# Expression of *STAT5*, *COX-2* and *PIAS3* in Correlation with NSCLC Histhopathological Features



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### Abstract

Signal transducers and activators of transcription (STATs), their inhibitors and cyclooxygenase-2 (COX-2) participate in transformations of many various types of cancers. The aim of the present study was to evaluate the relationship between STAT5A/B, COX-2, and PIAS3 mRNA expression and tumor staging, metastasis status, and histopathological subtype in 71 patients with confirmed non-small cell lung cancer (NSCLC) diagnosis. Total RNA was isolated from NSCLC tissue samples and the expression of the studied genes was assessed using TaqMan probes in real-time PCR assay. The expression levels of STAT5A, STAT5B, and COX-2 genes were increased in 69%, 79%, and 71% NSCLC samples respectively, while PIAS3 expression was decreased in the majority (69%) of the studied tissues. Statistically significant differences were observed between STATS isoforms (P = 0.0008), with higher expression of STAT5B. We found statistically significant positive correlation between STAT5B and COX-2 (rho = 0.045), and significant negative correlation between STAT5B and PIAS3 (rho = -0.049). The negative correlation between STAT5B and PIAS3 (rho = -0.43) was also observed in T2a+T2b tumor group. Additionally, STAT5B and COX-2 expression levels were significantly different between T1a+T1b and T2a+T2b tumors (P = 0.002 and P = 0.041, respectively), with higher expression of both genes in T2 tumor stage. PIAS3 expression was significantly lower in NSCC subtype as compared with SCC subtype (P = 0.017). Also, STAT5A and STAT5B immunoexpression was assessed, and the results indicated significantly higher protein levels in NSCLC patients as compared with controls (P = 0.048 and P = 0.034, respectively). High STAT5B immunoexpression was positively correlated with STAT5B gene expression in tumors (rho = 0.755). STAT5B protein level was also significantly higher in T2a+T2b tumors, reflecting high STAT5B gene expression in this group. There was no statistically significant association between mRNA and protein expression levels of the studied genes and patients' characteristics: age, gender, smoking. The obtained results highlight the importance of the genes STAT5B and COX-2 in lung cancer progression.

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### Introduction

It is documented that despite of more and more modern treatment options, lung cancer is one of the leading reason of cancer related deaths in the world. Non-small cell lung cancer (NSCLC) is recognized as the most common – accounting for 75–85% – among all lung malignant tumors [1]. With advances in molecular biology, the detection and analysis of changes in expression levels of many important genes involved in signaling pathways may supply predictive molecular markers harboring diagnostic and/or prognostic value for NSCLC.

In many human cancers, including NSCLC, one of the key pathways that promote cellular survival or cell growth is Janus kinase/signal transducers and activators of transcription (JAK/ STAT) pathway. It is one of the pleiotropic cascades of molecules involved in signal transduction for proliferation, development and apoptosis [2,3]. STAT (signal transducers and activators of transcription) protein family of transcription factors consists of seven members: 1–4, 5A, 5B, and 6 [2]. Among them, and beside STAT3, the oncogenic activity of STAT5 was documented both *in vitro* and *in vivo* [4,5]. After phosphorylation, the two STAT5 proteins, STAT5A and STAT5B, as homo- or heterodimer, may translocate to the nucleus and bind to specific STAT5 response elements of target gene promoters, thus influencing their expression [6]. It is recognized that *STAT5A* and *STAT5B* isoforms are encoded by two tandemly linked genes on chromosome 17q11.2 [7]. They act as independent transcription factors [8] and modulate important cellular processes in different manner in normal and malignant cells [9]. Generally, active STAT5 promotes cell cycle progression, proliferation, invasion, angiogenesis, and inhibits apoptosis.

Table 1. Histopathological verifications of NSCLC tissue samples.

Histopathological type of NSCLC	squamous cell carcinoma (SCC)	41 (57.75%)
	non-squamous cell carcinoma (NSCC)	30 (42.25%)
AJCC*	AJCC IA	14 (19.72%)
	AJCC IB	11 (15.49%)
	AJCC IIA	13 (18.31%)
	AJCC IIB	10 (14.08%)
	AJCC IIIA/IIIB	23 (32.39%)
pTNM*	T1a+T1b	19 (26.76%)
	T2a+T2b	33 (46.48%)
	T3+T4	19 (26.76%)

\*AJCC – American Joint Committee on Cancer Staging according to the IASCLC Staging Project 7th ed. (2010) Cancer.

\*pTNM — post-operative Tumor Node Metastasis classification according to the WHO Histological Typing of Lung Tumor.

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The overexpression of STAT5 has been recognized in several types of human tumors, mainly in breast and prostate cancers [10-13]. However, as so far, the data on the role of STAT5 in NSCLC cells, as well as on its activation status in NSCLC is still very limited. Recently, it has been documented that besides several cytokines, hormones and growth factors, EGF influences the STAT5 expression in human lung adenocarcinoma cell line thus leading to the increased cyclooxygenase-2 (COX-2) expression [14]. COX-2, belonging to the COX enzyme family (COX-1, COX-2 and COX-3) [15], is a key enzyme in the biosynthesis of prostaglandins (PG). COX-2 is involved in the initiation and progress of tumors in situ and its overexpression is frequently recognized in many tumor types, including NSCLC [16-18]. However, carcinogenic effect of COX-2 upregulation in relation to the expression level of STAT5 in NSCLC patients has not been investigated vet.

