Respiration

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Lung Ultrasound in the Emergency Department for Early Identification of COVID-19 Pneumonia

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Keywords

Lung ultrasound \cdot COVID-19 \cdot Pneumonia diagnostics \cdot Viral infection \cdot Emergency department

Abstract

Background: Coronavirus disease 2019 (COVID-19) is a pandemic overwhelming the health care systems worldwide. Lung ultrasound (LUS) use has been proposed to identify suspected COVID-19 patients and direct them to the isolation area in the emergency department (ED) or to discharge them for outpatient treatment. **Objective:** Our aim was to retrospectively investigate the use of LUS in the ED to identify COVID-19 pneumonia (CP). Methods: We performed a retrospective single-center study including all patients accessing the ED who underwent LUS examination for suspicion of COVID-19 during the initial outbreak. Demographics, clinical parameters, laboratory values, imaging features, and outcome variables were collected. The receiver operating characteristic (ROC) curve was used to evaluate diagnostic accuracy. Results: A total of 41% patients were COVID-19-positive; 67% of them were diagnosed with CP. The ROC curve of the LUS score showed an area under the curve of

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0.837 (95% CI 0.75–0.92) and with a cutoff value \geq 3 identified 28 of 31 patients with CP and 11 of 15 without (sensitivity 90%, 95% CI 74–97%; specificity 75%, 95% CI 56–76%). LUS in combination with nasopharyngeal swab has a sensitivity of 100% (95% CI 74–97%) and a specificity of 61% (95% CI 44–67%). **Conclusions:** LUS is a promising technique for early identification of CP in patients who accessed the ED in an active epidemic time. The LUS score shows a sensitivity of 90% for CP, allowing to quickly direct patients with CO-VID-19 to the ED isolation area or to discharge them for outpatient treatment. \otimes 2020 S. Karger AG, Basel

Introduction

Coronavirus disease 2019 (COVID-19) is a pandemic overwhelming the health care systems worldwide [1]. Since the beginning of March 2020, emergency departments (EDs) in South Tyrol, Italy had to address an ex-

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Giacomo Strapazzon Institute of Mountain Emergency Medicine Eurac Research, Viale Druso 1 IT--39100 Bolzano (Italy) giacomo.strapazzon @eurac.edu pected increasing wave of suspected COVID-19 patients to the ED, and different management and triage strategies were undertaken. History of exposure, clinical and laboratory characteristics, imaging tests including chest X-ray, computed tomography (CT), and lung ultrasound (LUS), and real-time polymerase chain reaction (RT-PCR) assay from specimens [2] have guided the emergency provider decision process of suspected COVID-19 patients.

The clinical presentation of COVID-19 ranges from completely asymptomatic cases to very severe pneumonia showing diffuse alveolar damage and airway inflammation [3], acute respiratory distress syndrome, and multiorgan failure. COVID-19 pneumonia (CP) showed a typical CT pattern involving peripheral areas of the lung [4]. Such changes on CT have been compared to LUS findings, showing an encouraging correlation [5–7]. Thanks to the broad diffusion of the technique in Italy [8], the potential role of LUS for early identification of CP was initially proposed by the colleagues in Lodi (Italy) via social media [9]. LUS shows a sensitivity and specificity of 85-93% and 86-93%, respectively in the diagnosis of pneumonia [10–14]. Lu et al. [15] described a sensitivity of 69-100% and a specificity of 86-93% of LUS in the diagnosis of CP. Compared to CT, LUS is safer, repeatable, radiation-free, cheap, and allows a point-of-care use. Soldati et al. [16] proposed an international standard LUS score for CO-VID-19 patients. Various perspective papers [17-19] and video tutorials [20] further contributed to theorize report and encouraged its use in the COVID-19 outbreak. Based on the difficulties to perform a high number of CT scans to diagnose CP due to the lack of resources and dedicated areas [21], we developed a specific protocol including LUS in the evaluation of patients in the ED.

