A Rare Case of *Cryptococcus gattii* Meningitis in Advanced HIV Disease, Sagittal Thrombosis, and Immune Reconstitution Syndrome, Resolved With Isavuconazonium

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Abstract

Cryptococcus gattii is a species that has received more recognition in the recent past as distinct from *Cryptococcus neoformans*. *C gattii* is known to cause meningeal disease in both immunocompetent and immunosuppressed hosts. Patients may be clinically asymptomatic until immunosuppressive conditions occur such as corticosteroid treatment or an HIV infection. HIV-associated cryptococcal infections are most often due to *C neoformans*. *C gattii* is found in a minority. Speciation and subtyping of *Cryptococcus* are not always accomplished. In many parts of the world, there is no availability for speciation of *Cryptococcus*. Travel history may provide a clue to the most probable species. This case demonstrates a case of *C gattii* meningitis with a multiplicity of complications. These include advanced HIV disease secondary to nonadherence, immune reconstitution inflammatory syndrome, and superior sagittal sinus thrombosis. The patient represented diagnostic and therapeutic dilemmas over time. Headache was the primary symptom in cryptococcal meningitis, immune reconstitution inflammatory syndrome, and superior sagittal sinus thrombosis. All are discussed in detail as potential etiologies for the primary disease. Isavuconazonium is a relatively new broad-spectrum antifungal azole that was used as salvage therapy.

Keywords

Cryptococcus gattii, cryptococcal meningitis, superior sagittal sinus thrombus, thrombus in HIV, IRIS, isavuconazonium, azoles

Introduction

There are approximately 1 million cases of cryptococcosis each year worldwide and an estimated 650 000 associated deaths. Patients with advanced HIV disease comprise the majority.[1-3]

Cryptococcus has 37 known species but only 2 have been identified as pathogenic for cryptococcosis: *C neoformans* and *C gattii*. In 1970, *C gattii* was first described in Australia as a subspecies of *C neoformans*. It is a fungus associated with several ecologic niches.[4-6] In 1999, *C gattii* emerged in British Columbia and the northwest United States. *C gattii* was recognized as a separate species from *C neoformans* in 2002.[7]

C neoformans is globally ubiquitous and accounts for nearly all meningeal infections in patients with HIV. *C gattii* has a more restricted distribution and accounts up to 15% of cryptococcal meningeal infections in endemic areas. The geographic range appears to be expanding with the continued study of *C gattii*.[6,8]

C neoformans and *C gattii* are encapsulated, heterobasidiomycetous fungi. The asexual form is recovered from clinical cases and is an encapsulated yeast that reproduces by budding.[9] The respiratory tract is the primary portal of entry.[9] Lymphohematogenous dissemination is responsible for seeding other organs including the meninges (see Figure 1).
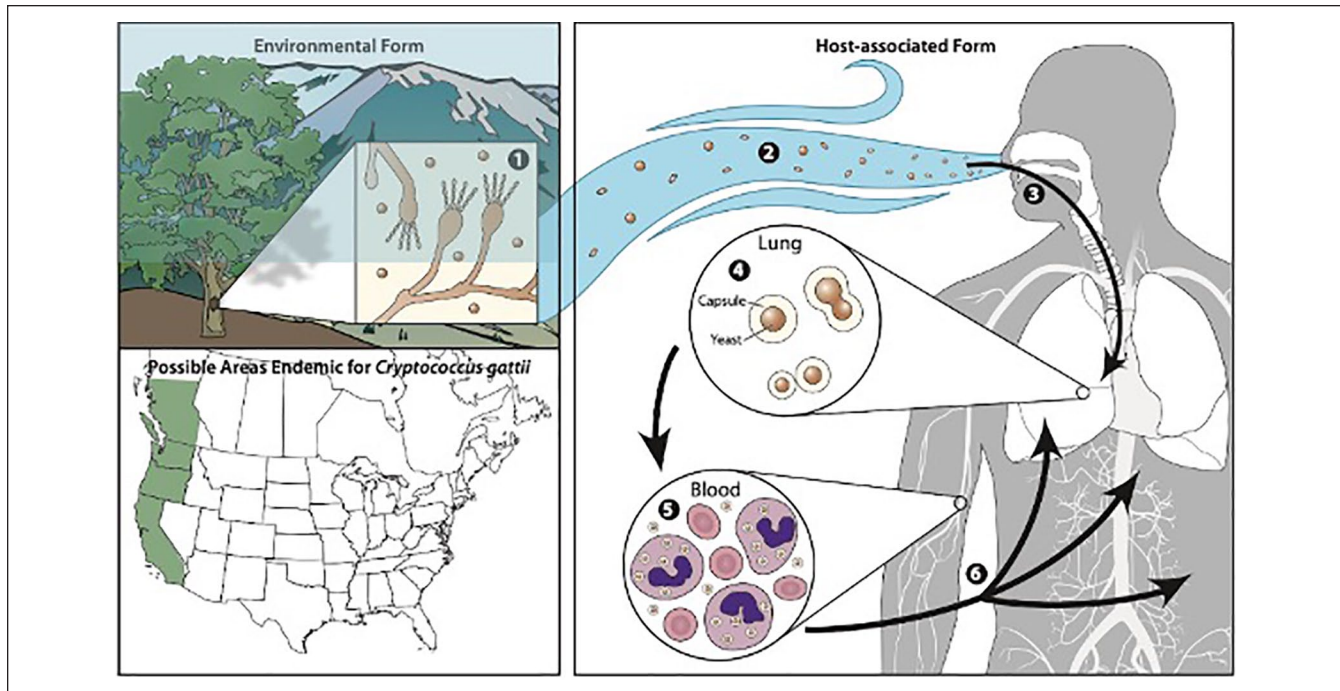
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Figure 1. Encapsulated yeast inhaled into the respiratory tract. Lymphohematogenous dissemination.[4]

