Expression of miR-146a-5p in breast cancer and its role in proliferation of breast cancer cells

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Abstract. In the present study, the expression level of microRNA-146a-5p (miR-146a-5p) in breast cancer tissue and cell lines was investigated and its effects on proliferation of breast cancer cells. miR-146a-5p expression was detected by reverse transcription-quantitative polymerase chain reaction in breast cancer tissues, paraneoplastic tissue (collected by The Department of Oncology of Changhai Hospital from January 2014 to June 2015), breast cancer cell line MCF-7 and normal breast epithelial cell line MCF 10A. Bioinformatics analysis was conducted to forecast target genes of miR-146a-5p, which was further verified by fluorescent reporter gene detection. The results demonstrated the expression level of miR-146a-5p in breast cancer tissue was significantly higher, compared with paraneoplastic tissue (P<0.01), and the expression level of miR-146a-5p in MCF-7 cells was significantly higher, compared with MCF 10A cells (P<0.01). Overexpression of miR-146a-5p in MCF-7 cells can promote the proliferation, and low expression miR-146a-5p in MCF-7 can inhibit the proliferation. BRCA1 was further identified as a target gene of miR-146a-5p by bioinformatics analysis and fluorescent reporter gene detection. It was concluded that miR-146a-5p is expressed in breast cancer tissue and breast cancer cell line and may regulate the proliferation of MCF-7 via BRCA1.

Introduction

Breast cancer derived from mammary gland epithelial tissue has had an increasing morbidity rate since the 1980s (1). Breast

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cancer is also the most common malignant tumor in China, and accounts for 12.2% of the novel diagnosed cases and 9.6% of mortalities (2).

A microRNA (miRNA) is a small non-coding single strand RNA containing ~22 nucleotides, with a function of combining the 3'untranslated region (UTR) of target genes. With the combination of intracellular proteins to form a RNA interference silencing ribonucleoprotein complex, the target mRNA is bound to and degraded via complete complementary sequence specific binding; however, it can inhibit translation by incomplete complementary combination, to regulate the expression of gene at the post transcriptional level (3,4). At present, nearly 1,000 miRNA sequences have been cloned *in vivo*, including those involved in: The development of the organism (5); cell differentiation and proliferation (6); cell apoptosis (7); tumorigenesis (8); inflammation (9); and numerous other physiological and pathological processes (3). It is speculated that 1/3 of gene expression may be regulated by miRNAs (10).

Numerous studies have indicated that miRNAs serve an important part in genesis and progression of breast cancer (11-15). In the previous experimental study (16), it was determined that the expression of miR-146a-5p in breast cancer tissue was significantly higher, compared with paraneoplastic tissue.

The present study further investigated the expression of miR-146a-5p in breast cancer and its association with the proliferation of breast cancer cells to search for the target gene and understand the molecular mechanism underlying miR-146a-5p, which regulates the proliferation of breast cancer cells (17), providing a novel strategy for the prevention and treatment of breast cancer.

Materials and methods

Breast cancer tissue and paraneoplastic tissue. The surgical specimens of carcinoma tissue and paraneoplastic tissue (<3 cm from the edge of tumor specimens) were collected from 32 clinical diagnosed female patients with breast cancer from January 2014 to June 2015 in The Department of Oncology, Changhai Hospital (Shanghai, China). The median age of the patients was 49 years, and the age ranged from 27-69 years. Any patient underwent chemotherapy, immunotherapy or

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radiotherapy were excluded. All specimens were frozen in liquid nitrogen rapidly and stored at -80°C.