No data were available about the sensitivity and specificity of LUS for early identification of suspected CP in the ED during an active epidemic. Our aim was to retrospectively investigate the use of LUS in the ED to identify CP. Our expectation was that LUS would be able to identify CP with a high sensitivity, and its use among ED management strategies could lead to early diagnosis and management of CP, avoiding unnecessary hospital admissions and enabling EDs to allocate resources more appropriately.

Methods

Study Population, Setting, and Data Collection

This retrospective single-center study included data of all suspected COVID-19 patients who underwent LUS examination in ED triage at the Regional Hospital of Bolzano, Italy between March 24 and April 14, 2020. Demographics, clinical parameters, laboratory values, imaging features, and outcome variables were collected, and researchers analyzed only anonymized data. This study was registered in the EudraCT database (No. 2020-001785-11).

Case Definition

Patients were defined as suspect for COVID-19 when presenting two or more of the following criteria: fever, cough, dyspnea, previous contact with positive cases within the last 14 days, or coming from or having been in at-risk areas. A confirmed case of CP was defined as follows: a patient with suggestive symptoms, with imaging features of pneumonia and positive RT-PCR assay, or one with suggestive symptoms, with or without arterial blood gas (ABG) impairment, with imaging features suggestive of CP even after a first negative RT-PCR nasopharyngeal swab after consulting with infectious disease consultant, in agreement with the WHO case definition statement [22].

Laboratory Examinations

Clinical specimens for COVID-19 diagnostic testing were obtained from nasopharyngeal swab and processed by RT-PCR assay, according to the guidelines of the National Institute of Health Care, Italy [23].

Blood samples were obtained for analysis of ABG and routine laboratory, including blood count, lactate dehydrogenase, D-dimer, and C-reactive protein. Additional tests were performed based on clinical course, including N-terminal prohormone of brain natriuretic peptide, procalcitonin, and troponin-I.

Radiologic Procedures

Chest X-ray was performed with patients in semi-recumbent position and, if made possible by the patient's conditions, orthostatic using analogic portable machines (Mobilett XP Hybrid, Siemens, Erlangen, Germany) and digital machines (Alara DR Max, Siemens; GM85, Samsung, Korea).

CT scan was performed with Somatom Definition Flash and Somatom Definition Edge (Siemens). High-resolution CT was performed with protocol of acquisition without intravenous contrast medium, at the end of inspiration and at the end of expiration, with craniocaudal acquisition; the system of automated modulation of exposition (CAREDose4D) was active. One-millimeter slices were reconstructed on the axial plane with a parenchymal window and 3-mm slices with a mediastinal window, followed by 3-mm multiplanar reconstructions in the coronal and sagittal plane.

LUS Scan

Ultrasound examinations were performed by four experienced operators using a portable ultrasound device (M-Turbo, Sonosite Fujifilm Europe, Amsterdam, The Netherlands) with a convex transducer (5–2 MHz). After abdominal preset selection (no compound; filters off; general frequency setting), focus was set on the pleural line, depth at 6 cm, and low gain (i.e., to evaluate pleural gliding). The ultrasound device was protected with a plastic film; the probe was covered by a single-use film and underwent full disinfection after every use.

Patients were examined in a sitting position (when possible) with continuous scan in all intercostal spaces, covering the whole chest surface. Based on Bouhemad et al. [24], LUS features were reported in a standard sheet that included ten fields for each lung

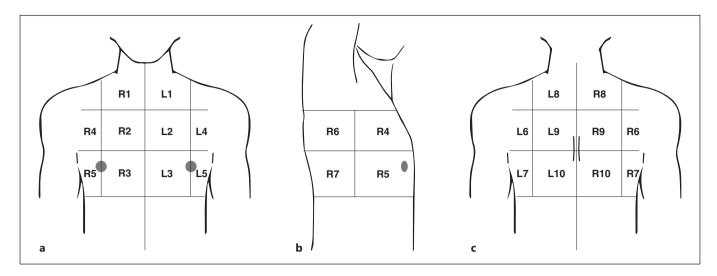


Fig. 1. Chest areas for LUS reporting. The areas allow the mapping of lung lesions to provide a panoramic vision and evaluation. For every area the operator will notice the highest score found. Each lung is divided into 10 areas. **a** Three parasternal (1 apical = apical segment of upper lobe; 2 medium = anterior segment of upper lobe; 3 lower = medial segment of medium lobe/lingular segment) for each side. **b** Two axillary anterior (4 upper = anterior segment