The lungs and the central nervous system (CNS) are the most common clinical sites of infection.[2,9] Involvement of the parenchyma of the brain and meninges occurs in 40% to 86% of patients.[10] Characteristic manifestations of CNS disease include meningitis, cryptococcomas, and spinal cord granulomas. Patients typically present with signs and symptoms of meningitis including fever, headache, nausea, vomiting, focal neurologic deficits, memory loss, or change in consciousness including cloudy/delirium, obtundation/stupor, and coma.[9,11]

One of the most critical determinants of outcome for cryptococcal meningoencephalitis is the control of intracranial pressure (ICP). Approximately 50% of HIV-infected patients with cryptococcal meningoencephalitis demonstrate an elevated baseline ICP >250 mm H_2O of CSF. Lumbar puncture (LP) is recommended. An opening pressure of ≥250 mm H_2O with symptoms warrants medical therapy. CSF drainage should be performed sufficiently to achieve a closing pressure of <200 mm H_2O or 50% reduction of initial opening pressure.[1,12]

Neuroimaging is performed in patients with suspected cryptococcal meningitis.[1,9] It does not alter management once cryptococcus is diagnosed. Magnetic resonance imaging (MRI) is preferred in patients with HIV disease and may demonstrate leptomeningeal enhancement or focal brain disease.[9] LP for high ICP is much more important to both follow and treat elevated ICP.

Cerebrospinal fluid (CSF) analysis in cryptococcosis typically reveals a lymphocytic pleocytosis with elevated protein and decreased glucose levels. Cell counts vary from <20 cells/cmm to >100 cells/cmm.[13,14] Low lymphocyte proliferative responses after stimulation with antigens are particularly noted in persons with active HIV disease. The CSF formula is that of chronic meningitis and is, therefore, the differential diagnosis.[15]

Mycologic diagnosis is accomplished by a variety of methods including positive India ink examination of CSF and blood, fungal cultures, and cryptococcal antigen (CrAg) detection in blood and spinal fluid.[9,14]

Management of *C gattii* meningitis is approached with consideration for 3 risk groups: HIV-infected individuals, organ transplant recipients, and ostensibly immune normal hosts. For each respective risk group, therapeutic algorithms have been articulated. This includes an induction phase, consolidation phase, and a maintenance phase. Induction therapy for meningoencephalitis involves fungicidal regimens, such as a polyene and flucytosine. It is followed by suppressive regimens using fluconazole. Recognition of increased ICP is critical and should be addressed as a separate problem from meningitis. See reference guidelines by the Infectious Diseases Society of America (IDSA).[1]

The host response to *C gattii* depends on the competency of the host's innate and adaptive immune systems. Host immunodeficiencies of the innate system may eventuate into clinical cryptococcal disease if there is macrophage failure. For the adaptive immune system, cryptococcal disease progression is facilitated in the setting of a predominately Th-2 response. There is a depletion of CD4^{+} T-helper (Th)-1 and Th-17 cells, as well as an increase in Th-2 cells. Failure to produce a Th-1 dominate response means failed host protection against cryptococcal infection.

