

Draft Whole-Genome Sequence and Annotation of *Xenorhabdus griffinae* Strain BMMCB Associated with the South African Entomopathogenic Nematode *Steinernema khoisanae* Strain BMMCB

Boipelo Mothupi,^a Jonathan Featherston,^b Vincent Gray^a

School of Molecular and Cell Biology, University of the Witwatersrand, Johannesburg, South Africa^a; Agricultural Research Council, Biotechnology Platform, Pretoria, South Africa^b

***Xenorhabdus griffinae* strain BMMCB (LDNM0000000) belongs to the family Enterobacteriaceae and was isolated from the South African entomopathogenic nematode *Steinernema khoisanae* strain BMMCB (GenBank accession no. KT027382). Here, we report the draft whole-genome sequence of *X. griffinae* strain BMMCB with a genome size of 4,183,779 bp and 44.7% G+C content. The NCBI Prokaryotic Automatic Annotation Pipeline (PGAAP) revealed 3,970 genes.**

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Address correspondence to Boipelo Mothupi, boipelo.mothupi@students.wits.ac.za.

Xenorhabdus species are symbiotic Gram-negative bacteria which belong to the *Enterobacteriaceae* family and have a specific obligatory association with *Steinernematid* entomopathogenic nematodes (EPNs) (1). The EPN-bacteria complex holds great potential as biocontrol agents of insect pests (2). The *Steinernematid* EPN infective juvenile (IJ) gains entry through the natural openings of insect host larvae and regurgitate the *Xenorhabdus* bacterial endosymbiont into the hemolymph (1). Following infection, *Xenorhabdus* spp. depress the insect host's immune system and release insecticidal metabolites, which are lethal to the insect (3). Bactericidal, fungicidal, and nematocidal compounds released by the bacteria facilitate the maintenance of a monoxenic environment within the infected host's hemocoel (4–6). Several compounds with antibiotic activity secreted by *Xenorhabdus* spp. include benzylacetone (7), xenorhabdins and xenocoumacin (8), phenethylamides (9), and cyclolipopeptide (10). In this study, we present the description of the *X. griffinae* BMMCB draft whole-genome sequence and annotation.

X. griffinae BMMCB was isolated from the hemolymph of *Galleria mellonella* larvae infected with the South African entomopathogenic nematode *Steinernema khoisanae* BMMCB (isolation methods are described in reference 11). Genomic DNA was extracted from colony bacterial cultures using the ZR Fungal/Bacteria DNA kit (catalogue D6005) and was purified with the DNA clean and concentrator – 5 kit (catalogue D4013). The strain was identified through the amplification and sequencing of the 16S rDNA. On NCBI BLAST, the results showed a 98% similarity percentage to *Xenorhabdus griffinae* and was named *Xenorhabdus griffinae* strain BMMCB.

Illumina libraries were generated using the Illumina Nextera DNA sample preparation kit (FC-121-1031) and sequenced using an Illumina MiSeq instrument (version 3, chemistry 300 × 300 bp). CLC Genomic Workbench v7 (CLC bio) was used for quality trimming of adapter sequences and merging of overlapping paired reads. The *X. griffinae* BMMCB genome was subse-

quently assembled *de novo* and 231 contigs (≥400 bp) were generated with an average coverage of 232×. The whole-genome sequence comprises 4,183,779 bp, with an N_{50} of 57,901 bp and G+C content of 44.7%, similar to those of other *Xenorhabdus* spp. (10). The assembly was submitted for annotation to the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) server (http://www.ncbi.nlm.nih.gov/genome/annotation_prok/). Annotation revealed a total of 3,970 genes (3,614 protein coding [CDs] and 271 pseudogenes). The genome has 12 rRNAs (5S, 16S, and 23S), 70 tRNAs, and 3 noncoding RNAs. The polyketide synthase genes involved in biosynthetic pathway of antibiotic compounds have been identified.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. LDNM00000000. The version described in this paper is LDNM01000000.

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REFERENCES

1. Stock PS, Goodrich-Blair H. 2008. Entomopathogenic nematodes and their bacterial symbionts: the inside out of a mutualistic association. *Symbiosis* 46:65–75.
2. Ciche TA, Darby C, Ehlers R, Forst S, Goodrich-Blair H. 2006. Dangerous liaisons: the symbiosis of entomopathogenic nematodes and bacteria. *Biol Contr* 38:22–46. <http://dx.doi.org/10.1016/j.biocontrol.2005.11.016>.
3. Fang XL, Li ZZ, Wang YH, Zhang X. 2011. *In vitro* and *in vivo* antimicrobial activity of *Xenorhabdus bovienii* YL002 against *Phytophthora capsici* and *Botrytis cinerea*. *J Appl Microbiol* 111:145–154. <http://dx.doi.org/10.1111/j.1365-2672.2011.05033.x>.
4. Ogier JC, Pagès S, Bisch G, Chiapello H, Médigue C, Rouy Z, Teyssier C, Vincent S, Tailliez P, Givaudan A, Gaudriault S. 2014. Attenuated virulence and genomic reductive evolution in the entomopathogenic bacterial symbiont species, *Xenorhabdus pionarii*. *Genome Biol Evol* 6:1495–1513. <http://dx.doi.org/10.1093/gbe/evu119>.
5. Wang Y, Fang X, An F, Wang G, Zhang X. 2011. Improvement of

- antibiotic activity of *Xenorhabdus bovienii* by medium optimization using response surface methodology. *Microb Cell Fact* 10:98. <http://dx.doi.org/10.1186/1475-2859-10-98>.
6. Bode HB. 2009. Entomopathogenic bacteria as a source of secondary metabolites. *Curr Opin Chem Biol* 13:224–230. <http://dx.doi.org/10.1016/j.cbpa.2009.02.037>.
 7. Ji D, Yi Y, Kang G-H, Choi Y-H, Kim P, Baek N-I, Kim Y. 2004. Identification of an antibacterial compound, benzylideneacetone, from *Xenorhabdus nematophila* against major plant-pathogenic bacteria. *FEMS Microbiol Lett* 239:241–248. <http://dx.doi.org/10.1016/j.femsle.2004.08.041>.
 8. McInerney BV, Taylor WC, Lacey MJ, Akhurst RJ, Gregson RP. 1991. Biologically active metabolites from *Xenorhabdus* spp., part 2. Benzopyran-1-one derivatives with gastroprotective activity. *J Nat Prod* 54:785–795. <http://dx.doi.org/10.1021/np50075a006>.
 9. Li J, Chen G, Webster JM, Czyzewska E. 1995. Antimicrobial metabolites from a bacterial symbiont. *J Nat Prod* 58:1081–1086. <http://dx.doi.org/10.1021/np50121a016>.
 10. Gualtieri M, Aumelas A, Thaler J-O. 2009. Identification of a new antimicrobial lysine-rich cyclolipopeptide family from *Xenorhabdus nematophila*. *J Antibiot* 62:295–302. <http://dx.doi.org/10.1038/ja.2009.31>.
 11. Kaya HK, Stock SP. 1997. Entomopathogenic nematodes, p 259–262. *In* Knowles BH, Blatt MR, Tester M, Horsnell JM, Carroll J, Menestrina G, Ellar DJ (ed), *Manual of techniques in insect pathology*. Academic Press, New York, NY.