of upper lobe; 5 lower = lateral segment of medium lobe), and two axillary posterior (6 upper = apical segment of lower lobe; 7 lower = posterior-lateral basal segments of lower lobe). **c** Three paravertebral (8 upper = posterior segment of upper lobe; 9 medium = apical segment of lower lobe; 10 lower = posterior-lateral basal segments of lower lobe). Areas are divided as right (R) or left (L) based on the side of the patient. LUS, lung ultrasound.

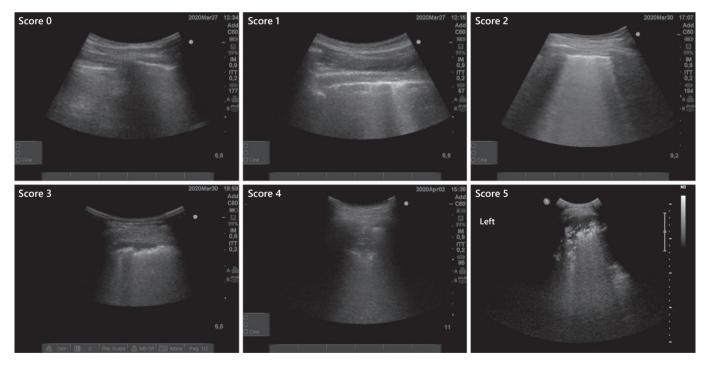


Fig. 2. LUS scoring. Score 0: normal lung surface with regular pleural line and no artifacts. Score 1: focal irregularities with vertical artifacts. Score 2: coalescent vertical artifact determining a focal white lung with mild surface irregularity. Score 3: subpleural con-

solidation causing a marked surface irregularity with underlying white lung. Score 4: focal consolidation of >2 cm. Score 5: large consolidation with air bronchograms visible in more than one area. LUS, lung ultrasound.

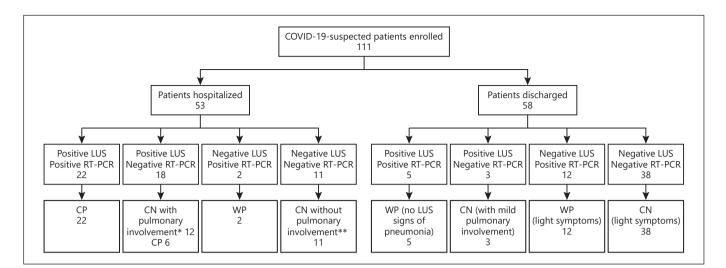


Fig. 3. Sample inclusion chart and outcome based on LUS and nasopharyngeal swab. * Congestive heart failure, carcinomatous lymphangitis, other viral pneumonia, bacterial pneumonia, pulmonary edema with renal failure, bilateral pleural effusions. ** Urinary infections, exacerbation of ulcerative rectocolitis, esophageal

varices with hematemesis, fever in chronic renal failure in dialysis, thoracic pain, legionellosis. CN, no coronavirus disease 2019; CO-VID-19, coronavirus disease 2019; CP, coronavirus disease 2019 pneumonia; LUS, lung ultrasound; RT-PCR, real-time polymerase chain reaction; WP, coronavirus disease 2019 without pneumonia.

(Fig. 1). The severity score for each field was classified in five stages, from 0 (normal A pattern) to 5 (large consolidations extended to more fields) (Fig. 2). Operators were trained for the detection of lung fissures and not to mistake them for pathologic findings [25]. LUS operators worked wearing double gloves, FFP2 or FFP3 masks, whole protection suits, as well as protective face shields and footwear, and none of them became infected during the study. The LUS score was initially aimed at describing COVID-19 lung lesions and at recording them in the clinical documentation for a possible follow-up.