HIV infection may be considered a failure of host cellular responses in the progressed phase of disease. Initially, however, HIV selectively infects CD4+ T-cells incognito and replicates intracellularly. It is undetected by the innate or the adaptive systems. There is pathogenic hyperactivation and proliferation of infected CD4$^+$ T cells specific for antigens other than HIV. Eventually, CD4+ T-cell depletion results and accelerates HIV disease progression.[16]

Immune reconstitution inflammatory syndrome (IRIS) is an immunologic exacerbation based on improved host response to antiretroviral therapy. Potential treatment challenges related to IRIS are well described in HIV and cryptococcal meningeal coinfection.[8,9] Similar to cryptococcal meningitis and HIV-encephalitis, the presenting symptoms commonly include headache, fever, and meningismus.[8] Increased ICP, hydrocephalus, and systemic inflammation can also occur.

Cerebral venous thrombosis (CVT) is the rarest complication of venous thrombosis in cryptococcal meningitis and is difficult to diagnose.[17,18] Complaints of headaches require cerebral vascular imaging for diagnosis. Evidence-based recommendations indicate anticoagulation treatment to reduce the risk of cardiovascular complications and death. The American Heart Association and American Stroke Association recommend anticoagulation for 3 to 6 months in provoked CVT, and 6 to 12 months in unprovoked CVT.[18]

CVT occurs at an annual incidence of 3 to 5 per million and tends to affect women and younger individuals.[17] Risk factors for CVT include thrombophilia, infection, trauma, immobilization, reproductive (especially puerperium and pregnancy), malignancy, medications, inflammatory, hematological, endocrine, systemic, and intracranial abnormalities.[18] Multiple risk factors of thrombosis may coexist as demonstrated in our case (advanced HIV disease, IRIS, and cryptococcus infection). The impact of multiple risk factors for venous thrombosis and its implications for management highlights the need for early comprehensive risk assessment.

Prophylactic strategies are considered. A heightened awareness of the potential difficulty in achieving adequate anticoagulation due to drug-drug interaction in patients on antiretroviral regimens is merited.

Methods

Kern Medical Institutional Review Board approval was obtained for review of the patient's record. A literature search was conducted on PubMed, ResearchGate, Google Scholar, and major journal databases including Infectious Diseases Society of America, Centers for Disease Control and Prevention, and Clinical Infectious Diseases. The following search terms were applied: *Cryptococcus gattii*, cryptococcal meningitis, superior sagittal sinus thrombus, cerebral venous thrombosis, thrombus in HIV, IRIS, isavuconazonium, azole side effects, ICP, and meningitis. Twenty-three reference articles were pulled.

Case Presentation

A 45-year-old Caucasian man was diagnosed with HIV disease at another institution. He was nonadherent with antiretroviral therapy. One month after diagnosis, he presented to our institution with a severe headache and vomiting for 2 days.

On physical examination, his temperature was 37.3 °C, the pulse was 63 beats per minute, the respiratory rate was 21 breaths per minute, and the blood pressure was 140/91 mm Hg. The oxygen saturation was 97%, while the patient was breathing ambient air. He was in mild distress secondary to his headache. The general physical examination was unremarkable except for mild meningismus. The neurologic examination revealed normal mental status, cranial nerves, motor, gait, and balance.

Complete blood count with differential revealed lymphocytopenia (0.9×10^3 cells/μL) and routine chemistry was all normal except for mild transaminitis (aspartate aminotransferase 56, alanine aminotransferase 141 units/L). His absolute CD4+ cell count was 38 cells/μL, and his HIV-1 RNA polymerase chain reaction revealed 35 284 copies/mL.

LP demonstrated an opening pressure of 340 mm H_2O. The CSF analysis revealed a red blood cell count of 10 cells/cmm, a white blood cell count of 24 cells/cmm, lymphocytes of 73%, monocytes of 25%, neutrophils of 1%, and eosinophils of 1%. CSF glucose was 56 mg/dL and CSF protein was 75 mg/dL. Analysis of the CSF with India ink was positive for *Cryptococcus* species. Latex agglutination assay of CSF revealed a CrAg titer of 1:2046. Subsequently, the blood was positive for yeast. Serum assay revealed a CrAg titer of 1:4096.