On the other hand, as a modulator of activity of STAT5, a protein inhibitor of activated STAT3 (PIAS3), which regulates different DNA binding transcription factors implicated in the immune response (e.g., NF $\kappa$ B, SMAD, and MITF), was recognized in breast cancer [19]. To date, only a small number of reports focused on PIAS3 participation in cancers involving lung tumors, has been published [20–23]. Additionally, although it is known that PIAS3 plays a vital role in oncogenic process influencing STAT3 protein, interaction between PIAS3 and STAT5 has not been fully recognized yet. The aim of our study was to determine the relationship between *STAT5*, *PIAS3* and also *COX-2* and their reciprocal relationship on transcriptional level in NSCLC patients. To achieve this goal, we assessed the mRNA expression of these genes and their association with histopathological features of NSCLC tumors as well as clinical characteristic of patients. The prespecified hypothesis tested was that *STAT5*, *COX-2* and *PIAS3* expression levels were modified in non-small cell lung cancer, playing a role in lung carcinogenesis. Additionally, we analyzed the levels of STAT5A and STAT5B proteins in the studied samples.

### **Materials and Methods**

The study has been approved by the Ethical Committee of the Medical University of Lodz, Poland no. RNN/64/11/KE. Written informed consent was obtained from each patient.

### 1. Characterization of the NSCLC tissue samples and patients clinical characteristics

Biological material (lung tissue) was obtained from 84 patients admitted to the Department of Thoracic Surgery, General and Oncologic Surgery, Medical University of Lodz, Poland, between July 2010–June 2012. Based on the results of preoperative cytological/histological assessment, the patients were qualified for surgery and were treated by either lobectomy or pneumonectomy.

**Table 2.** Characteristics of NSCLC patients in terms of tobacco addiction and consumption (the amount of cigarettes smoked per day and PYs).

Tobacco addiction and consumption		n = 71
Smokers		66 (92.96%)
Non-smokers		5 (7.04%)
The smoking period	<40 years	37 (52.11%)
	$\geq$ 40 years	29 (40.85%)
The amount of cigarettes smoked	10–15 cigarettes per day	6 (9.09%)
	20 cigarettes per day (1 pack)	43 (65.15%)
	30–40 cigarettes per day (1.5–2 packs)	17 (25.76%)
Pack Years (PYs)	up to 40 PYs	30 (45.45%)
	≥40 PYs	36 (54.54%)

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Gene	Histopathological NSCLC subtype	Mean RQ value (range)	Number (percentag	ge) of samples with
			RQ value>1	RQ value<1
STAT5A	SCC (n = 41)	3.25 (0.25-4.49)	31 (75.60%)	10 (24.39%)
	NSCC (n = 30)	2.155 (0.05–4.54)	18 (60.00%)	12 (40.00%)
	Total (n = 71)	2.75 (0.05-4.54)	49 (69.01%)	22 (30.99%)
STAT5B	SCC (n = 41)	8.92 (0.67–59.29)	35 (85.37%)	6 (14.63%)
	NSCC (n = 30)	9.26 (0.77–864.17)	21 (70.00%)	9 (30.00%)
	Total (n = 71)	9.13 (0.77-864.17)	56 (78.87%)	15 (21.13%)
СОХ-2	SCC (n = 41)	9.34 (0.10-58.21)	29 (70.73%)	12 (29.27%)
	NSCC (n = 30)	4.89 (0.03–97.53)	21 (70.00%)	9 (30.00%)
	Total (n = 71)	3.17 (0.24–20.89)	50 (70.42%)	21 (29.58%)
PIAS3	SCC (n = 41)	0.21 (0.16–1.74)	14 (34.15%)	27 (65.85%)
	NSCC (n = 30)	0.94 (0.02–1.12)	8 (26.67%)	22 (73.33%)
	Total (n = 71)	0.54 (0.02–14.91)	22 (30.99%)	49 (69.02%)

Table 3. The expression levels (RQ values) of the studied genes in individual histopathological NSCLC subtypes.

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Immediately after resection, lung tissue samples (100–150 mg) and the adjacent non-cancerous macroscopically unchanged tissues (100 mg; 10 cm distant from the primary lesion) obtained from the same patients were placed in a stabilization buffer RNAlater<sup>®</sup>. Each tissue sample was divided into smaller parts (30–50 mg) for individual analysis. All samples were frozen at  $-80^{\circ}$ C.

The resected lesions were post-operatively histhopathologically evaluated and classified according to the AJCC staging as well as TNM classification (pTNM). Based on the results, from the group of 84 patients, 12 of them were excluded due to a concomitant malignancy and the suspected metastasis to the lung, in the case of one patient, there was no full clinical information. The final study group included seventy one (n = 71) patients with confirmed NSCLC diagnoses. Histopathological assessments of tumor specimens were obtained from pathomorphological reports, and were as follows: squamous cell carcinoma (SCC, n = 41), nonsquamous cell carcinoma (NSCC, n = 30). Histopathological verifications of non-small cell lung carcinoma tissues are shown in Table 1. The studied group consisted of 25 women, mean age 63±8.717 and 46 men, mean age 65±8.234. All cases were primary tumors without chemo- or radiotherapy treatment. The smoking history was available for all patients: 5 patients were nonsmokers, and 66 were smokers or former smokers. They were divided into groups according to their smoking habits: time of tobacco addiction and amount of cigarettes smoked - the latter was presented as Pack Years (PYs) and was calculated according to the NCI Dictionary of Cancer Terms (1 Pack Year is equal to 20 cigarettes smoked per day for 1 year), see Table 2.

