, but they are in close proximity in the tertiary structure to initiate the catalytic reaction. In the following catalytic reaction, deprotonation of the catalytic cysteine (49 Cys) thiol group is facilitated by H-bonding with the threonine hydroxyl group, and the positive charge of the arginine side chain. Therefore, the conserved catalytic triad is critical to the antioxidant activity of BxPrx; the destruction of this tertiary structure could lead to the loss of antioxidative activity.

BxPrx also has two conserved motifs of Gly-Gly-Leu-Gly (GGLG) near the N-terminus and Tyr-Phe (YF) in the C-terminal arm. GGLG and YF are common features of Prx2s, and they can detect the fluctuation of hydrogen peroxide levels in the environment [33, 34]. The GGLG motif is a part of the ATP binding site in the complex structure of peroxiredoxin [33] and is located in close proximity to Cp in the BxPrx tertiary structure, which could provide a convenient energy supply for the catalytic reaction. The catalytic redox reaction of peroxiredoxin is an ATP-dependent reaction. As a H<sub>2</sub>O<sub>2</sub> sensitive motif, GGLG can react quickly and supply energy for the initiation of an antioxidant reaction for Prx2 when H<sub>2</sub>O<sub>2</sub> levels increase. Therefore, GGLG potentially plays a critical role in the antioxidative mechanism of BxPrx. On the other hand, a YF-containing C-terminal helix is observed to be located above the active site Cr in the tertiary structure of BxPrx to prevent unfolding, which will cause a longer reaction of the active site Cr. Consequently, Prx is prone to inactivation when challenged with high levels of H<sub>2</sub>O<sub>2</sub>, i.e., Prx is sensitive to increasing H<sub>2</sub>O<sub>2</sub> [34]. Therefore, by having two sensitive motifs for H<sub>2</sub>O<sub>2</sub>, BxPrx should also play a role in the signal transmission of *B. xylophilus*, in terms

of sensing the ROS defense of the host plant and detoxifying ROS as an antioxidant enzyme.

The phylogenetic analysis showed that Prx2 in mammals, arthropods and nematodes originated from the same root, which suggests that Prx2 could play a similar function in all these organisms. Among them, BxPrx had the shortest genetic distance to a Prx2 in *Ascaris suum* in the constructed NJ tree with bootstrap value of 58 on the root branch of the two Prxs; in MP tree, the bootstrap value on the root branch of the two Prxs is 46, so it is cutoff when values over 50 are shown. *Ascaris suum* (a large roundworm) is a parasitic nematode that causes ascariasis in pigs. Although the hosts of the two parasites belong to different taxonomic groups, the two parasites have similar transmission pathways and have similar life histories in their respective hosts. *Bursaphelenchus xylophilus* is transmitted by insect vectors such as the Japanese pine sawyer beetle, and *A. suum* also utilizes a dung beetle as an intermediary for distribution. They are both migratory parasites and they can move within the host to feed. Similar transmission pathways and living conditions result in similar parasite-host relationships, which suggests a similar function for their antioxidant Prx.

### Transmembrane mechanism of the leader-less BxPrx

Based on an array of web-based bioinformatics analyses, BxPrx, a leader-less peptide, is predicted to have transmembrane capabilities. The outcome of this *in silico* prediction was validated by both nuclear- and immunohistochemical studies. Unlike classical secretion via the endoplasmic reticulum (ER)/Golgi Body pathway for proteins with signal peptides, non-classical secretion of leaderless proteins requires a structural accommodation (e.g., transmembrane regions within the leader-less peptides) or a carrier (transmembrane proteins such as the ABC transporter). In this study, a stretch of 23 consecutive hydrophobic amino acids assembles the only transmembrane helix on the OFR of BxPrx, which potentially can assist BxPrx transmission through the hydrophobic lipid bilayer of the cell membrane. A domain swap or mutation at the transmembrane helix would provide a definitive answer for the involvement of this transmembrane mechanism in *B. xylophilus*.