Chest radiographs showed 2 irregular noncalcified densities of the right lower lobe with early central cavitation. Bronchoalveolar lavage confirmed *Cryptococcus*. Neuroimaging of the head was found to be negative.

Cryptococcal meningitis with pulmonary cryptococcoma was diagnosed. The patient was admitted and started on induction therapy with intravenous liposomal amphotericin B (AmB) and oral flucytosine. After a 14-day course, he was discharged home on consolidation therapy with fluconazole and antiretroviral therapy. He was followed closely by the infectious disease clinic.[1] Fungal isolates from blood and CSF were demonstrated to be *C gattii*.

The patient was hospitalized 9 times over a 12-month period for recurrence of fever, headache, and elevated ICP. Therapeutic LPs with CSF removal were intermittently required. A shunt was not placed due to inconsistent need for CSF drainage beyond a few days per episode.

The serum and CSF CrAg varied considerably over this time course. The maximum serum CrAg titer was 1:>4096
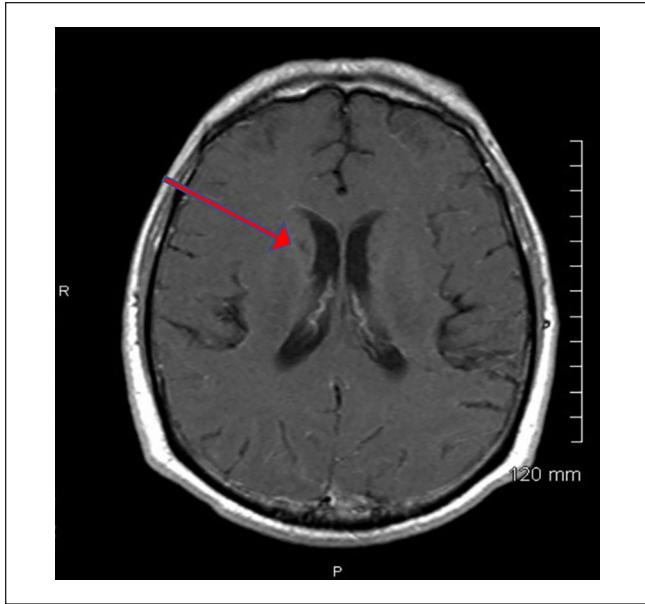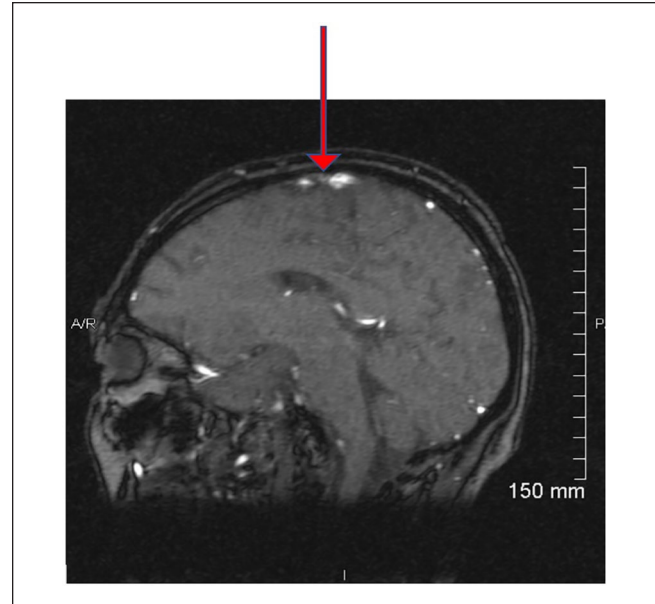
Figure 2. Magnetic resonance imaging of brain axial T1-weighted postcontrast with a red arrow showing the cryptococcoma.

Figure 3. Magnetic resonance venography of brain showing a red arrow at the region of signal void of the sagittal 2-dimensional echo sequence source images, indicating a superior sagittal sinus thrombosis.

and the minimum was 1:256. The maximum CSF CrAg titer was 1:>2048 and the minimum was 1:8. Fluconazole levels were judged to be therapeutic (41.4 μg/mL serum fluconazole).