### 2. RNA extraction, real-time PCR

Total RNA was extracted from lung tissues (cancer tissue obtained from the center of lung lesion and macroscopically unchanged lung tissue obtained from the most distant site from the resected lesion) using Universal RNA Purification Kit (Eurix, Poland), and according to the manufacturer's recommendations. The quality and quantity assessments of RNA samples were determined by minielectrophoreses in polyacrylamide gel (Agilent 2100 Bioanalyzer, Agilent, USA), using RNA 6000 Pico/Nano LabChip kit (Agilent Technologies, USA). Complementary DNA (cDNA) was transcribed from 100 ng of total RNA, using a High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems,

USA) in a total volume of 20 µl per reaction. Reverse transcription (RT) master mix contained: 10× RT buffer, 25× dNTP Mix (100 mM), 10× RT Random Primers, MultiScribe<sup>TM</sup> Reverse Transcriptase, RNase Inhibitor and nuclease-free water. RT reaction was performed in a Personal Thermocycler (Eppendorf, Germany) in the following conditions: 10 minutes at 25°C, followed by 120 minutes at 37°C, then the samples were heated to 85°C for 5 seconds, and hold at 4°C. The relative expression was assessed using TaqMan probes: Hs00234181\_m1, Hs00273500\_m1, Hs00153133\_m1, Hs00180666\_m1 for the studied genes STAT5A, STAT5B, COX-2 and PIAS3, respectively, as well as for  $\beta$ -actin (ACTB, Hs99999903\_m1), as the reference gene. The PCR mixture contained: cDNA (1 to 100 ng), 20× TaqMan<sup>®</sup> Gene Expression Assay, 2× TaqMan<sup>®</sup> Gene Expression Master Mix, RNase-free water in a total volume of 20 µl. The qPCR reactions were performed in Applied Biosystems 7900HT Fast Real-Time PCR System for 39 cycles, with annealing temperature of 60°C, repeated 3 times for each sample. The relative expression of the studied samples were assessed using the Comparative delta-delta C<sub>T</sub> method (TaqMan Relative Quantification Assay software) and presented as RQ value,



Figure 1. Box-and-whisker plots, representing *STAT5B* expression levels (mean RQ values) in the studied tumor groups classified according to pTNM staging in relation to their size. doi:10.1371/journal.pone.0104265.g001

**Table 4.** The results of Neuman-Keuls' multiple comparison test regarding differences in *STAT5B* RQ values in the studied tumor groups classified according to pTNM staging in relation to the tumor size.

	T1a+T1b	T2a+T2b	T3+T4
T1a+T1b		0.001782	100000
T2a+T2b	0.001782		0.723456
T3+T4	1.000000	0.723456	

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adjusted to  $\beta$ -actin expression level. Macroscopically unchanged lung tissue served as calibrator sample.

### 3. Tissue homogenization, ELISA assays

Lung tissue samples (30–50 mg) were rinsed in ice-cold PBS buffer (0.01 mol/l, pH 7.0–7.2) and homogenized in 5 ml of PBS. The resulting suspension was subjected to two cycles of freezing and thawing. Then, the homogenates were centrifuged for 5 minutes at 5000×g, the supernatant was removed and the suspension was aliquoted and stored at  $-80^{\circ}$ C until further analysis.

STAT5A and STAT5B immunoexpression levels in lung tissue homogenates were assessed using ELISA Kit for Signal Transducer And Activator Of Transcription 5A (SEB738Hu, Uscn Life Science Inc., China) and ELISA Kit for Signal Transducer And Activator Of Transcription 5B (SEB727Hu, Uscn Life Science Inc., China), according to the manufacturer's procedure. The intensity of the final colorimetric reaction, in proportion to the amount of protein bound, was measured in a plate reader (ELx800, BioTek) at 450 nm. The obtained results were compared to the standard solution of known concentrations (0.312–20 ng/ ml).

#### 4. Statistical analysis

ANOVA Kruskal-Wallis' test and U Mann-Whitney test, were used to compare the levels of relative expression values (RQs) of *STAT5A*, *STAT5B*, *COX-2* and *PIAS3* among NSCLC subtypes (SCC, NSCC).



Figure 2. Box-and-whisker plots, representing the expression levels (mean RQ values) of *COX-2* gene in the studied tumor groups classified according to pTNM staging in relation to their size.

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The Spearman's rank correlation coefficient, U Mann-Whitney's test and ANOVA Kruskal-Wallis' test were performed in order to evaluate the relationship between the expression levels (RQ values) of the studied genes (STAT5A, STAT5B, COX-2 and PIAS3), between RQs and the examined parameters (patient characteristics: age, gender and tumor staging according to pTNM and AJCC classification), as well as to find correlations regarding STAT5A and STAT5B protein levels in the studied samples. The accepted level of statistical significance was estimated at P<0.05. The results of relative expression levels of the studied genes are presented as RQ (relative quantification) means  $\pm$  SEM and RQ means  $\pm$  SD values. Statistica for Windows 10.0 program was applied for calculations.

### Results

### 1. Relative expression levels of *STAT5A*, *STAT5B*, *COX-2* and *PIAS3*

Relative expression levels of the studied genes (*STAT5A*, *STAT5B*, *COX-2*, *PIAS3*), expressed as RQ values, were determined using delta-delta  $C_T$  method, adjusted to the expression of  $\beta$ -actin (endogenous control) and relative to the expression level of calibrator (macroscopically unchanged lung tissue) for which RQ = 1.

The obtained RQ values for the individual genes were correlated with histopathological NSCLC subtypes (SCC, NSCC), tumor staging (pTNM, AJCC), patients' age, gender and smoking history.

STAT5A expression was increased (RQ value>1) in the majority (69%) of the studied samples. Regarding the individual histotypes, it was increased in a greater proportion in SCC group, but there were no statistically significant differences. STAT5B expression was increased in all studied histopathological NSCLC subtypes, in a range of 70%-85%, depending on a histotype. Expression levels of STAT5 isoforms (STAT5A and STAT5B) were statistically significant different (P = 0.0008; Spearman's rank correlation coefficient), with higher expression level displayed by STAT5B. The expression level of COX-2 was nearly uniformly increased in all NSCLC subtypes, at the level of 70%. In contrast, *PIAS3* expression was decreased (RQ value<1) and it was observed in all studied histopathological NSCLC subtypes, in a range of 66%-73%, depending on a histotype. Statistically significant differences were found between SCC and NSCC subtypes (P = 0.017; U Mann-Whitney's test), with significantly lower gene expression observed in NSCC samples. The results regarding genes' expression levels are summarized in Table 3.