Cell culture. Human breast cancer cell line MCF-7, normal breast epithelial cells MCF 10A and human embryonic kidney cell line, 293T, were purchased from Shanghai Bioleaf Biotech Co., Ltd. (Shanghai, China). Cells were maintained in RPMI-1640 medium (Thermo Fisher Scientific, Inc., Waltham, MA, USA) with 10% FBS in a 5% CO₂ incubator at 37°C. The medium was replaced every 2-3 days. When the cells were at 70-80% confluency, the transfection experiment was carried out.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA was extracted from breast cancer tissue, paraneoplastic tissue, human breast cancer cell line MCF-7 and normal breast epithelial cells MCF 10A. RT-qPCR was used to detect the expression of miR-146a-5p. Primers of miR-146a-5p and β-actin U6 were designed and synthesized by Sangon Biotech Co., Ltd. (Shanghai, China). RT primers were of an miRNA specific stem loop structure. OligodT20 were used as RT primers for reverse transcription. The relative expression of miRNA and U6 were detected by using in SYBR Premix Ex Taq enzyme (Takara Biomedical Technology (Beijing) Co., Ltd., Beijing, China) and specific qPCR primer. The primer sequences were as follows: miR-146a-5p forward, 5'-cgagtccagttttcccagga-3', and reverse, 5'-gtcgtatccagtgcaggg-3'; U6 forward, 5'-ctcgcttcggcagcaca-3', and reverse, 5'-aacgcttcacgaatttgcgt-3'.

RT-qPCR reaction system: 7.5 μ l 2X SYBR master Mix, 0.3 μ l forward Primer, 0.3 μ l reverse Primer, 1.0 μ l cDNA, 5.9 μ l ddH₂O and a total volume of 15 μ l.

RT-qPCR reaction conditions: 95°C for 10 min, then 95°C 15 sec, annealing temperature (55°C) depending on the primer for 20 sec and 72°C for 30 sec, for 40 cycles, this was following with 72°C for 10 min. The $2^{-\Delta\Delta Cq}$ method was used for quantification (18).

Transfection and experimental groups. The logarithmic growth phase cells were divided into blank control group, mock group, over expression group and suppression group. The blank control group had no intervention, whilst the mock, over expression and suppression group were transfected with mock (empty vector pSuper plasmid gifted from Shanghai Institutes for Biological Sciences), 100 nmol/l mimic miR-146a-5p and 100 nmol/l inhibitor miR-146a-5p (Shanghai GenePharma Co., Ltd, China), respectively, according to the Lipofectamine[®] 2000 (Invitrogen; Thermo Fisher Scientific, Inc. CA, USA) specification. Mimic miR-146a-5p and inhibitor miR-146a-5p were synthesized by Shanghai GenePharma Co., Ltd. (Shanghai, China). The effects of transfections were evaluated through RT-qPCR by detecting the miR-146a-5p expression level of each group.

MTT. MTT was used to detect the proliferation rate at 1 day (D1), 2 days (D2) and 3 days (D3) following transfection. The cells in the logarithmic growth phase were seeded into a 96 well plate ($5x10^3$ cells/well). A total of 20 μ l 5 μ g/ μ l MTT solution was added into each well at D1, D2 and D3 following transfection, then 150 μ l dimethyl sulfoxide (Henan Tianfu

Chemical Co., Ltd., Henan, China) was also added into each well following a 4-h incubation at room temperature, value the optical density of each group at detection wavelength of 490 nm. The proliferation ratio of the other groups were calculated based on comparison with the blank control group value and the experiment was repeated three times.

Bioinformatics prediction. Using TargetScan (http://www.targetscan.org/vert_71/), PicTar (http://www.pictar.org), MiRBase (http://www.mirbase.org) and BibiServ (https://bibiserv.cebitec.uni-bielefeld.de/index.html) software to predict the potential target gene of miR-146a-5p. The results were screened according to the matching condition between miRNA seed sequence and target sequence, RNA double chain free energy and miRNA target sequence conservation among different species.

Dual fluorescence reporter gene assay. The 3'UTR region of BRCA1 gene sequence that can interact with 3'UTR region of miR-146a-5p was cloned into the same region in reporter gene Luciferase of plasmid pGl3. If miR-146a-5p can act on the target gene sequence, then the translation process of report gene luciferase will be inhibited, the amount of luciferase protein with activity will be reduced, thus the catalytic substrate luminescence signal will be reduced; therefore, the inhibitory effect of miR-146a-5p on the target gene sequence can be reflected indirectly by the luminescence signal strength. Pre-miR-146a-5p was amplified with the 293T cell genome as a template, subcloned into pSuper plasmid H1 RNA promoter into multiple downstream cloning sites, and the recombinant pSuper plasmid expressing pre-miR-146a-5p was constructed. Cell lysis following transfection into 293T cells for 24 h, then the luciferase detection kit (Promega Corporation, Madison, WI, USA) was used to detect the relative expression of luciferase.