ABC transporters are members of a protein super-family that is one of the largest and most ancient families found in all extant phyla from prokaryotes to humans [35]. As transmembrane proteins, ABC transporters can carry a wide variety of substrates across extra- and intra-cellular membranes using the

energy from ATP hydrolysis [36]. Moreover, ABC transporters, which are prevalent among all nematodes, were considered as a possible transmembrane mechanism for the ER/Golgi-independent leader-less secretion pathway in a parasitic flatworm, liver fluke *Fasciola hepatica*, especially for the secretion of the leader-less *F. hepatica* antioxidants such as Prx2 [37]. Therefore, we propose that leader-less BxPrx is secreted either by its transmembrane region or by a specific ABC transporter or by both mechanisms working together in *B. xylophilus*.

### Characterization of recombinant BxPrx

Higher induction temperatures lead to a greater production of recombinant protein within a shorter time which typically results in the abnormal folding of a protein and the formation of an inclusion body. In this study a large quantity of active recombinant BxPrx was obtained by induction at optimal conditions (20 °C overnight with 1 mM IPTG). Zipfel *et al* [38] recombinantly expressed a Prx2 from the human parasitic nematode, *Onchocerca volvulus*. The detailed characterization of this recombinant Prx2 indicated that *O. volvulus* Prx2 predominantly expressed at the host-parasite interface, which was important for the infection by this parasitic nematode.

The different reactions on the immunoblots between *B. xylophilus* and *B. mucronatus* suggest differences at either the Prx2 genes or Prx2 expression profiles. *Bursaphelenchus mucronatus*, an indigenous species in China, is very similar to the morphology and biology of *B. xylophilus* [39]. However, *B. mucronatus* and *B. xylophilus* have distinctively different pathogenicities. *Bursaphelenchus mucronatus* has weak pathogenicity [40] and the disease prevalence of *B. mucronatus* is far less than that of *B. xylophilus* [3]. It was reported that the propagation of *B. xylophilus* was higher than *B. mucronatus*, and even higher under competitive conditions in laboratory cultures [39]. After more than two-decades of invasion, *B. xylophilus* has significantly increased its colonization capability and has replaced *B. mucronatus* in natural forest ecosystems in China [3]. This phenomenon might be the result of a differentially expressed gene associated with the pathogenicity and invasive capability of the two parasites. Most recently, a microRNA survey provided novel information on the regulation of growth, development, behavior, and stress response in *B. xylophilus* at the epigenetic level [25]. Combining genomic and epigenomic resources in *B. xylophilus* will shed light on the molecular underpinnings that govern the infection and distribution of *B. xylophilus*.

### Tissue localization of BxPrx

The immunohistolocalization study with a polyclonal antiserum against recombinant BxPrx suggest that: 1) BxPrx is fairly abundant, 2) this antioxidant can be found in muscle cells and particles, and 3) it is widely distributed and concentrated underneath the cuticle of the nematode. This localization profile coincides with Prxs in other parasitic nematodes. A 2-Cys Prx from a plant parasitic nematode, the yellow potato cyst nematode *Globodera rostochiensis*, was found on the cuticle surface and in the stylet secretions of infective and post-infective juveniles [41, 42]. The other 2-Cys Prx was localized in the hypodermis and cuticle in the infective larvae of *Onchocerca volvulus*, the nematode pathogen causing onchocerciasis or 'river blindness' disease in Africa [38]. Based on the abundance, distribution, and localization profile of BxPrx, it is germane to speculate that the secretion of this antioxidant in the cuticle region could be an adaptive response to protect this plant parasitic nematode against the host immune responses.