## 2. Statistical analysis of relationship between studied gene expression levels and clinical features of patients and tumor characteristics

There was no statistically significant correlation between RQ values of *STAT5A* and the clinical features of NSCLC patients,

**Table 5.** The results of Neuman-Keuls' multiple comparison test regarding differences in COX-2 RQ values in the studied tumor groups classified according to pTNM staging in relation to the tumor size.

	T1a+t1b	T2a+T2b	T3+T4
T1a+T1b		0.02459	1000000
T2a+T2b	0.02459		0.27776
T3+T4	1.000000	0.27776	

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i.e., patients' age (P=0.48), gender (P=0.54), and history of smoking assessed as PY (P=0.87), as well as histopathological features of tumor, i.e., pTNM classification (P=0.34), AJCC classification (P=0.83), NSCLC subtypes (SCC *vs* NSCC, P=0.62) (ANOVA Kruskal-Wallis test, U Mann-Whitney's test followed by Spearman's rank correlation coefficient).

Similarly, statistical analysis concerning STAT5B did not reveal significant correlations between its expression levels and patients' age (P=0.59), gender (P=0.45), history of smoking (P=0.66), NCSLC subtypes (P=0.85) and tumor AJCC staging (P=0.23) (ANOVA Kruskal-Wallis test, U Mann-Whitney's test followed by Spearman's rank correlation coefficient). However, statistically significant differences were found in relation to tumor size classified according to pTNM staging (P=0.034; ANOVA Kruskal–Wallis test), as presented in **Figure 1**. Neuman-Keuls' multiple comparison test revealed statistically significant differences between T1a+T1b vs T2a+T2b group (see **Table 4**).

There was no statistically significant correlation between RQ values of *COX-2* and patients' age (P = 0.09), gender (P = 0.44) and history of smoking (P = 0.34) (ANOVA Kruskal-Wallis test, U Mann-Whitney's test followed by Spearman's rank correlation coefficient). Similarly, no statistically significant correlations were found between RQ values of *COX-2* and histopthological NSCLC subtypes: SCC *vs* NSCC (P = 0.51) and tumor AJCC classification (P = 0.28) (U Mann-Whitney's test). However, the association between tumor size according to pTNM staging and *COX-2* expression level was statistically significant (P = 0.041; ANOVA Kruskal–Wallis test), as presented in **Figure 2**. Neuman-Keuls'



Figure 3. Box-and-whisker plots, representing *PIAS3* expression levels (mean RQ values) in the studied tumor groups classified according to the histopathological NSCLC subtypes. doi:10.1371/journal.pone.0104265.q003

multiple comparison test revealed statistically significant differences in RQ values between T1a+T1b and T2a+T2b group (see **Table 5**).

Regarding *PIAS3* expression, statistical analysis did not reveal any significant correlations between the RQ values and patients' age (P=0.48), gender (P=0.37), and history of smoking (P=0.58) (ANOVA Kruskal-Wallis test, U Mann-Whitney's test followed by Spearman's rank correlation coefficient), as well as tumor staging according to pTNM (P=0.22) and AJCC classification (P=0.12) (ANOVA Kruskal-Wallis test). Statistically significant differences were found between histopathological NSCLC subtypes (P=0.017; U Mann-Whitney's test), as presented in **Figure 3**.

### 3. Statistical analysis of the reciprocal relationship between the expression of the studied genes

Finally, we assessed the reciprocal relationship between the expression levels of the studied genes. Spearman's rank correlation coefficient revealed statistically significant negative correlation between *STAT5B* and *PIAS3* genes (rho = -0.049, P = 0.04; Spearman's rank correlation) and positive correlation between *STAT5B* and *COX-2* genes (rho = 0.045, P = 0.027; Spearman's rank correlation). Additionally, statistically significant negative correlation between *STAT5B* and *PIAS3* genes was observed in T2a+T2b tumor group (rho = -0.43, P = 0.041; Spearman's rank correlation).

#### STAT5A and STAT5B immunoexpression levels

A quantitative measurements of STAT5A and STAT5B in NSCLC and control samples were performed using ELISA method. The mean values (ng/ml) obtained in the studied samples are presented in **Table 6**. Statistical analysis revealed that both STAT5A and STAT5B protein immunoexpression levels were significantly higher in lung cancer samples as compared with normal (macroscopically unchanged, control) lung tissues (P=0.048 in case of STAT5A and P=0.034 in case of STAT5B; U Mann-Whitney's test). However, the differences between NSCLC histopathological subtypes (SCC *vs* NSCC) weren't statistically significant, neither for STAT5B (P=0.61; U Mann-Whitney's test) nor for STAT5A (P=0.81; U Mann-Whitney's test).

STAT5B immunoexpression levels were significantly different between samples grouped according to pTNM classification in relation to tumor size (P = 0.045; ANOVA Kruskal-Wallis test), as presented in **Figure 4**. Neuman-Keuls' multiple comparison test revealed statistically significant differences between T1a+T1b and T2a+T2b group (see **Table 7**).

We didn't find any significant correlations between STAT5B immunoexpression level and patients' age (P=0.61), gender (P=0.40), the history of smoking assessed as PY (P=0.63), and AJCC classification (P=0.17) (ANOVA Kruskal-Wallis test, U Mann-Whitney's test followed by Spearman's rank correlation coefficient).