Statistical analysis. SPSS 13.0 (SPSS, Inc. Chicago, IL, USA) statistical software was applied to determine to statistical significance. Data are presented as mean ± standard deviation. Two groups were compared by Student's t-test and the comparison between multiple groups was conducted by one-way analysis of variance followed by Student-Neuman-Keuls post hoc comparisons.

Results

Expression of miR-146a-5p in breast cancer and breast cancer cell lines. The results of RT-qPCR demonstrated that the expression of miR-146a-5p in breast cancer tissue was 3.2 ± 0.3 times more, compared with paraneoplastic tissue (Fig. 1A, P<0.01), the expression in MCF-7 cell lines was 2.9 ± 0.3 times more, compared with MCF 10A cells (Fig. 1B, P<0.01).

miR-146a-5p mimic and inhibitor transfection into MCF-7 cells. According to the miR mimic and inhibitor transfection introduction, the transfection concentration was 50 nM. The results indicated that the expression of miR-146a-5p in MCF-7 cells transfected with miR-146a-5p mimic was 80±7 times more, compared with original expression (Fig. 2A, P<0.01), whilst cells transfected with miR-146a-5p inhibitor decreased



Figure 1. Expression of miR-146a-5p in breast cancer tissue and breast cancer cell line. (A) The expression of miR-146a-5p in breast cancer tissues was significantly higher, compared with the paraneoplastic tissue. **P<0.01. (B) The expression of miR-146a-5p in breast cancer cell line MCF-7 was significantly higher, compared with the normal breast epithelial cells MCF 10A. **P<0.01. miR-146a-5p, microRNA-146a-5p.



Figure 2. Mimic miR-146a-5p and inhibitor miR-146a-5p transfected MCF-7 cells. (A) Transfected with mimic miR-146a-5p, the expression of miR-146a-5p in MCF-7 cells was significantly upregulated, compared with blank control group. **P<0.01. (B) Transfected with inhibitor miR-146a-5p, the expression of miR-146a-5p in MCF-7 cells was significantly downregulated, compared with the blank control group. **P<0.01. miR-146a-5p, microRNA-146a-5p.





Figure 3. Effects on the proliferation of MCF-7 following transfection of mimic miR-146a-5p (*P<0.05 vs. the blank control group) and inhibitor miR-146a-5p (*P<0.01 vs. the blank control group). miR-146a-5p, microRNA-146a-5p; OD, optical density.

Figure 5. The luciferase activity of transfected miR-146a-5p group was significantly reduced compared with the control group. **P<0.01.

to about 0.25 ± 0.04 times compared with original expression (Fig. 2B, P<0.01). These results indicated that the transfections were successful.

Effect of miR-146a-5p on the proliferation of breast cancer cell line MCF-7. The proliferation (OD 490) of MCF-7 cells increased gradually following D3. The proliferation of the overexpression group significantly increased following

3'-uuGGGUACCUUAAGUCAAGAGu-5' hsa-miR-146a-5p : I I I I I : I I I I I I I 489:5'-agUCCUUGGG---CAGUUCUCa-3' BRCA1

Figure 4. Bioinformatics analysis revealed that the 3'untranslated region of tumor suppressor gene BRCA1 existed at the acting site of miR-146a-5p. miR-146a-5p, microRNA-146a-5p.

transfection of the miR-146a-5p mimic at D2 (0.51 ± 0.03 , P<0.05) and D3 (0.73 ± 0.04 , P<0.01), whilst the mock group cells at D2 and D3 (D2, 0.42 ± 0.03 ; D3, 0.54 ± 0.05) had no significant difference (Fig. 3) with the blank control group (D2, 0.40 ± 0.04 ; D3, 0.56 ± 0.05). By contrast, the proliferation of suppression group following the transfection of miR-146a-5p inhibitor was significantly decreased, compared with the blank control group at D2 (0.30 ± 0.03 , P<0.05) and D3 (0.41 ± 0.04 , P<0.01). These results indicated that high expression of miR-146a-5p inhibited proliferation, and low expression of miR-146a-5p inhibited proliferation significantly (Fig. 3).