In the fourth hospitalization, a follow-up MRI of the brain demonstrated the development of cryptococcomas in the right posterior cerebellum and left temporal region. He was declared a therapeutic failure on fluconazole. Isolates were sent for sensitivity testing and results showed. Isolate sensitivity did not indicate drug resistance to any antifungal medications. Sensitivity results revealed AmB 0.5 μg/mL, fluconazole 4.0 μg/mL, natamycin 4.0 μg/mL, itraconazole 0.25 μg/mL, posaconazole 0.25 μg/mL, voriconazole 0.06 μg/mL, isavuconazole (ISA) 0.125 μg/mL. Induction therapy with AmB and flucytosine was reintroduced for 6 weeks. Subsequently, consolidation therapy with voriconazole 450 mg twice daily was initiated, at calculated dosing 6 mg/kg.

Salvage therapy with voriconazole, did not improve his fever, headache, or elevated ICP, prevent hospitalizations, or reduce the CSF or blood CrAg titers. He was adherent to therapy, but therapeutic levels were not achieved. After 3 months on voriconazole, the patient developed dysphagia that drew concern for possible onset of IRIS or worsening CNS cryptococcosis. *Candida* was not identified. The dysphagia self-resolved within 2 months, requiring no further workup.

On the ninth hospitalization, and 1 year into treatment, an MRI of the head, revealed a new cryptococcoma (see Figure 2) and evidence of CVT. Magnetic resonance venography of the brain revealed a superior sagittal sinus thrombosis (see Figure 3). Superior sagittal sinus thrombosis was diagnosed and treated with anticoagulation therapy[18] (see

Figure 4 for diagnoses, hospitalizations and therapeutic regiments).

Most of the patient's treatment was spent inpatient. He was started on multiple rounds of induction therapy, with regiment durations of 2 weeks or 6 weeks. Typically, he ended up in the emergency room with hospitalization for elevated ICP. Consolidation therapies included fluconazole and voriconazole, both of which failed, despite patient adherence. The therapeutic levels of antifungal medications were measured in the serum regularly throughout treatment. Therapeutic levels were achieved consistently with fluconazole antifungal treatment. Voriconazole was consistently identified at subtherapeutic in the serum despite patient adherence.

Isolate sensitivity did not suggest resistance, but his worsening condition warranted therapeutic modification. Antifungal therapy was switched from voriconazole to ISA 372 mg daily. Concern for IRIS was also increasing. Antifungal and antiretroviral therapies were continued in accordance to guidelines. No treatment holiday was taken. Treatment efforts were focused on ICP control, anticoagulation therapy for CVT, and continued adherence to antifungal therapy and medications for HIV.

The patient's cryptococcal disease significantly improved since his transition to ISA. His headache and ICP improved. His absolute CD4+ cell count was 308 cells/μL. HIV-1 RNA polymerase chain reaction was not detected (<20 copies/mL: "not detected") at his 1-year follow-up. The superior sagittal sinus thrombosis also resolved as demonstrated by magnetic resonance venography. There have not been any

Diagnoses, Hospitalizations & Therapeutic Regiments										
	Initial HIV Diagnosis (*CD4/ VL) 6/2017	**^KM HV1** 7/2017	**KM HV2** 10/2017	**KM HV3** 10/2017	**KM HV4** 10/2017	**KM HV5** 04/2018	**KM HV6** 06//2018	**KM HV7** 07/2018	**KM HV8** 07/2018	**KM HV9** 08/2018
+LOS	Outside facility	14 days	1 day	1 day	8 days	6 days	9 days	5 days	11 days	1 days
ART Therapy (ABC/DTG/3TC)	Initiated	Continued	Continued	Continued	Continued	Continued	Contin-ued	Continued	Continued	Continued
Adherent to ~ ART Therapy?	No	No	No	No	No	Yes	Yes	Yes	Yes	Yes
Antifungal Induction Therapy	Not Applicable	!AmB // Flucyt. 2-week regiment	None	None	AmB Flucyt. 6-week regiment	AmB Flucyt. 6-week regiment	None	None	AmB 6-week regiment	Continued AmB → 6-week regiment
Antifungal Consolidation Therapy	Not Applicable	=Flucon. started at discharge	Flucon. continued	Flucon. continued	Flucon. Failure -Isolates sent for sensitivity testing (Not Resistant) #Vori Started	Vori	Vori	Vori	Vori Failure	&ISA started
Adherent to Antifungal Therapy?	Not Applicable	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Therapeutic Antifungal levels reached?	Not Applicable	Not Applicable	Yes	Yes	Not Appli-cable	No	No	No	No	Not Applicable
Cryptococcosis Initial Diagnosed		X								
Pulmonary Cryptococcomas		X								New Lesions
CNS Cryptococcomas					X					New Lesions
ºSSS Cerebral Thrombosis								X		
IRIS									X	