Table 6. STAT5A a	ind STAT5B protein	levels (ng/ml) in th	e studied samples (	controls, NSCLC ar	d NSCLC histopath	ological subtypes).		
Protein	NSCLC (n = 71)		Control (n=20)		NSCLC subtypes			
	Mean [ng/ml]	Range [ng/ml]	Mean [ng/ml]	Range [ng/ml]	NSCC (n = 30)		SCC (n=41)	
					Mean [ng/ml]	Range [ng/ml]	Mean [ng/ml]	Range [ng/ml]
STAT5A	16.73	0.39–19.25	4.88	0.33-10.22	16.73	0.39–19.25	15.62	0.39–18.71
	P = 0.048 U Mann-Whi	tney's test			P>0.05 U Mann-Whitr	iey's test		
STAT5B	17.87	0.35-19.88	2.90	0.36-9.43	15.84	0.35-18.62	18.12	0.53-19.88
	P=0.034 U Mann-Whi	tney's test			P>0.05 U Mann-Whitr	ney's test		
doi:10.1371/journal.pone	.0104265.t006							

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Figure 4. Box-and-whisker plots, representing STAT5B protein levels (ng/ml) in the studied tumor groups classified according to pTNM staging in relation to the tumor size. doi:10.1371/journal.pone.0104265.g004

No significant correlations were found between pSTAT5A immunoexpression level and patients' age (P = 0.83), gender (P = 0.17), the history of smoking assessed as PY (P = 0.92), tumor size according to pTNM classification (P = 0.46), and AJCC classification (P = 0.98) (ANOVA Kruskal-Wallis test, U Mann-Whitney's test followed by Spearman's rank correlation coefficient).

## 5. Statistical analysis of the reciprocal relationship between *STAT5A/B* mRNA expression and STAT5A/B immunoexpression

The positive correlation was found between the expression levels of STAT5B gene and the immunoexpression levels of STAT5B (rho=0.755, P=0.04; Spearman's rank correlation coefficient) in NSCLC samples, see **Figure 5**. No significant correlation was found between STAT5A gene expression and STAT5A protein level (P=0.43, Spearman's rank correlation coefficient).

### Discussion

It is commonly known that deregulations of STAT signaling pathway can result in oncogenesis and the constitutive activation of *STAT5* is found in many human malignancies [10,12,24], including lung cancer [25]. Our study results confirmed the implication of *STAT5* isoforms in NSCLC development. We observed the increased expression levels of *STAT5* genes (A and B isoforms) in the majority of studied NSCLC tissues, both at mRNA and protein level. Our result are in accordance with observations of other investigators who have also reported STAT5A and STAT5B overexpression on mRNA and/or protein level in cancer tissue – prostate, breast, colorectal, esophageal and lung – when compared to normal tissue [10,24–31].

Interestingly, it is known that STAT5A and STAT5B isoforms have distinct biological functions [32–33] and the expression patterns of these proteins differ among many types of cancers cells. Both STAT5A and STAT5B are key tumor growth stimulators, but STAT5B overexpression seems to be more significant in head and neck and prostate cancers, while STAT5A – in breast cancer [31,34,35]. However, the exact role of STAT5A as opposed to STAT5B in malignant cell transformation is not well understood. **Table 7.** The results of Neuman-Keuls' multiple comparison test regarding differences in STAT5B immunoexpression levels in the studied tumor groups classified according to pTNM staging in relation to the tumor size.

	T1a+T1b	T2a+T2b	T3+T4
T1a+T1b		0.0046134	0.92834
T2a+T2b	0.0046134		0.107350
T3+T4	0.92834	0.107350	

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Recently, Tang et al. [36] have demonstrated differential regulation of mammary carcinoma cell behavior upon forced expression of STAT5A and STAT5B: STAT5B exhibited lower potency than STAT5A in enhancing survival and anchorageindependent growth of mammary carcinoma cells and exerted no effects on cell motility. In our study we assessed the immunoexpression of STAT5A and STAT5B proteins in lung cancer tissue and found it significantly higher than in normal lung tissue. The study performed by Sánchez-Ceja et al. [25] indicated the highest nuclear STAT5 expression in LCC (large cell carcinoma), lower in SCC and the lowest in AC (adenocarcinoma). In our study we included LCC and AC in one NSCC group due to the small number of LCC samples. However, we didn't find any significant differences between the groups. STAT5A and STAT5B protein levels were similar in the studied NSCLC subtypes, however, for procedural reasons, we could not analyze the location of proteins (nuclear or cytoplasmic).

The analysis of both gene isoforms on mRNA level in NSCLC samples revealed significantly higher expression of *STAT5B* isoform as compared with *STAT5A*. It might suppose different functions for both isoforms also in lung cancer cells, i.e., in result, different downstream target genes (including oncogenes) specifically induced by either STAT5A or STAT5B. The mechanism of lower *STAT5A* gene expression in NSCLC is not known, however in lymphomas *STAT5A* was found to be epigenetically silenced [37]. Consistent with the obtained results indicating high *STAT5B* immunoexpression, as we observed positive correlation between these two parameters.



Figure 5. Positive correlation between STAT5B protein levels (ng/ml) and gene expression levels (RQ values) in NSCLC samples.

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We documented also statistically significant differences in STAT5B expression levels depending on tumors size, i.e., significantly higher STAT5B expression in T2 group, which was additionally reflected by significantly higher STAT5B protein levels in this group. Thus, it could further suggest that STAT5B has more important implications than STAT5A in the pathogenesis of NSCLC. As so far there is no available published data concerning association of STAT5B expression with tumor progression and prognosis in NSCLC. The analysis of STAT5 immunohistochemical staining in NSCLC tissue, performed by Sánchez-Ceja et al. [25], suggested the association of STAT5 protein with advanced lung cancer stages. Regarding other tumor types the data are conflicting. The significant association of STAT5B protein expression with TNM stage was found in colorectal cancer [38], but not in gastric cancer [39]. Interesting results were obtained by Tang et al. [36], who proved a dual role of STAT5A in human mammary carcinoma cells: promotion of tumor formation by enhancing survival and anchorage-independent growth but simultaneous inhibition of tumor metastasis by suppression of cellular invasiveness.