Statistical Analysis

Univariate contributions of predictors and group comparisons were assessed by means of the χ^2 test for categorical data or the Kruskal-Wallis H test and the Mann-Whitney U test for continuous data. Therefore, we describe categorical data using frequencies or percentages, whereas means and standard deviations are given for continuous variables. To identify predictors that contribute independently to diagnosis, a stepwise logistic regression (forward Wald) was performed, and we computed predicted probabilities for each case. Receiver operating characteristic (ROC) curve analvsis and the estimated area under the curve (AUC) were used to quantify how accurately LUS and nasopharyngeal swabs, alone or in combination with other parameters (predicted probabilities from logistic regression), could discriminate between CP and other diagnoses, and ROC was also used to visually assess an optimal cutoff. Furthermore, sensitivity and specificity were evaluated and we calculated respective CIs using the Jeffreys method [26]. All given *p* values are two-sided and $p \le 0.05$ was regarded as significant. SPSS IBM version 26 (IBM Corp., Armonk, NY, USA) was used for statistical analysis.

Results

A total of 111 consecutive patients underwent ED evaluation with LUS assessment for suspected CP from March 24 to April 14, 2020 (Fig. 3) and were included in the study. The average duration of LUS examinations was 5–10 min.

Out of the 111 patients, 65 (59%) were COVID-19-negative and 46 (41%) were COVID-19-positive; 31 (67%) of them were diagnosed with CP and 15 (33%) had CO-VID-19 without pneumonia (WP). Demographics, clinical parameters, laboratory values, imaging features, and outcome variables are shown in Table 1 and Figure 3.

LUS pathological features in CP were mono- or bilateral, with distribution mainly in the posterior and lateral areas, consisting of pleural irregularities with increase in vertical artifacts, local and diffuse areas of "white lung" with surface irregularities, surface consolidations, and deeper confluent consolidations. No lobar or segmental distribution was observed as the lesions had often a patchy distribution. Pleural effusion was found only in 1 out of 31 CP patients. The lowest LUS score in CP patients was 3 (2 cases) and the highest score was 41 (1 case), with a mean score of 11 ± 10 in CP patients compared to 3 ± 6 in WP patients and 3 ± 6 in COVID-19-negative patients (Kruskal-Wallis test, p < 0.001).

Logistic regression analysis showed that the parameters fever (OR 6.5, 95% CI 1.1–37.3, p = 0.035), nasopha-