Detection of miR-146a-5p target genes. Through further analysis of bioinformatic methods, it was determined that the 3'UTR of tumor suppressor gene BRCA1 existed at the acting site of miR-146a-5p (Fig. 4). Furthermore, the interacting sequence was highly conserved between species. All examples indicated that BRCA1 may be the target gene of miR-146a-5p. The pGL3 plasmid was recombined with luciferase expression of BRCA1 3'UTR. The luciferase results demonstrated that the luciferase activity (0.40±0.04) of transfected miR-146a-5p group was significantly lower, compared with the control group transfected with mock (Fig. 5, P<0.01), indicating that BRCA1 was the target gene of miR-146a-5p.

Discussion

miRNA is a type of non-coding small single stranded RNA containing 20-24 nucleotides involved in gene transcription and expression (19). miRNA is not encoding protein, but has the ability to direct degrade the target mRNA or inhibit its translation by complete or incomplete complementary combination with the target mRNA (20). Numerous studies have demonstrated that miRNA is involved in the process of tumor growth, metastasis and angiogenesis by regulating the expression of oncogenesis, migration and other associated genes (21-23).

In 2005, Iorio *et al* (24) first reported the change of miRNAs expression profile in human breast cancer and determined that there were 29 kinds of miRNAs with an expression disorder, in which the expression of miRNA-21 and miRNA-155 were significantly upregulated, whilst the expression of miRNA-10b, miRNA-125b and miRNA-145 were significantly reduced. Additionally, a number were associated with the clinical and pathological features of breast cancer, including estrogen and progesterone receptor expression and vascular invasion. In 2007, Blenkiron *et al* (25) determined that there were 133 kinds of miRNA expression in normal breast tissue and/or cancer tissues, a number of which were associated with molecular subtype of breast cancer. Following this, the role of miRNAs in breast cancer gained more attention.

In the previous work, it was determined that the expression of miR-146a-5p in breast cancer tissue was significantly higher, compared with paraneoplastic tissue (16). It was confirmed through RT-qPCR in the present study and also discovered that the expression of miR-146a-5p in breast cancer cell line MCF-7 was also significantly higher, compared with control cells. High expression of miR-146a-5p in MCF-7 could significantly promote the proliferation, and low expression of miR-146a-5p could significantly inhibit it. BRCA1 was preliminarily confirmed as the target gene of miR-146a-5p by bioinformatics prediction and fluorescence reporter gene detection.

BRCA1 is a tumor suppressor gene, which frequently mutated in hereditary breast cancer (26). BRCA1 serves an important role in DNA repair, cell cycle control, transcriptional activation, ubiquitin and other processes (27,28). In the present study, it was determined that the proliferation of MCF-7 cells was enhanced following high expression of miR-146a-5p, whilst the proliferation of MCF-7 cells decreased following low expression of miR-146a-5p. It was hypothesized that the decreased expression of BRCA1, in consequence of the high expression of miR-146a-5p, weakened the anti-tumor ability and caused the increase of tumor cell proliferation. Following miR-146a-5p low expression, the expression of BRCA1 was increased, which caused the inhibition of tumor and the decrease of the proliferation ability.

In conclusion, high expression of miR-146a-5p in breast cancer and breast cancer cell lines may regulate the proliferation of MCF-7 by regulating the expression of BRCA1.

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Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

GW, HJ and YY designed this study. JZ, DJ, HZ, DY and LQ performed the experiments and interpreted the data. GW and HJ analyzed and interpreted the data. GW and HJ performed the histological examination of the breast and were major contributors towards the writing of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The Changhai Hospital Scholastic Ethics Committee approved the research. All methods were performed in accordance with the relevant guidelines and regulations.

Consent for publication

All subjects signed informed consent for publication.

Competing interests

The authors declare that they have no competing interests.

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