*CD4+ cell count was 38 cells/microliter, HIV-1 RNA polymerase chain reaction (PCR) revealed 35,284 copies/mL.
^KM HV#: Kern Medical Hospitalization Visit #
+LOS: Length of Stay
~ART Therapy (ABC/DTG/3TC): Antiretroviral therapy abacavir (ABC), lamivudine (3TC), dolutegravir (DTG)
!AmB: Liposomal Amphotericin B
//Flucyt: Flucytosine
=Flucon: Fluconazole
    Fluconazole therapeutic range is normally 5 - 20 mcg/mL, but some clinicians use range 1 - 40 mcg/mL.
#Vori: Voriconazole
    Voriconazole therapeutic range for treatment: trough 2.0 to 5.5 mcg/mL. Serum trough levels less than or equal to 1 mcg/mL are reported to be associated with lack of therapeutic response
&ISA: isavuconazonium
oSSS: Superior Sagittal Sinus

Figure 4. Diagnoses, hospitalizations, and therapeutic regiments.

Table 1. Literature Review of Cryptococcal Meningitis.

Study	Publishing type	Country	Age (years)	First symptom at presentation	Thrombosis location	IRIS	HIV disease (CD4+ count at time of diagnosis)	Cryptococcus meningitis and species
Thiansukhon et al[19]	Poster	Thailand	49	Headache	Straight sinus Transverse and sigmoid venous sinuses bilaterally	No	Yes CD4+ count: 70 cells/mm^3	Yes Cryptococcus neoformans
Alejandra et al[20]	Case report	Mexico	21	Headache	Left transverse sinus Bilateral sigmoid sinuses	No	Yes CD4+ count: 96 cells/mm^3	Yes No speciation
Senadim et al[21]	Case report	Turkey	19	Headache	Right transverse sinus	No	No	Yes Cryptococcus neoformans
Kulkarni et al[22]	Case report	India	37	Headache	Superior sagittal sinus Right transverse sinus Proximal internal jugular vein	No	Yes CD4+ count: 24 cells/mm^3	Yes No speciation
Ren et al[23]	Case report	China	31	Headache	Bilateral transverse sinus Superior sagittal sinus Inferior longitudinal sinus	No	No	Yes No speciation
Equiza et al[24]	Case report	Spain	45	Headache, dizziness, nausea	Right transverse sinus Right sigmoid sinus	No	Yes Unknown	Yes Cryptococcus neoformans
Mohamed et al[25]	Case report	Kenya	40	Headache, vomiting, blurred vision, painful Right eye	Distal superior sagittal sinus Left transverse sinus	No	Yes CD4+ count: 9 cells/mm^3	Yes No speciation
Kammeyer and Lehmann[26]	Case report	United States	61	Headache, chills, night sweats	Left transverse and sigmoid sinus	No	No	Yes Cryptococcus neoformans
Heidari 2020	Case report	United States	45	Headache	Superior sagittal sinus	Yes	Yes CD4+ count: cells/mm^3	Yes Cryptococcus gattii

Abbreviation: IRIS, immune reconstitution inflammatory syndrome.

further hospitalizations since ISA therapy. He is adherent to ISA maintenance therapy and antiretroviral therapy 12 months after the ninth hospital discharge.

Discussion

Eight cases were found in the medical literature that recognize concomitant cryptococcal meningitis and CVT (Table 1). Five of these 8 cases involve individuals with HIV disease. Specifically, CVT was identified in the transverse sinus, superior sagittal sinus, and sigmoid sinuses in these 8 cases. *C neoformans* was found in half of the recorded cases, while 3 did not distinguish the species. We were unable to identify any instances of CVT involving *C gattii*. Additionally, none of these cases demonstrated the multiplicity of problems, extensive hospitalizations, and specific therapy as reported here.