Variable	All (<i>n</i> = 111)	CP (<i>n</i> = 31)	WP (<i>n</i> = 15)	CN (<i>n</i> = 65)	p
Sex, female/male	54/57 (49/52%)	12/19 (39/61%)	8/7 (53/47%)	34/31 (52/48%)	0.430
Age, years	55±21	64±16	51±18	52±22	0.019
Patients with comorbidities	38 (41%, 25–43%)	7 (23%, 11–39%)	4 (27%, 10–52%)	27 (41%, 30–54%)	0.15
eneral symptoms		21 (600/ 50, 020/)	0 (530) 20 5(0)	40 (620/ 51 540/)	0.62
Cough Fever	69 (63%, 54–72%) 74 (68%, 59–77%)	21 (68%, 50–82%) 28 (90%, 76–97%)	8 (53%, 29–76%) 11 (73%, 48–90%)	40 (63%, 51–74%) 35 (56%, 44–68%)	0.63 0.004
Dyspnea	46 (41%, 33–51%)	13 (42%, 26–60%)	3 (20%, 6–44%)	30 (48%, 36–60%)	0.004
Weakness	14 (13%, 7–20%)	5(16%, 6-31%)	4(27%, 10-51%)	5 (8%, 3–16%)	0.11
Thoracic pain	12 (11%, 6–18%)	0	2 (13%, 3–36%)	10 (16%, 8–26%)	0.07
Jeurological symptoms					
Syncope	2 (2%, 1-6%)	1 (3%, 1–14%)	0	1 (2%, 1–7%)	
Smell/taste loss	6 (5%, 2–11%)	2 (7%, 1-20%)	2 (13%, 3-36%)	2 (3%, 1–10%)	0.54
Headache	4 (4%, 2–9%)	1 (3%, 1–14%)	0	3 (5%, 1–12%)	
Gastroenteric symptoms					
Vomiting	5 (5%, 2–10%)	2 (7%, 1–20%)	0	3 (5%, 1–7%)	0.84
Diarrhea	6 (5%, 2–11%)	4 (13%, 5–29%)	2 (13%, 3–36%)	6 (9%, 4–19%)	0.04
Others uncommon symptoms					
Confusion	2 (2%, 1–6%)	2 (7%, 1–20%)	0	0	
Paresthesia	1 (1%, 1–4%)	0	1 (7%, 1–27%)	0	0.43
Nasal obstruction	1 (1%, 1–4%)	0	0	1 (2%, 1–7%)	
Other non-COVID-19-related symptoms					
Leg edemas	4 (4%, 2–9%)	0	0	4 (7%, 2–14%)	
Abdominal pain	1(1%, 1-4%)	1 (3%, 1–14%)	0	0	0.87
Leg pain Hemoptysis	1 (1%, 1-4%) 1 (1%, 1-4%)	0	0 0	$1 (2\%, 1-7\%) \\1 (2\%, 1-7\%)$	
5pO ₂ , %	96±3	94±3	97±2	96±3	0.000
Blood gas analysis	JO <u>1</u> J	J 115	<i>)</i> / <u>-</u>	70 ±0	0.000
PaO ₂ , mm Hg	79±18	68±15	83±23	84±16	
PaO ₂ /FiO ₂	376±87	323±74	394±110	398±77	0.000
Lymphopenia (<1,100/µL)	36 (32%, 24-42%)	18 (58%, 41-74%)	3 (20%, 6-44%)	15 (23%, 14-35%)	0.002
Increased LDH (>230 U/L)	49 (46%, 37-56%)	22 (71%, 54-85%)	4 (29%, 10-54%)	23 (48%, 26-50%)	0.004
CRP, mg/dL	3.1±4.9	4.28±3.61	0.81±1.16	3.1±5.8	0.000
Other laboratory values					
NT-proBNP (>125 pg/mL)	11 (10%, 5–16%)	1 (3%, 1–14%)	0	10 (16%, 8–26%)	
Troponin-T (14 ng/L)	5 (5%, 2–10%)	1 (3%, 1–14%)	0	4 (7%, 2–14%)	0.16
Procalcitonin (>0.05 ng/mL)	12 (11%, 6–18%)	3 (10%, 2–24%)	1 (7%, 1–27%)	8 (12%, 6-23%)	0.10
D-dimer (500 ng/mL)	3 (3%, 1–7%)	0	0	3 (5%, 1–12%)	
Chest X-ray, <i>n</i> /total					
Positive ¹	33 (30%, 22–39%)	19 (63%, 45–79%)	2(14%, 3-38%)	12 (18%, 10–29%)	0.000
Negative	74 (68%, 59–76%)	10 (33%, 19–51%)	12 (86%, 61–97%)	52 (80%, 69–88%)	0.000
Uncertain	2 (2%, 1–6%)	1 (3%, 1–15%)	0	1 (2%, 1–7%)	
LUS	22 (200/ 22 200/)	25 (010/ (4 010/)	2(120/2)2(0/)	6 (00/ 4 100/)	
Positive ¹	33 (30%, 22–39%)	25 (81%, 64–91%)	2(13%, 3-36%)	6 (9%, 4-18%)	
Negative	62(56%, 47-65%)	4(13%, 5-28%)	13 (87%, 64–97%)	45 (69%, 57-79%) 14 (21%, 12, 23%)	0.000
Uncertain Mean score	16 (14%, 9–22%) 5.1±7.8	2 (7%, 1–19%) 11.1±9.6	0 1.5±2.8	14 (21%, 13–33%) 2.9±5.8	
Chest CT					
Performed in	14 (13%, 7-20%)	5 (16%, 6-32%)	1 (7%, 1–27%)	8 (12%, 6-23%)	
Positive ¹	6 (5%, 2–11%)	5 (16%, 6–32%)	(7/0, 1-27/0) 0	1(2%, 0-25%)	
Negative	7 (6%, 3–12%)	0	0	7 (11%, 5–20%)	0.002
Uncertain	1(1%, 1-4%)	0	1 (7%, 1–27%)	0	