In our study, we tried to establish – to our knowledge as a first research group – the relationship between *STAT5A* and *STAT5B* mRNA expression and patient's age, gender, and NSCLC histopathological subtype. Such an association could indicate an importance of *STAT5* mRNA expression level as a diagnostic tool in NSCLC. However, we didn't find any significant associations.

PIAS3 (protein inhibitor of activated STAT3), was originally identified as a specific inhibitor of the STAT3 signaling pathway [40]. Now it is well recognized that the PIAS family (PIAS1, PIAS3, PIASx and PIASy) regulates a variety of cytokine and transcription factors with downstream alterations in apoptosis, angiogenesis and a number of signaling pathways. Regarding PIAS3 and its role in lung tumorigenesis, it has been demonstrated that PIAS3 decreases lung cancer growth and increases the antitumor effects of EGFR inhibitors [20], activates the intrinsic apoptosis pathway via altered expression of Bcl-2 and Akt family members [21,23], and interacts with several other transcription factors, including: ETS, EGR, NR1I2 and GATA1 [21], all of them playing important roles in cancer development. The STAT3 independent effect of PIAS3 includes also the repression of STAT5 transcriptional activity, as demonstrated by Rycyzyn and Clevenger [19]. In our study the expression level of PIAS3 was negatively correlated with the expression level of STAT5B in the whole group of studied tumors, and especially, the negative correlation was observed in T2 tumor group.

In several cancers a decrease or loss of PIAS3 expression was demonstrated, indicating it as a protein with putative tumor suppressor function [22], but a paucity of studies were focused on lung cancer [20,21,23]. The results obtained in NSCLC cell lines indicated that overexpression of PIAS3 had growth inhibitory effect [20,23]. Our analysis revealed the decreased *PIAS3* expression in the majority of the studied NSCLC samples.

Moreover, it was significantly lower in NSCC as compared to SCC. Our results are similar to those obtained by Kluge et al. [20] who found *PIAS3* mRNA expression higher in SCC than in AC.

Cyclooxygenase (COX), is the key enzyme in the biosynthesis of the prostanoids and plays a central role in many important cellular processes, including inflammatory response, tumorigenesis, and tumor progression [41]. COX is comprised of three categories, including COX-1, COX-2 and COX-3 [15]. COX-2 converts arachidonic acid to bioactive lipids including prostaglandin E2 (PGE<sub>2</sub>), and their role in initiating and progressing tumors in situ was established. It was suggested that the overexpression of COX-2 and the resulting increase in PGE<sub>2</sub> levels may represent a tumor strategy to escape immunosurveillance [42]. COX-2 overexpression was found by immunohistochemical method in well and moderately differentiated carcinomas of the lung, colon, and breast [18,43]. In lung cancer, COX-2 overexpression was reported to inhibit apoptosis [16], promote angiogenesis [44] and metastasis [45]. In our study we confirmed increased expression of COX-2 gene in the majority of NSCLC samples, regardless the histopathological subtype. In contrast, it was found that COX-2 protein expression in adenocarcinoma was significantly higher than that in squamous cell carcinoma [43,46,47]. These differences might be caused by further posttranslational modifications of COX-2 protein in lung cancer cells.

Interestingly, we observed statistically significant differences in COX-2 expression with respect to tumor size – significantly higher gene expression levels were shown in T2a+T2b tumors as compared with T1a+T1b group. The meta-analysis combining 14 published studies, revealed a significant association between COX-2 expression and poor survival for stage I NSCLC (i.e., encompassing T1a, T1b and T2a tumors) [48]. However, the studies reporting the relationship between COX-2 expression and survival among lung cancer patients are inconsistent. The study performed by Groen et al. [49] confirmed that high COX-2 expression predicted poor prognosis in NSCLC, but the results of more recent study have suggested that the prognostic role of COX-2 in NSCLC needs to be confirmed by further high-quality prospective studies [50].

In human lung adenocarcinoma cells it was found that COX-2 expression was stimulated *via* the activation of the STAT5

#### References

- Spira A, Ettinger DS (2004) Multidisciplinary management of lung cancer. N Engl J 350:379–392.
- Bowman T, Garcia R, Turkson J, Jove R (2000) STATs in oncogenesis. Oncogene 19:2474–2488.
- Bromberg J, Darnell JE (2000) The role of STATs in transcriptional control and their impact on cellular function. Jr Oncogene 19:2468–2473.
- Barash I (2012) Stat5 in breast cancer: potential oncogenic activity coincides with positive prognosis for the disease. Carcinogenesis 33:2320–2325.
- deGroot RP, Raaijmakers JA, Lammers JW, Koenderman L (2000) STAT5-Dependent CyclinD1 and Bcl-xL expression in Bcr-Abl-transformed cells. Mol Cell Biol Res Commun 3:299–305.
- Ihle JN (2001) The Stat family in cytokine signaling. Curr Opin Cell Biol 13: 211–217.
- Lin JX, Mietz J, Modi WS, John S, Leonard WJ (1996) Cloning of human Stat5B. Reconstitution of interleukin-2-induced Stat5A and Stat5B DNA binding activity in COS-7 cells. J Biol Chem 271(18):10738–10744.
- Hennighausen L, Robinson GW (2008) Interpretation of cytokine signaling through the transcription factors STAT5A and STAT5B. Genes Dev 22:711– 721.
- Furth PA, Nakles RE, Millman S, Diaz-Cruz ES, Cabrera MC (2011) Signal transducer and activator of transcription 5 as a key signaling pathway in normal mammary gland developmental biology and breast cancer. Breast Cancer Res 13:220.
- Gu L, Vogiatzi P, Puhr M, Dagvadorj A, Lutz J, et al. (2010) Stat5 promotes metastatic behavior of human prostate cancer cells in vitro and in vivo. Endocr Relat Cancer 17: 481–493.
- Tan SH, Nevalainen MT (2008) Signal transducer and activator of transcription 5A/B in prostate and breast cancers. Endocr Relat Cancer 15:367–390.