A failed induction or consolidation therapy requires a modification of dose, duration, or change of the therapeutic agent. Relapse of signs and symptoms during treatment should be carefully assessed to decipher between a failure to control fungal growth from drug resistance, or adherence, or IRIS. IRIS is an additional problem seen in HIV and can mimic therapeutic failure. Secondary resistance to fluconazole is an emerging problem in some geographical locations. This was not case in our patient as demonstrated by the sensitivity results (AmB 0.5 µg/mL, fluconazole 4.0 µg/mL, natamycin 4.0 µg/mL, itraconazole 0.25 µg/mL, posaconazole 0.25 µg/mL, voriconazole 0.06 µg/mL, isavuconazole 0.125 µg/mL). In persistent and relapse infections, isolates should be submitted for sensitivity testing and alternative agents should be considered.[1]

Low CD4+ counts (<200 cells/mm^3), high viral load, and nonadherence to antiretroviral therapies are all risk factors for venous thrombosis. The most common risk factor is an ongoing infection. Proinflammatory cytokines, interluekin-6 and TNF-α, and endothelial activation have been implicated as underlying pathogenesis.[27]

Patients with advanced HIV disease with active *C gattii* meningitis appear to be at a higher risk for complications than persons without HIV disease. Individuals with *Cryptococcus* and HIV disease are at substantial risk for IRIS. IRIS is most probable in patients who have a very low CD4 count (<200 cells/mm^3) when antiretroviral therapy is initiated. HIV therapy was difficult in this patient because he was not adherent at both the first institution and at our institution with initial therapies. With varying timelines of poor adherence and monitoring of his ART medications, all may have contributed to the development of IRIS. The timing of initiating antiretroviral therapy remains challenging.

Minor IRIS manifestations will typically resolve spontaneously in days to weeks. Major IRIS complications, such as CNS inflammation with increased ICP may require corticosteroids.[1] The exactness of when to initiate antiretroviral therapy in the setting of cryptococcus and HIV coinfection to side skirt IRIS remains indeterminate. Recommendations propose a wide range of 2 to 10 weeks should be undertaken to accommodate this uncertainty.[1]

Conclusion

Patients with complex cryptococcal disease may have a multiplicity of complications that include increased ICP, hydrocephalus, and CVT. An additional complication is IRIS more common in HIV patients. The clinical presentation of these symptoms may overlap significantly. Significant effort is required to discern the appropriate therapeutic intervention to achieve resolution.

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References

1. Perfect JR, Dismukes WE, Dromer F, et al. Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the Infectious Diseases Society of America. *Clin Infect Dis*. 2010;50:291-322. doi:10.1086/649858
2. Pappas PG. Cryptococcal infections in non-HIV-infected patients. *Trans Am Clin Climatol Assoc*. 2013;124:61-79.
3. MacDougall L, Fyfe M, Romney M, Starr M, Galanis E. Risk factors for *Cryptococcus gattii* infection, British Columbia, Canada. *Emerg Infect Dis*. 2011;17:193-199. doi:10.3201/eid1702.101020
4. Centers Disease Control Prevention. Emergence of *Cryptococcus gattii*—Pacific Northwest, 2004-2010. *MMWR Morb Mortal Wkly Rep*. 2010;59:865-868.
5. Harris JR, Lockhart SR, Debess E, et al. *Cryptococcus gattii* in the united states: clinical aspects of infection with an emerging

pathogen. *Clin Infect Dis*. 2011;53:1188-1195. doi:10.1093/cid/cir723

6. Sorrell TC. *Cryptococcus neoformans* variety *gattii*. *Med Mycol*. 2001;39:155-168. doi:10.1080/mmy.39.2.155.168

7. Kwon-Chung KJ, Boekhout T, Fell JW, Diaz M. (1557) Proposal to conserve the name *Cryptococcus gattii* against *C hondurianus* and *C bacillisporus* (*Basidiomycota, Hymenomycetes, Tremellomycetidae*). *Taxon*. 2002;51(4):804-806. doi:10.2307/1555045

8. Sun HY, Alexander BD, Huprikar S, et al. Predictors of immune reconstitution syndrome in organ transplant recipients with cryptococcosis: implications for the management of immunosuppression. *Clin Infect Dis*. 2015;60:36-44. doi:10.1093/cid/ciu711

9. Perfect JR. Cryptococcosis (*Cryptococcus neoformans* and *Cryptococcus gattii*). In: Bennett JE, Dolin R, Blaser MJ, eds. *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. 9th ed. Elsevier; 2019:3146-3161. doi:10.1016/B978-1-4557-4801-3.00264-2

10. Aydin H, Kaya I. Raised intracranial pressure. In: Turgut M, Challa S, Akhaddar A, eds. *Fungal Infections of the Central Nervous System*. 1st ed. Springer; 2019:255. doi:10.1007/978-3-030-06088-6

11. Posner JB, Saper CB, Schiff ND, Plum F, eds. *Plum and Posner's Diagnosis and Treatment of Stupor and Coma*. 5th ed. Oxford University Press; 2019:5.