### Summary

Prx has been found in a wide range of organisms, including bacteria, plants, invertebrates and mammals [18], and has been shown to play important roles in parasite-host interactions. In this study, a 2-cysteine peroxiredoxin gene in *B. xylophilus* (BxPrx) was successfully cloned and its primary structure was thoroughly analyzed. BxPrx, a conserved Prx2 protein, has two conserved cysteine active sites, a catalytic triad, and two H<sub>2</sub>O<sub>2</sub> sensitive motifs. The spatial configuration and the putative functions of these conserved domains were analyzed using a tertiary structure simulated by the SWISS-MODEL. Two plausible transmembrane mechanisms are discussed in light of the presence of this leader-less BxPrx in the cytoplasm and under the *B. xylophilus* cuticle. The induction of recombinant BxPrx is optimized, and the purified recombinant protein exhibited specific Prx activity neutralizing H<sub>2</sub>O<sub>2</sub>. The combined results from the gene discovery, protein localization, and recombinant protein characterization suggest that BxPrx potentially plays a key role in combating the oxidative burst engineered by the ROS defense system in host plants during the infection process. In short, BxPrx is a putative genetic factor facilitating *B. xylophilus* infestation of pine tree hosts.

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### Author Contribution

ZL, YC, and YW conceived the experiments; ZL, XL, QZ, and XZ designed the study; ZL, QZ, and XZ analyzed the data; and ZL, QZ, and XZ wrote the manuscript. All authors read and approved the final version of manuscript.

### Conflict of Interests

The authors have declared that no conflict of interest exists.

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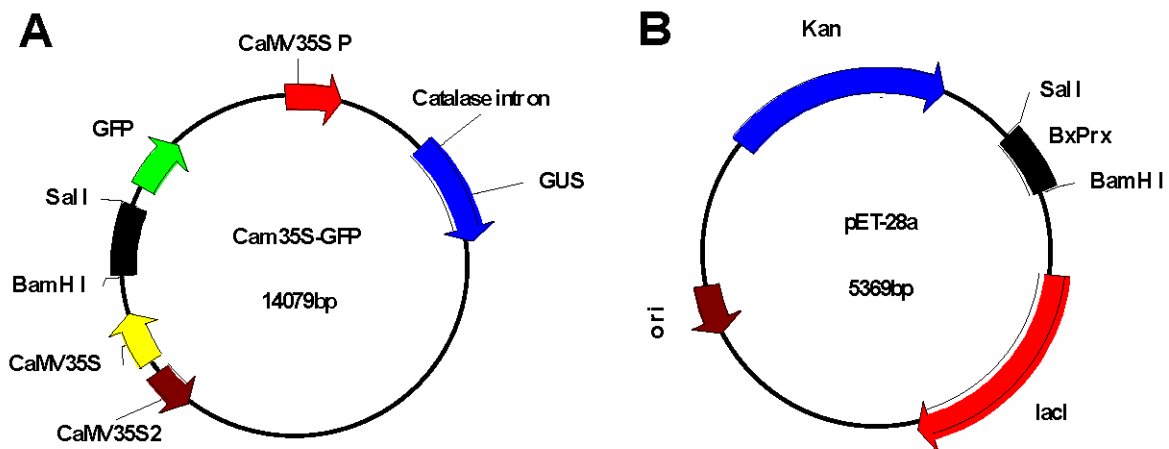
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## Additional Tables and Figures

**Table S1.** Primer sequences for the expression of recombinant BxPrx

Primer ID	Primer Sequence (5'-3')
BxPrx3S	TTYGTNTGYCCNACNGAR
BxPrx5R	NGCNGGRCANACYTCNCC
BxPrxSB	CGGGATCCATGTCCAAGGCTTTCATT*
BxPrxRS	ACGCGTCGACTTAATGTTTGTGAAATA**

“\*”: BamH I recognition site is underlined . “\*\*”: Sal I recognition site is underlined.



**Figure S1.** Construction of recombinant plasmid vectors. Recombinant plasmids of Cam35S-GPS-BxPrx (A) and pET-28a-BxPrx (B), respectively, were constructed by ligating the entire ORF of *BxPrx* into the Cam35S-GFP between restriction sites BamH I and Sal I.