Table 1. Descriptive statistics of the aggregated data of the entire population (111 patients), divided into CP, WP, and CN

Table 1 (continued)

Variable	All (<i>n</i> = 111)	CP (<i>n</i> = 31)	WP (<i>n</i> = 15)	CN (<i>n</i> = 65)	p
RT-PCR on swab					
Positive	36 (32%, 24-42%)	25 (81%, 64-91%)	11 (73%, 48-90%)	0	
Negative	72 (65%, 57–75%)	6 (19%, 7-36%)	3 (20%, 6-44%)	63 (97%, 88-99%)	0.000
Uncertain	3 (3%, 1–7%)	0	1 (7%, 1–27%)	2 (3%, 1–10%)	
Outcome					
Hospitalized	53 (48%, 37-57%)	28 (90%, 76-97%)	2 (13%, 3-36%)	23 (35%, 25-47%)	0.000
Discharged	58 (52%, 43-61%)	3 (10%, 3-24%)	13 (87%, 64–97%)	42 (65%, 53-75%)	

Data are shown as n (%), mean ± standard deviation, or n (%, 95% CI). CN, no coronavirus disease 2019; COVID-19, coronavirus disease 2019; CP, coronavirus disease 2019 pneumonia; CRP, C-reactive protein; CT, computed tomography; LDH, lactate dehydrogenase; LUS, lung ultrasound; NT-proBNP, N-terminal prohormone of brain natriuretic peptide; RT-PCR, real-time polymerase chain reaction; WP, coronavirus disease 2019 without pneumonia. ¹ Considered as presence of any pathological findings.

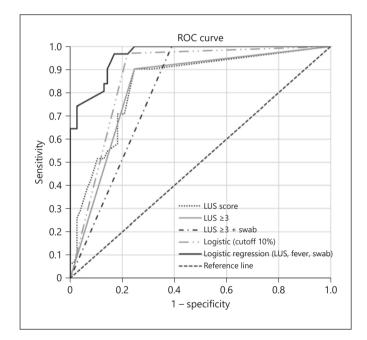


Fig. 4. ROC-AUC for CP diagnosis. Shown are ROC curves for the LUS score with an AUC of 0.84 (95% CI 0.75–0.92), applying the cutoff LUS \geq 3 with an AUC of 0.83 (95% CI 0.74–0.91), combining LUS \geq 3 with a positive or inconclusive swab with an AUC of 0.81 (95% CI 0.73–0.88), calculated probabilities from logistic regression using LUS score, fever, and swab (positive, inconclusive) with an AUC of 0.96 (95% CI 0.93–0.99), and applying the cutoff of 10% to calculated probabilities from logistic regression with an AUC of 0.87 (95% CI 0.81–0.94). AUC, area under the curve; CP, coronavirus disease 2019 pneumonia; LUS, lung ultrasound; ROC, receiver operating characteristic; ROC-AUC, area under the corresponding receiver operating characteristic curve.

ryngeal swab (OR 27, 95% CI 6–122, p < 0.001), and LUS score (OR 1.3 per unit, 95% CI 1.1–1.5, p < 0.001) were independently related to CP diagnosis.

The area under the corresponding ROC curve (ROC-AUC) for CP diagnosis using nasopharyngeal swab (positive or inconclusive) was 0.80 (95% CI 0.70–0.90) (Fig. 4) and accurately identified 25 of 31 CP patients and 14 of 15 WP patients (sensitivity 81%, 95% CI 74-97%; specificity 80%, 95% CI 44-67%). The ROC-AUC of LUS score was 0.84 (95% CI 0.75-0.92) and, when using a cutoff value \geq 3, accurately identified 28 of 31 patients with CP and 11 of 15 with WP (sensitivity 90%, 95% CI 74-97%; specificity 75%, 95% CI 56-76%). A ROC-AUC of LUS in combination with nasopharyngeal swab (one of both positive) of 0.81 (95% CI 0.73-0.89) accurately identified all 31 patients with CP and 14 of 15 WP patients (sensitivity 100%, 95% CI 74–97%; specificity 61%, 95% CI 44–67%). The ROC-AUC using the predicted probabilities from logistic regression with the parameters LUS score, swabs, and fever was 0.96 (95% CI 0.93-0.99) and, when using a cutoff of 10%, accurately identified 30 of 31 patients with CP and 12 of 15 patients with WP (sensitivity 97%, 95% CI 74-97%; specificity 78%, 95% CI 44-67%).