12. Chayakulkeeree M, Perfect JR. Cryptococcosis. In: Hospenthal DR, Rinaldi MG, eds. *Diagnosis and Treatment of Human Mycoses*. 1st ed. Springer; 2007:273.

13. Satishchandra P, Mathew T, Gadre G, et al. Cryptococcal meningitis: clinical, diagnostic and therapeutic overviews. *Neurol India*. 2007;55:226-232. doi:10.4103/0028-3886.35683

14. Lee SJ, Choi HK, Son J, Kim KH, Lee SH. Cryptococcal meningitis in patients with or without human immunodeficiency virus: experience in a tertiary hospital. *Yonsei Med J*. 2011;52:482-487. doi:10.3349/ymj.2011.52.3.482

15. Bennett JE, Hoover SE. Chronic meningitis. In: Bennet J, Dolin R, Blaser MJ, eds. *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. Vol 1. 9th ed. Elsevier; 2019:1220-1225. doi:10.1016/B978-1-4557-4801-3.00090-4

16. Deeks SG, Walker BD. The immune response to AIDS virus infection: good, bad, or both? *J Clin Invest*. 2004;113:808-810. doi:10.1172/JCI21318

17. Koopman K, Uyttenboogaart M, Vroomen PCAJ, Van Der Meer J, De Keyser J, Luijckx GJ. Risk factors for cerebral venous thrombosis and deep venous thrombosis in patients aged between 15 and 50 years. *Thromb Haemost*. 2009;102:620-622. doi:10.1160/TH09-06-0346

18. Rosenwasser RH, Weigele JB. Diagnosis and management of cerebral venous and dural sinus thrombosis. *Neurointerventional Manag Diagnosis Treat 2E*. 2012;18:469-484. doi:10.3109/9781841848075.023

19. Thiansukhon E, Potigumjon A, Smitasin N. Cerebral venous sinus thrombosis: a rare complication in cryptococcal meningitis. *Int J Infect Dis*. 2014;21(suppl 1):291. doi:10.1016/j.ijid.2014.03.1023

20. Alejandra G, Carla T, Laura M. Cerebral venous sinus thrombosis associated with cryptococcal meningitis in an HIV-positive patient. *J Clin Case Rep*. 2014;4:421. doi:10.4172/2165-7920.1000421

21. Senadim S, Alpaydin Baslo S, Tekin Güveli B, et al. A rare cause of cerebral venous thrombosis: cryptococcal meningoencephalitis. *Neurol Sci*. 2016;37:1145-1148. doi:10.1007/s10072-016-2550-9

22. Kulkarni A, Kumar M, Philip VJ. Cerebral venous sinus thrombosis: a rare complication of cryptococcal meningitis in HIV/AIDS patient. *IOSR J Dent Med Sci*. 2018;17:22-24.

23. Ren B, Guo Y, Chen K, Chen L. An anti-human immunodeficiency virus-negative pregnant woman was diagnosed with both cerebral venous sinus thrombosis and intracranial *Cryptococcus* simultaneously. *J Emerg Crit Care Med*. 2018;2:55. doi:10.21037/jeccm.2018.05.12

24. Equiza J, Fernandez-Eulate G, Rodriguez-Antigüedad J, et al. Cryptococcal meningoencephalitis presenting as cerebral venous thrombosis. *J Neurovirol*. 2020;26:289-291. doi:10.1007/s13365-019-00813-3

25. Mohamed A, Ali S, Kanyi J, Gardner A. Cerebral venous thrombosis associated with recurrent cryptococcal meningitis in an HIV infected patient. *J Neurol Sci*. 2019;405(suppl):51. doi:10.1016/j.jns.2019.10.861

26. Kammeyer J, Lehmann N. Cryptococcal fungemia leading to cerebral venous thrombosis in a multiple sclerosis patient on fingolimod. (4951). *Neurology*. 2020;94(15 suppl):4951. http://n.neurology.org/content/94/15_Supplement/4951.abstract

27. Crum-Cianflone NF, Weekes J, Bavaro M. Review: thromboses among HIV-infected patients during the highly active antiretroviral therapy era. *AIDS Patient Care STDS*. 2008;22:771-778. doi:10.1089/apc.2008.0010