pathway thus showing that STAT5 plays a certain role, affecting the expression of COX-2 [14]. In our study the increased expression of *STAT5* was accompanied by increased expression of *COX-2* in nearly 70% of NSCLC samples. Moreover, expression levels of *STAT5B* and *COX-2* showed a significant positive correlation.

We didn't observe any statistically significant associations between expression values of the studied genes and clinical patients' characteristics: age, gender as well as tobacco addiction and consumption. Unfortunately, we can't refer our results to those obtained by others, because the available studies were carried out on lung cancer cell lines, rather than lung tissue derived from NSCLC patients.

### Conclusions

In our study we found significantly higher expression levels of STAT5B (both on mRNA and protein level) and COX-2 gene in T2a+T2b as compared to T1a+T1b tumors. In the same tumor group, the negative correlation between STAT5B and PIAS3 expression was statistically significant. It could indicate the role of STAT5B isoform and COX-2 in lung cancer progression. In the absence of other reports on STAT5 gene expression in lung cancer, the results require confirmation in a larger group of NSCLC patients. It seems, however, that the role of STAT5B in the development of lung cancer is more similar to that in prostate cancer than in breast cancer. Such knowledge is extremely important in the context of inhibition of STAT5 in the potential treatment strategy of lung cancer. Additionally, the targeted therapy toward STAT activity might be based on the data regarding PIAS3 and/or COX-2 expression in lung cancer cells. Reassuming, our data confirmed the justification of ongoing clinical studies focused on selective inhibitors of COX and/or STAT in lung malignant cells.

### **Author Contributions**

Conceived and designed the experiments: DPL DD EB. Performed the experiments: DD EN MMS. Analyzed the data: DPL EB PG. Contributed reagents/materials/analysis tools: J. Kordiak KHC J. Kisza łkiewicz AA. Wrote the paper: DPL EB.

- Wagner KU, Rui H (2008) Jak2/Stat5 signaling in mammogenesis, breast cancer initiation and progression. J Mammary Gland Biol Neoplasia 13: 93– 103.
- Buettner R, Mora LB, Jove R (2002) Activated STAT signaling in human tumors provides novel molecular targets for therapeutic intervention. Clin Cancer Res 8:945–954.
- Cao S, Yan Y, Zhang X (2011) EGF stimulates cyclooxygenase-2 expression through the STAT5 signaling pathway in human lung adenocarcinoma A549 cells. Int J Oncol 39:383–391.
- Smith WL, DeWitt DL, Garavito RM (2000) Cyclooxygenases: structural, cellular, and molecular biology. Annu Rev Biochem 69: 145–182.
- Pöld M, Zhu LX, Sharma S, Burdick MD, Lin Y, et al. (2004) Cyclooxygenase-2-dependent expression of angiogenic cxc chemokines ena-78/cxc ligand (cxcl) 5 and interleukin-8/cxcl8 in human non-small cell lung cancer. Cancer Res 64:1853–1860.
- Hosomi Y, Yokose T, Hirose Y, Nakajima R, Nagai K, et al. (2000) Increased cyclooxygenase 2 (COX-2) expression occurs frequently in precursor lesions of human adenocarcinoma of the lung. Lung Cancer 30:73–81.
- Soslow RA, Dannenberg AJ, Rush D, Woerner BM, Khan KN, et al. (2000) COX-2 is expressed in human pulmonary, colonic, and mammary tumors. Cancer 89:2637–2645.
- Rycyzyn MA, Clevenger CV (2002) The intranuclear prolactin/cyclophilin B complex as a transcriptional inducer. Proc Natl Acad Sci U S A 99:6790–6795.
- Kluge A, Dabir S, Vlassenbroeck I, Eisenberg R, Dowlati A (2011) Protein inhibitor of activated STAT3 expression in lung cancer. Mol Oncol 5:256–264.
- Dabir S, Kluge A, Aziz MA, Houghton JA, Dowlati A (2011) Identification of STAT3-independent regulatory effects for protein inhibitor of activated STAT3 by binding to novel transcription factors. Cancer Biol Ther 12: 139–151.