Using a LUS score \geq 3, we identified 40 out of 53 (76%) patients who needed hospital admission and 50 out of 58 (87%) who did not (Fig. 3; Table 2). From the total of 48 LUS-positive patients, 27 (57%) had a positive nasopharyngeal swab, of whom 22 (81%) had been hospitalized. LUS was positive in 18 (86%) out of the 21 (43%) patients with a negative swab; within 6 cases, a first swab was negative but LUS suggested CP; conversely, LUS was positive and swab negative in cases with a diagnosis other than CP requiring hospital admission (Fig. 3). Eight patients with positive LUS were discharged because hospital admission

Table 2. Descriptive statistics of the aggregated data of the entire population (111 patients), divided into patient outcome (discharged/hospitalized) based on LUS score and RT-PCR assay of nasopharyngeal swab

LUS score	<3			≥3			
RT-PCR assay	negative	positive	total	negative	positive	total	
Discharged Hospitalized	38 (60.4%) 11 (17.4%)	12 (19%) 2 (3.2%)	50 (79.4%) 13 (20.6%)	3 (6.3%) 18 (37.5%)	5 (10.4%) 22 (45.8%)	8 (16.7%) 40 (83.3%)	
Total	49 (77.8%)	14 (22.2%)	63 (100%)	21 (43.8%)	27 (56.2%)	48 (100%)	

was clinically not required, although 5 of them had a positive swab. Patients with a LUS score \geq 3 had lower SpO₂ values (median 95 vs. 97%, *p* < 0.001), lower pO₂ (median 69 vs. 88 mm Hg, *p* < 0.001), and lower pO₂/FiO₂ (median 323 vs. 413, *p* < 0.001).

Discussion

LUS seems to be a promising technique for early identification of CP in patients with suspected COVID-19 accessing the ED in an active epidemic time. The LUS score shows a sensitivity of 90% for CP, allowing to quickly refer patients with COVID-19 to the ED isolation area or to discharge them for outpatient treatment.

LUS in CP showed a distribution of pathological features mainly involving both lungs, with multiple lung lobes and segments affected, similarly to the pattern already described [15]. These patterns are consistent with CT features [15, 27, 28]. The retrospective analysis allowed us to identify a cutoff value of the LUS score, and ROC curve analysis of LUS score ≥ 3 identified patients with CP with a sensitivity and specificity of 90 and 75%, respectively. The low cutoff value of the score means that minimum findings (i.e., two focal areas of "white lung" with a score of 2 + 2) can be diagnostic for CP. Such a low value of the score suggests that LUS may really improve early diagnosis of pneumonia and ED management strategies. Specifically, during an acute epidemic, in presence of organizational difficulties in performing CT scans, LUS could allow early diagnosis of CP, limiting the risk of spreading the virus when adequate protective paths are still not available. Nevertheless, further prospective studies are needed to validate this LUS score cutoff value.

LUS features are not specific for CP, as LUS acts like a lung densitometry. LUS interstitial syndrome gives infor-

mation about a surface dysventilation of the lung, and a consolidative pattern indicates the areas of major air loss [29, 30]. These patterns can also be found in many different lung diseases like other viral or bacterial pneumonias, carcinomatous lymphangitis, interstitial lung diseases, and lung contusions, but when LUS features and topographic distribution are combined with clinical and laboratory data, the diagnosis may be reliable. The combination of LUS, fever, and nasopharyngeal swab showed 97% sensitivity and 78% specificity in identifying patients with CP, suggesting a possible simplified diagnostic strategy with limited costs and time. Conversely, the combination of LUS and nasopharyngeal swab showed 100% sensitivity and 61% specificity in identifying patients with CP. Such combination allowed to correctly manage patients triaged for hospital admission.

Patients with CP in the ED were older males with fever and cough and a higher prevalence of lymphopenia, increased lactate dehydrogenase and C-reactive protein, impaired ABG values, and abnormal imaging features compared to WP patients or a diagnosis other than CO-VID-19. These findings are consistent with those of other recent studies [31, 32] and, compared to previous studies reporting CT findings and LUS findings of CP [15, 27], we describe imaging findings in combination with symptoms and laboratory data. Specifically, a linear regression between LUS score and SpO₂, pO₂, and pO₂/FiO₂ suggests a significant correlation between increasing LUS score and worsening O₂ exchanges. Even if LUS assessment is limited to lung surface and does not allow the evaluation of deep parenchyma, such findings further support the already reported consistency in CP between LUS and CT scan [15]. Although our aim was not to stratify CP severity, our data suggest the evaluation of such application of LUS in further studies and its usefulness also to establish the correct level of respiratory support [33].

Outpatient follow-up of COVID-19 patients without imaging features of pneumonia and with absence of other "red flags" is being implemented by general practitioners and infection control and prevention units [34, 35]. Primary care providers must be informed, and patients must be both instructed and motivated to follow the quarantine measures and react in case of clinical worsening. LUS was very useful to exclude pulmonary involvement early: this allowed early discharge of patients without pneumonia and still waiting for the RT-PCR result. With such a management strategy, the workload of the ED was reduced and dedicated to patients waiting to be addressed to COVID or not-COVID wards based on their RT-PCR result, and those needing more attention and assistance.

Our study has some limitations. First, our data should be contextualized in an active epidemic time (e.g., March to April 2020 in Italy, when the high number of subjects suspected to be infected increased the pretest probability), and further studies are needed to investigate its usefulness for the diagnosis of CP as the epidemic curve is decreasing. Second, as it was not possible to guarantee complete covering of medical wards with LUS-trained personnel, our data are based on cases consecutively recruited only in LUS-powered sessions. Multicenter studies are needed for a more precise definition of sensitivity and specificity of LUS in CP. Finally, as the patients were directed to different wards and hospitals, only the ED documentation was included in the analysis, so a correlation between LUS pattern and outcome was not possible. Nevertheless, we found a correlation between increasing LUS score and worsening hypoxemia.

LUS is a promising technique for early identification of CP in patients who access the ED with suspicion of COVID-19. Standardization of the LUS technique and further prospective multicenter studies, even in other pandemic phases, can help improve the reliability of LUS and the proposed cutoff score for its use for early diagnosis of CP during a pandemic.

Acknowledgments

We thank the personnel of the South Tyrolean Health Care System, Italy, for their relentless efforts and invaluable work during the COVID-19 pandemic.

Statement of Ethics

This study was conducted in accordance with the amended Declaration of Helsinki. It was approved by the local institutional review board (BZ 34-2020). Informed consent was not required.

Conflict of Interest Statement

No conflict of interest exists for any of the authors.

Funding Sources

None.

Author Contributions

A. Zanforlin takes responsibility for the content of the manuscript and data analysis. A. Zanforlin, G. Strapazzon, M. Falk, N. Vezzali: conceptualization. A. Zanforlin, G. Strapazzon, F. Ferro, N. Vezzali: project administration and supervision. A. Zanforlin, V. Gallina, L. Pagani, A. Viteritti, L. Valzolgher, M. La Guardia, F. Ferro, N. Vezzali: methodology. A. Zanforlin, V. Gallina, L. Pagani, A. Viteritti, L. Valzolgher, M. La Guardia, F. Ferro, N. Vezzali: investigation. A. Zanforlin, G. Strapazzon, M. Falk: data analysis and interpretation. A. Zanforlin, G. Strapazzon, M. Falk, L. Pagani: original draft. A. Zanforlin, G. Strapazzon, M. Falk, V. Gallina, L. Pagani, A. Viteritti, L. Valzolgher, M. La Guardia, F. Ferro, N. Vezzali: review and editing.

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