- Brantley EC, Nabors LB, Gillespie GY, Choi YH, Palmer CA, et al. (2008) Loss of protein inhibitors of activated STAT-3 expression in glioblastoma multiforme tumors: implications for STAT-3 activation and gene expression. Clin Cancer Res 14:4694–4704.
- Ogata Y, Osaki T, Naka T, Iwahori K, Furukawa M, et al. (2006) Overexpression of PIAS3 suppresses cell growth and restores the drug sensitivity of human lung cancer cells in association with PI3-K/Akt inactivation. Neoplasia 8:817–825.
- Sung YH, Park J, Choi B, Kim J, Cheong C, et al. (2005) Hematopoietic malignancies associated with increased Stat5 and Bcl-x(L) expressions in Ink4a/ Arf-deficient mice. Mech Ageing Dev 126:732–739.
- Sánchez-Ceja SG, Reyes-Maldonado E, Vázquez-Manríquez ME, López-Luna JJ, Belmont A, et al. (2006) Differential expression of STAT5 and Bcl-xL, and high expression of Neu and STAT3 in non-small-cell lung carcinoma. Lung Cancer 54:163–168.
- Creamer BA, Sakamoto K, Schmidt JW, Triplett AA, Moriggl R, et al. (2010) Stat5 promotes survival of mammary epithelial cells through transcriptional activation of a distinct promoter in Akt1. Mol Cell Biol 30:2957–2970.
- Liu JR, Wang Y, Zuo LF, Li FL, Wang Y, et al. (2007) Expression and clinical significance of COX-2, p-Stat3, and p-*Stat5* in esophageal carcinoma. Ai Zheng 26:458–462.
- Clevenger CV (2004) Roles and Regulation of Stat Family Transcription Factors in Human Breast Cancer Am J Pathol 165:1449–1460.
- Li H, Ahonen TJ, Alanen K, Xie J, LeBaron MJ, et al. (2004) Activation of signal transducer and activator of transcription 5 in human prostate cancer is associated with high histological grade. Cancer Res 64:4774–4782.
- Nevalainen MT, Xie J, Torhorst J, Bubendorf L, Haas P, et al. (2004) Signal transducer and activator of transcription-5 activation and breast cancer prognosis. J Clin Oncol 22:2053–2060.
- Xi S, Zhang Q, Gooding WE, Smithgall TE, Grandis JR (2003) Constitutive activation of Stat5b contributes to carcinogenesis in vivo. Cancer Res 63:6763– 6771.
- Lin JX, Leonard WJ (2000) The role of Stat5a and Stat5b in signaling by IL-2 family cytokines. Oncogene 19:2566–2576.
- Grimley PM, Dong F, Rui H (1999) Stat5a and Stat5b: fraternal twins of signal transduction and transcriptional activation. Cytokine Growth Factor Rev 10:131–157.
- Sultan AS, Xie J, LeBaron MJ, Ealley EL, Nevalainen MT, et al. (2005) Stat5 promotes homotypic adhesion and inhibits invasive characteristics of human breast cancer cells. Oncogene 24:746–760.
- Li H, Zhang Y, Glass A, Zellweger T, Gehan E, et al. (2005) Activation of signal transducer and activator of transcription-5 in prostate cancer predicts early recurrence. Clin Cancer Res 11:5863–5868.

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- Tang JZ, Zuo ZH, Kong XJ, Steiner M, Yin Z, et al. (2010) Signal transducer and activator of transcription (STAT)-5A and STAT5B differentially regulate human mammary carcinoma cell behavior. Endocrinology 151:43–55.
- Zhang Q, Wang HY, Liu X, Wasik MA (2007) STAT5A is epigenetically silenced by the tyrosine kinase NPM1-ALK and acts as a tumor suppressor by reciprocally inhibiting NPM1-ALK expression. Nat Med 13:1341–1348.
- Du W, Wang YC, Hong J, Su WY, Lin YW, et al. (2012) STAT5 isoforms regulate colorectal cancer cell apoptosis via reduction of mitochondrial membrane potential and generation of reactive oxygen species. J Cell Physiol 227:2421–2429.
- Kim DY, Cha ST, Ahn DH, Kang HY, Kwon CI, et al. (2009) STAT3 expression in gastric cancer indicates a poor prognosis. J Gastroenterol Hepatol 24:646–651.
- Chung CD, Liao J, Liu B, Rao X, Jay P, et al. (1997) Specific inhibition of Stat3 signal transduction by PIAS3. Science 278:1803–1805.
- Greenhough A, Smartt HJ, Moore AE, Roberts HR, Williams AC, et al. (2009) The COX-2/PGE2 pathway: key roles in the hallmarks of cancer and adaptation to the tumour microenvironment. Carcinogenesis 30:377–386.
- Harris SG, Padilla J, Koumas L, Ray D, Phipps RP (2002) Prostaglandins as modulators of immunity. Trends Immunol 23:144–150.
- 43. Li F, Liu Y, Chen H, Liao D, Shen Y, et al. (2011) EGFR and COX-2 protein expression in non-small cell lung cancer and the correlation with clinical features. J Exp Clin Cancer Res 30:27.
- Leahy KM, Koki AT, Masferrer JL (2000) Role of cyclooxygenases in angiogenesis. Curr Med Chem 7:1163–1170.
- Dohadwala M, Luo J, Zhu L, Lin Y, Dougherty GJ, et al. (2001) Non-small cell lung cancer cyclooxygenase-2-dependent invasion is mediated by CD44. J Biol Chem 276:20809–20812.
- Khuri FR, Wu H, Lee JJ, Kemp BL, Lotan R, et al. (2001) Cyclooxygenase-2 overexpression is a marker of poor prognosis in stage I non-small cell lung cancer. Clin Cancer Res 7:861–867.
- Hida T, Yatabe Y, Achiwa H, Muramatsu H, Kozaki K, et al. (1998) Increased expression of cyclooxygenase 2 occurs frequently in human lung cancers, specifically in adenocarcinomas. Cancer Res 58:3761–3764.
- Mascaux C, Martin B, Paesmans M, Berghmans T, Dusart M, et al. (2006) Has Cox-2 a prognostic role in non-small-cell lung cancer? A systematic review of the literature with meta-analysis of the survival results. Br J Cancer 95:139–145.
- 49. Groen HJ, Sietsma H, Vincent A, Hochstenbag MN, van Putten JW, et al. (2011) Randomized, placebo-controlled phase III study of docetaxel plus carboplatin with celecoxib and cyclooxygenase-2 expression as a biomarker for patients with advanced non-small-cell lung cancer: the NVALT-4 study. J Clin Oncol 29:4320–4326.
- Jiang H, Wang J, Zhao W (2013) Cox-2 in non-small cell lung cancer: a metaanalysis. Clin Chim Acta 419:26–32.